

Compilation of tRNA sequences and sequences of tRNA genes

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Received September 5, 1995; Accepted September 19, 1995

INTRODUCTION

The 1995 compilation contains 2700 sequences of tRNAs and tRNA genes. The last edition which appeared two years ago (1) was supplemented by 694 new sequences. The sequences of tRNA mutants and of tRNAs originating from transformed or differentiated cells were not considered.

The tRNAs included in the compilation are listed in Table 1. Each tRNA or tRNA gene is specified by the (abbreviated) name of the organism from which it was isolated and a four digit code: the first three digits identify the organism, the last digit specifies the particular isoacceptor. The amino acid specificity of the tRNA is indicated by a one-letter amino acid code. The tRNAs coding for selenocysteine were annotated with the letter Z. Initiator tRNAs are annotated with the letter X.

The sequences, references and footnotes of tRNAs and tRNA genes listed in Table 1 are deposited in the European Bioinformatics Institute (EBI) Data Library. The references are restricted to the first complete publication of the sequence unless additional information (e.g. base modification, corrections, etc.) was later obtained. In such cases additional references were added.

In order to facilitate a computer analysis an alignment is used which is most compatible with the tRNA phylogeny and known three-dimensional structures of tRNA. The corresponding numbering system is shown in Figure 1.

As was the case in the previous edition (1), this publication does not contain a sequence printout. Instead, the sequences have been deposited in the EBI Data Library. This publication should be therefore quoted as a reference for data obtained from the electronically accessible database. Information on how to access the sequence files can be obtained by electronic mail: send email to Netserv@ebi.ac.uk containing the command 'HELP TRNA.' The help file will contain all the information needed to obtain the requested sequence. The tRNA database is also available via anonymous FTP from ftp.ebi.ac.uk in the directory pub/databases/tRNA. It is also distributed on the EBI CD-ROM. Contact the EBI Data Library, Hinxton Hall, Hinxton, Cambridge CB10 1RQ, UK (Fax: +44 1223 494468, email: DataLib@EBI.AC.UK).

Researchers who wish to obtain the sequence information on a floppy disk, or as a hardcopy should contact M. Sprinzl, Laboratorium für Biochemie, Universität Bayreuth, D-95440 Bayreuth, Germany, Fax: +49 921 552432, email: Ma-

thias.Sprinzl@uni-bayreuth.de. Software allowing search for tRNA sequences according to several criteria, e.g. source, partial sequence, modified nucleoside, anticodon, amino acid specificity and printout of sequences in a form of tables or cloverleaves, is also available.

Presentation of sequences

The sequences in the database are divided into three parts. The first two parts contain the sequences of the tRNA genes and tRNAs, respectively, which can be fitted into the canonical tRNA alignment. The third part contains tRNA and tRNA gene sequences, mainly of animal mitochondria, whose secondary structures differ from most tRNAs and could not be aligned according to Figure 1.

An example for sequence presentation in the database is given in Table 2. Each sequence in the compilation occupies two consecutive lines. The first line begins with the letter 'D' or 'R' and contains the six-position identification code of the sequence ('D' or 'R' for DNA or RNA, respectively; a one-letter code for the amino acid, X for methionine-initiator, Z for selenocysteine; and the four-digit code specifying the organism and isoacceptor. After this, the sequence of the anticodon (in the case of tRNA sequences in its modified form) is given, followed by the name and the kingdom of organism (Table 1), and the sequence (99 standard positions). The second line begins with the sign '+' and contains the information about base-pairing (double helical regions only, tertiary interactions are not annotated). All other lines in the compilation begin with signs other than 'D,' 'R' or '+' (usually '*') and contain comments.

Nucleotides involved in Watson–Crick pairs are marked with '=', the G·U pairs are indicated with the sign '*'. Nucleotides 26 and 44 are considered to form a base pair included in the anticodon stem (Fig. 1).

The sequences in original publications denoted as 'yeast' are assigned to *Saccharomyces cerevisiae*. The user should be aware, however, that some of these organisms have possibly been misclassified and that the original literature should be consulted.

This compilation uses a one-letter code for all nucleotides including modified ones. For standard nucleotides, adenosine, cytidine, guanosine, thymidine and uridine the usual abbreviations, A, C, G, T and U, respectively, are used. To designate

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HUMAN	588	ACDEEFGHKLINPQRSSSTVWXY
ADYPCEROS MBLAMPUS	580	FV
DIOSPHALPHUS TRADOC.	581	FV
CEPHALOPHUS MAJOR.	582	FV
DAMALIGUS GORCAS	583	FV
GAZELLA THOMSON	584	FV
EDULS GLIPSPRYM.	585	FV
MADOCUA NRO	586	FV
OTFX GAZELLA	587	FV
TRACELAPHUS IMBER.	588	FV
EUKARYOTIC CYTOPLASM 600-699		
SINGLE CELL ORGANISMS AND FUNGI		
FLASMODIUM FALSE.	603	ALMNRRTV
TRYPANOSOMA BRUCI	605	IKENACQRRRTVVY
TETRAHYMENA PYRF.	606	NCG
LEISHMANIA TARRPI.	609	GKLGRTVW
DICTYOSTELIUM DIS.	616	ARRHKXLMNQRSSSTTVVWXY
PHYCARIUM POLYCEPH.	618	X
NEUROSPORA CRASSA	620	PLN
CANDESA ALECANIS	621	LSG
RHYTOPTHORA PAR.	622	C
PODOSPORA ANSERINA	624	SS
SACCHAROMYCES CBR.	628	AAACDEEFGGHHKILLMNQOPRRSSSTTVVWXXYZ
SCHIZOSACCHA. POM.	632	ADGFRHKRRSSSVXX
CANDIDA CYLINDRA.	637	S
PLANTS 670-749		
CHLAMYCIA TRACHOM.	673	TW
ARA BIDOPSIS THAL.	674	AFSSSSSSVWXXYYZ
GLYCI ME MAX	680	DWV
PHASEOLUS VULGARIS	698	LFP
NICOTIANA RUSTICA	700	SSSSSSYY
PETUNIA SP.	710	N
HILANTHUS ANNUUS	712	L
SORGHUM BICOLOR	714	G
ORYZA SATIVA	719	G
TRITICUM AESTIVUM	720	Y
TRITICUM VULGARE	724	F
SOYBEAN	730	C
ANIMALS 750-999		
CARINORHABI. ELGG.	758	DKLRRRWKZ
BOMBYX MORI	758	ASGE
DROSOPHILA MELANO.	774	ADSEFQDHHKILLMNPRSSSTVvxyZ
DROSOPHILA SIMUL.	780	S
SOUD	785	K
XENOPUS LAEVIS	792	AFELMXXYYZ
PODOCORYNE CARNEA	793	DFSS
CHICKEN	804	AACDFPPWZ
MOUSE	810	ACDCCGHHKLLPWXZ
RAT	816	DCFFFRHHKLLP
BOVINE	820	SZ
HUMAN	880	EEGDKKLLMNPPQOPRRSSSTTVVWXXYZ

PHASEOLUS VULGARIS	440	RLLMPTWXY
ANIMALS 460-699		
ASCARIS SUUM	464	RMS
ARDES ALBOPCTUS	480	DEGQORSVX
HAMSTER	524	GRS
RAT LIVER	528	DRKLLLVVW
BOVINE LIVER	538	DKKLLRSSTVWXX
HUMAN	588	S
EUKARYOTIC CYTOPLASM 600-699		
SINGLE CELL ORGANISMS AND FUNGI		
EUGLENA GRACILIS	604	DF
TETRAHYMENA THOMA.	608	DDDX
SCENODERMUS OBLIQ.	612	FXV
NEUROSPORA CRASSA	620	FK
SACCHAROMYCES CER.	628	ACDFDFGHHKILLMNPRSSSTTVVWXXYZ
SCHIZOSACCHA. POM.	632	DFV
TORULOPSIS UTILIS	636	ALPVXY
CANDIDA CYLINDRA.	637	LLSSSSS
PLANTS 670-749		
HORDEUM VULGARE	676	DF
WHEAT GERM	682	PQKRWXY
BRASSICA NAPUS	685	F
LUPINUS LUTRUS	694	FFGHMPPVXY
PHASEOLUS VULGARIS	698	LILX
PSUM SATIVUM	702	F
SPRACIA OLERACEA	704	S
NICOTIANA RUSTICA	705	SSSSYY
SOLANUM TUBEROSUM	707	LW
CUCUMIS SATIVUS	708	L
ANIMALS 750-999		
CARINORHABI. ELGG.	758	L
ASTERNA ANUREMIS	760	X
BOMBYX MORI	760	AAFTGG
DROSOPHILA MELANO.	774	SHKSSSTVWXY
EUPHALSIA GREDA	780	X
XENOPUS LAEVIS	792	DFX
SALMON LIVER	798	X
CHICKEN	804	W
MOUSE	810	EFFRKKMOPVXZ
RAT	816	DEKLLNQQSSVXX
RABBIT LIVER	822	DFXMX
BOVINE LIVER	828	DRLNRRRTVWYZ
CALF LIVER	834	F
COW MAMMARY GLAND	840	L
SHEEP LIVER	846	HL
HUMAN	880	AAEFGHLMNPPQOPRRSSSTTVVWXXYZ

PART THREE: tRNA and rRNA gene sequences that differ from the conventional alignment

PART TWO: tRNA Sequences

Source	Code	tRNA
VRUSES 090-029		
AVIAN ONCO. VIRUS	010	M
CHICKEN ASYMMYRUS	014	W
MOUSE M-MULV	018	RP
PHAGE T3	022	GLPQST
PHAGE T5	026	DLNRQ
ARCHAEBACTERIA 030-109		
HALOBACTERIUM CUT.	038	AGHNRSTVVX
HALOPLEX VILCAMI	050	AAACDEEFGGHHKILLMNPPQOPRRSSSTTVVWXXYZ
HALOCOCCLUS MGRPHUA	054	X
METHANOBAC. THORM.	082	GN
SULFOLOBUS ACID.	082	F
THERMOPLASMA ACID.	090	NX
BACTERIA 110-239		
MYCOPLASMA CAPRIC.	114	ACDEFGHIKILLMNPRSSSTTVVWXXYZ
MYCOPLASMA MYCOID.	118	AGPSTVX
SPROPLASMA CITRI	120	WV
STREPTOMYCES GRIS.	130	X
STREPTOMYCES COEL.	131	Q
STAPHYLOCC. EPID.	138	DD
MYCOBAC. SMAR.	142	X
BACILLUS STEARO.	146	RLVY
BACILLUS SUBTILIS	154	APGKXLMPPSSSTTVVWXXYZ
THERMUS THERMOPHI.	168	DFWXX
E.COLI	186	AAACDEEFGGHHKILLMNQOPRRSSSTTVVWXXYZ
SALMONELLA Typh.	170	GHLPPP
RHODOPIRRA. RUB.	202	PL
AGROMYCELLUM OLIVAR.	206	F
ANACYSTIS NIGULANS	210	LLX
SYNECHOCYSTIS SP.	214	E
ORGANELLES		
CHLOROPLASTS 240-359		
CHLAMYDOMONAS REIN.	244	E
EUGLENA GRACILIS	262	F
COELUM FRAGE	253	GEMR
SCENODERMUS OBLIQ.	259	MXV
LUPINUS ALBUS	263	V
HORDEUM VULGARE	264	DDXD
TRITICUM AESTIVUM	268	E
ZEA MAYS	272	I
GLYCI ME MAX	280	LL
NICOTIANA TABACUM	292	W
PHASEOLUS VULGARIS	316	RLLWVX
SPINACIA OLERACEA	320	RLLMPTWXY
MITOCHONDRIA 360-699		
SINGLE CELL ORGANISMS AND FUNGI 360-419		
TETRAHYMENA PYRF.	380	FV
TETRAHYMENA THOMA.	384	W
NEUROSPORA CRASSA	382	ALLTVWXY
SACCHAROMYCES CBR.	400	PQKLMPPSSSTVWXY
PLANTS 420-459		
SOLANUM TUBEROSUM	431	BL
ONOTHERA SP.	436	F

Source	Code	tRNA/rRNA gene
MITOCHONDRIA 360-699		
SINGLE CELL ORGANISMS 360-419		
AND FUNGI		
TRICHOPHYTON MENT. 403 C		
ANIMALS 750-999		
AFIS MELLIFERA	482	T
DAPHNIA PULEX	483	C
BALACHOPTERA PHYS.	534	MS5
BALACHOPTERA MUSC.	536	S
HALICHTERUS URYPIUS	537	E
PHOCA VITULINA	538	S
SAMBARO	542	S
CHIMPANZEE	542	S
PYGMY CHIMPANZEE	570	S
GORELLA	580	S
ORANG UTAN	584	S

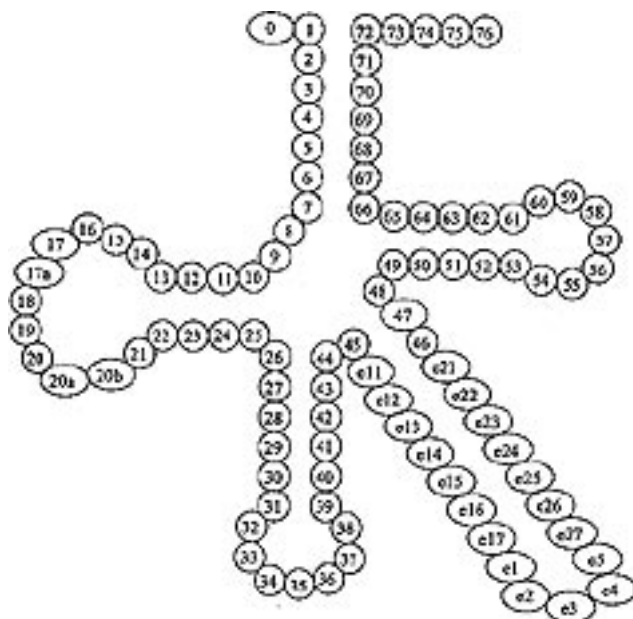


Figure 1. Numbering of nucleotides in tRNAs. Circles represent nucleotides which are always present; the ovals, nucleotides which are not present in each structure: these are nucleotides before the position 1 on the 5'-end, before and after the two invariant GMP residues 18 and 19 in the D-loop, and the nucleotides in the variable loop. The nucleotide to be added at a given site is indicated by the number of the preceding nucleotide followed by a colon and a letter in alphabetical order. The nucleotides in the variable stem have the prefix 'e' and are located between positions 45 and 46 obeying the base-pairing rules. The nucleotides in the 5'-strand and the 3'-strand are numbered by e11, e12, e13, ... and e21, e22, e23, ... respectively; and the second digit identifies the base-pair. In the case of a long variable region, the loop can be formed by up to five nucleotides: e1, e2, e3, e4 and e5.

modified nucleotides, the other ASCII signs are employed as defined in Table 3. Terminology and structure of the modified nucleosides occurring in tRNAs were used according to (2). Positions in particular sequence which are not filled (gaps in the generalised structure, Fig. 1) are indicated by a dash. All nucleotide insertions are denoted by underlining at the place of insertion.

Numbering and alignment of the variable region

The alignment of the variable region has been done in accordance with Steinberg and Kisselev (3). The extra arm is placed between nucleotides 45 and 46. The extra arm includes two double helical strands forming a stem and a loop. The annotations of the nucleotides in the extra arm positions begin with the letter 'e' (extra) followed by a one- or two-digit number. We have reserved a space for 7 base pairs in the stem and 5 nucleotides in the loop. The nucleotides in the loop are numbered from 1 to 5, whereas the

nucleotides in the stem are numbered from 11 to 17 (5'-branch) and from 27 to 21, in reverse order, (3'-branch), to indicate base pair formation between nucleotides 11–21, 12–22, etc. (Fig. 1). In the tRNAs where the extra arm position 45 is empty but where the nucleotides 46–48 between the extra arm and T-domain are present, the positions will be filled in the order 48, 46, 47, i.e. tRNAs use position 48, 46 and 47 for the first, second and third nucleotide, respectively, depending on the length of the sequence in this region. A similar situation occurs in tRNAs without a long extra arm, where the most variable position 47 is deleted in many sequences.

Alignment of animal mitochondrial tRNAs

In properly aligned tRNA sequences, nucleotides occupying the same position in different tRNA sequences should play a comparable structural or functional role. Most animal mitochondrial tRNAs cannot be easily aligned with other tRNAs mainly because of the absence of information about their three-dimensional structure. Experimental data, however, point to the existence of tertiary interactions in these tRNAs. In this compilation, we use an alignment which accounts for these interactions as much as possible. Where we could do so, the animal mitochondrial tRNAs were included in Parts I and II. The alignment of animal mitochondrial tRNA is, however, not yet unambiguous.

Some animal mitochondrial tRNAs have completely unusual secondary structure and cannot be fitted in the tRNA alignment used here (Parts I and II). We treated these sequences separately including them in Part III. Here, each particular sequence has its own alignment. To this group belong the tRNAs from: (i) mitochondria of a parasitic worm lacking the T-, or D-domain; (ii) mitochondria of molluscs, insects and echinoderm, with extended anticodon and T-stems; (iii) mammalian mitochondria, lacking the D-domain.

For some tRNA genes the secondary structure pattern cannot be clearly established. We have also included these sequences in Part III. It is possible that post-transcriptional modifications of these tRNAs will result in improvement of the secondary structure.

ACKNOWLEDGEMENTS

This project was supported by Fonds der Chemischen Industrie, Deutsche Forschungsgemeinschaft, Project Sp 243/5-1, and Molekulargenetischer Arbeitskreis Rhein/Main e.V.

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Table 2. Format of tRNA sequences in EBI databank

*PART ONE: SEQUENCES OF tRNA-GENES

*Number	Anticodon	Organism	Kingdom	accept stem 0123456789	D-domain 1111111111122222222222222333333344444	anticodon domain 0123456789001234567890123456789012345	variable region 1234567	T-loop 654321	T-domain 44455555555555566666666667777777	accept stem 77777777
DA0260	TGC	PHAGE T5	VIRUS	-GGGCGAATAGTGTGAGC	-GGG--AGCACACCGACTTGC	AAATCTGGTA		G-GGAGGTTGAGTGCCTCTT	TGCCACCA	
DA0340	TGC	ARCHAEOLOBUS FULG.	ARCHAE	-GGGCTCCTAGCTCAGC	-GGG--AGAGCGCCGCTTTGC	AGGCGGAG		GCCCGGGTTCAAATCCCG	CGAGTCCA	
DA0380	TGC	HALOBACTERIUM CUT.	ARCHAE	-GGGCCATAGCTCAGT	-GGT--AGAGTGCTCTCTTGC	AAGGAGGAT		GCCCTGGTTCGAAATCCCA	GTTGGTCCA	
DA0420	TGC	HALOBACTERIUM HAL.	ARCHAE	-GGGCCATAGCTCAGT	-GGT--AGAGTGCTCTCTTGC	AAGGAGGAT		GCCCTGGTTCGAAATCCCA	GTTGGTCCA	
DA0580	TGC	METHANOBAC. FORMI.	ARCHAE	-GGGCCGCTAGCTCAGACTGGG	-AGAGCGCCGCTTGC	AAGCGGAG		GCCCCGGTTCAAATCCCG	TGGTCCA	
DA0620	TGC	METHANOBAC. THERM.	ARCHAE	-GGGCCGCTAGCTCAGACTGGG	-AGAGCGCCGCTTGC	AAGCGGAG		GCCCCGGTTCAAATCCCG	TGGTCCA	
DA0660	TGC	METHANOCOCC. VANI.	ARCHAE	-GGGCCGCTAGCTCAGT	-GGG--AGAGCGCTCCCTTGC	AAGGCGAG		GCCCTGGTTCAAATCCCG	CGGTTCCA	
DA0670	TGC	METHANOTRIX SOEH.	ARCHAE	-GGGCTGTAGCTCAGCT	-GGT--AGAGCGCCCTTGC	AAGGCGAG		GCCCTGGTTCGAAATCCCA	GAAGTCCA	
DA0680	TGC	METHANOTHERM. FER.	ARCHAE	-GGGCCATAGCTCAGCTGGG	-AGAGCGCCGCTTGC	AAGCGGAG		GCCCCGGTTCAAATCCCG	TGGTCCA	
DA0780	TGC	METHANOSPIR. HUNG.	ARCHAE	-GGGCTCCTAGCTCAGCT	-GGA--AGAGCGCCGCTTGC	AAGCGGAG		GCCCTGGTTCGAAATCCCA	GAAGTCCA	
DA0940	TGC	THERMOCOCCUS CELER	ARCHAE	-GGGCCGCTAGCTCAGCTGGG	-AGAGCGCCGCTTGC	AAGCGGAG		GCCCCGGTTCGAAATCCCG	CGGTTCCA	
DA0980	TGC	THERMOPROT. TENAX	ARCHAE	-GGGCCGCTAGCTCAGC	-GGA--AGAGCGCCGCTTGC	CGCGGAG		ATCCCGGTTTCAATCCCG	CGGTTCCA	
DA0981	CGC	THERMOPROT. TENAX	ARCHAE	-GGGCCGCTAGCTCAGC	-GGA--AGAGCGCCGCTTGC	CGCGGAG		ATCCCGGTTTCAATCCCG	CGGTTCCA	
DA1140	TGC	MYCOPLASMA CAPRIC.	EUBACT	-GGGCCCTAGCTCAGCT	-GGG--AGAGCACCTGC	CCGAGGGG		GTCCAGGTTTTCGATCCCT	TAGGGTCCA	
DA1180	TGC	MYCOPLASMA MYCOID.	EUBACT	-GGGCCCTAGCTCAGCT	-GGG--AGAGCACCTGC	CCGAGGGG		GTCCAGGTTTTCGATCCCT	TAGGGTCCA	
DA1200	TGC	MYCOPLASMA PNEU.	EUBACT	-GGGATGTAGCTCAACT	-GAT--AGAGCACCTGATTTGC	CACTCAGGAG		GTTGAGGTTTTCGATCCCT	TAGGGTCCA	
DA1230	TGC	ACHOLEPLASMA LAID.	EUBACT	-GGGGCTTAGCTCAGCT	-GGG--AGAGCGCCGCTTGC	CCGAGGAG		GTCCAGGTTTTCGATCCCT	TAGGGTCCA	
DA1231	TGC	ACHOLEPLASMA LAID.	EUBACT	-GGGGCTTAGCTCAGCT	-GGG--AGAGCGCCGCTTGC	CCGAGGAG		GTCCAGGTTTTCGATCCCT	TAGGGTCCA	
DA1260	TGC	SPIROPLASMA MELIF.	EUBACT	-GGGCCGCTAGCTCAGCT	-GGG--AGAGCACCTGCTTGC	CCGAGGGG		GTCCAGGTTTTCGATCCCT	TAGGGTCCA	
DA1280	TGC	BORRELIUM BURGDORF.	EUBACT	-GGGGCTTAGCTCAGT	-GGCT--AGAGCATCGGCTTTC	AAGCGGAG		GTCCAGGTTTTCGATCCCT	TAACTCCA	
DA1470	TGC	ENTEROCOCCUS HIRAE	EUBACT	-GGGGCTTAGCTCAGCT	-GGG--AGAGCGCCGCTTTC	CCGAGGAG		GTCCAGGTTTTCGATCCCT	TAGGGTCCA	
DA1480	TGC	STAPHYLOCOCC. AURE.	EUBACT	-GGGGCTTAGCTCAGCT	-GGG--AGAGCGCCGCTTTC	CCGAGGAG		GTCCAGGTTTTCGATCCCT	TAGGGTCCA	

(Continued in the EBI databank. See text for instructions.)

Table 3. Modified nucleosides in tRNA and their abbreviations

one-letter code of nucleotides	symbol [2]	name [2]			
V	V		:	G	unknown modified guanosine
			K	m1G	1-methylguanosine
			L	m2G	N ² -methylguanosine
			#	Gm	2'-O-methylguanosine
			R	m22G	N ² ,N ² -dimethylguanosine
				m22Gm	N ² ,N ² ,2'-O-trimethylguanosine
			7	m7G	7-methylguanosine
			[fs7d7G	archaeosine
			Q	Q	queuosine
			8	manQ	mannosyl-queuosine
			9	galQ	galactosyl-queuosine
			Y	yW	wybutosine
			W	o2yW	peroxywybutosine
			N	?U	unknown modified uridine
			[mnm5U	5-methylaminomethyluridine
			2	s2U	2-thiouridine
			J	Um	2'-O-methyluridine
			4	s4U	4-thiouridine
			&	ncm5U	5-carbamoylmethyluridine
			1	mcm5U	5-methoxycarbonylmethyluridine
			S	mnm5s2U	5-methylaminomethyl-2-thiouridine
			3	mcm5s2U	5-methoxycarbonylmethyl-2-thiouridine
			V	omo5U	uridine 5-oxyacetic acid
			5	mo5U	5-methoxyuridine
			!	cmnm5U	5-carboxymethylaminomethyluridine
			\$	cmnm5s2U	5-carboxymethylaminomethyl-2-thiouridine
			X	acp3U	3-(3-amino-3-carboxypropyl)uridine
			.	mcm5U	5-(carboxylhydroxymethyl)uridine methyl ester
				cmnm5Um	5-carboxymethylaminomethyl-2'-O-methyluridine
			-	ncm5Um	5-carbamoylmethyl-2'-O-methyluridine
			D	D	dihydrouridine
			P	Y	pseudouridine
]	m1Y	1-methylpseudouridine
			Z	Ym	2'-O-methylpseudouridine
			T	m5U	ribosylthymine
			F	m5s2U	5-methyl-2-thiouridine
			\	m5Um	5, 2'-O-dimethyluridine
H	?A	unknown modified adenosine			
-	m1A	1-methyladenosine			
/	m2A	2-methyladenosine			
+	i6A	N ⁶ -isopentenyladenosine			
*	ms2i6A	2-methylthio-N ⁶ -isopentenyladenosine			
=	m6A	N ⁶ -methyladenosine			
6	t6A	N ⁶ -threonylcarbamoyladenosine			
E	m6t6A	N ⁶ -methyl-N ⁶ -threonylcarbamoyladenosine			
	ms2t6A	2-methylthio-N ⁶ -threonylcarbamoyladenosine			
:	Am	2'-O-methyladenosine			
!	I	inosine			
O	m1I	1-methylinosine			
-	Ar(p)	2'-O-ribosyladenosine (phosphat)			
.	io6A	N ⁶ -[cis-hydroxyisopentenyl]adenosine			
<	7C	unknown modified cytosine			
%	s2C	2-thiocytidine			
B	Cm	2'-O-methylcytosine			
M	ac4C	N ⁴ -acetylcytosine			
?	m5C	5-methylcytosine			
.	m3C	3-methylcytosine			
}	k2C	lysidine			
>	f5C	5-formylcytidin			