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Regulation of bone mass, bone loss and osteoclast activity by cannabinoid receptors

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Abstract

Accelerated osteoclastic bone resorption plays a central role in the pathogenesis of osteoporosis and other bone diseases. Identifying the molecular pathways that regulate osteoclast activity provides a key to understanding the causes of these diseases and to the development of new treatments. Here we show that mice with inactivation of cannabinoid type 1 (CB₁) receptors have increased bone mass and are protected from ovariectomy induced bone loss. Pharmacological antagonists of CB₁ and CB₂ receptors prevented ovariectomy induced bone loss *in vivo* and caused osteoclast inhibition *in vitro* by promoting osteoclast apoptosis and inhibiting production of several osteoclast survival factors. These studies show that the CB₁ receptor plays a role in the regulation of bone mass and ovariectomy induced bone loss and that CB₁ and CB₂ selective cannabinoid receptor antagonists are a novel class of osteoclast inhibitors that may be of value in the treatment of osteoporosis and other bone diseases.

Introduction

Osteoclasts are cells derived from the monocyte – macrophage lineage that play an important role in modelling bone during skeletal growth and in remodelling bone during adult life(1). Increased osteoclast activity or uncoupling of osteoclastic bone resorption from bone formation results in focal or generalised bone loss and is a characteristic feature of bone diseases such as osteoporosis, Paget's disease of bone, and cancer associated bone disease(2). The importance of osteoclastic bone resorption in the pathogenesis of these disease is reflected by the fact that the most successful drug treatments for bone disease work by inhibiting bone resorption(3). Osteoclastic bone resorption is regulated by a complex interplay between circulating calcitropic hormones like parathyroid hormone, calcitriol and sex hormones; and local regulators of bone cell activity like receptor activator of nuclear factor kappa B ligand (RANKL), macrophage colony stimulating factor (M-CSF) and osteoprotegerin(4). Recent work has shown that neuroendocrine pathways and neurotransmitters also play a key role in the regulation of bone remodelling(5–9). In view of this, we investigated the role of the endocannabinoid system in the regulation of bone mass and bone turnover by studying the skeletal phenotype in mice with targeted inactivation of cannabinoid type 1 (CB₁) receptors (CB₁ KO mice) and by studying the effects of cannabinoid receptor ligands on bone cell function *in vitro* and ovariectomy induced bone loss *in vivo*.

Results

Skeletal Phenotype of CB₁ KO mice

We found that CB₁ KO mice had significantly increased bone mineral density (BMD) when compared with wild type (WT) littermates. For example, the levels of BMD assessed by dual X-ray absorptiometry (DXA) at the femur were 18% higher in CB₁ KO animals than WT and values at the spine were 10% higher (Figure 1, panel a). Further evaluation using peripheral quantitative computed tomography (pQCT) showed that trabecular BMD values at the tibial metaphysis in CB₁ KO mice were 16% higher than in WT (Figure 1, panel b) and this difference was clearly evident on low power photomicrographs at this site (Figure 1 panel c). Bone histomorphometry showed that CB₁ KO mice had significantly increased trabecular bone volume at the tibial metaphysis compared with wild type mice, consistent with the BMD measurements, but no difference was observed between CB₁ KO mice and wild type littermates in terms of osteoclast numbers, eroded surfaces or osteoblast numbers (Supplementary Table 1). This indicates that absence of the CB₁ receptor regulates BMD but does not influence bone turnover appreciably in normal adult mice. In order to determine whether the CB₁ receptor plays a role in the regulation of bone loss, we studied the effects of ovariectomy in CB₁ KO and WT mice. We found that CB₁ KO mice were completely resistant to trabecular bone loss induced by ovariectomy when compared with WT; analysis by pQCT showed no significant change in total BMD of the tibial metaphysis in CB₁ KO mice following ovariectomy, compared with a 12% reduction in WT (Figure 2, panel a). Analysis by micro-computed tomography (μ CT) showed a 40% reduction in trabecular bone volume following ovariectomy in WT mice (Figure 2, panel b); a 12% reduction in trabecular thickness (Figure 2, panel c) and a 30% reduction in trabecular number (Figure 2, panel d) whereas none of these variables changed significantly in CB₁ KO mice. Uterine weight (mean \pm standard deviation) fell to a similar degree after ovariectomy in WT mice (0.38 ± 0.03 vs. 0.08 ± 0.01 g; $p < 0.01$) and CB₁ KO mice (0.43 ± 0.02 vs. 0.09 ± 0.01 g; $p < 0.01$). These data indicate that the CB₁ receptor plays an essential role in regulating bone loss that results from estrogen deficiency, but that the gonadal response to ovariectomy is unaffected by CB₁ deficiency.

Effects of CB receptor ligands on osteoclast function

In order to further explore the mechanisms by which the CB₁ pathway regulates bone mass and bone loss, we studied the effects of various cannabinoid receptor agonists and antagonists on bone cell function *in vitro* using primary mouse osteoblast cultures and RANKL-generated mouse osteoclast cultures. None of the CB ligands that we tested significantly affected osteoblast growth or viability at concentrations of up to 20 μ M (data not shown), but significant effects on osteoclast activity were observed using ligand concentrations in the nanomolar range. The CB₁-selective antagonist AM251(10) and the CB₂-selective antagonists SR144528 and AM630(10) significantly inhibited osteoclast formation in RANKL and M-CSF stimulated mouse bone marrow cultures in a concentration dependent manner with IC₅₀ values of 700nM for AM251; 850nM for SR144528 and 100nM for AM630 (Figure 3, panel a). Conversely, the endogenous cannabinoid receptor agonist anandamide (AEA) and the non-selective synthetic agonist CP55940 stimulated osteoclast formation in a concentration dependent manner between 100nM and 5 μ M (Figure 3, panel a). Representative photomicrographs from cultures treated with vehicle, AEA, AM251 and SR144528 are shown in Figure 3, panel b. Moreover, AEA reversed the inhibitory effect of AM251 on osteoclast formation (Figure 3, panel c), consistent with a receptor mediated effect.

Expression and functional role of CB₁ and CB₂ receptors on osteoclasts

Both the CB₁ and CB₂ receptor subtypes were expressed on mouse osteoclasts as assessed by immunohistochemical staining (Supplementary Figure 1, panel a), western blotting (Supplementary Figure 1, panel b) and RT-PCR (Supplementary Figure 1, panel c), indicating

that CB₁, CB₂ or both receptors could mediate the effects of CB ligands on osteoclast activity. In order to clarify the relative role of the CB₁ and CB₂ receptors as targets for the regulation of osteoclast activity, we compared the effects of AM251 and AM630 on RANKL induced osteoclast formation in cultures generated from CB₁ KO mice and wild type littermates. Cultures prepared from CB₁ KO mice were resistant to the inhibitory effects of AM251 on osteoclast formation when compared with WT cultures (Figure 4, panel a). The CB₂ selective antagonist AM630 inhibited osteoclast formation to a similar extent in cultures prepared from CB₁ KO and WT mice and was an order of magnitude more potent than AM251 (Figure 4, panel b). Taken together these data indicate that the CB₁ receptor contributes in part, to the osteoclast inhibition which occurs *in vitro* when these cultures are exposed to CB antagonists but that osteoclast inhibition can also occur through antagonism of CB₂ receptors.

Effects of CB antagonists on ovariectomy induced bone loss

Since CB₁ KO mice were resistant to ovariectomy induced bone loss, we wanted to determine if pharmacological blockade of CB receptors could prevent the bone loss that occurs as the result of ovariectomy. We studied the effects of AM251 in doses of 0.3–3.0 mg/kg/day on ovariectomy induced bone loss in wild type mice and SR144528 at a single dose of 3 mg/kg/day. These doses were chosen on the basis of previous work, which has shown that AM251 and the related compound SR141716 prevent diet induced obesity in wild type mice in doses of 3–30 mg/kg/day (11–13). We found that AM251 protected against ovariectomy induced bone loss in a concentration-dependent manner at doses as low as 0.3 mg/kg/day (Figure 5, panels a – c). Bone histomorphometry showed that osteoclast numbers and active resorption surfaces were increased following ovariectomy in vehicle treated animals, whereas no significant increase in either variable was observed in AM251 treated animals (Supplementary Table 2). Osteoblast numbers were unaffected by AM251 treatment, indicating that the protective effect of CB receptor blockade on ovariectomy induced bone loss was primarily mediated by inhibiting bone resorption, rather than by stimulating bone formation. Administration of SR144528 to ovariectomized mice in a dose of 3 mg/kg/day also completely prevented ovariectomy induced bone loss, with results almost identical to those observed with AM251 at 3 mg/kg/day (data not shown).

Effects of CB ligands on osteoclast apoptosis and signalling pathways

We investigated the molecular mechanisms by which CB receptor blockade inhibits bone resorption, by studying the effects of CB receptor antagonists on apoptosis and key intracellular signalling pathways involved in osteoclast survival using purified rabbit osteoclasts and RANKL generated mouse osteoclasts. We found that AM251 and SR144528 stimulated apoptosis of rabbit and mouse osteoclasts by 12–17 fold above control values (Supplementary Figure 2, panel a), as detected by deoxynucleotidyl transferase-mediated dUTP nick end labelling (TUNEL) and typical changes in nuclear morphology on DAPI stained cultures (Supplementary Figure 2, panel d) Activation of caspase 3 was observed in both rabbit and mouse osteoclasts that were exposed to CB antagonists (Supplementary figure 2 panel b) and this was accompanied by a reduction in osteoclast numbers, and inhibition of bone resorption (Supplementary figure 2, panel c; data from rabbit osteoclasts shown). The only significant difference between the response of mouse and rabbit osteoclasts to CB antagonists was that maximal stimulation of apoptosis occurred 2 μM in mouse osteoclasts compared with 10 μM in rabbit osteoclasts, presumably because of species differences in affinity of the receptors for the ligands tested. We also investigated the effects of AM251 on activation of key survival factor in osteoclasts including extracellular signal regulated kinase (ERK), the p65 component of NFκB, *NFATc1*; *c-jun* and *c-fos* (14–16). These studies showed that AM251 at a concentration of 2 μM abolished RANKL induced ERK phosphorylation (Figure 6, panel a) and significantly inhibited RANKL induced nuclear translocation and DNA binding of

NFATc1, *c-jun* and *c-fos* (Figure 6, panels b, c and d). In contrast, AM251 did not inhibit RANKL induced NFκB (p65) activation in this assay (data not shown).

Discussion

The studies presented here indicate that the CB₁ receptor plays a hitherto unrecognised role in the regulation of bone mass and bone loss resulting from estrogen deficiency. Adult CB₁ KO mice had significantly higher BMD than WT littermates at several skeletal sites and CB₁ KO mice were completely protected against ovariectomy induced bone loss. The synthetic cannabinoid receptor antagonists AM251, SR144528 and AM630 inhibited osteoclast formation and bone resorption *in vitro* and both AM251 and SR144528 protected against ovariectomy induced bone loss *in vivo* by inhibiting osteoclastic bone resorption. Conversely, the endogenous CB agonist anadamide, and the synthetic agonist CP55940 enhanced osteoclast formation *in vitro* and anadamide reversed the inhibitory effects of AM251 osteoclast formation, consistent with a receptor mediated effect. Osteoclasts generated from CB₁ KO mice were resistant to the inhibitory effects of AM251 compared with WT mice, consistent with the hypothesis that the osteoclast inhibition we observed with CB antagonists was mediated in part, by the CB₁ receptor. We also gained evidence to show CB₂ receptor mediated osteoclast inhibition can also occur *in vitro*, from the observation that the CB₂-selective antagonist AM630 inhibited osteoclast formation equally well in cultures prepared from CB₁ KO and WT mice. The inhibitory effects on osteoclast formation that we observed at high AM251 concentrations (>1000nM) in CB₁ KO cultures were probably attributable to blockade of the CB₂ receptor by this compound(10). The extent to which CB₁ versus CB₂ receptors contributed to the prevention of ovariectomy induced bone loss that we observed with CB antagonist treatment *in vivo* remains to be determined, as does the relative contribution of central versus peripheral CB receptors in mediating this effect. Further work will also be required to determine whether the osteoclast inhibitory effects we observed were due to antagonism of the action of endogenous CB receptor ligands or to inverse agonism, since AM630, AM251 and SR144528 have all been shown to elicit effects at CB receptors that are converse to those produced by CB agonists, even in the absence of agonist binding(17).

The mechanism of osteoclast inhibition resulting from CB receptor blockade was most likely due to osteoclast apoptosis since AM251 and SR144528 markedly enhanced apoptosis of mature rabbit osteoclasts and of RANKL generated mouse osteoclasts. Exposure of mouse osteoclasts to AM251 blocked RANKL-induced ERK phosphorylation and prevented activation of several key osteoclast stimulatory factors including *NFATc1*, phospho *c-jun* and *c-fos*. Interestingly however, AM251 had no inhibitory effect NFκB activation suggesting that cannabinoid receptor antagonists may cause osteoclast inhibition through the ERK – AP1 pathway, rather than the NFκB pathway.

The phenotype that we observed in CB₁ KO mice indicates that the CB₁ receptor plays an essential role in regulating BMD and bone loss that results from oestrogen deficiency. Furthermore, the pharmacological studies *in vitro* show that the CB₁ and CB₂ receptors may have overlapping functions in the regulation of osteoclast activity. Further studies of the skeletal phenotype in CB₂ KO mice will be required to fully evaluate the role of the CB₂ receptor subtype on bone metabolism and osteoclast function *in vivo*.

The results of our studies are important since we have shown that pharmacological antagonists of CB receptors represent a novel class of osteoclast inhibitors, whereas CB receptor agonists act as stimulators of bone resorption. This suggests that CB receptor antagonists may represent a promising new class of antiresorptive drugs for the treatment of osteoporosis and other bone diseases associated with increased osteoclast activity. Our data also raised the possibility that

recreational or therapeutic use of cannabis derivatives that act as agonists at CB receptors might enhance bone loss and predispose to osteoporosis.

Materials and Methods

Cell culture—Mouse osteoclast cultures were generated by isolating mononuclear cells from the bone marrow of mouse long bones essentially as described by Takahashi(18). The cells were cultured in α MEM with 10% fetal calf serum and Macrophage colony stimulating factor (M-CSF) (100ng/ml) for 3 d. Adherent cells were collected and plated at 0.5×10^4 cells/well in 96-well plates in 125 μ l of α MEM supplemented with 10% FCS, M-CSF (25ng/ml) and RANKL (100ng/ml) for 4 d. Test substances were added on d 7 and the cultures were terminated on d 10. Rabbit osteoclast cultures were performed as described(19) and exposed to test substances for 48 h. Osteoclast formation was assessed by counting Tartrate resistant acid phosphatase positive multinucleated cells and bone resorption was assessed by measuring resorption pit area using reflected light microscopy as described(20). Apoptosis was detected by the characteristic changes in nuclear morphology following 4,6-diamidino-2-phenylindole (DAPI) staining as previously described(20), and by detection of fragmented DNA by terminal deoxynucleotidyl transferase-mediated dUTP nick end labelling (TUNEL) using the ApopTag Plus Fluorescein In Situ Apoptosis Detection Kit (Intergen). Cells with clearly fragmented DNA or positive ApopTag labelling were counted and expressed as a percentage of total cell number.

Immunofluorescence—Immunofluorescent staining of mouse osteoclasts for CB receptors was performed using a rabbit antibody to the CB₁ receptor(21) (Caymen Chemical, catalog number 101500) and a rabbit antibody to the CB₂ receptor(22) (Caymen Chemical, catalog number 101550) followed by Alexa-Fluor 488 goat-anti-rabbit IgG (Molecular Probes). Controls were performed by omitting the primary antibody. Osteoclasts were identified by dual labelling with a mouse anti-CD61 antibody directed against the vitronectin receptor (Pharmagen) followed by Alexa-Fluor 594 goat-anti-mouse IgG.

Western Blotting—Intact (p-30) and activated (cleaved, p-19) caspase-3 levels were detected by western blotting using a goat polyclonal anti-caspase-3 antibody (Santa Cruz Biotechnology, Inc), and a rabbit polyclonal anti-cleaved caspase-3 antibody (Cell Signalling Technology). Detection of native and phosphorylated ERK was by western blotting using antibodies to ERK and phospho-ERK (Cell Signalling Technology). Detection of CB₁ and CB₂ receptors was by western blotting using the same primary antibodies as for immunofluorescence, with an anti-rabbit secondary antibody linked to horse radish peroxidase (Caymen Chemical). The chemiluminescent signal was developed using SuperSignal (Pierce).

Molecular biology—Activation and DNA binding of *NFATc1*, NF κ B (p65), phospho *c-jun* and *c-fos* were assessed by analysis of nuclear extracts using TRANS-AM transcription factor assay kits (Active Motif). Detection of CB₁ and CB₂ receptor mRNA by reverse transcription polymerase chain reaction (RT/PCR) was performed using Qiagen Quantitect custom primers for the CB₁ receptor (primers 5'-CCTTGCAGATACAACCTT-3' and 5'-TGCCATGTCTCCTTTGATA-3'; predicted product size 94bp) and the CB₂ receptor (primers 5'-CATAAGCCGATCTCTCAA-3' and 5'-CCAAAGCTGGTGCAGGAA-3'; predicted product size 82bp). The PCR protocol consisted of an initial incubation at 95°C for 15 min., followed by 40 cycles of 94°C for 15 s., 56°C for 30 s. and 76°C for 30 s. All reactions were run with no template and no RT controls.

Drugs

All compounds were dissolved in dimethylsulfoxide and added to the cultures in such that the final concentration was 0.1%. Control cultures were treated with dimethylsulfoxide alone at the same concentrations. AM251 and AM630 were obtained from Tocris. SR144528 was synthesised by the coupling of 1-(4-Methylbenzyl)-5-(4-chloro-3-methylphenyl)pyrazole-3-carboxylic acid chloride and (1S)-2-endo,exo-amino-1,3,3-trimethylbicyclo[2.2.1]heptane. The identity and purity of SR144528 was verified using nuclear magnetic resonance spectroscopy.

Animals

Animal experiments were approved by the ethical review board of the University of Aberdeen and were conducted in accordance with UK Home Office regulations. The strain of CB₁ KO mice and wild type littermates used in this study were as described by Pryce and colleagues (23). Experiments were also performed using C57BL/6 mice obtained from Harlan Laboratories. Ovariectomy or sham ovariectomy was performed in 9 week old adult female mice as previously described(24). Treatment with AM251 and SR144528 was commenced 2 days after ovariectomy or sham ovariectomy by intraperitoneal administration of the drug in corn oil. Controls received corn oil alone. The treatment was continued for 19 d and the experiment terminated on d 21. Eight mice were studied per group. Bone mineral density was measured by DXA at the spine and femur using a PIXIMUS scanner; and at the tibial metaphysis by pQCT using a Stratec Research M scanner or by micro computed tomography (μ CT), using a Skyscan 1072 scanner. Following BMD scanning the limbs were embedded and processed for bone histomorphometry as previously described(24).

Statistics

Between group comparisons were by ANOVA with Dunnet's post-test. IC₅₀ values were calculated from concentration response curves using the GraphPad PRISM computer program (GraphPad Software).

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

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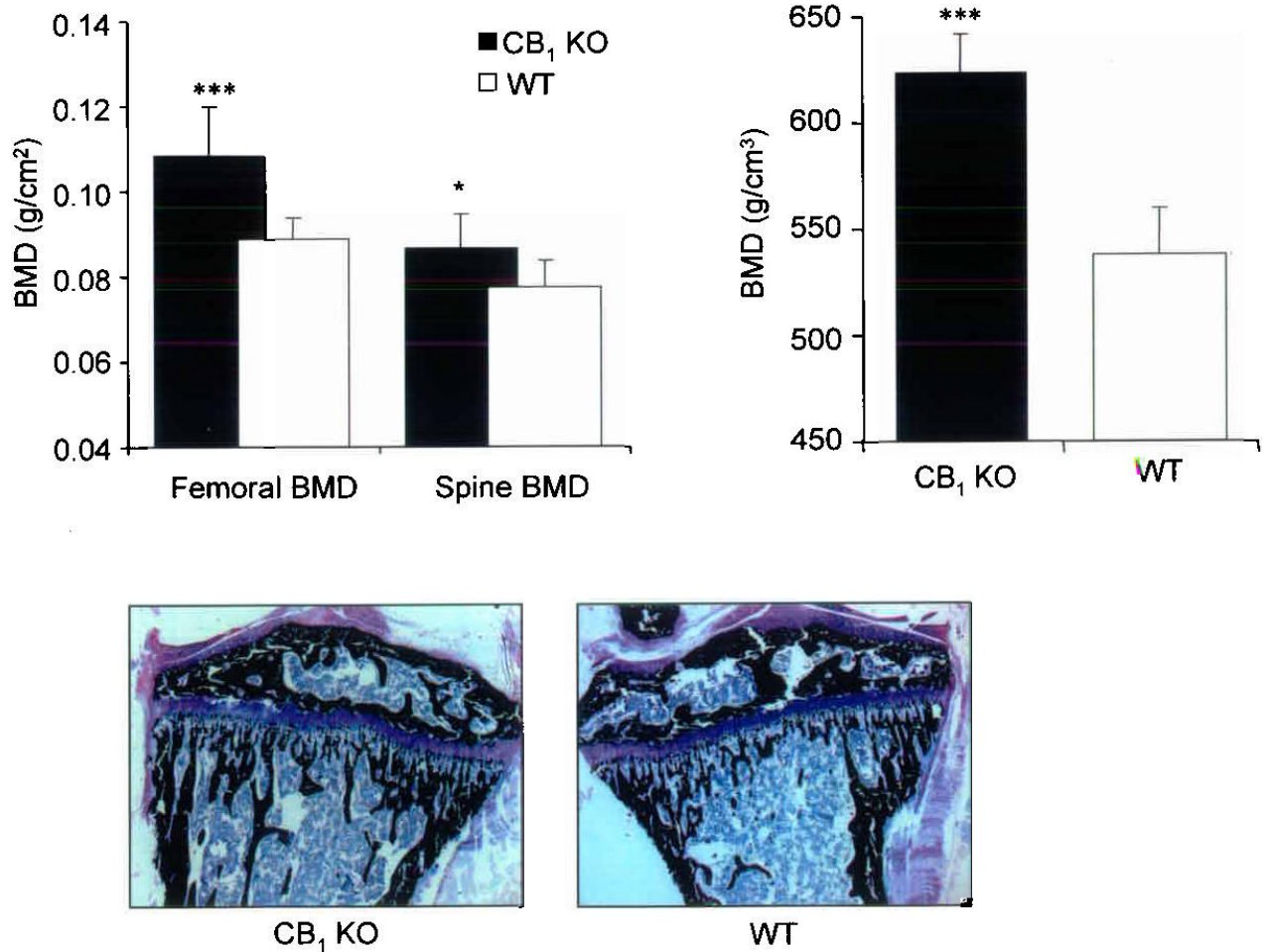


Figure 1. CB₁ KO mice have increased bone mass

a, Bone mineral density at the spine and femur assessed by DXA in CB₁ KO mice and wild type littermates; **b** trabecular bone mineral density at the tibial metaphysis assessed by pQCT in CB₁ KO mice and wild type littermates; **c**, representative photomicrograph of the proximal tibia from and CB₁ KO mice (left panel) and wild type mice (right panel) Values in the bar charts are means and standard errors. Significant differences between CB₁ KO and wild type mice are indicated by: *** $p < 0.001$; * $p < 0.02$.

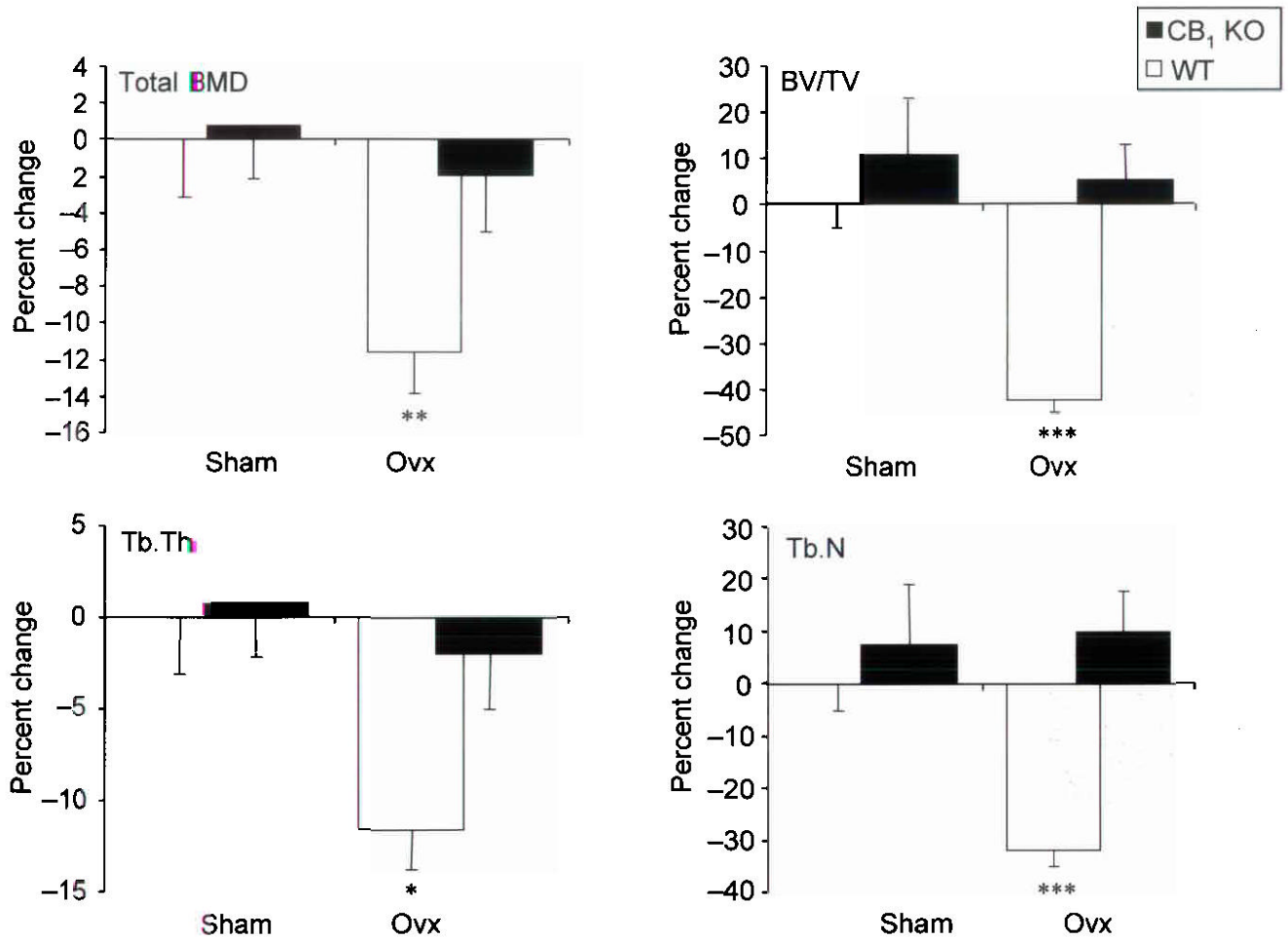


Figure 2. CB₁ KO mice are protected against ovariectomy induced bone loss

a, Total BMD at the tibial metaphysis in CB₁ KO mice and wild type littermates before and after sham operation or ovariectomy (ovx); **b**, Bone volume / total volume (BV/TV) assessed at the same site by μ CT; **c**, Trabecular thickness (Tb.Th) assessed by μ CT; **d** trabecular number (Tb.N) assessed by μ CT. Values in the bar charts are expressed as the percent change relative to the value in sham operated wild type animals and are means and standard errors. Significant differences between CB₁ KO and wild type mice are indicated by: *** p<0.001; ** p<0.01.

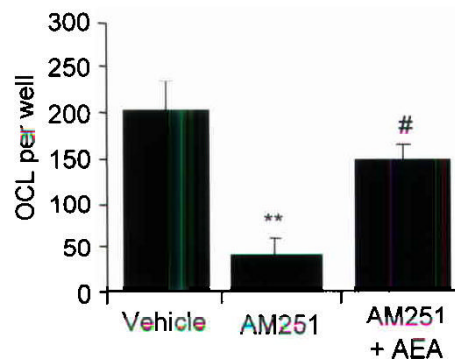
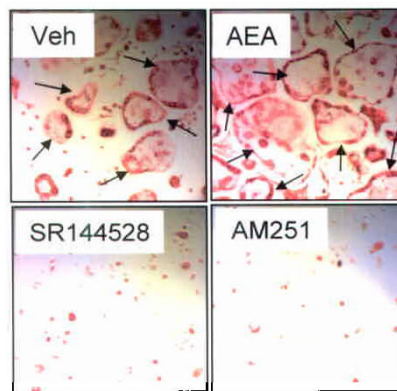
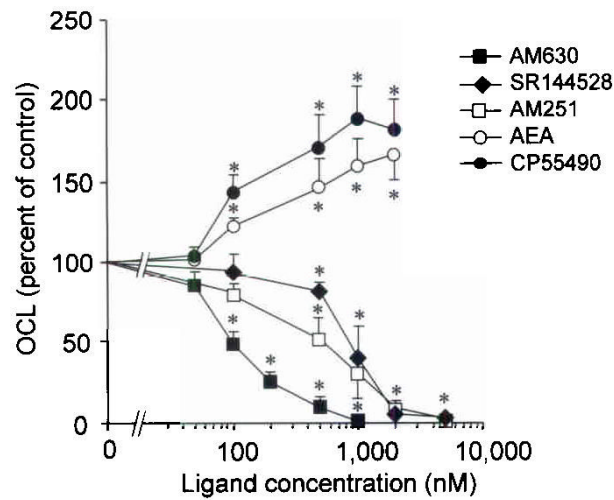


Figure 3. Regulation of osteoclast formation by cannabinoid receptor ligands
a, Effects of the CB ligands on osteoclast formation in C57BL/6 bone marrow cultures stimulated with RANKL and M-CSF. The data shown are from three independent experiments and are expressed as a percent of values in control cultures.. Significant inhibition or stimulation of osteoclast formation when compared with control is indicated by * $p < 0.05$ or below. **b**, representative photomicrographs of vehicle treated culture and cultures treated with anandamide (AEA), SR144528 and AM251 stained for TRAcP. Osteoclasts (arrows) were larger and more numerous in the AEA treated cultures when compared with vehicle and were virtually absent from the SR144528 and AM251 treated cultures. **c**, effect of AM251 alone (1 μ M) and AM251 in combination with AEA (5 μ M) on osteoclast formation. Values in the

bar charts and line graphs are means and standard deviations. Significant differences are indicated by: *** $p < 0.001$ (vehicle vs. AM251); # $p < 0.02$ (AM251 vs. AM251 + AEA).

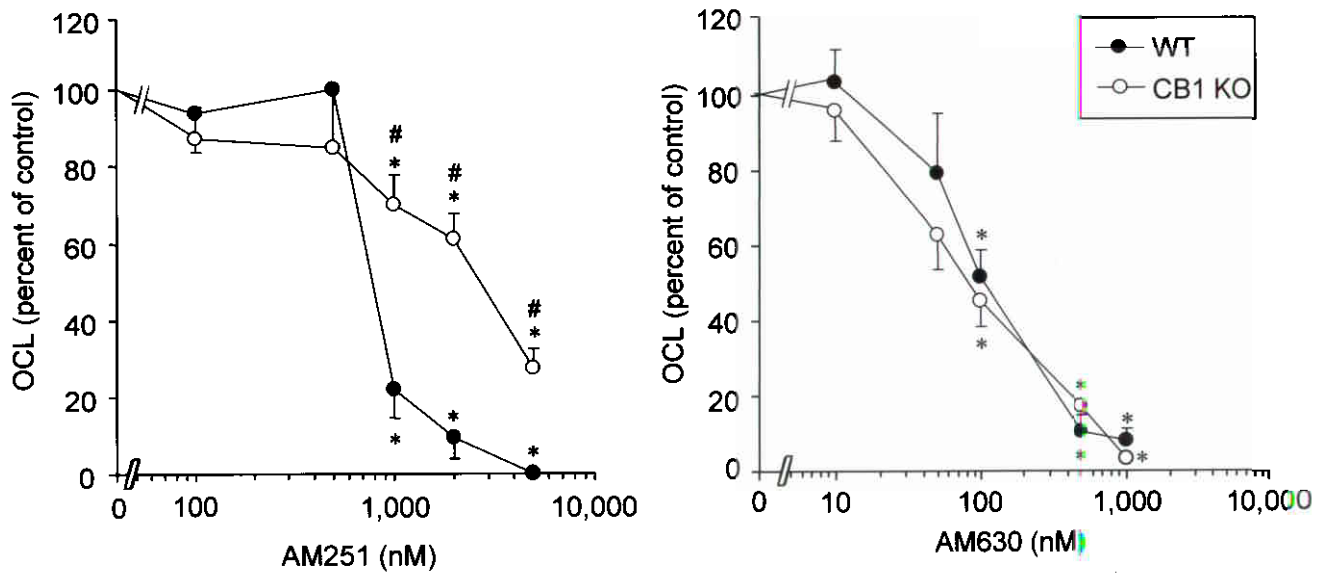


Figure 4. Osteoclasts generated from CB1 KO mice are resistant to the inhibitory effects of CB₁ selective, but not CB₂ selective receptor antagonists
a, effect of AM251 on osteoclast formation in RANKL / M-CSF stimulated bone marrow cultures prepared from CB₁ KO mice and wild type littermates. *b*, effect of AM630 on osteoclast formation in bone marrow mononuclear cells cultured from wild type and CB₁ KO mice. Values are means and standard deviations. Significant differences from control cultures are indicated by * p<0.05. Significant differences between CB₁ KO and wild type cultures are indicated by # p<0.05.

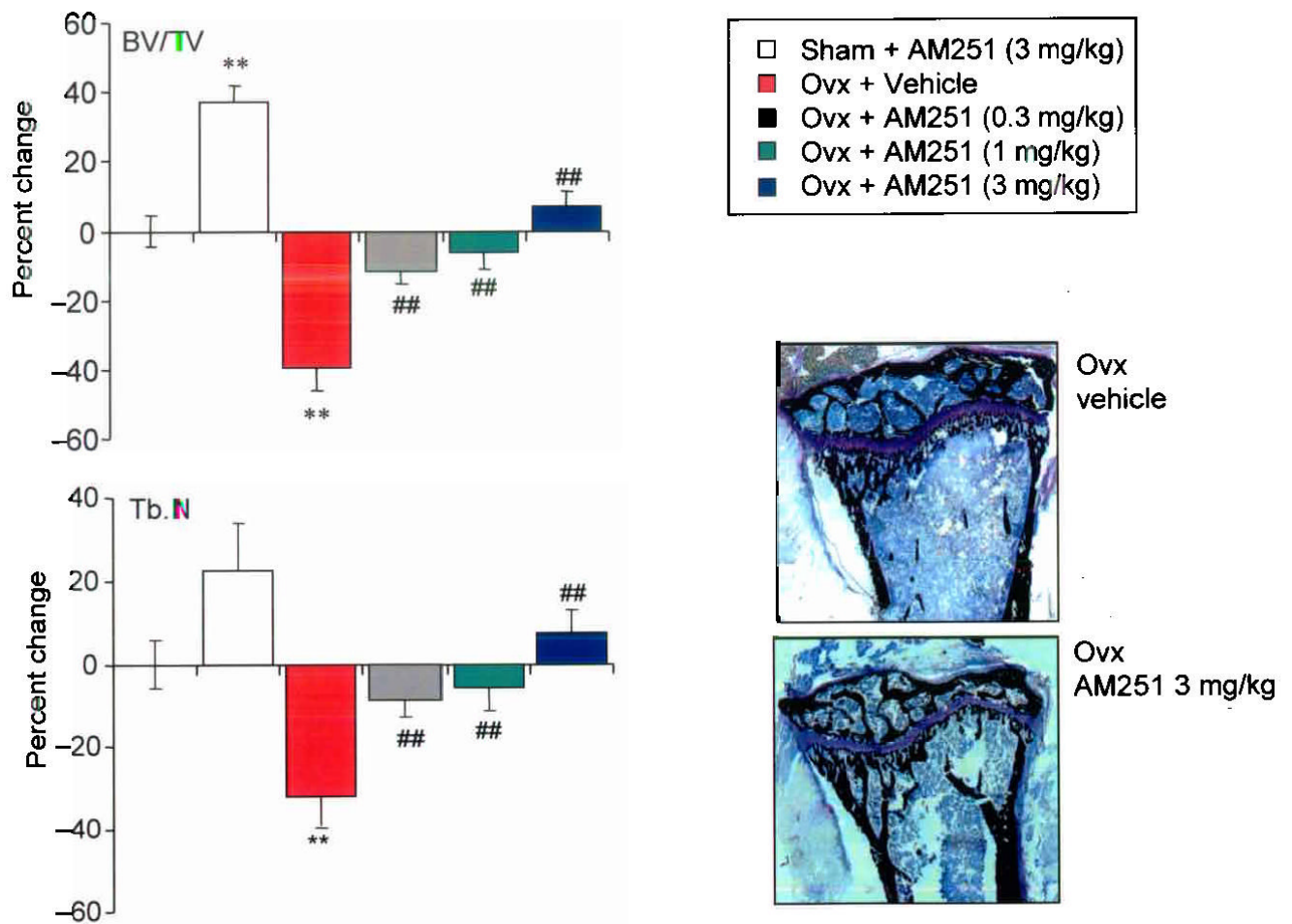


Figure 5. Cannabinoid receptor antagonists prevent ovariectomy induce bone loss
a. Effects of AM251 on trabecular bone density in sham operated (sham) and ovariectomized (ovx) C57BL/6 mice; **b.** Effects of AM251 on trabecular number in sham operated (sham) and ovariectomized (ovx) C57BL/6 mice. Values are expressed as percent change relative to sham operated vehicle treated control mice. **c.** representative photomicrograph of the tibial metaphysis 21 days after ovariectomy in a vehicle treated mouse (top panel) and an AM251 treated mouse (bottom panel). Values in the bar charts are means and standard errors. Significant differences between sham OvX and sham OvX + AM251 and between sham OvX and OvX are indicated by ** $p < 0.001$; significant differences between OvX and OvX + AM251 are indicated by ## $p < 0.002$.

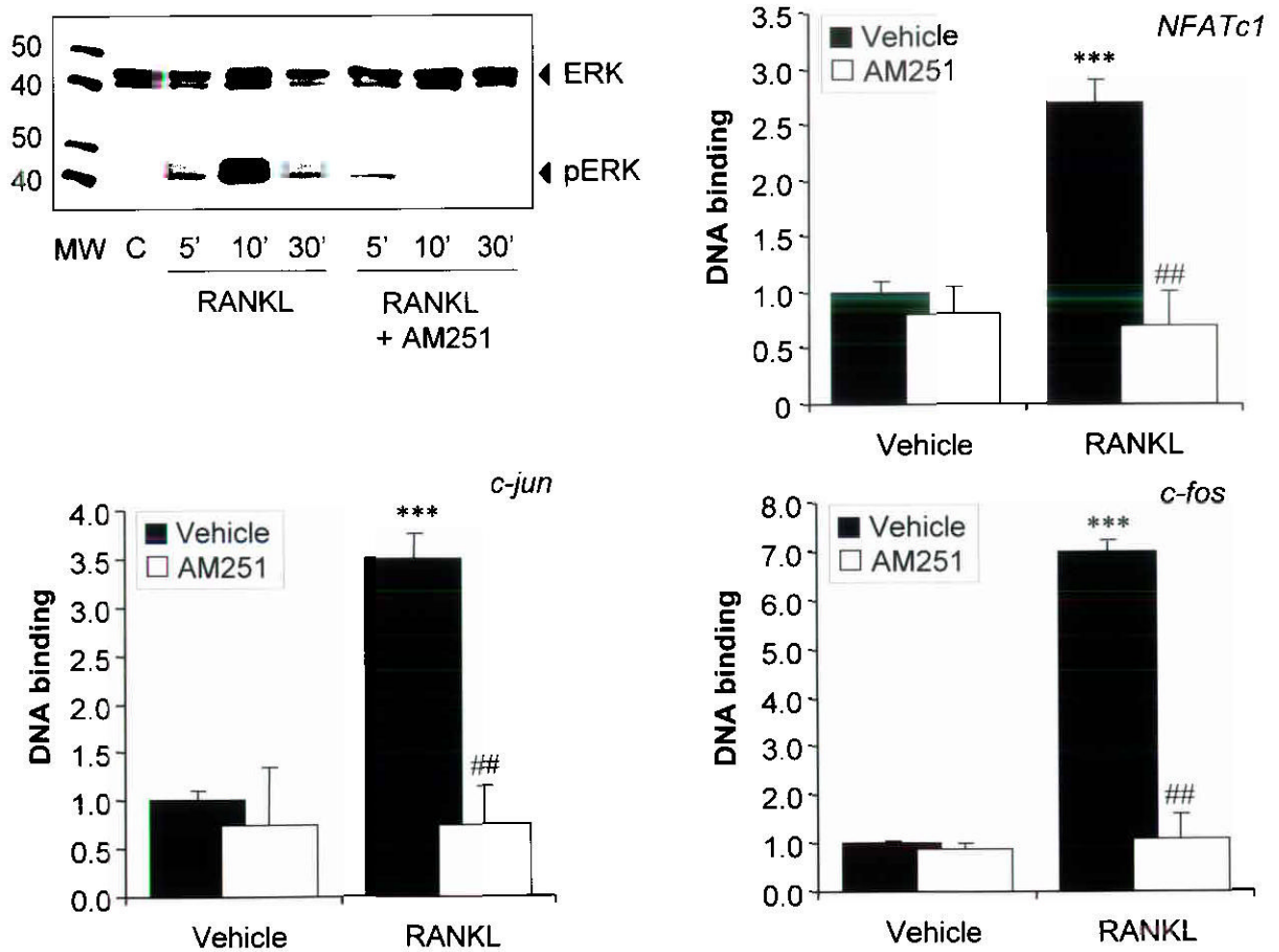


Figure 6. Cannabinoid receptor antagonists inhibit ERK, *c-fos*, *c-jun* and *NFATc1* activation in mouse osteoclasts

a, Effects of the CB antagonist AM251 (5 μ M) on ERK phosphorylation as detected by western blot in vehicle (C) and RANKL (100ng/ml) treated mouse osteoclasts. The numbers refer to the time (min) after RANKL stimulation. MW is molecular weight marker and the size of the marker bands is indicated in kDa. **b**, Effects of AM251 on nuclear translocation and DNA binding of phospho *NFATc1* in mouse osteoclasts treated with RANKL (100ng/ml) in the presence of vehicle or AM251 (2 μ M). **c**, Effects of AM251 on nuclear translocation and DNA binding of *c-jun* in mouse osteoclasts treated with RANKL (100ng/ml) in the presence of vehicle or AM251 (2 μ M). **d**, Effects of AM251 on DNA binding of *c-fos* in mouse osteoclasts treated with RANKL (100ng/ml) in the presence of vehicle or AM251 (2 μ M). Values are expressed as the change relative to vehicle treated cultures. Values in the bar charts are means and standard deviations. Significant differences between vehicle and RANKL treated cultures are indicated by: *** $p < 0.001$; and differences between AM251 and vehicle treated cultures are indicated by: ## $p < 0.001$. The results shown are representative of three independent experiments.