Immunoglobulin G Antibodies against the Endosymbionts of Filarial Nematodes (*Wolbachia*) in Patients with Pulmonary Dirofilariasis

F. Simón,¹ G. Prieto,¹ R. Morchón,¹ C. Bazzocchi,² C. Bandi,² and C. Genchi^{2*}

Laboratorio di Parassitologia, Facultad de Farmacia, Universidad de Salamanca, 37007 Salamanca, Spain,¹ and Dipartimento di Patologia Animale, Igiene e Sanità Pubblica Veterinaria, Sezione di Patologia Generale e Parassitologia, 20133 Milan, Italy²

Received 28 May 2002/Returned for modification 7 August 2002/Accepted 19 September 2002

The dog parasite *Dirofilaria immitis* can infect humans. Patients with pulmonary dirofilariasis were tested for immunoglobulin G (IgG) antibodies against the surface protein of *Wolbachia*, the bacterial endosymbiont of *D. immitis*. These patients showed significantly higher IgG titers than healthy individuals from areas in which *D. immitis* was endemic as well as areas in which it was not endemic. Titration of anti-*Wolbachia* surface protein IgG could become useful for diagnostic applications.

Dirofilaria immitis is the causative agent of heartworm disease in dogs and cats. The disease is found worldwide in subtropical and temperate areas (8). In dogs and cats, the parasite develops to the adult stage in the pulmonary arteries and in the right cardiac chambers (6, 10). The infection is transmitted by several species of mosquitoes that are frequently able to bite both humans and animals. People living in areas of endemicity are thus at risk of infection. Humans are, however, "dead-end hosts," since larvae do not normally develop into the adult stage in humans. Human pulmonary dirofilariasis develops when the nematode dies, embolizes and travels to the lung, and lodges in a small branch of the pulmonary artery. In these cases, chest radiography shows well-circumscribed, noncalcified or calcified nodules (5, 9, 12).

In areas in which heartworm infection in dogs is endemic, clinically healthy people are frequently found positive for antibodies against D. immitis antigens. For example, Prieto et al. (14) recently recorded seroprevalence values ranging from 26 to 37% in three areas of southern Europe. The high percentage of seroprevalence in healthy people in areas of endemicity hampers the serological diagnosis of pulmonary dirofilariasis. Two methods have been proposed for the experimental diagnosis of this disease; they are based on the use of recombinant or native D. immitis proteins (13, 18). However, neither method allows clear-cut distinction between healthy humans from areas of endemicity and patients with pulmonary nodules. Serological studies have also shown different antibody profiles in humans: immunoglobulin G (IgG), IgM, and predominantly IgE antibodies against D. immitis antigens were detectable in healthy individuals, while in patients with pulmonary lesions, the IgE response was not observed (7, 17). Evaluation of immunoglobulin profiles could aid in diagnosis. However, the IgG, IgM, and IgE responses in healthy individuals vary throughout the year (7, 11).

Filarial nematodes, including D. immitis, harbor obligate,

intracellular, gram-negative bacteria belonging to the genus *Wolbachia* (*Rickettsiales*). *Wolbachia* is a stable and abundant component of the body of filarial nematodes (1, 2). It was recently shown that *Wolbachia* surface protein (WSP) induces a specific IgG response in cats infected with *D. immitis* (3) and in monkeys infected with lymphatic filariae (15). In addition, *Wolbachia* appears to play a role in the immunopathogenesis of filarial diseases (4, 16).

So far, all studies which have shown a specific antibody response against *Wolbachia* proteins have been performed with natural hosts of filarial nematodes, with hosts in which the parasite can develop to the adult stage, or after inoculation of hundreds of infective larvae (e.g., see reference 3). Whether an antibody response against *Wolbachia* develops in dead-end hosts under natural conditions (such as for *D. immitis* in humans) is not known. The aim of this study was to investigate the IgG response against a *Wolbachia* protein in humans living in areas in which dog heartworm disease is endemic.

Forty-two serum samples from humans were assigned to the following groups. Group 1 (G1) contains 10 serum samples from patients with pulmonary nodules due to D. immitis infection (these samples were kindly supplied by Patrick Lammie, Centers for Disease Control and Prevention, Atlanta Ga.; diagnosis was made by bioptisy sampling). Group 2 (G2) contains 18 serum samples from clinically healthy humans living in areas in which heartworm infection is endemic (Po River Valley, northern Italy: 10 samples; Colombian Amazonia, South America: 8 samples) and previously found by an enzyme-linked immunosorbent assay (ELISA) to be IgG positive for D. im*mitis* by use of both somatic and excretory or secretory antigens from adult nematodes (14, 19). Group 3 (G3) contains 14 serum samples from healthy humans living in a mountainous area of the province of Salamanca, Spain, where D. immitis infection in dogs and mosquitoes has not been recorded; these donors were found by the ELISA to be seronegative for D. immitis infection.

The WSP of *D. immitis*, produced in recombinant form and purified as described by Bazzocchi et al. (3), was used as an antigen in an ELISA for the detection of IgG in the 42 serum samples. The ELISA was performed as described by Perera et al. (13), with minor modifications. Briefly, microplate wells

^{*} Corresponding author. Mailing address: Dipartimento di Patologia Animale, Igiene e Sanità Pubblica Veterinaria, Sezione di Patologia Generale e Parassitologia, Via Celoria 10, 20133 Milan, Italy. Phone: 39 02 50318101. Fax: 39 02 50318095. E-mail: claudio.genchi @unimi.it.

TABLE 1. ELISA results for recombinant WSP in human sera

Group		OD	
	Avg	Range	SD
G1	0.87	0.58-1.35	0.26
G2	0.35	0.26-0.58	0.11
G3	0.03	0.20-0.39	0.07

were coated with 0.8 μ g of recombinant WSP; serum samples were analyzed at a 1:30 dilution, and anti-human peroxidaseconjugated IgG was diluted 1:4,000. The optical density (OD) was measured at 492 nm. The cutoff (0.5) was the OD arithmetical average plus three standard deviations for the 14 serum samples from the clinically healthy blood donors living in the *D. immitis*-free area.

The ELISA results are reported in Table 1. All of the serum samples from patients with pulmonary nodules due to *D. immitis* (G1) showed high ODs that were consistently above the cutoff. Serum samples from healthy donors found serologically positive for *D. immitis* (G2) and from donors living in areas of nonendemicity (G3) had significantly lower ODs (one-way analysis of variance; *F*, 49.04; *P*, <0.0001; degrees of freedom, 39).

Our results show that the IgG response against the WSP of D. immitis is consistently detectable only in patients with pulmonary nodules due to the parasite. In healthy blood donors from areas in which D. immitis is endemic and who have IgG against somatic and excretory or secretory antigens of adult parasites, the IgG levels against WSP are lower. Only in 3 cases out of 14 were the IgG titers in this group above the cutoff of our ELISA. This result suggests that the surface protein of Wolbachia endosymbionts stimulates the host immune system only after the death of preadult worms in the small branches of pulmonary arteries, or at least when the development of D. *immitis* has progressed to a stage at which nematode death can lead to the release of a sufficient amount of bacteria. In any case, our results provide further evidence for the immunological role of Wolbachia in filarial infection, with special reference to humans, and also show that IgG titers are related in some way to the clinical status of the patient.

Our results may suggest an interesting method for the serodiagnosis of pulmonary dirofilariasis. Differential diagnosis (e.g., to exclude pulmonary cancer) requires surgical biopsy, and even a reliable serological assay would not necessarily exclude the need for histological analysis. However, a reliable serologial test for pulmonary dirofilariasis would aid in the final diagnosis, allowing better treatment of patients and better planning of invasive diagnostic procedures, such as thoracoscopy. Based on our results, titration of anti-WSP IgG appears to be a very promising tool for the diagnosis of pulmonary dirofilariasis, possibly in combination with titration of IgE against *D. immitis* antigens. Of course, examination of additional serum samples from healthy individuals, from patients with pulmonary nodules, and from patients with different parasitoses is needed before an anti-WSP assay can be proposed as a test for the diagnosis of human dirofilariasis.

We thank Patrick Lammie for providing serum samples from humans with pulmonary nodules, Laura Kramer for critical reading of the manuscript, and Stefano Novati for suggestions.

This work was supported by Consejería de Educación y Cultura Junta de Castilla y Léon (grant SA01/00F given to F. Simón) and by MIUR-COFIN 2001 (support given to C. Genchi).

REFERENCES

- Bandi, C., T. J. C. Anderson, C. Genchi, and M. L. Blaxter. 1998. Phylogeny of *Wolbachia* in filarial nematodes. Proc. R. Soc. Lond. B 265:2407–2413.
- Bandi, C., A. J. Trees, and N. W. Brattig. 2001. Wolbachia in filarial nematodes: evolutionary aspects and implications for the pathogenesis and treatment of filarial diseases. Vet. Parasitol. 98:215–238.
- Bazzocchi, C., F. Ceciliani, J. W. McCall, I. Ricci, C. Genchi, and C. Bandi. 2000. Antigenic role of the endosymbionts of filarial nematodes: IgG response against the *Wolbachia* surface protein in cats infected with *Dirofilaria immitis*. Proc. R. Soc. Lond. B 267:2511–2516.
- Brattig, N. W., U. Rathjens, M. Ernst, F. Geisinger, A. Renz, and F. W. Tischendorf. 2000. Lipopolysaccharide-like molecules derived from *Wolbachia* endobacteria of the filaria *Onchocerca volvulus* are candidate mediators in the sequence of inflammatory and anti-inflammatory responses of human monocytes. Microbes Infect. 2:1147–1157.
- Cordero, M., M. R. Muñoz, A. Muro, F. Simón, and L. Perera. 1992. Small calcified nodule: an undescribed radiologic manifestation of human pulmonary dirofilariasis. J. Infect. Dis. 165:398–399.
- Dillon, R. 1984. Feline dirofilariasis. Vet. Clin. N. Am. Small Anim. Pract. 14:1185–1199.
- Espinoza, E., M. Cordero, A. Muro, F. Lorente, and F. Simon. 1993. Anti-Dirofilaria immitis IgE: seroepidemiology and seasonal variation in an exposed human population. Trop. Med. Parasitol. 44:172–176.
- Genchi, C., L. H. Kramer, and G. Prieto. 2001. Epidemiology of canine and feline dirofilariasis: a global view, p. 121–133. *In* F. Simon and C. Genchi (ed.), Heartworm infection in humans and animals. Ediciones Universidad Salamanca, Salamanca, Spain.
- Harrison, E. G., and J. H. Thompson. 1971. Dirofilariasis, p. 903–916. In R. A. Marcial-Rojas (ed.), Pathology of protozoal and helminthic diseases. The Williams & Wilkins Co., Baltimore, Md.
- Kume, S., and S. Itagaki. 1955. On the life-cycle of *Dirofilaria immitis* in the dog final host. Br. Vet. J. 111:16–24.
- Muro, A., M. Cordero, A. Ramos, and F. Simón. 1991. Seasonal changes in the levels of anti-*Dirofilaria immitis* antibodies in an exposed human population. Trop. Med. Parasitol. 42:371–374.
- Orihel, T. C., and M. L. Eberhard. 1998. Zoonotic filariasis. Clin. Microbiol. Rev. 11:366–381.
- Perera, L., A. Muro, M. Cordero, E. Villar, and F. Simón. 1994. Evaluation of a 22 kDa *Dirofilaria immitis* antigen for the immunodiagnosis of human pulmonary dirofilariosis. Trop. Med. Parasitol. 45:249–252.
- Prieto, G., G. Cancrini, A. Muro, C. Genchi, and F. Simon. 2000. Seroepidemiology of *Dirofilaria immitis* and *Dirofilaria repens* in humans from three areas of southern Europe. Res. Rev. Parasitol. 60:95–98.
- Punkosdy, G. A., V. A. Dennis, B. L. Lasater, G. Tzertzinis, J. M. Foster, and P. J. Lammie. 2001. Detection of serum IgG antibodies specific for *Wolbachia* surface protein in rhesus monkeys infected with *Brugia malayi*. J. Infect. Dis. 184:385–389.
- Saint André, A., N. M. Blackwell, L. R. Hall, A. Hoerauf, N. W. Brattig, L. Volkmann, M. J. Taylor, L. Ford, A. G. Hise, J. H. Lass, E. Diaconu, and E. Pearlman. 2002. The role of endosymbiotic *Wolbachia* bacteria in the pathogenesis of river blindness. Science 295:1892–1895.
- Simon, F., A. Muro, M. Cordero, and J. Martin. 1991. A seroepidemiologic survey of human dirofilariosis in western Spain. Trop. Med. Parasitol. 42: 106–108.
- Sun, S., and K. Sugane. 1992. Immunodiagnosis of human dirofilariosis by enzime linked immunosorbent assay using recombinant DNA-derived fusion protein. J. Helminthol. 66:220–226.
- Viera, C., I. D. Vélez, M. N. Montoya, S. Agudelo, C. Genchi, and F. Simón. 1998. *Dirofilaria immitis* in Tikuna Indians and their dogs in the Colombian Amazon. Ann. Trop. Med. Parasitol. 92:123–125.