Prostaglandin E_2 is a mediator of 5-hydroxytryptamine induced water and electrolyte secretion in the human jejunum

L K MUNCK, A MERTZ-NIELSEN, H WESTH, K BUKHAVE, E BEUBLER, AND J RASK-MADSEN

From Section of Gastroenterology, Department of Medicine G, Bispebjerg Hospital, University of Copenhagen and Department of Biochemistry and Nutrition, The Technical University of Denmark, Denmark and Department of Experimental and Clinical Pharmacology, University of Graz, Graz, Austria

SUMMARY Studies in the rat jejunum *in vivo* have shown that 5-hydroxytryptamine (5-HT) causes secretion of fluid and luminal release of prostaglandin (PG) E_2 . These effects can be blocked by indomethacin and ketanserin, which suggests that PGE₂ may be an important intermediate in the transduction mechanism leading to 5-HT induced fluid secretion. To test this hypothesis in man 'steady state' perfusions (9 ml/min) were done in eight healthy volunteers using the triple lumen technique. The proximal jejunum was perfused with Ringer's solution which contained ⁵¹Cr-EDTA as a non-absorbable marker. Before and after the administration of indomethacin (1·0 mg/kg *iv*) the effects of exogenous 5-HT (10 μ g/kg/min *iv*) on jejunal net transport of fluid and electrolytes and jejunal flow rate (JFR) of PGE₂ were measured in 15-min periods for 2×60 minutes after a 60 minute control period. 5-HT reversed fluid and electrolyte absorption into profuse secretion (p<0·01, Duncan's multiple range test) and significantly increased JFR of PGE₂ (p<0·01). Indomethacin partly restored fluid and electrolyte absorption (p<0·01) and inhibited JFR of PGE₂ (p<0·05). These results provide further evidence in favour of the theory that PGs are involved in 5-HT induced intestinal fluid secretion.

It is generally accepted that 5-hydroxytryptamine (5-HT) induces calcium dependent intestinal fluid and electrolyte secretion,¹⁻⁷ possibly through activation of phosphatidylinositol turnover,⁸ but without altering mucosal adenylate cyclase activity^{9 10} or cyclic nucleotide contents.¹¹ Furthermore, prostaglandins (PGs) have been proposed to be important intermediates in 5-HT induced fluid secretion⁶ and recent experiments in animals have shown that jejunal fluid secretion and luminal release of PGE₂, mediated by 5-HT, are inhibited by the cyclooxygenase inhibitor, indomethacin, in addition to the 5-HT₂ receptor antagonist, ketanserin.⁶

5-hydroxytryptamine has been implicated as a

Received for publication 25 April 1988.

cause of diarrhoea in patients with carcinoid syndrome¹²⁻¹⁴ and triple lumen perfusions in such patients have shown jejunal secretion of fluid, which could be reduced by the 5-HT receptor antagonist, methysergide.¹⁵ Also excess intestinal PG formation has been reported to be responsible for secretion in a variety of diarrhoeal diseases^{12 to} and enteral and parenteral administration of PGs elicits intestinal secretion.¹⁷⁻¹⁹

To clarify further the role of PGs in 5-HT induced intestinal secretion 'steady state' perfusions of the proximal jejunum were carried out in eight healthy volunteers. The relevance of the above observations in animals for the understanding of diarrhoeal disease in man were evaluated by determining the influence of exogenous 5-HT on the transfer of fluid and electrolytes and the flow rate of PGE₂, in addition to the effects of indomethacin on this response.

Address for correspondence: Dr Jørgen Rask-Madsen, Section of Gastroenterology, Department of Medicine G, Bispebjerg Hospital, DK-2400 Copenhagen NV, Denmark.

Methods

SUBJECTS

Ten healthy volunteers (three women, median age 22 years, range 20–29) free of medication consented to the study protocol, which was carried out in accordance with the Helsinki Declaration II and approved by the regional ethics committee and the Danish National Health Service.

In two cases the study was stopped prematurely: one participant experienced headache, dizziness, and blurred vision before the start of 5-HT infusion; he had a blood pressure of 160/110 mmHg; another had nausea and vomiting, which became intolerable after start of the 5-HT infusion.

EXPERIMENTAL DESIGN

'Steady state' perfusions were carried out as described by Cooper et al.²⁰ After an overnight fast the jejunum was intubated with a triple lumen polyvinyl tube (Meadox Surgimed Ltd, Denmark), the position of which was checked fluoroscopically before and after the perfusion. The infusion port was placed 5-10 cm aborally to the ligament of Treitz. The mixing segment was 10 cm and the test segment was 30 cm. Another tube was positioned in the antrum of the stomach, which was continously emptied. The jejunum was perfused at a rate of 9 ml/min (LKB 2115 Multiperspex pump, Bromma, Sweden) with an isotonic solution containing Na⁺ 145 mmol/l, K⁺ 5 mmol/l. Cl 140 mmol/l. HCO₃ 10 mmol/l. and as a non-absorbable marker ⁵¹Cr-EDTA 30 µCi/l. Proximal aspiration was by siphonage (range 0.5-2.0ml/min), while distal aspiration was by intermittent suction (range 0.8-4.0 ml/min; Pump AB, Einar Engell, Sweden).

After a 60 minute equilibration period samples were collected every 15 minutes and immediately stored at -20° C for analysis within three weeks. After a 60 minute control period 5-HT (5hydroxytryptamine creatininesulphate (Serva Chemicals, FRG) dissolved in saline) was infused into a cubital vein at a rate of approximately 100 ml/h equal to 5-HT 10 µg/kg/min. Sixty minutes after start of 5-HT infusion an intravenous bolus injection of indomethacin (Dumex, Copenhagen, Denmark) 1.0 mg/kg was administered over 10 minutes and the 5-HT infusion was continued for another 60 minutes. Because of the lag phase of drug response, and because no equilibration period was interposed between the test periods, the results of the first 15 minute collections in each test period were added to those obtained in the previous period for calculation of the average transport/flow rate in that period. The participants were monitored in a cardioscope and their blood pressure and heart rate were determined regularly and at the start of 5-HT infusion each minute for 10 minutes.

ANALYTICAL PROCEDURES

Net fluid and electrolyte transfer was determined as previously described.³⁰ Sodium and potassium were determined by flame photometry (Lange, Berlin, FRG) and chloride was measured in a chloride meter (Chloro-counter, Mark II, Labo International, Utrecht, Belgium). ⁵¹Cr-activity was determined by gammaspectrometry (Model 1185, Searle Nuclear Chicago Division, Chicago, USA). The values were expressed as ml/h or meq/h/cm length of jejunum. Positive values represent absorption and negative values secretion.

PGE₂ concentrations in aspirates were measured by a radioimmunological method validated by gas chromatography-mass spectrometry as previously described,^{21,22} because the mucosal 'overflow' of PGE₂ presently appears to provide the most reliable index of the balance between intestinal PGE₂ synthesis and degradation *in vivo*.²³ The jejunal flow rate (JFR) of PGE₂ – that is, the amount of PGE₂ passing a cross section of the jejunum/minute – was calculated as the concentration of PGE₂ in the aspirate from the proximal aspiration port times the flow rate of fluid passing this port. Transepithelial secretion rates of PGE₂ are not given because the differences between the amounts passing the



Fig. 1 Net fluid transfer in proximal human jejunum during control conditions (C), in response to 5-HT infusion (10 $\mu g/kg \times min iv$) and following the administration of indomethacin (I, 1 mg/kg iv). **=p<0.01 v control and \$\$=p<0.01 v 5-HT.

proximal and the distal aspiration ports were negligible and not significantly different from zero (p>0.05). This observation suggests that degradation of luminal PGE₂ occurs rapidly and/or that the ratio between PGE₂ synthesis and degradation varies along the jejunum. The values of PGE₂ determinations are given as ng/min.

STATISTICAL ANALYSES

All results are given as mean (SE). The data were analysed by Duncan's multiple range test and probability values less than 0.05 were considered significant.

Results

FLUID TRANSFER

Fluid was absorbed in all control periods. 5-HT reversed (p<0.01) net fluid absorption (+1.99 ml/h× cm (0.31)) into profuse secretion (-0.91 ml/h×cm (0.22)). Indomethacin significantly (p<0.01) inhibited this effect of 5-HT, but fluid absorption (+0.73 ml/h×cm (0.27)) was not completely restored (Fig. 1).

ELECTROLYTE TRANSFER

Electrolytes were absorbed during the control period (Na⁺: +29·1 mmol/h×cm (6·3); Cl⁻: +24·7 mmol/h×cm (5·5); K⁺: +1·7 mmol/h×cm (0·4)). 5-HT elicited net secretion of electrolytes (Na⁺: -17·8 mmol/h×cm (9·4); Cl⁻: -18·3 mmol/h×cm (6·9); K⁺: -0·8 mmol/h×cm (0·3); p<0·01) and this effect was partially reversed (Na⁺: +8·5 mmol/h×cm (7·9); Cl⁻: +7·0 mmol/h×cm (6·5); K⁺: +0·5 mmol/h×cm (0·3); p<0·01) by indomethacin (Fig. 2).

FLOW RATE OF PROSTAGLANDIN E2

The fasting jejunal flow rate of PGE₂ at the proximal aspiration port was low during control periods (0.130 ng/min (0.031)) and 5-HT markedly increased JFR of



Fig. 2 Net transfer of electrolytes (mean \pm SE) in proximal human jejunum during control conditions (C), in response to infusion of 5-HT (10 µg/kg×min iv) and following the administration of indomethacin (I, 1 mg/kg iv). **=p<0.01 v control, *=p<0.05 v control, and §§=p<0.01 v 5-HT.



Fig. 3 Jejunal flow rate of PGE_2 in proximal human jejunum during control conditions (C), in response to infusion of 5-HT (10 $\mu g/kg \times min iv$) and following the administration of indomethacin (I, 1 mg/kg iv). **=p<0.01 v control and §§=p<0.01 v 5-HT.

PGE₂ (0.494 ng/min (0.131); p<0.01). Indomethacin significantly inhibited the stimulatory effect of 5-HT on JFR of PGE₂ (0.183 ng/min (0.041); p<0.05) (Fig. 3).

ADVERSE EFFECTS

The infusion of 5-HT caused tenderness and redness at the infusion site and in a single case venous spasm, which necessitated a change of the infusion site. The alterations in systolic blood pressure observed during 5-HT infusion were transient and did not exceed 10 mmHg. No significant effect on heart rate was observed. All participants, but three, experienced nausea and slight abdominal discomfort. The administration of indomethacin transiently intensified nausea in all subjects and provoked vomiting in a few cases, but within a few minutes the abdominal symptoms were relieved.

Discussion

5-hydroxytryptamine has been shown to induce intestinal secretion in animals¹⁻⁷ by eliciting chloride secretion, in addition to decreasing the electroneutral absorption of sodium chloride.^{2,4,24} Furthermore, enterochromaffine cells¹⁴ and nerves in the myenteric plexus²⁵ contain 5-HT. Although specific 5-HT receptors are present in the submucosal ganglia,²⁶ enterocytes do not seem to possess specific receptors for 5-HT.^{27,28} These observations suggest that the action of the secretagogue is an indirect one and that 5-HT requires a paracellular mediator to elicit the final secretory response.

In the morphine withdrawal syndrome observed in the rat the intestinal secretion is mediated by 5-HT^{*29} through stimulation of local PG formation in the absence of increased mucosal cyclic AMP contents.⁹ The secretion of fluid and the release of PGE₂ are inhibited by the specific 5-HT_2 -receptor antagonist, ketanserin, and the cyclooxygenase-inhibitor, indomethacin, without affecting the release of 5-HT.⁹ Intestinal fluid secretion mediated by 5-HT appears to be calcium dependent^{23,5,7} and the secretory effects of physiologically low doses of exogenous PGE₂ are prevented by the calcium antagonist, verapamil.⁶ Therefore, PGE₂ may be an important intermediate in the transduction mechanism leading to fluid secretion.¹⁶

Although the application of intestinal steady state perfusion in man may only add little to the elucidation of the mechanisms responsible for 5-HT induced secretion, the relevance of the above observations for the understanding of diarrhoeal disease should be judged from data obtained in man. We chose the triple lumen technique to avoid the influence of mechanical stimulation, if any, and/or the possible activation of local PG formation through nervous reflex mechanisms provoked by occluding balloons.

The results of the present study unequivocally show that exogenous 5-HT induces profuse jejunal secretion of fluid and electrolytes in man, in addition to increasing JFR of PGE₂. These effects were only partially inhibited by indomethacin. The infusion rate of 5-HT used in this study was chosen to obtain a local mucosal concentration similar to that reported in previous *in vivo* experiments on the rat jejunum.⁶ The local jejunal plasma concentration of 5-HT was calculated roughly to be approximately 85 ng/ml, assuming pulmonary and hepatic clearances of 40%³⁰ and 100%, respectively, and a cardiac output of 5 l/min.

The apparent discrepancy between the effects of conventional doses of indomethacin and those previously obtained in the rat⁶ is explained by the use of a 10-fold higher dose in the latter experiments. We would tentatively conclude, therefore, that the secretory action of 5-HT observed in the present study provides further evidence for the hypothesis that PGE₂ is involved in 5-HT induced intestinal secretion and may prove relevant for the understanding of the pathogenesis of certain diarrhocal disorders, such as cholera³¹ and the carcinoid syndrome.¹³

This work was presented, in part, at the Jubilee Meeting of the British Society of Gastroenterology in London on September 15–18, 1987 (*Gut* 1987; **28**:

A1396), and was supported by the Danish Medical Research Council (No 12-4508), the World Health Organization (D-82138), the Commission of the European Communities (TSD-397 DK), and the Austrian Scientific Research Funds (No P-6087M). The authors thank Anne Hallander, Inge-Lise Løffval, and Lene Persson for technical and Lisbeth Aas for secretarial assistance.

References

- Powell DW, Field M. Pharmacological approaches to treatment of secretory diarrhea. In: Field M, Fordtran JS, Schultz SG, eds. *Secretory diarrhea*. Bethesda: American Physiological Society, 1980: 187–207.
- 2 Donowitz M, Asarkof N, Pike G. Calcium dependence of serotonin-induced changes in rabbit ileal electrolyte transport. J Clin Invest 1980; 66: 341–52.
- 3 Donowitz M. Ca²⁺ in the control of active intestinal Na and Cl transport: involvement in neurohumoral action. *Am J Physiol* 1983; 245: G165–77.
- 4 Hardcastle J, Hardcastle PT, Redfern JS. Action of 5hydroxytryptamine on intestinal ion transport in the rat. *J Physiol* 1981; **320**: 41–55.
- 5 Hardcastle J, Hardcastle PT, Noble JM. The involvement of calcium in the intestinal response to secretagogues in the rat. *J Physiol* 1984; **355**: 465–78.
- 6 Beubler E, Bukhave K, Rask-Madsen J. Significance of calcium for the prostaglandin E₂-mediated secretory response to 5-hydroxytryptamine in the small intestine of the rat in vivo. *Gastroenterology* 1986; **90**: 1972–7.
- 7 Zinner MJ, McFadden D, Sherlock D, Jaffe BM. Verapamil reversal of scrotonin-induced jejunal secretion of water and electrolytes in awake dogs. *Gastroenterology* 1986; **90:** 515–9.
- 8 Berridge MJ, Irvine RF. Inositol triphosphate, a novel second messenger in cellular signal transduction. *Nature* 1984; **312**: 315–21.
- 9 Beubler E, Bukhave K, Rask-Madsen J. Colonic secretion mediated by prostaglandin E₂ and 5hydroxytryptamine may contribute to diarrhoea due to morphine withdrawal in the rat. *Gastroenterology* 1984; 87: 1042-8.
- 10 Brunsson I, Sjöqvist A, Jodal M, Lundgren O. Mechanisms underlying the small intestinal fluid secretion caused by arachidonid acid, prostaglandin E₁, and prostaglandin E₂ in the rat *in vivo*. Acta Physiol Scand 1987; **130**: 633–42.
- 11 Brastitus TA, Field M, Kimberg DV. Intestinal mucosal cyclic GMP: regulation and relation to ion transport. *Am J Physiol* 1976; 231: 275–82.
- 12 Jaffe BM. Prostaglandins and serotonin: Nonpeptide diarrhcagenic hormones. World J Surg 1979; 3: 565–78.
- 13 Antonsen S, Hansen MG, Bukhave K, Rask-Madsen J. Influence of a new 5-HT₂ receptor antagonist (ketanserin) on jejunal PGE₂ release and ion secretion due to malignant carcinoid syndrome [Abstract]. *Gut* 1982; **223**: A887.
- 14 Ahlman H. Scrotonin and carcinoid tumors. J Cardiovasc Pharmacol 1985; 7: S79–85.

- 15 Donowitz M, Binder HJ. Jejunal fluid and electrolyte secretion in carcinoid syndrome. *Dig Dig* 1975; 20: 1115-22.
- 16 Rask-Madsen J. Eicosanoids and their role in the pathogenesis of diarrhoeal diseases. *Clin Gastroenterol* 1986; 15: 545-66.
- 17 Karim SMM, Filshie GM. Therapeutic abortion using PGF_{2α}. Lancet 1970; i: 157–9.
- 18 Matuchansky C, Bernier J-J. Effect of prostaglandin E₁ on glucose, water, and electrolyte absorption in the human jejunum. *Gastroenterology* 1973; 64: 1111–8.
- 19 Milton-Thompson GJ, Cummings JH, Newman A, Billings JA, Misiewicz JJ. Colonic and small intestinal response to intravenous prostaglandin $F_{2\alpha}$ and E_2 in man. Gut 1975; 16: 42–6.
- 20 Cooper H, Levitan R, Fordtran JS, Ingelfinger FJ. A method for studying absorption of water and solute from the human small intestine. *Gastroenterology* 1966; 50: 1–7.
- 21 Bukhave K, Rask-Madsen J. Prostaglandin E_2 in jejunal fluids and its potential diagnostic value for selecting patients with indomethacin-sensitive diarrhoea. *Eur J Clin Invest* 1981; **11**: 191–7.
- 22 Bukhave K, Gréen K, Rask-Madsen J. Comparison of radioimmunological determinations with gas chromatography mass spectrometry dosage. A study of PGE₂ and PGF₂ in gastrointestinal fluids. *Biomed Mass Spectrom* 1983; **10:** 265–8.
- 23 Rask-Madsen J, Bukhave K. The difficulties of estab-

lishing the pathophysiological role of prostaglandins in secretion. In: Skadhauge E, Heintze K, eds. *Intestinal absorption and secretion*. Lancaster: MTP Press, 1983: 453-68.

- 24 Donowitz M. Tai YH, Asarkof N. Effect of scrotonin on active electrolyte transport in rabbit ileum, gallbladder and colon. Am J Physiol 1980; 239: G463-72.
- 25 Furness JB, Costa M. Neurons with 5hydroxytryptamine-like immunoreactivity in the enteric nervous system. Their projections in the guinea-pig small intestine. *Neuroscience* 1982; 7: 341–9.
- 26 Gershon MD, Takaki M, Tamir H, Brancheck T. The enteric neural receptor for 5-hydroxytryptamine. *Experimentia* 1985; **41**: 863–8.
- 27 Gaginella TS, Rimele TJ, Wietecha M. Studies on rat intestinal epithelial cell receptors for serotonin and opiates. J Physiol 1983; 335: 101–11.
- 28 Cooke HF. Neurobiology of the intestinal mucosa. *Gastroenterology* 1986; **90:** 1057–81.
- 29 Gintzler AR. Serotonin participation in gut withdrawal from opiates. *J Pharmacol Exp Ther* 1979; **211:** 7–12.
- 30 Zinner MJ, Kasher F, Jaffe BM. The hemodynamic effect of intravenous infusions of serotonin in conscious dogs. J Surg Res 1983; **34:** 171–8.
- 31 Beubler E, Kollar G, Saria A, Bukhave K, Rask-Madsen J. The role of 5-hydroxytryptamine and prostaglandin E₂ in cholera toxin-induced fluid secretion in the small intestine of the rat in vivo. *Gastroenterology* 1987; 92: 1315.