The effects on human gastric secretion of prolonged continuous intravenous infusions of maximal and supramaximal doses of histamine acid phosphate and pentagastrin

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SUMMARY The effects of continuous intravenous infusions of 'maximal' and 'supramaximal' doses of histamine acid phosphate and pentagastrin were assessed in a normal human volunteer. The secretory patterns in respect of the two drugs were indistinguishable over two and a quarter hours. During the steady states of acid secretion, outputs of pepsin and of electrolytes were also constant. The output of acid during the first hour of the steady state is a valid measurement of the maximal secretory capacity. The constant output of pepsin during the steady state of acid secretion suggests that both drugs are true stimulants of pepsin secretion.

After a single subcutaneous injection of histamine acid phosphate, as in the augmented histamine test (Kay, 1953), the output of acid rises towards a peak, which is transient and occurs at a variable time during the first hour after the administration of the drug (Baron, 1963). The transient nature of this response leaves considerable room for error in the estimation of the maximal output of acid, particularly in states characterized by low secretion (Lawrie, Smith, and Forrest, 1964). When a maximal dose of histamine or pentagastrin is administered by continuous intravenous infusion the output of acid rises to reach a plateau or steady state (Lawrie et al. 1964; Konturek and Lankosz, 1967). The calculation of the maximal secretory capacity is not then dependent on the observation and interpretation of a peak response, and hence there is less room for error in calculating the maximal output of acid than in the augmented histamine test.

The study reported now was undertaken to determine respectively (1) whether the maximal gastric secretory outputs of acid, pepsin, and electrolytes were sustained when histamine acid phosphate and pentagastrin were administered, by continuous intravenous infusion, for prolonged periods, and (2) whether the outputs of these constituents in the first hour of steadystate conditions were a valid assessment of this state.

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Methods

TESTS OF SECRETION

Twelve tests of gastric secretion were performed on one subject, a normal healthy male volunteer aged 52 years. He received in random sequence, on three occasions each, 40 and 80 μ g/kg/hr of histamine acid phosphate and 6.0 and 12.0 μ g/kg/hr of pentagastrin. Each test was preceded by a fast of at least 12 hours, and at least three days intervened between tests.

In the tests using histamine, a prior injection of mepyramine maleate was given to counteract side effects; 50 mg was given before a dose of 40 μ g/kg/hr and 100 mg before a dose of 80 μ g/kg/hr.

The stimulants were diluted in 0.9% saline, and their concentrations in solution were dependent on the dose required according to body weight. The rates of infusion were controlled by a Palmer constant infusion pump and varied from 5.37 to 11.74 ml per hour.

The technique of performing the secretory tests was identical to that described previously by Lawrie and his colleagues (1964). After emptying the stomach, basal (unstimulated) gastric juice was collected for 30 minutes, and then the stimulant was administered, by continuous intravenous infusion, for two and a quarter hours. Throughout the test, gastric juice was continuously collected in 15-minute samples.



Fig. The mean outputs of the components in gastric juice, per 15 minutes, in one normal subject, before and during stimulation by histamine acid phosphate and pentagastrin. Each 15-minute output is the mean of three correspondingly timed outputs in three tests. The infusions were begun after the second 15-minute period.

The concentrations of the hydrogen (H^+) , sodium (Na⁺), potassium (K⁺), and chloride (Cl-) ions, and of pepsin were determined, in duplicate, in each 15-minute sample of gastric juice, and the means of the duplicate values were taken. The H⁺ concentration was determined on aliquots of 10 ml of undiluted gastric juice. by titration with 0.1 N NaOH to a pH of 7.0, using a Radiometer automatic titrator (type PPPIC). The concentration of pepsin was determined on aliquots of 0.2 ml of undiluted gastric juice, using radioiodinated human albumin as the substrate, by the method of Klotz and Duvall (1957). The concentrations of the Na⁺ and K⁺ were determined, by flame photometry, using 0.1 ml aliquots of gastric juice, appropriately diluted with deionized water. The concentration of the Cl- was determined on undiluted gastric juice by means of a chloride meter. The output of a constituent of the gastric juice was calculated as the product of the volume of the sample and the concentration of the constituent in it.

Results

The pattern of secretion in response to each dose of both drugs is shown in the Figure. Each 15minute output is the mean of three correspondingly timed responses in three separate tests. The outputs of acid, chloride, and potassium increased until plateau values were reached. The output of sodium fell, but became approximately constant when the steady state of acid secretion was achieved. There was an initial transient peak output of pepsin, followed by a lower, more constant output which was in excess of that found in basal secretions.

In each test, 60-minute outputs of each constituent of the gastric juice were calculated from four successive 15-minute outputs, beginning 15 minutes after the start of the infusion. For each constituent, these 60-minute outputs were clearly similar. Those of acid and pepsin are shown in Table I.

Stimulant	Test	Acid Output (m-Equiv/60 minutes)					Pepsin Output (mg/60 minutes)					
		Time after	Time after the Start of the Infusions (minutes)									
		15 to 75	30 to 90	45 to 105	60 to 120	75 to 135	15 to 75	30 to 90	45 to 105	60 to 120	75 to 135	
Histamine	1	35.66	34.84	35.79	35.75	36.27	83·88	73.05	74.46	74·86	77.73	
40 μ g/kg/hr	2	29.08	28·16	31.20	32.27	29.47	69-01	65.75	73.36	77.62	77.19	
	3	34.74	36-35	37.38	39.22	39.83	77·22	73·33	77·71	81-11	78 ·66	
Histamine	1	33.76	34.00	33.05	32.74	32·19	83.56	78.52	75.03	76·92	73·48	
80 μg/kg/hr	2	34.52	36.20	37.01	36.96	36.66	93·13	94·32	88.00	79 ·59	81.08	
	3	36.26	38.46	38.50	37.81	37-27	98·58	94·29	91.20	86.33	87.43	
Pentagastrin	1	38.83	37.76	37-27	38.62	38·79	71·01	66.80	66·24	75.23	78.51	
6 μg/kg/hr	2	43.58	42.60	39.75	38.45	37.12	76.52	72·31	68.65	63·23	58.22	
	3	27.03	26.45	27.22	27.57	27.76	73·92	73-55	74.25	74·19	74·43	
Pentagastrin	1	41.83	39.74	38.12	37.31	37.09	84·09	79 .66	77.10	79 ·77	78·20	
12 μg/kg/hr	2	39.37	39.14	39.25	40.14	39.39	96 ∙75	100.95	100-19	104-25	99·64	
	3	40.09	41.42	40.34	40.08	39.39	81.84	82·84	80.72	76.97	80·59	

Table 1 Acid and pepsin outputs in a normal subject after stimulation by histamine and pentagastrin

Test	Histamine						Pentagastrin						
	40 µg/kg/hr			80 µg/kg/hr			6 μg/kg/hr			12 μg/kg/hr			
	1	2	3	1	2	3	1	2	3	1	2	3	
Acid	0.19	0.12	0.26	-0.06	0.13	0.02	-0.04	-0·29	0.03	-0.30	0.00	0.01	
Pepsin	0.41	-0.43	-0.11	-0-59	-1·06²	-1·12 [*]	-0.03	-0·95ª	0.33	0·29	0.46	0.01	
Sodium	0.04	0.08	0.07	-0.60	-0.02	0.04	0.06	-0.02	0.03	0.01	-0.05	0.04	
Potassium	0.06	0.01	0.02	0-04	-0.01	0.04	0.02	-0.02	-0.02	-0.04	-0.06	-0.03	
Chloride	0·65ª	1·03²	0.58	-0.12	0-09	0-03	-0.12	-0.43	0.11	-0.36	0.04	0.07	

Table II Regression coefficients of the outputs of the different constituents of the gastric juice, on time, for the last eight \times 15-minute estimations in each test ¹

¹There is a statistically significant trend when the regression coefficient > 0.6. ³Indicates that there is a statistically significant trend.

In order to determine whether there was a statistically significant alteration in the outputs of the different constituents of the gastric juice, after the first 15 minutes of stimulation, the regression coefficient of each variable, on time, was calculated in each test. Each analysis was therefore based on the last eight \times 15-minute outputs in each test. A regression coefficient larger than 0.6 would be statistically significant and would imply that there was a trend for the variable to change with time. The regression coefficients obtained in these analyses are shown in Table II. In only three of the groups of analyses were they greater than 0.6. This was in the cases of the chloride output stimulated by 40 μ g/kg/hr of histamine and the pepsin output stimulated by 80 μ g/kg/hr of histamine and 6 μ g/kg/hr of pentagastrin. In all of the remaining analyses, there was no tendency for the outputs of the various constituents to alter, with time, during the last eight \times 15-minute periods of stimulation. It is therefore justifiable to select any 60-minute period during this time to calculate the plateau hour output of any of the constituents which were measured.

Comment

There was no difference in the gastric secretory pattern when 'maximal' and 'supramaximal' doses of histamine acid phosphate and pentagastrin were administered by continuous intravenous infusion for two and a quarter hours. With both drugs, the steady state of gastric acid secretion was maintained, without evidence of fatigue, at the end of this period of time.

The output of acid in the first hour of the steady state was a valid measurement of this state. The outputs of the electrolytes and generally those of pepsin were also sustained throughout the steady state of acid secretion. The steady state, therefore, is not only one of acid secretion tut also of electrolyte and of pepsin secretion.

From the establishment of steady-state conditions with either drug, the output of pepsin was sustained at levels more than twice that found in resting gastric juice. This finding would support the suggestion that a washout of preformed pepsin does not totally explain the pepsin response following stimulation with these drugs (Bucher, Ivy, and Gray, 1941; Hirschowitz, London, and Pollard, 1957; Gillespie and Bowen, 1962; Makhlouf, McManus, and Card, 1967). The initial peak response could be compatible with such a washout effect, but the continuing steady state of pepsin secretion would suggest that both drugs are true stimulants of the pepsin-forming cells.

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