Measurement of autoantibodies against human eye muscle plasma membranes in Graves' ophthalmopathy

MARTA FARYNA, JANUSZ NAUMAN, ANDRZEJ GARDAS

Abstract

Antibodies that reacted with plasma membranes of human eye muscle but showed no binding to plasma membranes of human skeletal muscle were identified in serum of patients with Graves' ophthalmopathy. Rabbit antithyroglobulin serum at a dilution of 1×10^{-3} to 20×10^{-3} had no effect on the binding of these antibodies to eye muscle membrane antigens. There was no correlation between antihuman eye muscle plasma membrane antibodies and antihuman thyroid membrane antibodies or antibodies against thyroglobulin.

It is suggested that specific antibodies against eye muscle membranes are present in Graves' ophthalmopathy and that this disease might represent a distinct autoimmune disorder.

Introduction

The aetiology of Graves' ophthalmopathy is not fully understood, but substantial evidence indicates that it is a separate organ specific autoimmune disorder. Though cell mediated immunity seems to be well documented,1 2 the presence of specific autoantibodies directed against retro-orbital antigens has only recently been reported. Kodama et al showed serum autoantibodies reacting with soluble antigens isolated from human eye muscle cytosol.3 Kendall-Taylor et al also showed autoantibodies in the serum of patients with severe infiltrating ophthalmopathy that specifically react with antigens localised in porcine eye muscle membranes.4 In the present study we measured antihuman eye muscle plasma membrane antibodies in the serum of patients with Graves' ophthalmopathy, using the ability of protein A from Staphylococcus aureus to interact with the Fc fragment of IgG. We also attempted to study the possible correlation between these autoantibodies and antihuman thyroid membrane antibodies and antithyroglobulin antibodies.

Patients and methods

PATIENTS

We studied 20 patients (seven men, 13 women) aged 22-70 with infiltrating Graves' ophthalmopathy. The severity of the eye signs ranged from class 2 to class 6 in the classification of the American Thyroid Association.5 6 At the time of the study five patients were hyperthyroid, nine were euthyroid but had been hyperthyroid in the

Department of Biochemistry, Medical Centre of Postgraduate Education, 01-813 Warsaw, Poland MARTA FARYNA, MD, senior assistant

JANUSZ NAUMAN, MD, PHD, professor of medicine ANDRZEJ GARDAS, MSC, PHD, associate professor of biochemistry

Correspondence to: Professor J Nauman,

past, and six had no signs of thyroid disease. Nineteen healthy blood donors served as controls.

ESTIMATION OF ANTIEVE MUSCLE PLASMA MEMBRANE ANTIBODIES

Protein A (Pharmacia Fine Chemicals, Sweden) was labelled with iodine-125 by means of iodogen (1, 3, 4, 6-tetrachloro, 3α , 6α , diphenylglycouril; Pierce Co, USA) and purified as described by Gardas et al.7

Crude human eye muscle or skeletal muscle plasma membranes were prepared by the method of Amir et al.8 Both eye muscles and skeletal muscles were obtained at necropsy from subjects without a history of autoimmune disease within 24 hours of death. Antieve muscle membrane antibodies were estimated by a slight modification of the method of Gardas *et al.*⁷ The crude fraction of human eye muscle membrane (0.5 mg protein) diluted in 250 µl 10 mM phosphate buffer, pH 7.4, containing 50 mM sodium chloride and ovine albumin (1 g/l) was first preincubated with 50 μ l serum for two hours at 37°C and then left overnight at 4°C. After preincubation 1 ml phosphate buffer was added and the tubes mixed and centrifuged at 10 000 g for 10 minutes at 4° C. The supernatant was discarded and the pellet washed with 1 ml cold buffer. Eye muscle plasma membranes preincubated with serum (100 μ g protein) were then incubated with ¹²⁵I protein A (50 000 cpm) for two hours at 4°C. The reaction was stopped by adding 1 ml cold phosphate buffer, and samples were centrifuged at 10 000 g for 10 minutes at 4° C. The supernatant was discarded and the radioactivity in the resulting pellet counted in a 80 000 Wallac-LKB counter.

The final results were expressed as an antieye muscle membrane antibody index. The index was calculated by dividing the percentage binding of ¹²⁵I protein A to plasma membranes preincubated with test serum minus non-specific binding by the percentage binding of ¹²⁵I protein A to plasma membranes preincubated with normal serum minus non-specific binding. Non-specific binding was the binding of ¹²⁵I protein A to membranes preincubated with buffer only.

OTHER PROCEDURES

Antithyroid plasma membrane antibodies were estimated by the method of Gardas et al.7 Further studies have shown that an antithyroid plasma membrane antibody index above 1.6 should be considered to be positive for the presence of antithyroid plasma membrane antibodies.⁹

Antithyroglobulin antibodies were estimated by radioimmunoassay as described by Kielczynski.10 Antithyroglobulin antibodies with a titre of $1/4 \times 10^6$ were obtained by immunisation of rabbits with highly purified human thyroglobulin.10 The Cochran-Cox test was used to evaluate the significance of the results.

Results

Values of the antihuman eye muscle plasma membrane antibody index in patients with Graves' ophthalmopathy varied from 0.81 to 2.17 and in controls from 0.53 to 1.55 (fig 1). The upper limit of normal for the index was therefore chosen as 1.55. The mean index in patients and controls was significantly different (1.66 (SE 0.33) v1.00 (0.29), p < 0.001).

When human skeletal muscle plasma membranes were used as a source of antigen no significant difference in antihuman eye muscle plasma membrane antibody index was found between the patients with Graves' ophthalmopathy and the controls (1.04 (0.22) v 1.00 (0.11)).

To exclude a possible role of thyroglobulin in the binding of antihuman eye muscle plasma membrane antibodies to putative antigens, preincubation mixtures of pooled serum from patients and from

controls were enriched with rabbit antithyroglobulin antibodies at dilutions from 1×10^{-3} to 20×10^{-3} . At these dilutions the thyroglobulin antibodies had no effect on ¹²⁵I protein A binding to membranes preincubated with serum from either the patients or the controls (fig 2).



FIG 1—Antihuman eye muscle plasma membrane antibody (AEMA) index in patients with Graves' ophthalmopathy and controls.



FIG 2-Influence of antithyroglobulin antibodies on ¹²⁵I protein A binding to eye muscle plasma membranes preincubated with pooled serum from healthy blood donors (O---O), or buffer alone (---).

To investigate whether antibodies against eye muscle plasma membranes and thyroid plasma membranes represent the same type of IgG the antihuman eye muscle plasma membrane antibody and antihuman thyroid membrane antibody indexes obtained in the same patients were compared, but no correlation could be demonstrated (r = -0.07). Similarly, there was no correlation between the antihuman eye muscle plasma membrane antibody index and titres of thyroglobulin antibodies in the same patients (r = 0.22).

Discussion

Using the ability of protein A to interact with Fc fragment of IgG we were able to show the presence of antibodies that react with human eye muscle plasma membranes in most patients with Graves' ophthalmopathy. We also showed that these antibodies are specific as they did not react with human skeletal muscle plasma membranes. The presence of such antibodies was recently reported by Kendall-Taylor et al, who also emphasised their high specificity.4 We detected these antibodies in 14 (70%) of patients with Graves' ophthalmopathy. Similar results were obtained by Kodama et al, who used soluble human eye muscle antigens to investigate specific antibodies and obtained positive results in 75% of patients studied.3 Two possibilities might explain the "negative" values of the antihuman eye muscle plasma membrane antibody index in some of our patients. As some of the patients had a long history of eye disease they may possibly have achieved an immunological remission. Alternatively, a negative index might be due to the relatively low sensitivity of the assay as a result of using tissues obtained at necropsy.

In previous studies Konishi et al reported an affinity of eye muscle for thyroglobulin.11 Mullin et al also suggested that putative antigens present in retro-orbital tissues contain either thyroglobulin or material like thyroglobulin.12 Using specific antibodies against human thyroglobulin in a wide range of concentrations we were unable to alter the binding of IgG from patients or controls to human eye muscle plasma membranes. These results exclude the presence of thyroglobulin in human eye muscle membranes as well as the possibility that thyroglobulin, within the eye muscle membrane, could be a component of the antigen or antigens that react with autoantibodies present in the serum of patients with Graves' ophthalmopathy.

The studies of Kodama et al and Kendall-Taylor et al strongly suggest that Graves' ophthalmopathy is a separate organ specific autoimmune disorder that for reasons yet unknown commonly occurs in association with hyperthyroidism.³ ⁴ In our previous studies we have shown that antithyroid plasma membrane antibodies are present in almost all patients with Graves' disease.13 The lack of correlation between indexes of antihuman eye muscle plasma membrane antibodies and antihuman thyroid membrane antibodies in individual patients with Graves' ophthalmopathy also provides support for the possibility that ophthalmopathy is an independent autoimmune disorder. Further studies are necessary to investigate the possible correlations of the level of orbital antibodies with the duration of the eye disease and with the severity of the disease.

We thank Dr J Kupryjanczyk, from the department of pathology, for collecting human eye muscles, and Professor R Hall, of Cardiff, for his advice in the preparation of this manuscript.

References

- 1 Manhieu R, Winand R. Demonstration of delayed hypersensitivity to retrobulbar and thyroid tissues in human exophthalmos. J Clin Endocrinol Metab 1972;34:

- Manhieu R, Winand R. Demonstration of delayed hypersensitivity to retrobulbar and thyroid tissues in human exophthalmos. J Clin Endocrinol Metab 1972;34: 1090-2.
 Munro RE, Lamki L, Row VV, Volpe R. Cell mediated immunity in the exophthalmos of Graves' disease as demonstrated by the migration inhibition factor (MIF) test. J Clin Endocrinol Metab 1973;37:286-92.
 Kodama K, Sikorska H, Bandy-Dafoe P, Bayly R, Wall JR. Demonstration of a circulating autoantibody against a soluble eye-muscle antigen in Graves' ophthalmopathy. Lancet 1982;ii:1353-6.
 Kendall-Taylor P, Atkinson S, Holcombe M. A specific IgG in Graves' ophthalmopathy and its relation to retro-orbital and thyroid autoimmunity. Br Med J 1984;289:1183-6.
 Werner SC. Classification of the eye changes in Graves' disease. J Clin Endocrinol Metab 1969;29:982-4.
 Werner SC. Modification of the classification of the eye changes of Graves' disease: recommendations of the ad hoc committee of the American Thyroid Association. J Clin Endocrinol Metab 1977;44:203-4.
 Gardas A, Czarnocka B, Nauman J. New simple and sensitive method of estimation of anti-thyroid diseases. Endokrynol Pol 1982;33:295-301.
 Amir SM, Carraway TF, Kohn LD, Winand RJ. The binding of thyrotropin to isolated bovine thyroid plasma membrane antibodies in serum of patients with autoimmune thyroid diseases, comparison with other assays. Acta Endocrinol (Copenh) 1984;105:492-9.
 Kielczynski K. Radioimmunological determination of thyroglobulin and anti-bodies against thyroglobulin in serum. Endokrynol Pol 1982;33:246-55.
 Konish J, Herman MM, Kriss JP. Binding of thyroglobulin and thyroglobulin-antithyroglobulin in normal human orbital muscle. Endocrinology 1977;97:100: 351-66.
 Mulin BR, Levinson RE, Freidman A, Henson DE, Winand RJ, Kohn LD. Delayed hypersensitivity in Graves' 198-9.
 Mullin BR, Levinson RE, Freidman A, Henson DE, Winand RJ, Kohn LD.

- 13 Gardas A, Czarnocka B, Nauman J. The presence of autoantibodies directed to thyroid plasma membrane antigens in sera of patients with thyroid disorders, estimated by the reaction with labelled protein A. Acta Endocrinol (Copenh) 1984;105:500-4.

(Accepted 26 September 1984)