

Anastomotic recurrence of colorectal cancer – a biological phenomenon or an avoidable calamity?

Local recurrence after radical surgical excision of a large bowel neoplasm is nothing short of a catastrophe, because it heralds the onset of a progressive, painful, debilitating condition which untreated, has an average survival time of 11 months.¹ As many patients do not have disseminated disease, death is often slow and attended by considerable morbidity.² Most local recurrences occur after resection of a carcinoma in the rectum or sigmoid colon.³ Perineal pain, tenesmus, diarrhoea, incontinence and eventually obstruction supervene.

Once an anastomotic recurrence is diagnosed, treatment is palliative in most patients and options are limited. Surgery offers the only realistic chance of cure, but is rarely feasible. Radiotherapy relieves symptoms in half the patients without altering mean survival.¹ Palliation of obstruction by constructing a stoma, or of urinary obstruction by long term catheterisation⁴ causes further distress in an already limited life of poor quality.

The cause of this distressing complication of either restorative resection, or rectal amputation is the subject of considerable debate. The wide variation in the reported incidence of anastomotic recurrence⁵⁻¹⁵ may reflect differing surgical techniques, and in particular inadequate local excision of the tumour or its adjacent lymphatics, and inappropriate choice of low restorative resection. Supporters of the theory of exfoliated cell implantation would argue that omission to wash out the bowel ends intraoperatively with a cytotoxic agent is associated with a high incidence of local luminal recurrence.^{16,17}

Some of these dogmatic surgical views have been seriously questioned in the last decade and there is increasing evidence that there may be a more widespread biological abnormality in a vicinity of the tumour, predisposing some patients to a high risk of local recurrence.

Inadequate excision

Adequacy of local tumour clearance is fundamental to the prevention of locally recurrent disease, but emphasis has previously been placed on the importance of adequate distal clearance, particularly of rectal cancer. There is now substantial evidence that a 5cm distal clearance margin is unnecessary and that submucosal infiltration, lymphatic metastases and islands of tumour tissue are rarely present more than 2cm away from the macroscopic edge of the growth.^{18,19} Furthermore, although some authorities have implied higher local recurrence rates after restorative resection, particularly with the widespread use of stapling devices and a decreasing frequency of abdominoperineal excision, there is not a shred of evidence that sphincter saving surgery is accompanied by a higher rate of local recurrence.²⁰⁻²⁵

Adequate excision of the mesorectum is probably more important. The

presence of micrometastases in the mesorectum was highlighted by Heald and others in 1982²⁶ and a recent review of his own 2.6% incidence of local recurrence supports the view that wide excision of the mesorectum is probably a most important factor in achieving these commendable results.²⁷

In our view inadequate lateral clearance has received insufficient recognition. Durdey and colleagues found that 38% of patients had microscopic involvement of the lateral margins of the resected specimen. After a median follow up of 23 months, 75% of those with microscopic evidence of lateral spread had developed a local pelvic recurrence.²⁸ We have found that cytological smears from the four quadrants of the pelvis and imprint cytology of the tumour bed are a reliable indicator of inadequate tumour clearance. Of 60 patients studied, 12 had positive malignant cytology, and after a mean follow up of 16.5 months (range 1–38 months), seven of those developed a local recurrence (personal communication). We feel that cytology can identify patients with a high probability of local recurrence, in whom early postoperative radiotherapy may be advisable.

Implantation of exfoliated malignant cells

A further contentious cause of local recurrence is the possible implantation of exfoliated malignant cells into the anastomosis.^{30–32} Large bowel cancer may recur as nodules in the abdominal or perineal wound, as well as at the colonic anastomosis, suggesting that the mechanism of implantation of viable tumour cells might be an important cause of recurrence.^{33,34} Some but not all centres,^{35,36} have shown exfoliated tumour cells to be capable of excluding vital dyes such as trypan blue³⁷ and fluorescein.³⁸ Exclusion of vital dyes and fluorescein however, does not mean that the cells are capable of active division to produce new tumours *in vivo*. Malignant cells harvested from the lumen of the bowel are capable of growth in cell culture, but the cultures are self limiting and regress at seven to 10 days.³⁹

The only convincing test for replication is the ability of exfoliated malignant cells to produce tumours in immuno deprived mice. Only one study has claimed that injected suspensions of malignant cells into the tail vein of immuno deprived mice produced metastases, and even then the tumours did not display the histological pattern of the parent colorectal neoplasm.⁴⁰

In vitro studies have shown that certain solutions, particularly chlorhexidine-cetrimide and povidone-iodine rapidly kill exfoliated malignant cells, again using exclusion of vital dyes as the criterion of viability (personal communication).⁴¹ Hypochlorite solution, or mercuric perchloride have been traditionally advised for operative washout before fashioning intestinal anastomoses after tumour resection,⁴³ but this has never been tested scientifically. Until the results of a randomised clinical trial comparing preoperative cytotoxic washout with a placebo washout are available, clinicians will never know whether this time honoured practice is justified.

Altered biological properties at large bowel anastomoses

An alternative and largely unexplored mechanism for local recurrence is the possibility of some biological change at the anastomotic site, which increases

the susceptibility to cancer. Animal studies have consistently shown an increased yield of bowel tumours at the site of transection, stoma formation, anastomosis after resection, or merely the placement of a non-absorbable suture,⁴⁴ irrespective of whether the insult occurs before or after administration of one of the common chemical carcinogens, such as DMA, DMH, azoxymethane and MNNG*.⁴⁵⁻⁵³ A paper in this issue of *Gut* provides further evidence of proliferative instability in close proximity to a colonic anastomosis.⁵⁴

Using the azoxymethane rat model, Roe and colleagues from Bristol have reaffirmed an increased yield of tumours at the site of a surgical insult. They have shown that tumours occur irrespective of the timing of the carcinogen in relation to the operation and within a maximum time studied: three months. It is interesting that the greatest number of tumours were found when the carcinogen was given immediately postoperatively, whether in the sham, or in the transection group. Perhaps their hypothesis of immediate postoperative hyperaemia carrying high concentration of the carcinogen to the anastomosis is responsible in the transected animals, but it is not applicable to the controls. By measuring crypt cell height and mitotic index they have shown disordered cell kinetics in the 10 crypts adjacent to the anastomosis, even when histological evidence of re-epithelialisation of the anastomosis was complete. The crypt cell height had in fact returned to normal at 12 weeks and the labelling index was tending to fall. It would have been interesting to prolong the experiment and see when this variable also returned to normal. The fact that the tumour yield in the animals having the carcinogen 12 weeks after surgery was still highest at the site of the anastomosis must indicate, however, a continuing susceptibility to malignant change. The techniques used in the study do not account for cell cycle time, nor provide dynamic assessment of the birth rate of crypt cells. Technically more difficult stathmokinetic methods are better estimators of cellular proliferation. We believe these methods should be applied to examine the correlation between hyperplasia and neoplasia.

Matthews, Cooke and coworkers⁵⁵ have done a preliminary study of this kind with a few animals and highlighted some of the difficulties of the method. Their early data support the view that there is a close association between reparative hyperplasia and neoplasia, perhaps because of the selective action of chemical carcinogens on the stem cells.^{16 17}

Other reports suggest that there are altered biological properties in the mucosa adjacent to a large bowel neoplasm. Sulphomucin staining of colonic mucosal goblet cells is associated with increased cell kinetics and is common adjacent to a large bowel tumour.^{56 57} Sulphomucin staining at the site of an anastomosis is associated with an increased risk of local recurrence after apparently curative resection.^{58 59} Flow cytometric analysis has also shown that aneuploid tumours not only carry a poor prognosis, but are associated with increased risk of local recurrence.⁶⁰⁻⁶²

Finally, we must ask the question: are the tumours at anastomoses in the animal model the same as those in man? We think not. The true mucosal intraluminal anastomotic recurrence is not common. Large bowel cancer is often associated with polyps adjacent to a tumour and also present elsewhere in the colon.⁴³ In some cases local recurrence may be because of malignant change in a residual adenoma. Some apparently 'local'

* DMA-2,3-dimethyl-4-aminobiphenyl; DMH-1,2-dimethylhydrazine; MNNG-N-methyl-N-nitro-N-nitrosoguanidine.

recurrences are missed synchronous tumours,⁹ whilst late recurrences may arise from a metachronous tumour developing close to the anastomosis. In most patients local recurrence is largely extrarectal and the lesion seen and biopsied through the endoscope is merely the tip of the iceberg. In our view, the term 'anastomotic recurrence' is misleading. These recurrences rarely appear to be mucosal, but are mostly pelvic, suggesting that mechanisms other than local intramural reparative processes are important in their pathogenesis.

M R B KEIGHLEY AND CHRISTINE HALL

*Clinical Teaching Block,
General Hospital,
Steelhouse Lane,
Birmingham B4 6NH.*

References

- 1 Welch JP, Donaldson GA. Detection and treatment of recurrent cancer of the colon and rectum. *Am J Surg* 1978; **135**: 505–11.
- 2 Manson PN, Corman ML, Collier SA, Veidenheimer MC. Anastomotic recurrence after anterior resection for carcinoma. *Dis Colon Rectum* 1976; **19**: 219–24.
- 3 Hojo K. Anastomotic recurrence after sphincter saving resection for rectal cancer. *Dis Colon Rectum* 1986; **29**: 11–14.
- 4 Reid JDS, Robins RE, Atkinson KG. Pelvic recurrence after anterior resection and EEA stapling anastomosis for potentially curable carcinoma of the rectum. *Am J Surg* 1984; **147**: 69–32.
- 5 Wilking N, Herrera L, Petrelli NJ, Mittelman A. Pelvic and perineal recurrences after abdominoperineal resection for adenocarcinoma of the rectum. *Am J Surg* 1985; **150**: 561–3.
- 6 Pihl E, Hughes ESR, McDermott FT, Price Ann B. Recurrence of carcinoma of the colon and rectum at the anastomotic suture line. *Surg Gynecol Obstet* 1981; **153**: 495–6.
- 7 Kennedy HL, Langevin JM, Goldberg SM, *et al.* Recurrence following stapled coloproctostomy for carcinomas of the mid portion of the rectum. *Surg Gynecol Obstet* 1985; **160**: 5136.
- 8 Welch JP, Donaldson GA. The clinical correlation of an autopsy study of recurrent colorectal cancer. *Ann Surg* 1979; **189**: 496–502.
- 9 Enker WE, Dragsacevic S. Multiple carcinomas of the large bowel: a natural experiment in aetiology and pathogenesis. *Ann Surg* 1978; **187**: 8–11.
- 10 Hickey RC, Romsdahl MM, Johnson DE, *et al.* Recurrent cancer and metastases. *World J Surg* 1982; **6**: 585–95.
- 11 Anderson JM. Chemoradiotherapeutic prevention of local recurrence after stapled anastomoses in rectal cancer. *Scott Med J* 1981; **26**: 21–23.
- 12 Rosen CB, Beart RW. Jr., Istrup DM. Local rectal carcinoma after hand sewn and stapled anastomoses. *Dis Colon Rectum* 1985; **28**: 305–9.
- 13 Adloff M, Arnaud JP, Schloegel M, *et al.* Factors influencing local recurrence after abdominoperineal resection for cancer of the rectum. *Dis Colon Rectum* 1985; **28**: 413–5.
- 14 Hurst PA, Prout WG, Kelly JM, *et al.* Local recurrence after low anterior resection using the staple gun. *Br J Surg* 1982; **69**: 275–6.
- 15 Anderberg B, Enblad P, Sjö Dahl R, Wettfors J. Recurrent rectal carcinoma after anterior resection and rectal stapling. *Br J Surg* 1983; **70**: 1–4.
- 16 Lockhart-Mummary HE, Ritchie Jean K, Hawley PR. The results of surgical treatment for carcinoma of the rectum at St. Mark's Hospital from 1948 to 1972. *Br J Surg* 1976; **63**: 673–7.
- 17 Jones PF, Thomson HJ. Long term results of a consistent policy of sphincter preservation in the treatment of carcinoma of the rectum. *Br J Surg* 1982; **69**: 564–8.
- 18 Williams NS, Dixon MF, Johnston D. Reappraisal of the 5 centimetre rule of distal excision for carcinoma of the rectum: a study of distal intramural spread and of patients' survival. *Br J Surg* 1983; **70**: 150–4.
- 19 Williams NS, Durdey P, Johnston D. The outcome following sphincter-saving resection and abdominoperineal resection for low rectal cancer. *Br J Surg* 1985; **72**: 595–8.

- 20 Leff EI, Shaver JO, Hoexter B, *et al.* Anastomotic recurrences after low anterior resection: stapled vs hand-sewn. *Dis Colon Rectum* 1985; **28**: 164–7.
- 21 Gillen P, Peel ALG. Comparison of the mortality, morbidity and incidence of local recurrence in patients with rectal cancer treated by either stapled anterior resection or abdominoperineal resection. *Br J Surg* 1986; **73**: 339–41.
- 22 Pheils MT, Chapuis PH, Newland RC, Colquhoun K. Local recurrence following curative resection for carcinoma of the rectum. *Dis Colon Rectum* 1983; **26**: 98–102.
- 23 Williams NS. The rationale for preservation of the anal sphincter in patients with low rectal cancer. *Br J Surg* 1984; **71**: 575–81.
- 24 Phillips RKS, Hittinger Rosemary, Blesovsky Lynda, *et al.* Local recurrence following 'curative' surgery for large bowel cancer: I The overall picture. *Br J Surg* 1984; **71**: 12–6.
- 25 McDermott FT, Hughes ESR, Pihl E, *et al.* Local recurrence after potentially curative resection for rectal cancer in a series of 1008 patients. *Br J Surg* 1985; **72**: 34–7.
- 26 Heald RJ, Husband EM, Ryall RDH. The mesorectum in rectal cancer surgery – the clue to pelvic recurrence? *Br J Surg* 1982; **69**: 613–6.
- 27 Heald RJ, Ryall RDH. Recurrence and survival after total mesorectal excision for rectal cancer. *Lancet* 1986; *i*: 1479–82.
- 28 Durdey P, Quirke P, Dixon MF, Williams NS. Lateral spread of rectal cancer, the key to local recurrence [Abstract]. *Br J Surg* 1986; **73**: 1042.
- 30 Oakland DJ. The diagnosis of carcinoma of the colon by exfoliative cytology. *Proc R Soc Med* 1964; **57**: 279–82.
- 31 Keynes WM. Implantation from the bowel lumen in cancer of the large intestine. *Ann Surg* 1961; **153**: 357–64.
- 32 McGrew Elizabeth A, Laws JF, Cole WH. Free malignant cells in relation to recurrence of carcinoma of the colon. *JAMA* 1954; **154**: 1251–4.
- 33 Killingback M, Wilson E, Hughes ESR. Anal metastases from carcinoma of the rectum and colon. *Aust NZ J Surg* 1965; **34**: 178–87.
- 34 Norgren J, Svensson JO. Anal implantation metastasis from carcinoma of the sigmoid colon and rectum – a risk when performing anterior resection with the EEA stapler? *Br J Surg* 1985; **72**: 602.
- 35 Rosenburg IL, Russell CW, Giles GR. Cell viability studies on the exfoliated colonic cancer cell. *Br J Surg* 1978; **65**: 188–90.
- 36 Rosenburg IL. The aetiology of colonic suture line recurrence. *Ann R Coll Surg Engl* 1979; **61**: 251–7.
- 37 Tennant Judith R. Evaluation of the trypan blue technique for determination of cell viability. *Transplantation* 1964; **2**: 685–94.
- 38 Thomas D, Mosedale Betty, Zola H. The use of the indirect fluorescent antibody technique in assessing the activity of antilymphocytic sera and antilymphocytic globulins. *Clin Exp Immunol* 1971; **8**: 987–91.
- 39 Skipper D, Cooper AJ, Marston JE, Taylor I. Exfoliated malignant cells and local recurrence of colorectal carcinoma. *SRS Abs Nov 86* (in press).
- 40 Symes MO, Fermor B, Umpleby HC, *et al.* Cells exfoliated from colorectal cancers can proliferate in immune deprived mice. *Br J Cancer* 1984; **50**: 423–5.
- 41 Umpleby HC, Williamson RCN. The efficacy of agents employed to prevent anastomotic recurrence in colorectal carcinoma. *Ann R Coll Surg Engl* 1984; **66**: 192–4.
- 43 Umpleby HC, Bristol JB, Rainey JB, Williamson RCN. Survival of 727 patients with single carcinomas of the large bowel. *Dis Colon Rectum* 1984; **27**: 803–10.
- 44 Pozharisski KM. The significance of nonspecific injury for colon carcinogenesis in rats. *Cancer Res* 1975; **35**: 3824–30.
- 45 Williamson RCN, Davies PW, Bristol JB, Wells M. Intestinal adaptation and experimental carcinogenesis after partial colectomy. Increased tumour yields are confined to the anastomosis. *Gut* 1982; **23**: 316–25.
- 46 Harte PJ, Steele G Jr., Rayner AA, Munroe AE, King VP, Wilson RE. Effects of major small bowel resection on dimethylhydrazine-induced bowel carcinogenesis. *J Surg Oncol* 1981; **18**: 87–93.
- 47 Williamson RCN, Bauer FLR, Oscarson JEA, Ross JS, Malt RA. Promotion of azoxymethane-induced colonic neoplasia by resection of the proximal small bowel. *Cancer Res* 1978; **38**: 3212–7.
- 48 Williamson RCN, Bauer FLR, Ross JS, Watkins JB, Malt RA. Enhanced colonic carcinogenesis with azoxymethane in rats after pancreaticobiliary diversion to mid small bowel. *Gastroenterology* 1979; **76**: 1386–92.
- 49 Zedeck MS, Grab DJ, Sternberg SS. Differences in the acute response of the various

- segments of the rat intestine to treatment with the intestinal carcinogen methylazoxymethanol acetate. *Cancer Res* 1977; **37**: 32–6.
- 50 Kanazawa K, Yamamoto T, Sato S. Experimental induction of colonic carcinomas in rats. *Jpn J Exp Med* 1975; **45**: 439–56.
- 51 Gennaro AR, Villanueva R, Sukonthaman T, *et al.* Chemical carcinogenesis in transposed intestinal segments. *Cancer Res* 1973; **33**: 536–41.
- 52 Lamont JTK, O'Gorman TA. Experimental colon cancer. *Gastroenterology* 1978; **75**: 1157–9.
- 53 Rubio CA, Nylander G, Wallin B, *et al.* Carcinogenesis at colonic anastomotic sites. *Dis Colon Rectum* 1984; **27**: 468–70.
- 54 Roe R, Fermor B, Williamson RCN. Proliferative instability and experimental carcinogenesis at colonic anastomoses. *Gut* 1987; **28**: 808–15.
- 55 Matthews J, Carpenter R, Strachan J, Gregory P, Cooke T. Does cell proliferation play a part in anastomotic recurrence of colonic tumours [Abstract]. *Br Assoc Surg Oncol* Dec 1986.
- 56 Colacchio TA, Chabot JA, Zimmerman BW. Differential mucin staining in colorectal neoplasms. *Am J Surg* 1984; **147**: 666–9.
- 57 Filipe MI, Cooke KB. Changes in composition of mucin in the mucosa adjacent to carcinoma of the colon as compared with the normal: a biochemical investigation. *J Clin Pathol* 1974; **27**: 315–8.
- 58 Wood CB, Dawson PM, Habib NA. The sialomucin content of colonic resection margins. *Dis Colon Rectum* 1985; **28**: 260–1.
- 59 Habib NA, Salem R, Lucj RJ, *et al.* A histochemical method that predicts local recurrence after curative resection in carcinoma of the colon and rectum. *Surg Gynecol Obstet* 1984; **159**: 436–8.
- 60 Armitage NC, Robins RA, Evans DF, *et al.* The influence of tumour cell DNA abnormalities on survival in colorectal cancer. *Br J Surg* 1985; **72**: 828–30.
- 61 Forsslund G, Cedermark B, Ohman U, *et al.* The significance of DNA distribution pattern in rectal carcinoma. *Dis Colon Rectum* 1984; **27**: 579–84.
- 62 Melamed MR, Enker WE, Banner P, *et al.* Flow cytometry of colorectal carcinoma with three year follow up. *Dis Colon Rectum* 1986; **29**: 184–6.