Pharmacokinetics of ranitidine in critically ill patients

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1 The plasma pharmacokinetics of ranitidine (50 mg i.v.) have been studied in 17 critically ill patients in an intensive care unit. Measurements of gastric aspirate pH were also made in 16 of these patients. Ranitidine therapy was part of the patients' normal drug regimen.

2 Ranitidine plasma concentration was measured by high performance liquid chromatography and appropriate polyexponential equations were fitted to concentration-time data to enable calculation of relevant pharmacokinetic parameters. Values of the volume of the initial dilution space (median = 89 ml kg^{-1}) and volume of distribution at steady state (median = 1.54 l kg^{-1}) were about 60% of corresponding mean literature values for healthy controls. Plasma clearance (median = $4.22 \text{ ml min}^{-1} \text{ kg}^{-1}$) and terminal half-life (median = 4.7 h) were about 2–3 fold less and 2–3 fold greater, respectively, than values for healthy controls. There was wide interpatient variation in all the pharmacokinetic parameters. Renal impairment was considered to be largely responsible for the low plasma clearance.

3 Gastric aspirate pH was measured at 0, 1 and 7 h after ranitidine administration and 58% of samples were found to be above pH 4. Four patients had gastric pH values which were consistently below pH 4 despite average trough plasma ranitidine concentrations equal to or greater than those required for a 50% suppression of gastric acid secretion in normal volunteers.

Keywords ranitidine plasma pharmacokinetics critically ill patients gastric aspirate pH

Introduction

Ranitidine is a potent histamine H_2 -receptor antagonist which is used in the treatment of peptic ulcer (Brogden *et al.*, 1981) and in the prevention of stress-induced ulceration in critically ill patients (Barth *et al.*, 1984), although the use of histamine H_2 -receptor blockers in this latter patient group is controversial (Greene *et al.*, 1984). After intravenous administration ranitidine is largely excreted by the kidneys (McNeil *et al.*, 1981). The pharmacokinetics of the drug have been studied extensively in normal volunteers (Garg *et al.*, 1981; Lebert *et al.*, 1981a; McNeil *et al.*, 1981; Van Hecken *et al.*, 1982; McGonigle *et al.*, 1982; Young *et al.*, 1982; Chau *et al.*, 1982) and in selected patient groups with renal (Meffin *et al.*, 1983; McFadyen *et al.*, 1983) or hepatic (Young *et al.*, 1982; Smith *et al.*, 1984) disease. There are, however, no published data on the pharmacokinetics of ranitidine in the critically ill and thus the present study was

* Present address: School of Pharmacy, South Australian Institute of Technology, Adelaide, South Australia. Correspondence: Dr K. F. Ilett, Department of Pharmacology, University of Western Australia, Nedlands, Western Australia 6009. undertaken to assess ranitidine plasma kinetics and gastric pH effects within this patient group.

Methods

Seventeen critically ill patients being cared for in the Intensive Care Unit took part in the study which was approved by the Human Rights Committee of The University of Western Australia. Ranitidine therapy was part of the patients' normal drug regimen. Their clinical and biochemical details are summarised in Tables 1 and 2. Most had multiple diagnoses and all were receiving several other medications (range 7–16) in addition to ranitidine. Ranitidine was the sole anti-ulcer therapy in all but patient IT who also received antacid via a nasogastric tube.

Ranitidine administration and collection of blood and gastric fluid samples

Patients were receiving intermittent intravenous infusions of ranitidine. One patient was studied after the first dose of ranitidine while the remaining patients were studied after being on continuous ranitidine therapy for a mean of 5.5 days (range 1–15 days). Prior to study, 14 patients received 50 mg ranitidine (Zantac, Glaxo, Australia) intravenously every 8 h while two patients received 25 mg ranitidine 8 and 12 hourly, respectively. On the study day, 50 mg ranitidine (in 10 ml normal saline) was administered as an intravenous infusion over $3.93 \pm$ 0.20 min (mean \pm s.e. mean) by means of a constant rate infusion pump.

Arterial blood samples (2.5 ml) were collected from each patient through an indwelling catheter already located in the radial artery for measurement of blood pressure and blood gases. Samples were taken immediately before the ranitidine dose and at 1, 3, 4 (or end of infusion), 6, 10, 15, 20 and 30 min and at 1, 2, 3, 4, 6 and 8 h after dosage. The samples were collected into heparinised polypropylene tubes and after being separated by centrifugation, plasma was stored at -20° C until assayed for ranitidine. In all patients except IT, gastric fluid was aspirated from an in situ nasogastric tube at 0, 1 and 7 h after dosage and its pH was measured using a digital pH meter. In six patients, 2-5 additional pH measurements were made on gastric aspirates obtained at various times from 2 to 8 h after dosage.

Ranitidine analysis

Ranitidine was measured by high performance liquid chromatography (h.p.l.c.). Aliquots of

plasma (1 ml) were mixed with $0.16 \mu g$ of internal standard (AH20480; N-methyl-N'-[3-[(3-dimethylaminomethyl)phenoxy[propyl]-2-nitro-l, 1-ethylenediamine hydrochloride) and 0.05 ml of 2 M NaOH and then extracted with 4 ml of ether: chloroform: isopropyl alcohol (2:1:1) by vortexing for 1 min. After centrifugation, 3.5 ml of the organic phase was removed and backextracted into 0.1 ml of 5% acetic acid by vortexing for 1 min. After a further centrifugation, the organic phase was discarded and 40 µl aliquots of the acetic acid extract were injected onto the h.p.l.c. column. Approximate retention times for AH20480, desmethylranitidine and ranitidine were 3.5, 4.0 and 5.4 min, respectively, and there was no interference from other drugs which the patients received. Despite their relatively close retention times, baseline resolution between ranitidine and N-desmethylranitidine was achieved largely because of the very narrow peaks which are characteristic of the procedure. The coefficient of variation for the assay was 3.5% at 0.043 mg l^{-1} and 3.1% at 1.30 mg l^{-1} (n = 6) with a limit of detection for ranitidine of about 0.01 mg l⁻¹ plasma.

Pharmacokinetic and statistical analysis

With the exception of the one occasion on which the first dose of the ranitidine regimen was studied, the contribution to the plasma ranitidine concentrations from previous doses was subtracted by use of the superposition principle. The following equation was then fitted to the plasma ranitidine concentration-time data using the nonlinear least squares programme NONLIN (Metzler, 1974):

$$C = \sum_{i=1}^{n} \frac{C_{i}R_{o}(e^{\lambda_{i}T}-1)}{\lambda_{i}} \cdot e^{-\lambda_{i}t}$$

where C = plasma ranitidine concentration at time *t*, the time after start of infusion, $R_o =$ infusion rate, $C_i =$ dose-independent coefficient for the ith exponential, $\lambda_i =$ the ith exponential and T = infusion time.

The polyexponential equation that best described the data for each subject was chosen on the basis of the distribution of residuals and on a visual inspection of the goodness of fits. The coefficients and exponents of the chosen equation were used for the calculation (Wagner, 1976) of the following parameters: volume of the initial distribution space (V_1) , volume of distribution at steady state (V_{ss}) , total systemic plasma clearance (CL) and terminal disposition half-life $(t_{1/2})$.

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Concomitant medication*	owel infarction 4, 5, 12, 14, 15, 16, 17, 21, 23, 33, 41. ia rtest I vascular disease heart disease	iis 2, 5, 13, 14, 19, 23, 30, 33, 40, 41. ve jaundice ure nia	aft 2, 5, 12, 13, 14, 15, 33, 38, 42. rrest y oedema ative bleeding	ccal empyema 2, 4, 5, 9, 12, 13, 14, 17, 21, 23, 27, 28, 33, 42. mpairment sry failure	al infarction 4, 5, 9, 10, 14, 15, 16, 22, 25, 32, 33, 42. n pneumonitis nonary collapse	nia 1, 11, 13, 14, 20, 23, 27, 29, 31, 33, 36, 41. renal failure ve cardiac falure obstructive airways
Diagnosi	Embolic Septicaer Cardiac Diabetes Peripher Ischaemi	Pancreat Obstruct Renal fa Septicae	Aortic g Cardiac Pulmoná Post-ope	Streptoc Hepatic Respirat	Myocard Aspirati Left pul	Septicae Celluliti Chronic Congest Chronic
Weight (kg)	61	105	62	54	83	85
Age (years)	73	4	67	45	59	7
Ser	ш	W	W	ц	W	X
Subject	۲ ۲	RW	AM	MC	DP	HL

* Key to concomitant medications: 1 = acetazolamide, 2 = adrenaline,

7 = cefoxitin, 8 = chloramphenicol, 9 = clindamycin, 10 = clonazepam, 11 = cloxacillin, 12 = digoxin, 13 = dopamine, 14 = folic acid, 15 = frusemide, 16 = gentamicin, 17 = heparin, 18 = hydrocortisone, 19 = insulin, 20 = ipratropium, 21 = metoclopramide, 22 = metronidazole, 23 = morphine, 24 = moxalactam, 25 = nitroglycerin, 26 = nystatin, 27 = pancuronium, 28 = paracetamol, 29 = penicillin V, 30 = pethidine, 31 = phenoperidine, 32 = phenytoin, 33 = phytomenadione, 34 = potassium chloride, 35 = prochlorperazine, 36 = salbutamol, 37 = tobramycin, 38 = total parenteral nutrition, 39 = trace elements, 40 = verapamil, 41 = vitamin B1, 42 = vitamin B complex. 3 = a aluminium hydroxide/magnesium hydroxide mixture, 4 = amoxycillin, 5 = ascorbic acid, 6 = cefotaxime,

				Plasma co	ncentration*			
								$Estimated^{\dagger}$
		:		Alkaline	Aspartate amino	ł		creatinine
Patient	I otal protein (g l ⁻¹ ; 63–78)	Albumin (g l ⁻¹ ; 35–45)	Bilirubin (µmol l ⁻¹ ; 3–17)	phosphatase (u l ⁻¹ ; 100–350)	transferase (u l ⁻¹ ; 6–42)	Urea (µmol 1 ⁻¹ ; 3–8)	Creatinine (µmol 1 ⁻¹ ; 50–120)	clearance (ml min ^{_l})
AB	65	32	78	195	25	15.1	106	67
KK	67	\$	29	249	58	48.6	493	18
RB	71	41	105	172	56	10.7	73	59
BB	56	33	108	59	33	31.0	265	23
)) I	99	36	62	157	1518	17.1	122	37
RR	99	31	22	124	40	23.3	161	55
AR	99	39	81	106	56	34.2	242	25
ΜM	75	42	27	259	48	13.9	117	80
T	99	29	10	148	22	11.8	62	76
AS	58	41	16	68	18	23.1	358	13
HB	74	42	80	1040	4	16.2	74	55
JR	48	22	17	139	12	17.5	83	56
RW	62	37	19	158	67	30.8	707	18
AM	65	33	520	241	88	17.9	118	49
MC	72	39	159	252	4	11.7	4	144
DP	62	37	30	156	105	5.6	78	112
HI	68	25	56	324	45	44.3	656	11
* Units ai	nd laboratory rel ed by the metho	ference range in d of Biornsson	n parenthesis. et al. (1983).					
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Table 2

Group data were expressed either as median (and range) or as mean \pm s.e. mean unless otherwise indicated. Differences between group means were assessed by use of Student's *t*-test. Statistical association between variables was assessed by linear regression analysis. A probability value of less than 0.05 was considered significant.

Results

Analysis of the plasma concentration-time profiles indicated that in 13 of the studies, a triexponential curve was required to best represent the data while a biexponential curve was chosen in the remaining four studies. A typical profile is shown in Figure 1. At the end of the infusion, the actual mean arterial plasma ranitidine concentration was 4.59 mg l⁻¹ (range 1.47–8.39 mg l⁻¹) and at 8 h after dosage the mean concentration was 0.26 mg l⁻¹ (range 0.04–1.76 mg l⁻¹). Mean ratio of the actual plasma drug concentration at the end of infusion to that at 8 h was 37 with a range from 2 to 124.

The pharmacokinetic parameters describing the disposition of ranitidine are summarized in Table 3. There was a wide inter-patient variation in all parameters which reflects the diversity of the group with respect to factors such as age, concurrent pathological states and other medications. Correlations between the various pharmacokinetic parameters and age were tested but none was significant with or without correction of the parameters for body weight. Without correction for body weight, the correlation between ranitidine plasma clearance and estimated creatinine clearance was not significant (r = 0.439, P > 0.05). There was, however, a weak but significant correlation between weight corrected ranitidine plasma clearance and estimated creatinine clearance $(y = 0.028 \times$ +2.89; r = 0.537, P < 0.05).

Figure 2 shows gastric aspirate pH in the six patients in whom more than three pH measurements were made during the dose interval. Subject TH was studied after the first dose of ranitidine whereas the other five patients shown in Figure 2 were studied after being on a thricedaily ranitidine regimen for between 2 and 15 days. In the remaining 10 patients in the study group, gastric aspirates were only obtained at 0, 1 and 7 h after dosage. Considering only the 0, 1 and 7 h pH data from all 16 patients (excluding the 0 h pH for subject TH), 58% of gastric aspirate samples had a pH above 4 with a corresponding mean actual plasma ranitidine concentration of 0.41 mg l^{-1} , while the remaining 42% were below pH 4 and the corresponding mean plasma ranitidine concentration was 0.38 mg l^{-1} . Patients KK, AR, HB and RW had gastric pH values which were consistently less than 4 despite trough (average of the 0 and 8 h concentrations) plasma ranitidine concentrations during the



Figure 1 Typical plasma ranitidine concentration (log scale) vs time profile (Subject DP). The solid line is the computer-generated triexponential fit to the data.

Patient	V, (l)	$\frac{V_l}{(mg \ kg^{-l})}$	V _{ss} (l)	$(l \ kg^{-l})$	CL (ml min ⁻¹)	CL (ml min ⁻¹ kg ⁻¹)	t _{1/2} (h)
AB	8.5	106	148.8	1.86	737	9.21	3.5
KK	13.9	193	151.6	2.11	206	2.86	10.6
RB	1.9	33	58.5	1.01	253	4.36	4.0
BB	1.1	17	137.8	2.19	69	1.10	23.6
JJ	1.8	41	43.9	0.99	153	3.48	3.9
RR	13.2	155	384.3	4.52	313	3.68	17.0
AR	9.4	106	158.0	1.78	323	3.64	6.7
MW	7.6	89	72.7	0.86	435	5.12	2.6
IT	69.3	924	170.5	2.27	381	5.08	6.0
AS	27.1	484	86.0	1.54	328	5.86	3.5
HB	4.8	83	34.6	0.60	245	4.22	1.9
JR	3.5	57	60.1	0.99	210	3.45	5.1
RW	14.5	138	88.8	0.85	192	1.83	5.7
AM	1.6	26	268.5	4.33	326	5.25	12.3
MC	6.4	119	87.1	1.61	305	5.65	4.4
DP	6.0	72	73.6	0.89	509	6.13	2.8
ТН	4.7	55	88.5	1.04	263	3.09	4.7
Median Range	6.4 1.1–69.3	89 17–924	88.5 34.6–384.3	1.54 0.60–4.52	305 69–737	4.22 1.10–9.21	4.7 1.9–23.6

 Table 3 Pharmacokinetic parameters for critically ill patients receiving ranitidine (50 mg i.v.)



Figure 2 Gastric aspirate pH at various times during the ranitidine dose interval for subjects HB (Δ), AR (\blacktriangle), AM (\Box), MC (∇), DP (\bullet) and TH (\circ).

study dose interval of 0.097, 0.21, 0.26 and 0.45 mg l^{-1} , respectively. Three of these four patients had common diagnoses of renal impairment, pancreatitis and septicaemia (Table 1).

Discussion

Our critically ill patients had V_{ss} values ranging from 34.6 to 384.3 l (median = 88.5 l) or from 0.60 to $4.52 \, \mathrm{l \ kg^{-1}}$ (median = $1.54 \, \mathrm{l \ kg^{-1}}$) compared with published mean values for normal volunteers of 1.87 l kg⁻¹ (Garg et al., 1981) and 106 l (Young et al., 1982). Volume of the initial dilution space ranged from 17–924 ml kg⁻¹ (median = 89 ml kg⁻¹) compared to the mean value of 163 ml kg⁻¹ in normal volunteer subjects (Chau *et al.*, 1982). Half-life in the critically ill patients (median = 4.7 h) was some two to three fold longer than the range of means of 1.7–2.2 h in normal volunteers (McNeil et al., 1981; Garg et al., 1981; Lebert et al., 1981a; Van Hecken et al., 1982; Young et al., 1982; McGonigle et al., 1982) but was closer to that for patients with renal impairment (5.3 h, Meffin et al., 1983; 7.3 h, McFadyen et al., 1983). Plasma clearance in the critically ill (median = $4.22 \text{ ml min}^{-1} \text{ kg}^{-1}$) was about one half that for normal volunteer subjects (range of means = 8.9-10.4 ml min⁻¹ kg⁻¹; Lebert et al., 1981a,b; Chau et al., 1982; Garg et al., 1981) and again was closer to the mean of 5 ml min⁻¹ kg⁻¹ which can be calculated from the data of Meffin et al. (1983) in patients with renal impairment. However, it should be noted that our critically ill patients showed wide inter-patient variability in both clearance and half-life with an approximate 8-12 fold range for these parameters. In renal disease, clearance has been reported to vary over a 5 fold range (Meffin et al., 1983) while in other studies half-life varied over a 1.4-5.2 fold range (McGonigle et al, 1982; McFadyen et al, 1983). In hepatic cirrhosis, Smith et al., 1984) found a 2 fold variation in clearance values for their patients.

On the basis of clinical laboratory test data, patients KK, BB, JJ, RR, AR, AS, RW and TH were known to have renal impairment. Mean clearance for ranitidine in these patients was 3.39 ± 0.55 ml min⁻¹ kg⁻¹ and was significantly lower (t = 2.23, P < 0.05) than in the remaining patients (5.2 ± 0.58 ml min⁻¹ kg⁻¹). Considering the whole group of patients, all but two had estimated creatinine clearance values which were below or at the lower end of the range for healthy subjects. Hence, it would seem that impaired renal excretion is an important factor in the low ranitidine clearance in our patients.

The correlation between ranitidine clearance

and estimated creatinine clearance in the present study is in agreement with previous data (Meffin et al., 1983) but should be interpreted cautiously as there may be variability in estimation of creatinine clearance in critically ill patients. Renal clearance accounts for about 70% of the total body clearance of ranitidine in healthy volunteers (Chau et al., 1982) while metabolic (hepatic) clearance presumably accounts for the balance of ranitidine clearance. Our data suggest that reduced renal clearance is an important factor in the low total body clearance observed in our patients. Reduced hepatic metabolism is less likely to be a major factor but cannot be entirely excluded since recent studies have shown that hepatic cirrhosis in patients with normal renal function results in a mild to moderate impairment of ranitidine plasma clearance (Young et al., 1982; Smith et al., 1984).

Although a gastric pH of 5 is the suggested goal of antacid therapy, a cut-off point of pH 4 was chosen in the present study on the basis that Hastings et al., (1978) found that a pH of 3.5 was adequate to control stress bleeding, and on the finding that pH levels of 5-6 are not readily obtainable with the related histamine H₂receptor blocking drug cimetidine (Priebe et al., 1980; Stothert et al., 1980). The data in Figure 2 indicate that pH measurements at 0, 1 and 7 h after ranitidine dosage are representative of the pH profile throughout the dose interval. The question of whether ranitidine maintenance dose should be reduced in this patient group is not easy to answer since overall only 58% of gastric aspirate samples had pH values which were compatible with effective suppression of acid secretion (i.e., pH > 4). This response rate occurred despite trough plasma ranitidine concentrations which were similar to (median = 0.11 mg l^{-1} , range = 0.05-1.44 mg l^{-1}) those required to cause a 50% suppression of gastric acid secretion (range of means = 0.073 - 0.2 mg l^{-1}) in healthy volunteers (Lebert *et al.*, 1981b; Mignon et al., 1982; Holloway et al., 1984). Moreover, four of our patients failed to achieve significant elevation of gastric pH at any of the times at which measurements were made. At first glance these results appear to conflict with a study by Macchi et al. (1982) in a similar group of patients in which both 10 and 20 mg doses of intravenous ranitidine given 8 hourly kept gastric pH above 5 for a mean of 18–19 h day⁻¹ with a range of between 2 and 23 h. However, More et al. (1985) also reported considerable difficulty in maintaining gastric pH above 4 in critically ill patients despite a total daily dosage of between 200 and 600 mg ranitidine. It is interesting to to note that mean plasma ranitidine concentration in our patients was similar irrespective of whether corresponding gastric pH was above or below 4. Further work is therefore necessary to quantify the plasma concentration-response relationship for ranitidine in critically ill patients and to identify possible subgroups of non-responders who may require additional anti-ulcer prophylaxis.

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