

Role of ammonia in the pathogenesis of the gastritis, hypergastrinaemia, and hyperpepsinogenaemia I caused by *Helicobacter pylori* infection

A M El Nujumi, P A Rowe, S Dahill, C A Dorrian, W D Neithercut, K E L McColl

Abstract

Studies were performed in patients with and without renal failure to investigate the role of bacterial ammonia production in the pathogenesis of the mucosal abnormalities caused by *Helicobacter pylori*. The high rate of *H pylori* ammonia production in uraemic patients should accentuate any ammonia induced effects. The median (range) gastric juice ammonium concentration in the *H pylori* positive patients with renal failure was 19 mmol/l (11-43) compared with 5 mmol/l (1-11) in the *H pylori* positive patients without renal failure ($p < 0.005$). In the *H pylori* negative patients the values were 3 mmol/l (0.5-11) and 0.7 mmol/l (0.1-1.4) respectively in the patients with and without renal failure ($p < 0.01$). Despite the much higher ammonia production in the *H pylori* positive uraemic patients, the nature and severity of their gastritis was the same as that in the *H pylori* positive non-uraemic patients. The median (range) fasting serum gastrin concentration was raised in the uraemic patients compared with the non-uraemic patients but was similar in the uraemic patients with (95 pmol/l (52-333)) or without (114 pmol/l (47-533)) *H pylori* infection. The median (range) serum pepsinogen I concentration was also high in the uraemic compared with the non-uraemic patients and was significantly higher in uraemic patients with *H pylori* (352 ng/ml, range 280-653) than in those without *H pylori* infection (165 ng/ml, range 86-337) ($p < 0.01$). These findings indicate that the gastritis and hypergastrinaemia associated with *H pylori* infection are not the result of mucosal damage induced by the organism's ammonia production.

(Gut 1992; 33: 1612-1616)

It is now generally acknowledged that *Helicobacter pylori* infection is the major cause of antral gastritis.¹ The bacterium, however, does not penetrate the gastric epithelium and the mechanism by which it induces inflammation of the underlying mucosa is unknown. The organism has very high urease activity² and it has been suggested that the production of high concentrations of ammonia at the epithelial surface could cause mucosal damage.³ Though the acidic gastric juice will rapidly convert ammonia to less toxic ammonium ions, the high pH at the site of ammonia production underneath the mucus layer could allow it to remain in its unionised noxious state.^{4,5} It has also been postulated that ammonia produced by *H pylori* predisposes to

mucosal damage by denaturing the structure of the protective mucus layer.⁶

H pylori infection has also been shown to increase the serum concentrations of gastrin⁷⁻¹² and pepsinogen I.^{7,13} Levi *et al* proposed that the hypergastrinaemia was related to bacterial ammonia production which raised the antral surface pH,⁸ but studies from our own group do not support this.¹⁴⁻¹⁸

To investigate the role of ammonia in the histological and biochemical changes that accompany *H pylori* infection, we have examined patients with chronic renal failure. The high intragastric urea concentrations in these patients greatly increase *H pylori* ammonia production which should accentuate any ammonia related effects.

Patients and methods

Gastric juice ammonium concentration, antral histology, and serum concentrations of gastrin and pepsinogen I were examined in nine *H pylori* positive and nine *H pylori* negative uraemic patients. These results were compared with those from age and sex matched control patients with normal renal function with (n=9) and without (n=9) *H pylori* infection. Each of the patients examined had been referred for upper gastrointestinal endoscopy to investigate dyspeptic symptoms. None was receiving anti-biotics or had taken acid inhibitory agents within the previous week. In addition, none of the patients had been treated with bismuth preparations or with immunosuppressive therapy.

The median ages of the uraemic patients were 51 years (range 34-73) in those who were *H pylori* positive and 49 years (range 27-67) in those who were *H pylori* negative. Three of the *H pylori* positive patients were on maintenance haemodialysis, four on continuous ambulatory peritoneal dialysis, and two had not yet begun dialysis. One of the *H pylori* negative patients was on maintenance haemodialysis and eight were on continuous ambulatory peritoneal dialysis. The median duration of dialysis was similar in the *H pylori* positive (15 months, range 0-100) and negative (20.5 months, range 0-142) patients.

All patients were examined between 0900 hours and 1100 hours after an overnight fast. Upper gastrointestinal endoscopy was performed after a venous blood sample had been taken for determination of serum concentrations of urea, gastrin, and pepsinogen I. Immediately after passing the instrument, 10 ml of gastric juice were collected by means of a trap in the suction line. After inspection of the upper

University Departments
of Medicine and
Therapeutics,
Pathological
Biochemistry, and
Pathology, Western
Infirmary, Glasgow
A M El Nujumi
P A Rowe
S Dahill
C A Dorrian
W D Neithercut
K E L McColl

Correspondence to:
Dr K E L McColl, University
Department of Medicine and
Therapeutics, Western
Infirmary, Glasgow G11 6NT.

Accepted for publication
31 May 1992

gastrointestinal tract, two biopsy specimens were taken from the greater curvature of the antrum, 2 cm from the pylorus.

The *H. pylori* status of patients and controls was determined by microscopy of antral biopsy for *Helicobacter* like organisms, rapid urease slide test (CLO test) of antral biopsy,¹⁹ and ¹⁴C-urea breath test. These tests have been shown to be reliable in detecting *H. pylori* infection in patients with and without renal failure.²⁰

ANALYSES

In the patients with chronic renal failure the pH of the gastric juice was determined using a combined glass electrode (Radiometer ETS 822) before storage at -20°C. Gastric juice urea and ammonium concentrations were determined in all subjects. For this the samples were thawed and centrifuged at 3000 g for 10 minutes to remove the mucus. The concentration of ammonium was measured in the supernatant after dilution in 0.2 M phosphate buffer pH 7.4, using an enzymatic method (Sigma, Dorset, UK) adapted for the Cobas Bio (Roche, Welwyn Garden City, UK) as previously described.²¹

The antral biopsy specimens were fixed in formalin and processed routinely. Paraffin sections were cut at three levels and stained with haematoxylin and eosin. An extra section from level two was stained with Cresyl fast violet for detection of *H. pylori*.²² They were examined by a single pathologist (SD) who was unaware of the patients' clinical details. The severity of antral gastritis was scored using the method of Rauws *et al*,²³ which we have found to be a sensitive method for assessing the severity of *H. pylori*-induced gastritis.^{9,16,21} Chronic inflammatory infiltrate in the lamina propria was scored as 0, 1, or 2; lamina propria polymorph infiltrate as 0, 1, 2, or 3; intraepithelial polymorph infiltrate

as 0, 1, 2, or 3; and mucosal erosions as 0, 1, or 2. The scores for these individual components of *H. pylori* related gastritis are then added to give a cumulative gastritis score ranging from 0-10.

The intraobserver variation in the scoring of the severity of the gastritis was assessed. This was performed by randomly selecting 15 slides scored by the pathologist (SD) at least one year earlier and having him rescore them unaware that he had previously examined them. The mean cumulative gastritis score for the group of 15 slides was 3.2 (range 0-6) when first scored compared with 3.3 (range 0-8) when rescored. The mean absolute difference in the cumulative gastritis score between the two assessments was 0.66 (range 0-2) and the coefficient of variation was 21%. This indicates that our pathologist would have a greater than 95% chance of detecting a difference in mean cumulative gastritis score of 1 when comparing two groups of nine subjects.

The serum gastrin concentration was determined by radioimmunoassay using antibody R98.²⁴ This detects both G17 and G34 and uses G17 as standard. Serum pepsinogen I was measured using commercial radioimmunoassay kits obtained from Incstar Ltd (Berkshire).

The statistical significance of differences between groups was assessed by the Mann-Whitney U test.

The study was approved by the Western Infirmary Ethical Committee and all patients gave written, informed consent.

Results

The serum urea concentration was raised to a similar extent in the renal failure patients with (median 20 mmol/l, range 12-31) or without (23, 11-32) *H. pylori* infection (normal range 2.5-7.5 mmol/l).

The median (range) gastric juice ammonium concentration in the *H. pylori* positive uraemic patients was 19 mmol/l (11-43) which was approximately four times greater than that in the *H. pylori* positive non-uraemic patients (5, range 1-11) ($p < 0.005$) (Fig 1). Gastric juice ammonium concentrations were similar in the *H. pylori* negative uraemic patients (median 3, range 0.5-11 mmol/l) and *H. pylori* positive non-uraemic patients (5, range 1-11) ($p = 0.2$), and both were significantly higher than the values for the *H. pylori* negative non-uraemic patients (0.7, range 0.1-1.4) ($p < 0.02$ for each). Intra-gastric pH was similar in the *H. pylori* positive and negative uraemic patients, with a median value of 3.0 (range 1.0-7.1) in the former and 2.1 (1.2-6.6) in the latter.

In spite of the marked difference in intra-gastric ammonium concentration, the severity of histological gastritis was similar in the *H. pylori* positive patients with (median cumulative gastritis score = 5, range 3-6) or without (5, range 3-7) renal failure (Fig 2). Combining the *H. pylori* positive patients with and without renal failure provided a wide range of gastric juice ammonium concentrations (1-43 mmol/l) but there was no correlation between this and the severity of the *H. pylori* related antral gastritis (Table). In spite of the *H. pylori* negative uraemic patients having

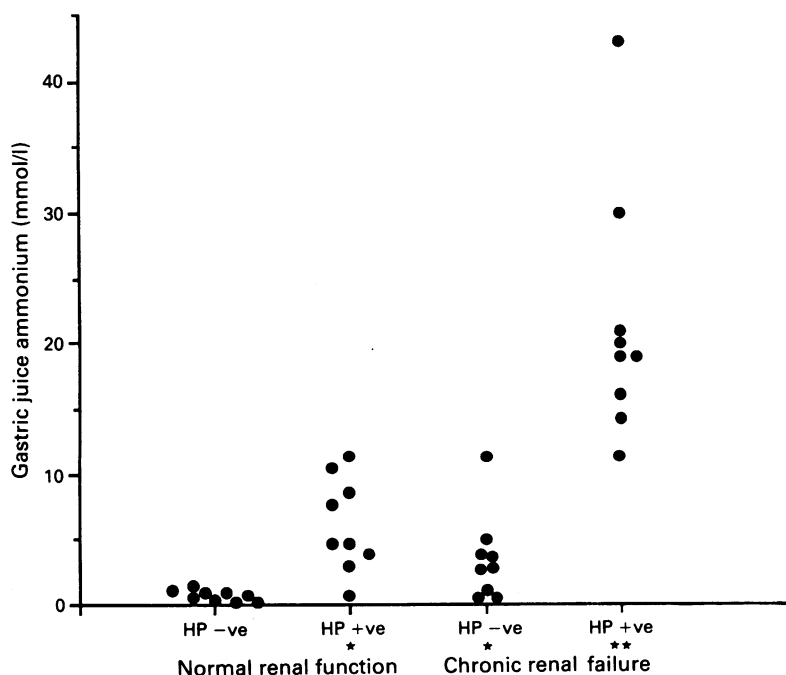


Figure 1: Effect of uraemia and *Helicobacter pylori* (HP) status on intragastric ammonium concentrations. *Indicates higher than *H. pylori* -ve non-uraemic patients at $p < 0.02$. **Indicates higher than the *H. pylori* +ve non-uraemic patients at $p < 0.005$.

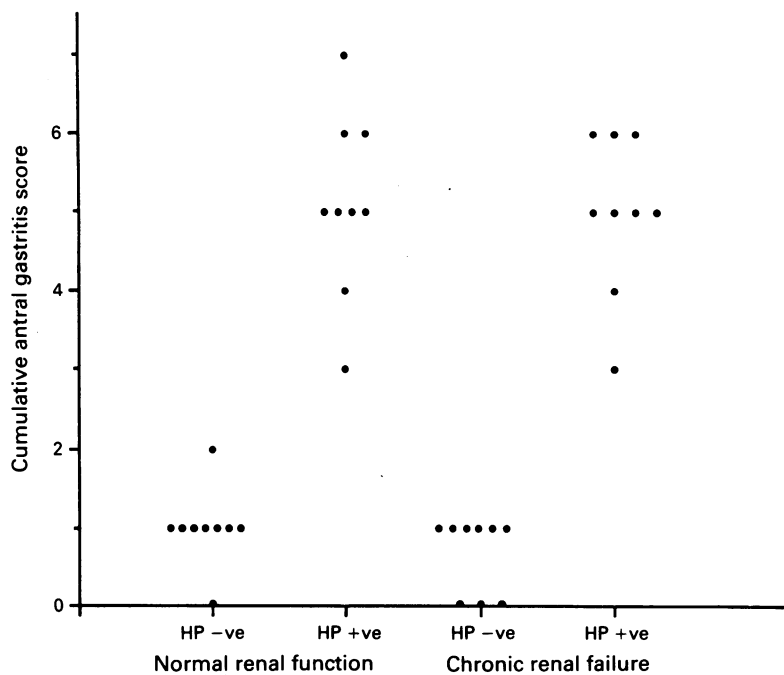


Figure 2: Cumulative antral gastritis scores in patients with and without chronic renal failure and of different *Helicobacter pylori* (HP) status.

intra-gastric ammonium concentrations similar to the *H pylori* positive non-uraemic patients, the former had cumulative gastritis scores of 1 or less which were equivalent to those in the *H pylori* negative patients with normal renal function.

There was no difference in the endoscopic appearance of the upper gastrointestinal tract in the renal failure patients with and without *H pylori* infection. Of the nine *H pylori* positive uraemic patients, two had oesophagitis, two had scattered petechiae in the stomach and duodenum, and one had erosive duodenitis. Of the nine *H pylori* negative uraemic patients, two had oesophagitis and one had scattered petechiae in stomach and duodenum. Endoscopy in the nine *H pylori* positive non-uraemic patients showed oesophagitis in one patient and active duodenal ulcer in another. In the nine *H pylori* negative non-uraemic patients, one had erosive duodenitis and another a deformed duodenum.

Gastric juice ammonium concentrations and antral gastritis scores in the *Helicobacter pylori* positive subjects with (+) and without (-) chronic renal failure. They are listed in descending order of gastric juice ammonium concentration

Gastric juice ammonium (mmol/l)	Chronic renal failure	Lamina propria chronic infiltrate score (0-2)	Intra-epithelial polymorph infiltrate score (0-3)	Lamina propria polymorph infiltrate score (0-3)	Epithelial erosions score (0-2)	Cumulative gastritis score (0-10)
43	+	2	1	1	0	4
30	+	2	1	2	0	5
21	+	2	2	1	0	5
20	+	2	2	2	0	6
19	+	2	2	1	0	5
19	+	2	2	2	0	6
16	+	1	1	1	0	3
14	+	2	2	2	0	6
11	+	1	1	1	2	5
11	-	2	2	1	0	5
10	-	2	2	2	1	7
8	-	2	1	2	0	5
7	-	2	0	1	0	3
5	-	2	1	2	0	5
5	-	2	1	1	0	4
4	-	2	1	2	0	5
3	-	2	2	2	0	6
1	-	2	2	2	0	6

The median serum pepsinogen I concentration (ng/ml) was higher in the *H pylori* positive uraemic patients (352, range 280-653) than in the *H pylori* negative uraemic patients (165, range 86-337) ($p < 0.01$) and both these groups had higher values than the non-uraemic patients of corresponding *H pylori* status ($p < 0.05$ for each) (Fig 3). Serum pepsinogen I values were similar in the non-uraemic patients with (median = 103, range 40-170) or without (92, 35-127) *H pylori* infection ($p = 0.3$). There was no relationship between the serum pepsinogen I concentration and type of renal replacement treatment.

The median serum gastrin concentration (pmol/l) in the non-uraemic patients was higher in those with *H pylori* (17, range 7-24) than in those without the infection (10, range 7-14) ($p < 0.05$) (Fig 4). Compared with these non-uraemic patients, the gastrin concentrations were noticeably high in the uraemic patients and there was no difference between the latter patients with (median=95, range 52-333) or without (114, range 47-533) *H pylori* infection. There was no relationship between the serum gastrin concentration and type of renal replacement treatment.

Discussion

This study shows that the gastric juice ammonium concentration is noticeably affected by both uraemia and *H pylori* infection. The extremely high intra-gastric ammonium concentration in the renal failure patients with *H pylori* infection can be explained by the combination of their high gastric juice urea concentration²⁵ and the high urease activity of the organism. We have previously shown that the intra-gastric production of ammonia by *H pylori* is controlled by the availability of urea in gastric juice.¹⁴

The reason for the higher gastric juice ammonium concentration in the *H pylori* negative uraemic patients than in the *H pylori* negative non-uraemic patients is not clear. However, a variety of urease producing bacteria are present in the mouth and swallowed in the saliva.²⁶ Though such bacteria have lower urease activity than *H pylori*, they may produce significant amounts of ammonia in the presence of high gastric juice concentrations of urea. In addition, it is possible that the high gastric juice urea concentration encourages the colonisation of the upper gastrointestinal tract by urease positive organisms. We have previously observed that *H pylori* negative uraemic patients have higher urease activity assessed by the ¹⁴C-urea breath test than *H pylori* negative non-uraemic patients.²⁰

Some in vitro and experimental animal studies have suggested that the antral gastritis induced by *H pylori* may be caused by the ammonia produced by the organism's high urease activity.^{3 27 29} Murakami *et al* showed that ammonia produced by the administration of urea plus urease to rats can cause microscopic injury to their gastric mucosa.³ In vitro studies by Smoot *et al* showed that ammonia produced by *H pylori* is cytotoxic to cultures of human gastric epithelial cells²⁷ and similar studies by Xu *et al*

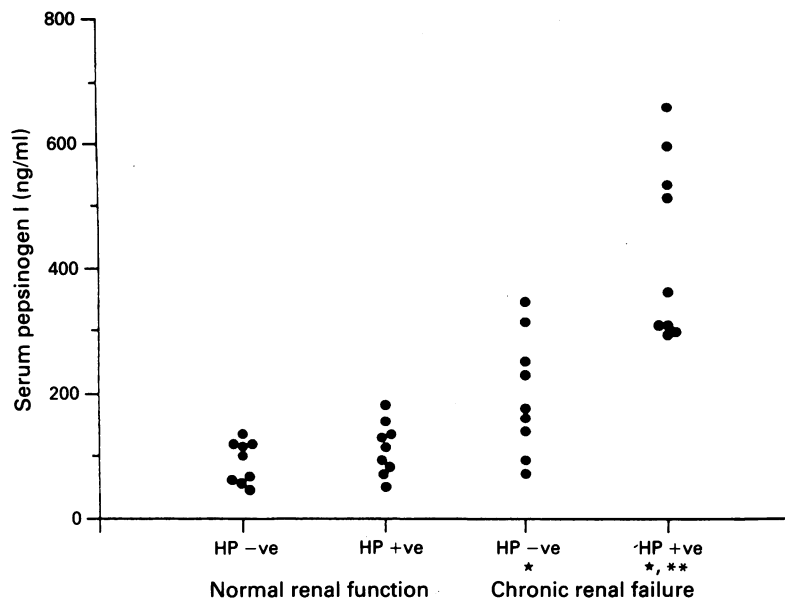


Figure 3: Serum pepsinogen I concentrations in patients with and without chronic renal failure and of different *Helicobacter pylori* (HP) status. *Indicates higher than non-uraemic patients of corresponding *H pylori* status at $p < 0.05$. **Indicates higher than *H pylori* -ve uraemic patients at $p < 0.001$.

found that *H pylori* ammonia production causes vacuolisation of Vero cell lines.²⁸ In addition to a direct toxic effect of ammonia, it has been proposed that ammonium ions may interact with neutrophil produced hypochlorous acid to produce the highly toxic mono-N-chloramine, NH_2Cl .²⁹ It has also been proposed that *H pylori* ammonia production will lead to mucosal damage by denaturing the protective mucus layer.⁶ In spite of the findings with animal models and in vitro experiments, the present study does not support a causal role for ammonia production in antral gastritis in man. This was demonstrated by the absence of any correlation between the severity of the gastritis or epithelial surface damage and the bacterial ammonia production. Examining patients with and without

uraemia enabled us to look at the gastric mucosal damage over a very wide range (43 fold) of in vivo ammonia production making it unlikely that any association was missed. The abnormalities related to ammonia administration noted in the earlier in vitro studies and animal models may be explained by the high pH employed which would have increased the proportion of unionised ammonia.

Trieblich *et al* have recently claimed that ammonia production does play a pathogenic role in the development of *H pylori* related gastritis in man.³⁰ However, this conclusion is not supported by their own data. They measured gastric juice ammonium and the severity of gastritis in five patients with chronic renal failure and *H pylori*, before and after treatment with ampicillin, and in five uninfected patients with renal failure. A positive correlation was observed between the gastric juice ammonium and severity of gastritis. This correlation, however, can be explained simply by the presence of absence of *H pylori* in their patient group and does not provide evidence of a causal association between *H pylori* ammonia production and gastritis. Within their small group of five patients with *H pylori*, there was no correlation between gastric juice ammonium and severity of gastritis, which is consistent with the findings of our present study in which we examined 18 such patients. Our conclusion that ammonia does not play a pathogenic role in the development of *H pylori* related gastritis in man is also consistent with the recent ultrastructural studies by Thomsen *et al*.³¹ They noted that there was no correlation between the location of the organism and the morphological damage to adjacent epithelial cells or the degree of subjacent inflammatory cell infiltrate.

The noticeably high serum pepsinogen I concentration in the patients with chronic renal failure is consistent with previous reports³² and can be explained by its impaired renal clearance.³³ The increase in serum pepsinogen I was particularly marked in the renal failure patients with *H pylori*, with its concentration being double that in the renal patients without the infection. It has previously been reported that *H pylori* infection raises serum pepsinogen I in non-uraemic patients but only by about 25%.^{7,13} The fact that *H pylori* infection raises serum pepsinogen I to a greater extent in uraemic than non-uraemic subjects would be consistent with bacterial ammonia production raising the serum concentration of the zymogen. However, little is known about the mechanism by which pepsinogen I reaches the serum or about the mechanism of its renal excretion.³³ If it is excreted by a saturable process then in patients with renal failure the same degree of increased delivery of pepsinogen I into the serum could produce a more marked increase in its serum concentration.

As previously reported, the serum gastrin concentration was found to be high in the renal failure patients compared with those with normal renal function.³⁴⁻³⁷ Serum gastrin concentration is also known to be raised in patients with *H pylori* infection,⁷⁻¹² and this is seen in the non-uraemic patients in the present study. It has been

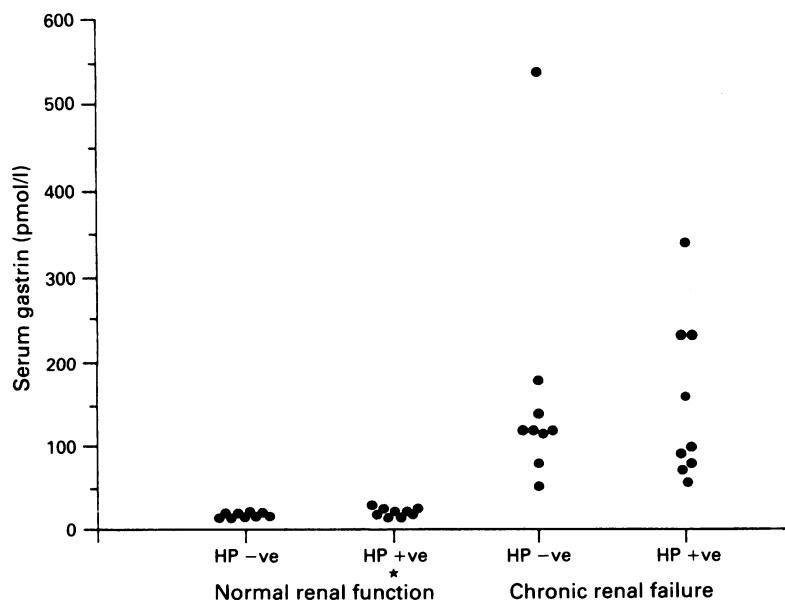


Figure 4: Effect of chronic renal failure and *Helicobacter pylori* (HP) status on fasting serum gastrin concentrations. *Indicates higher than *H pylori* -ve non-uraemic patients at $p < 0.05$.

suggested that the hypergastrinaemia induced by *H pylori* is caused by the ammonia produced by its urease raising antral surface pH and thereby blocking the suppression of gastrin release by luminal acid.⁸ However, we have previously found that neither increasing,¹⁴ inhibiting,¹⁵ nor completely abolishing¹⁶ *H pylori* urease activity in man alters serum gastrin. We have also found that *H pylori* related hypergastrinaemia cannot be explained by ammonia altering mucosal surface pH.^{17,18} The finding that the gastrin concentration is similar in the *H pylori* positive and negative uraemic patients, despite the much higher rate of ammonia production in the former, is further evidence against the hypergastrinaemia being due to bacterial ammonia production.

Ala-Kaila *et al* recently observed that patients with chronic renal failure could be divided into two statistically distinct groups according to their degree of hypergastrinaemia.³⁷ The differences in gastrin could not be explained by differences in acid secretion or severity of renal failure and they postulated an unknown mechanism causing enhanced synthesis of gastrin. The present study indicates that *H pylori* infection is not the unknown factor.

In conclusion, this study shows that the very high rate of *H pylori* ammonia production in uraemic patients is not associated with more marked gastritis or hypergastrinaemia. These findings support our previous work showing that *H pylori* release of ammonia is unlikely to be responsible for either the hypergastrinaemia or the gastritis caused by this bacterium.

This work was supported by a grant from the Biomedical Research Committee of the Scottish Home and Health Department and from the Research Support Group of the Greater Glasgow Health Board. The technical assistance of Mrs Devina Filmore, Department of Medicine, Queen's University, Belfast; of Miss Jennifer Harwood, Department of Pathological Biochemistry, Western Infirmary; and the secretarial assistance of Mrs Dorothy Ronney is gratefully acknowledged.

This work was presented at the 1991 Autumn Meeting of the British Society of Gastroenterology and published in abstract form in *Gut*.

- 1 McNulty CAM. Pathogenicity of *Campylobacter pylori* – a causative factor in gastritis? *J Gastroenterol* 1989; 24 (suppl 160): 3–6.
- 2 Marshall B, Langton S. Urea hydrolysis in patients with *Campylobacter pyloridis* infection. *Lancet* 1986; i: 965–6.
- 3 Murakami M, Yoo JK, Inada M, Miyake T. Effect of ammonia on the gastric mucosa in rats: pathophysiological importance of urease in gastric ulcer disease. *Jpn J Pharmacol* 1988; 47: 330–2.
- 4 Visek WJ. Diet and cell growth modulation by ammonia. *Am J Clin Nutr* 1978; 31: S216–20.
- 5 Visek WJ. Some aspects of ammonia toxicity in animal cells. *J Dairy Sci* 1968; 51: 286–95.
- 6 Sidebotham RL, Baron JH. Hypothesis: *Helicobacter pylori*, urease, mucus, and gastric ulcer. *Lancet* 1990; 335: 193–5.
- 7 Oderda G, Vaira D, Holton J, Ainley C, Altare F, Ansaldi N. Amoxicillin plus tinidazole for *Campylobacter pylori* gastritis in children: assessment by serum IgG antibody, pepsinogen I, and gastrin levels. *Lancet* 1989; i: 690–2.
- 8 Levi S, Beardshall K, Haddad G, Playford R, Ghosh P, Calam J. *Campylobacter pylori* and duodenal ulcers: the gastrin link. *Lancet* 1989; i: 1167–8.
- 9 McColl KEL, Fullarton GM, Chittajallu R, El Nujumi AM, Macdonald AMI, Dahill SW, *et al*. Plasma gastrin, daytime intragastric pH, and nocturnal acid output before and at 1 and 7 months after eradication of *Helicobacter pylori* in duodenal ulcer subjects. *Scand J Gastroenterol* 1991; 26: 339–46.
- 10 Graham DY, Opekum A, Lew GM, Evans DJ, Klein PD, Evans DG. Ablation of exaggerated meal stimulated gastrin release in duodenal ulcer patients after clearance of *Helicobacter (Campylobacter) pylori* infection. *Am J Gastroenterol* 1990; 85: 394–8.
- 11 Smith JTL, Pounder RE, Nwokolo CU, Lanzon-Miller S, Evans DG, Graham DY, *et al*. Inappropriate hypergastrinaemia in asymptomatic healthy subjects infected with *Helicobacter pylori*. *Gut* 1990; 31: 522–5.
- 12 Chittajallu RS, Ardill JE, McColl KEL. The degree of hypergastrinaemia induced by *Helicobacter pylori* is the same in duodenal ulcer patients and asymptomatic volunteers. *Eur J Gastroenterol Hepatol* 1992; 4: 49–53.
- 13 Chittajallu RS, Dorrian CA, Ardill JES, McColl KEL. Effect of *Helicobacter pylori* on serum pepsinogen I and plasma gastrin in duodenal ulcer patients. *Scand J Gastroenterol* 1992; 27: 20–5.
- 14 Chittajallu RS, Neithercut WD, Macdonald AMI, McColl KEL. Effect of increasing *Helicobacter pylori* ammonia production by urea infusion on plasma gastrin concentrations. *Gut* 1991; 32: 21–4.
- 15 Nujumi AM El, Dorrian CA, Chittajallu RS, Neithercut WD, McColl KEL. Effect of inhibition of *Helicobacter pylori* urease activity by acetohydroxamic acid on serum gastrin in duodenal ulcer subjects. *Gut* 1991; 32: 866–70.
- 16 Chittajallu RS, Dorrian CA, Neithercut WD, Dahill S, McColl KEL. Is *Helicobacter pylori* associated hypergastrinaemia due to the bacterium's urease activity or the antral gastritis? *Gut* 1991; 32: 1286–90.
- 17 Chittajallu RS, Neithercut WD, Ardill JES, McColl KEL. *Helicobacter pylori*-related hypergastrinaemia is not due to elevated antral surface pH. Studies with antral alkalinisation. *Scand J Gastroenterol* 1992; 27: 218–22.
- 18 McColl KEL, Nujumi AM El, Dorrian CA, Macdonald AMI, Fullarton GM, Harwood J. *Helicobacter pylori* and hypergastrinaemia during proton pump inhibitor therapy. *Scand J Gastroenterol* 1992; 27: 93–8.
- 19 Marshall BJ, Warren R, Francis GJ, Langton SR, Goodwin CS, Blincow ED. Rapid urease test in the management of *Campylobacter pyloridis*-associated gastritis. *Am J Gastroenterol* 1987; 82: 200–10.
- 20 Nujumi AM El, Rowe P, Dorrian CA, McColl KEL. Value of ¹⁴C-urea breath test to diagnose *Helicobacter pylori* in uraemic patients. *Gut* 1991; 32: A1220.
- 21 Neithercut WD, Milne A, Chittajallu RS, Nujumi AM El, McColl KEL. Detection of *Helicobacter pylori* infection of the gastric mucosa by measurement of gastric aspirate ammonium and urea concentrations. *Gut* 1991; 32: 973–6.
- 22 Burnett RA, Brown IL, Findlay J. Cresyl fast violet staining method for *Campylobacter*-like organisms. *J Clin Pathol* 1987; 40: 353.
- 23 Rauws EAJ, Langenberg W, Houthoff HJ, Zanen HC, Tytgat GNJ. *Campylobacter pyloridis*-associated chronic acute antral gastritis. *Gastroenterology* 1988; 94: 33–40.
- 24 Ardill JES. The measurement of gastrin by radioimmunoassay (PhD Thesis). Belfast: Queen's University, 1973.
- 25 Lieber CS, Lefevre A. Ammonia as a source of gastric hypoacidity in patients with uremia. *J Clin Invest* 1959; 38: 1271–7.
- 26 Bowden GHW, Ellwood DS, Hamilton IR. Microbial ecology of the oral cavity. In: Alexander M, ed. *Advances in microbial ecology*. Vol 3. New York: Plenum Press, 1979.
- 27 Smoot DT, Mobley HLT, Chippendale GR, Lewison JF, Resau JH. *Helicobacter pylori* urease activity is toxic to human gastric epithelial cells. *Infect Immun* 1990; 58: 1992–4.
- 28 Xu JK, Goodwin CS, Cooper M, Robinson J. Intracellular vacuolisation caused by the urease of *H pylori*. *J Infect Dis* 1990; 161: 1302–4.
- 29 Hazell SL. Urease and catalase as virulence factors of *Helicobacter pylori*. In: Menge H, ed. *Helicobacter pylori* 1990. Berlin: Springer-Verlag, 1991: 3–14.
- 30 Trieblich AJ, Korstein MA, Dlugosz JW, Paronetto F, Lieber C. Severity of *Helicobacter*-induced gastric injury correlates with gastric juice ammonia. *Dig Dis Sci* 1991; 36: 1089–96.
- 31 Thomsen LL, Gavin JB, Tasman-Jones C. Relation of *Helicobacter pylori* to the human gastric mucosa in chronic gastritis of the antrum. *Gut* 1990; 31: 1230–6.
- 32 Samloff IM, Liebman WM, Panitch NM. Serum group I pepsinogens by radioimmunoassay in control subjects and patients with peptic ulcer. *Gastroenterology* 1975; 69: 83–90.
- 33 Waldrum HL, Jorde R, Gunnes P. Renal excretion of, and the effect of, posture on serum group I pepsinogens. *Scand J Gastroenterol* 1982; 17: 253–5.
- 34 Muto S, Murayama N, Asano Y, Hosado S, Moyata M. Hypergastrinaemia and achlorhydria in chronic renal failure. *Nephron* 1985; 40: 143–8.
- 35 El-Ghonaimey E, Barsoum R, Soliman M, El-Fikky A, Rashwan S, El-Rouby O, *et al*. Serum gastrin in chronic renal failure: morphological and physiological correlations. *Am J Surg* 1981; 141: 334–8.
- 36 Wesdorp RI, Falcao HA, Banks PB, Martino J, Fisher JE. Gastrin and gastric acid secretion in renal failure. *Am J Surg* 1981; 141: 334–8.
- 37 Ala-Kaila A, Kekki M, Paronen I, Poakkalo T. Serum gastrin in chronic renal failure: its relation to acid secretion, G-cell density, and upper gastrointestinal findings. *Scand J Gastroenterol* 1989; 24: 939–48.