

Identification of Quantitative Trait Loci Across Recombinant Inbred Lines and Testcross Populations for Traits of Agronomic Importance in Rice

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ABSTRACT

This study was conducted to determine whether quantitative trait loci (QTL) controlling traits of agronomic importance detected in recombinant inbred lines (RILs) are also expressed in testcross (TC) hybrids of rice. A genetic map was constructed using an RIL population derived from a cross between B5 and Minghui 63, a parent of the most widely grown hybrid rice cultivar in China. Four TC hybrid populations were produced by crossing the RILs with three maintaining lines for the widely used cytoplasmic male-sterile (CMS) lines and the genic male-sterile line Peiai64s. The mean values of the RILs for the seven traits investigated were significantly correlated to those of the F₁ hybrids in the four TC populations. Twenty-seven main-effect QTL were identified in the RILs. Of these, the QTL that had the strongest effect on each of the seven traits in the RILs was detected in two or more of the TC populations, and six other QTL were detected in one TC population. Epistatic analysis revealed that the effect of epistatic QTL was relatively weak and cross combination specific. Searching publicly available QTL data in rice revealed the positional convergence of the QTL with the strongest effect in a wide range of populations and under different environments. Since the main-effect QTL is expressed across different testers, and in different genetic backgrounds and environments, it is a valuable target for gene manipulation and for further application in rice breeding. When a restorer line that expresses main-effect QTL is bred, it could be used in a number of cross combinations.

HETEROISIS has been very successfully exploited in diverse plants and animals. In agriculture, hybrid varieties contribute strongly worldwide to the production of many crop species, including the most important food crops, such as maize and rice (STUBER 1994; YUAN 1998; KHUSH 2001). Hybrid rice has a yield advantage of ~15–20% over the best commercial rice varieties. The area planted to hybrid rice in China accounts for >50% of the total rice area of the country at present. The cultivation of hybrid rice has started on a large scale in many Asian countries.

Several hypotheses have been proposed to explain the genetic basis of heterosis. The dominance hypothesis (BRUCE 1910) proposes that dominant factors from either parent mask deleterious recessive mutations from the other parent in the heterozygous F₁ population. In contrast, the overdominance hypothesis (SHULL 1908) holds that heterozygosity at single loci confers properties that are superior to either homozygote. The two hypotheses have been verified with molecular biology experiments (STUBER *et al.* 1992; XIAO *et al.* 1995). A third hypothesis suggests that heterosis may arise from epistasis between alleles at different loci (YU *et al.* 1997;

GOODNIGHT 1999). More recently, further results have suggested that epistasis is the primary genetic basis of heterosis. It is suggested that separate efforts should be taken for breeding high-yielding inbred and hybrid cultivars in rice (LI *et al.* 2001; LUO *et al.* 2001).

In hybrid rice breeding programs in China, the breeders have made intense efforts to improve the traits of inbred lines and have obtained a number of elite lines, for example, Minghui 63, a restorer line of the most popular hybrid rice variety Shanyou 63, and 9311, a restorer line of the first super hybrid rice. The characteristics of the parental lines have a profound effect on those of the F₁ offspring. Once an elite restorer line or male-sterile line has been developed, it is used to breed a series of hybrid varieties with strong heterosis that can be applied in rice production. The fact that the superior parental lines favorably enhance the performance of hybrid rice derived from many combinations in practice suggests that some common quantitative trait loci (QTL) may affect the performance of both the parental lines and the hybrids. Therefore, the detection of QTL controlling traits of the inbred lines and that of their hybrids is needed to understand the underlying genetic basis of the hybrid performance and to facilitate marker-aided breeding of hybrid rice.

The development of molecular markers in quantitative genetics greatly facilitates the study of quantitatively

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inherited complex traits related to F_1 heterosis and has made it possible to dissect the polygenes associated with such traits into individual Mendelian factors (PATERSON *et al.* 1988; STUBER *et al.* 1992). Molecular linkage genetic maps and QTL mapping techniques have been used to investigate the relationships between inbred lines and their hybrids and to identify QTL controlling agronomic traits and crop yields. STUBER *et al.* (1992) mapped QTL contributing to grain yield in two maize backcross (BC) populations derived from crosses between the F_3 progeny from a B73 \times Mo17 cross and their parental lines. The BC to B73 showed at least six QTL and the BC to Mo17 showed at least eight QTL for grain yield, and three of the QTL were detected in both of these BC populations. XIAO *et al.* (1995) investigated QTL controlling grain yield components in two rice BCF_1 populations between 198 F_7 recombinant inbred lines (RILs) and their parents. In all, 37 QTL were detected in the two populations: 27 QTL were detected in only one BCF_1 population and the other 10 were detected in two BCF_1 populations. LI *et al.* (2001) constructed five related rice mapping populations, including one RIL, two BC populations, and two testcross (TC) populations, which they analyzed in an attempt to detect QTL and found no correlation between the F_1 populations and their maternal RILs in terms of biomass yield and grain yield. Using data from the same experiment, LUO *et al.* (2001) found no correlations between the F_1 populations and their maternal RILs for the grain yield components PP (panicles per plant) and GP (grains per panicles). However, there was a significant correlation for GW (1000-grain weight). In current hybrid rice breeding, the male-sterile lines are crossed with elite restorer lines that are unrelated in pedigree, rather than backcrossed. The genetic relationships between the parental lines and F_1 hybrids remain to be elucidated.

In the study reported in this article, we introduced an experimental design that produced TC populations by mating the RILs with maintaining lines for currently popular improved male-sterile lines with different types of cytoplasmic male sterilities and a genic male-sterile line, which are unrelated by pedigree to the RILs. The RILs were derived from a cross between Minghui 63, a parent of the most widely grown hybrid variety, Shanyou 63, and B5, a breeding line with superior resistance developed at Wuhan University. The RILs and their corresponding TC hybrids were evaluated for seven traits of agronomic importance. The objectives of this study were to detect and evaluate the QTL controlling the agronomic traits in RILs and the performance of TC hybrids and to understand the genetic relationship between the inbred lines and their F_1 hybrids of improved modern rice.

MATERIALS AND METHODS

Plant materials: Five related mapping populations were used in this study. One was an RIL population composed of

187 F_8 lines derived by single-seed descent from a cross between Minghui 63 and B5 (Minghui 63/B5). Minghui 63 is a restorer line for many hybrid rice varieties. Three maintaining lines (Zhenshan 97B, II-32B, and YuetaiB) were selected to cross with the RILs because they have the same genomic composition as the corresponding cytoplasmic male-sterile (CMS) lines that are currently in common use. These lines are the maintaining lines for Zhenshan 97A, II-32A, and YuetaiA, which have WA-, IA-, and HL-type cytoplasmic male sterility, respectively. The other selected line was Peiai64s, a photo-thermo-sensitive genic male-sterile (PTGMS) line in which sterility is controlled by recessive nucleic genes. Four TC populations were developed, consisting of 160 Zhenshan 97B F_1 hybrids (Zhenshan97B/RILs, TCP_1), 181 II-32B F_1 hybrids (II-32B/RILs, TCP_2), 187 YuetaiB F_1 hybrids (YuetaiB/RILs, TCP_3), and 187 Peiai64s F_1 hybrids (Peiai64s/RILs, TCP_4).

Phenotypic evaluation: The 902 F_1 TC lines and 187 F_8 RILs were laid out in a field in a randomized complete block design with two replications (plots) for phenotypic evaluation in the summer of 2002 at the experimental farm of the Hubei Academy of Agricultural Sciences (Wuhan, China). Each plot consisted of three rows, each with 10 hills. Seedlings, 30 days old, of all experimental materials were transplanted in the field with a spacing of 16.7 cm between plants within each row and 26.7 cm between the rows. The middle six plants in the central row of each plot were sampled for analysis. The seven quantitative traits investigated were: heading date (HD; in days), plant height (PH; in centimeters), panicles per plant (PPP), spikelets per panicle (SPP), grains per panicle (GPP), GW (in grams), and grain yield per plant (GYPP; in grams). For SPP, GPP, and GYPP, all panicles in a plant were counted. Means over replications, for each trait and for each population, were used for QTL and other analyses.

Molecular markers and linkage maps: Preparation of genomic DNA from the parents and RILs followed the CTAB method as described by MURRAY and THOMPSON (1980). Two types of markers, RFLPs and SSRs, were used to survey DNA polymorphisms in the RILs. RFLP analyses, including restriction digestion, Southern blotting, and hybridization, were essentially as described by HUANG *et al.* (2001). Six restriction enzymes (*Apa*, *Bam*HI, *Hind*III, *Eco*RI, *Eco*RV, and *Dra*I) were used for surveying RFLPs. The RFLP probes were kindly provided by the Japanese Rice Genome Research Project and S. D. Tanksley and S. McCouch, Cornell University. In addition, 300 primer pairs from published data were used to survey SSR polymorphisms between the parents. The analysis, including PCR reactions and detection, essentially followed the methods of WU and TANKSLEY (1993). The DNA markers that detected polymorphisms between the parents were used to assay the entire population of 187 RILs. Molecular marker linkage maps were constructed using MAP-MAKER/EXP version 3.0 (LINCOLN *et al.* 1992).

Data analysis: For mapping main-effect and epistatic QTL, QTLMapper version 1.0 (WANG *et al.* 1999) was employed to identify loci affecting quantitative traits on the basis of composite interval analysis. Here, the main-effect QTL and epistatic QTL were defined as QTL with main effect and interaction between a pair of QTL, respectively (LI *et al.* 2001). A LOD score of 3.0 was selected as the threshold for the presence of a main-effect QTL on the basis of the total map distance and the average distance between markers; a LOD of 5.0 was used for declaring the existence of a putative pairs of epistatic QTL. With such a threshold, a false-positive QTL would be detected anywhere in the entire genome with a probability of ~ 0.05 (LYNCH and WALSH 1998). An independence test in which the initial scan suggested that two or more QTL were located on the same chromosome was performed, as described by PATERSON *et al.* (1988) and LANDER and BOTSTEIN (1989). The total phenotypic variation explained by

TABLE 1
Summary statistics for the seven quantitative traits measured in RILs and four TC populations

Population	Items	PH (cm)	HD (days)	PPP	SPP	GPP	GW (g)	GYPP (g)
RILs	Mean ± SD	99.7 ± 5.57	90.3 ± 2.97	12.0 ± 1.30	89.9 ± 13.68	69.9 ± 13.65	26.4 ± 1.88	20.8 ± 8.4
	Heritability ^a	73.1	89.3	47.3	86.7	82.5	80.3	42.3
TCP ₁	Mean ± SD	105.5 ± 6.47	88.6 ± 3.34	14.5 ± 3.45	131.4 ± 16.21	108.5 ± 15.89	26.0 ± 1.26	34.2 ± 9.08
	Heritability	67.2	87.5	38.6	84.9	65.7	68.8	45.8
TCP ₂	Mean ± SD	106.3 ± 5.90	92.6 ± 4.51	12.9 ± 3.61	140.1 ± 16.33	122.6 ± 16.78	25.9 ± 1.23	33.1 ± 11.13
	Heritability	69.3	84.3	35.2	85.2	64.6	67.7	53.6
TCP ₃	Mean ± SD	109.2 ± 3.51	85.0 ± 1.42	14 ± 2.46	136.3 ± 14.80	101.6 ± 14.00	24.5 ± 1.11	32.4 ± 5.56
	Heritability	58.1	79.6	37.4	75.9	67.4	72.7	51.7
TCP ₄	Mean ± SD	107.1 ± 3.04	86 ± 1.44	12.7 ± 2.42	142.4 ± 12.90	114.6 ± 12.44	23.5 ± 1.05	31.8 ± 4.37
	Heritability	65.3	82.7	45.5	79.8	77.2	76.5	48.2

PH, HD, PPP, SPP, GPP, GW, and GYPP indicate plant height, heading date, panicles per plant, spikelets per panicle, grains per panicle, 1000-grain weight, and grain yield per plant, respectively.

TCP₁, TCP₂, TCP₃, and TCP₄ indicate the TC populations comprising the F₁ offspring of Zhenshan 97B × RILs, II-32B × RILs, YuetaiB × RILs, and Peiai64s × RILs, respectively.

^aHeritability was broad-sense Heritability (h_B^2), $h_B^2 = V_G / (V_G + V_E) \times 100\%$.

all QTL was estimated by fitting a multiple regression model into the QTLMapper program.

RESULTS

Molecular marker linkage map: A molecular marker linkage map was constructed on the basis of the RILs of Minghui 63/B5 with 187 lines that served as the base population for generating the four TC populations employed in this study. A total of 244 molecular markers, including 190 RFLP and 54 SSR loci, were mapped on 12 linkage groups, covering 1478 cM according to the Kosambi function with an average interval of 6.1 cM between adjacent markers. The markers distributed relatively evenly among the chromosomes, and marker orders on the maps were in good agreement with those on previously published maps (CAUSSE *et al.* 1994; HARUSHIMA *et al.* 1998). Genotype segregation ratios of Minhui 63 and B5 followed the expected Mendelian ratio of 1:1 for most of the markers, except 12 markers (RM233, R2724, Y0193, R569, Y143D, S1520, R1679, RM242, R562, S1559B, R1506, and R2672) displayed distorted segregation ratios. This map is suitable for QTL analysis.

The performance of the populations: The means, SDs, and heritability of seven quantitative traits measured in the RILs and the four TC progenies are listed in Table 1. The measurements of seven traits varied widely in the RILs and the four TC progenies. The values for all of the traits were approximately normally distributed (data not shown), indicating the feasibility of QTL mapping for all these traits in the RILs and the four TC populations. The mean values showed that the grain yield and its components, except for GW, of the TC populations were higher than the corresponding values of the RIL population. The TC plants were also taller than the RIL plants. The heritability was high for PH,

HD, SPP, GPP, and GW and low for PPP and GYPP in the RIL and the four F₁ populations.

Relationships between the trait values of RILs and F₁ populations: Table 2 shows the phenotypic correlation coefficients between the values of individual F₁ hybrids and the values of their paternal RILs for the seven traits investigated. For all traits evaluated, there was a significant correlation between the means of the RILs and their F₁ performance in four TC populations, and the performance of TC hybrids was related to that of the RILs. The correlation coefficients were high for PH, HD, and GW; intermediate for PPP, SPP, and GPP; and low for GYPP. The presence of significant correlation between RILs and F₁'s for the investigated seven traits differs from the findings of LUO *et al.* (2001), possibly because of differences in the experimental materials and design between the two studies.

The contributions of six component traits to GYPP: The partial R^2 of six component traits to the total variances of grain yield per plant in the five populations are listed in Table 3. Regression analyses indicated that the three main yield traits PPP, GPP, and GW had high contributions (partial R^2) to the total variances of GYPP and that the contributions of the other traits were very low in the five populations. The partial R^2 in the testcross populations (TCPs) TCP₁, TCP₂, TCP₃, and TCP₄ were 65.4, 42.3, 38.0, and 65.6% for PPP; 23.6, 8.8, 84.4, and 77.0% for GPP; and 1.6, 3.8, 7.5, and 9.7% for GW, respectively. For the RILs, the partial R^2 was 24.5, 72.3, and 9.8% for PPP, GPP, and GW. For the three traits, the observed levels of the partial R^2 were higher for PPP and GPP than for GW.

Main-effect QTL across RILs and TC progeny: A total of 67 main-effect QTL affecting the seven traits in the RILs and the four TC populations were identified (Table 4, Figure 1). Seventeen main-effect QTL, dispersed among all 12 chromosomes, were mapped for

TABLE 2
Phenotypic correlation (*r*) coefficients for traits of agronomic importance between the mean trait values of RILs and F₁ performance

Population	Traits						
	PH	HD	PPP	SPP	GPP	KGW	GYPP
TCP ₁	0.412**	0.761**	0.468**	0.468**	0.430**	0.594**	0.222*
TCP ₂	0.465**	0.851**	0.671**	0.671**	0.685**	0.680**	0.321**
TCP ₃	0.604**	0.576**	0.350**	0.350**	0.495**	0.725**	0.234*
TCP ₄	0.610**	0.765**	0.285**	0.285**	0.453**	0.750**	0.356**

* Significance levels of $P < 0.05$; **significance levels of $P < 0.01$.

PH, which explained 50.1, 37.1, 53.0, 40.2, and 55.7% of the variance of this trait in the RILs, TCP₁, TCP₂, TCP₃, and TCP₄, respectively. Five of these main-effect QTL were detected in the RILs. The main-effect QTL *ph1*, which is bordered by markers S1501-C904 on chromosome 1 and had the strongest effect in the RILs, was detected in three TC populations (TCP₂, TCP₃, and TCP₄). The other 5 main-effect QTL detected in the RILs were not detected in any TC populations. In all, 12 main-effect QTL were detected in TC populations but not in RILs: *ph12b* (C2808-RG543) was detected in all four TC populations, *ph6* (C688-S1520) and *ph10a* (RM304-G2155) were detected in two TC populations [(TCP₁ and TCP₄) and (TCP₁ and TCP₂), respectively], while each of the other 9 main-effect QTL was detected in only one TC population. For the common main-effect QTL, for example, *ph1*, which was detected in the RILs and three TC progenies, the direction of the parental (B5) contribution was the same. However, the magnitude of *ph1*'s effect was not consistent across the RILs and the three TC progenies (4.2, 3.0, 2.1, and 3.0 in the RILs, TCP₂, TCP₃, and TCP₄, respectively; see Table 4); the difference in main effect was significant between RILs and TCP₃, and not among other populations. The same tendencies were observed for the other common main-effect QTL.

Only four main-effect QTL were detected for HD, accounting for 61.9, 67.1, 71.6, 21.5, and 36.5%

of the total variance of the trait in the RILs, TCP₁, TCP₂, TCP₃, and TCP₄, respectively. Two main-effect QTL were detected in the RILs. *Hd6a*, which is bordered by markers C688-S1520 on chromosome 6 and has the largest effect in the RILs, was also identified in four TC populations. This allele contributed a considerable portion to the total heading date variation in the TCPs (47.9% on average, ranging from 21.5 to 69.0%). Another main-effect QTL detected in RILs, *hd1*, was also identified in TCP₂. The other two main-effect QTL (*hd6b* and *hd7*) were identified in only one TC population. For the common main-effect QTL (*hd6a*), detected across the RILs and four TC progenies, the source of parental contribution was the same (Minghui 63).

Six main-effect QTL affecting the number of PPP were revealed, explaining 28.6, 14.1, 11.6, 10.3, and 30.9% of the total variance of this trait in the RILs, TCP₁, TCP₂, TCP₃, and TCP₄, respectively. Three of these main-effect QTL were detected in the RILs. The QTL *ppp2*, which is bordered by markers RM341-RM327 on chromosome 2 and has the largest effect in the RILs, was detected in three TC populations (TCP₂, TCP₃, and TCP₄). The other two main-effect QTL (*ppp4* and *ppp12*) found in the RILs were not detected in any TC populations. Four main-effect QTL (*ppp3*, *ppp6a*, *ppp6b*, and *ppp10*) were identified in TC populations, but not in the RILs. The direction of parental contribution and

TABLE 3
The partial R^2 of six traits to the total variances of grain yield per plant in the five populations

Population	Traits					
	PH	HD	PPP	SPP	GPP	GW
RILs	6.27 E-04	1.87E-05	0.245**	4.52E-05	0.723**	0.098**
TCP ₁	2.72 E-03**	1.55 E-04	0.654**	6.19 E-03**	0.236**	0.016**
TCP ₂	0.034**	1.96 E-03	0.423**	0.097**	0.088**	0.038**
TCP ₃	1.95 E-03	1.24 E-03	0.380**	8.08 E-03**	0.844**	0.075**
TCP ₄	1.8E-05	6.62 E-03**	0.656**	7.14 E-03**	0.770**	0.097**

* Significance levels of $P < 0.05$; **significance levels of $P < 0.01$.

the magnitude of the effect for the common QTL on the number of panicles per plant displayed the same trends as those affecting PH and HD.

We identified nine main-effect QTL affecting SPP, which explained 47.5, 39.9, 30.5, 54.0, and 40.7%, respectively, of the total variance of this trait in the five related populations. Three of these QTL were detected in the RILs. The QTL *spp1*, which is flanked by markers S1501-C904 on chromosome 1 and has the largest effect in the RILs, was also detected in four TC populations. Main-effect QTL *spp7* was identified in only one TC population (TCP₂) and *spp4* was not identified in any TC populations. Six main-effect QTL were revealed in TC populations, but not in RILs. Of these, *spp5* (RM164-C624) was identified in two TC populations (TCP₃ and TCP₄), and each of the other five was detected in only one of the four TC populations.

Eight main-effect QTL related to the number of GPP were found, which explained 49.0, 21.5, 29.2, 38.2, and 49.0% of the total variance of this trait in the five related populations, respectively. Three main-effect QTL were detected in the RILs: *gpp1* (which is flanked by markers S1501-C904 on chromosome 1 and has the largest effect in RILs) was detected in two TC populations (TCP₃ and TCP₄); the QTL *gpp6a* (C688-S1520) was identified in one TC population (TCP₂); and *gpp7* (R1789-RM242) was not detected in any TC populations. Five main-effect QTL were revealed in TC populations, but not in RILs: *gpp8b* and *gpp10* were identified in two of the four TC populations [(TCP₃ and TCP₄) and (TCP₁ and TCP₂)], respectively, while each of the other three was detected in only one of the four TC populations.

As many as 16 main-effect QTL affecting GW were detected, which explained 70.1, 20.2, 15.2, 59.1, and 48.4% of the total variance of this trait in the five related populations, respectively. Eight main-effect QTL were detected in the RILs: *gw1a*, which mapped between markers C904 and R596 on chromosome 1, had the largest effect in RILs and was also detected in three TC populations (TCP₁, TCP₃, and TCP₄); *gw8a* and *gw11* were also identified in TCP₃ and TCP₄; *gw1b*, *gw5*, and *gw6* were each identified in just one TC population. Two main-effect QTL (*gw3a* and *gw7*) detected in the RILs were not identified in any TC populations. Eight main-effect QTL were found in TC populations but not in the RILs. Of these, *gw3b* was detected in three TC populations, and both *gw2a* and *gw3c* were detected in two TC populations. The other 5 main-effect QTL were each detected in only one of the four TC populations.

Seven main-effect QTL affecting GYPP were identified, which explained 42.5, 27.6, 37.4, 22.6, and 23.2% of the total variance of this trait in the five populations, respectively. Three of these main-effect QTL were detected in the RIL population; *gypp1*, which is located between S1501 and C904 on chromosome 1 and has the largest effect in RILs, was also detected in three TC populations, while the other two (*gypp3a* and *gypp4*) were

not identified in any TC populations. Four main-effect QTL that were not detected in the RIL population were found in TC populations: *gypp6* and *gypp10* were both identified in two TC populations, while each of the other two (*gypp3b* and *gypp8*) were found in only one TC population.

Epistatic QTL detected in the RILs and TC progenies: Table 5 shows 29 digenic epistatic QTL pairs identified in the RIL and four TC populations. No common digenic epistatic QTL pairs were detected in the RILs and all of the TC populations, and no epistatic QTL were identified for HD in the five populations.

For plant height, four pairs of epistatic QTL (one in both the RILs and TCP₄, two in TCP₂) were identified, which explained 6.1, 8.9, and 2.8% of the total variation of this trait in the RILs and the two TC F₁ populations, respectively. For panicles per plant, six pairs of epistatic QTL (one in each of the RILs, TCP₁, and TCP₃, and three in TCP₄) were detected, explaining 5.9, 11.0, 13.5, and 18.5% of the total variation of this trait in the RILs and three TC populations, respectively. For spikelets per panicle, five pairs of epistatic QTL (one in TCP₁, two in both the RILs and TCP₄) were found, accounting for 12.1, 3.7, and 11.1% of the total variation of this trait in the RILs and two TC F₁ hybrids, respectively. Only two pairs of epistatic QTL (one in both TCP₁ and TCP₄) were revealed for grain number per panicle, explaining 14.4 and 6.9% of total phenotypic variation in two TC F₁ hybrids, respectively. For 1000-grain weight, six pairs of epistatic QTL (one in TCP₁, two in TCP₄, three in TCP₃) were detected, which explained 10.1, 12.7, and 5.7% of total phenotypic variation in three TC F₁ hybrids, respectively. For grain yield per plant, six pairs of epistatic QTL (one in TCP₃, two in TCP₂, and three in TCP₄) were identified, explaining 10.7, 5.3, and 19.0% of total phenotypic variation in the three TC F₁ populations, respectively. In summary, only a small number of epistatic QTL were detected and the degrees of variation that they explained were relatively small compared to the main-effect QTL.

DISCUSSION

Selection of TC populations and improved modern rice as experiment materials: It is essential to use appropriate experimental designs and materials for QTL mapping, and strenuous efforts have been made to construct experimental populations for detecting and analyzing QTL in the last decade. Plant populations with various genetic structures have been developed for the purpose, mainly consisting of F₂/F₃, BC, double haploids (DHs), RILs, and backcross inbred lines (BILs). In rice, permanent populations, such as DHs and RILs, are used most often, because of their inherent advantages of providing a permanent DNA supply and phenotyping opportunities for many different studies. Allelic differences are limited in these populations, since only two

TABLE 4
Main-effect QTL affecting traits of agronomic importance detected in RILs and four TC populations

Main-effect QTL ^a	Chromosome	Marker interval	RILs		TCP ₁		TCP ₂		TCP ₃		TCP ₄		Significant test ^c		
			LOD ^b	Effect ^c	LOD	R ² % ^d	LOD	R ² %	LOD	R ² %	LOD	R ² %		LOD	R ² %
<i>ph1</i>	1	SI501-C904	15.67	-4.2	23.4										
<i>ph2</i>	2	C920-C1769	4.12	-2.6	5.6										
<i>ph3a</i>	3	SI0656-r273	6.26	3.0	9.3										
<i>ph3b</i>	3	RM168-yl171													
<i>ph4</i>	4	yl03D-C2807	3.60	-2.4	4.7										
<i>ph5</i>	5	RM274-w143D													
<i>ph6</i>	6	C688-S1520													
<i>ph7</i>	7	C615-R565				4.95	-3.6	14.1						**	
<i>ph8</i>	8	R2662-RZ572													
<i>ph9a</i>	9	C1257-G1085													
<i>ph9b</i>	9	R1562-C570													
<i>ph10a</i>	10	RM304-G2155													
<i>ph10b</i>	10	S2083-RZ649A				7.74	-3.8	16.8	15.63	-3.8	30.0				
<i>ph11a</i>	11	RG2-C1172	4.99	2.8	7.1										
<i>ph11b</i>	11	R2918-S1559B													
<i>ph11c</i>	11	C6-G376													
<i>ph12</i>	12	C2808-RG543				3.56	-2.7	6.2	4.62	-2.5	8.0			NS	
<i>hd1</i>	1	y517-S2139	6.71	-2.2	7.2				3.37	-1.9	2.6			NS	
<i>hd6a</i>	6	C688-S1520	12.52	4.3	54.7	14.27	3.9	57.9	15.09	5.7	69.0			NS, **	
<i>hd6b</i>	6	SI1809-RM3				3.28	1.7	3.3							
<i>hd7</i>	7	R565-C1521				5.82	1.9	5.9							
<i>ppp2</i>	2	RM341-RM327	5.99	0.6	13.2				4.78	0.8	11.6			NS	
<i>ppp3</i>	3	R2404-R2443													
<i>ppp4</i>	4	R1496-C820	3.88	0.4	7.7										
<i>ppp6</i>	6	C688-S1520				3.75	-1.2	14.1							
<i>ppp10</i>	10	RZ649A-G1084													
<i>ppp12</i>	12	S826-C751A	3.83	-0.4	7.7										
<i>spp1</i>	1	SI501-C904	9.64	-15.1	26.1	13.52	-8.5	32.8	6.02	-8.4	22.1	10.85	-5.3	18.4	NS, *, **
<i>spp2a</i>	2	C1769-RM6				3.22	-3.9	7.1							
<i>spp2b</i>	2	RM327-S2068										8.52	-4.5	13.0	
<i>spp4</i>	4	yl03D-C2807	4.27	-5.4	8.5										
<i>spp5</i>	5	RM164-C624													
<i>spp6</i>	6	RI1679-C751B													
<i>spp7</i>	7	RI1789-RM242	4.29	7.0	12.9				9.36	7.0	8.4				NS
<i>spp8a</i>	8	C309-S2108													
<i>spp8b</i>	8	RM342-G1073													
<i>gpp1</i>	1	SI501-C904	10.40	-18.2	27.6										NS, **
<i>gpp3</i>	3	RM203-R2404													
<i>gpp6a</i>	6	C688-S1520	4.29	-13.0	12.9										**
<i>gpp6b</i>	6	RM3-R2654				3.23	-3.0	4.4				3.21	3.4	5.6	

(continued)

TABLE 4
(Continued)

Main-effect QTL ^a	Chromosome	Marker interval	RILs		TCP ₁		TCP ₂		TCP ₃		TCP ₄		Significant test ^c	
			LOD ^b	Effect ^c	LOD	R ² % ^d	LOD	R ² %	LOD	R ² %	LOD	R ² %		LOD
<i>gpp7</i>	7	R1789-RM242	4.27	12.4	8.5									
<i>gpp8a</i>	8	G278-C166				3.51	-8.2	4.0						
<i>gpp8b</i>	8	C309-S2108							4.52	-3.9	7.6	8.55	-4.4	12.3
<i>gpp10</i>	10	RM304-G2155				7.89	-9.7	21.5	20.8					NS
<i>gva1a</i>	1	C904-R596	10.6	-1.1	16.2	5.51	-0.5	13.1	9.38	-0.4	6.3	5.38	-0.3	4.5
<i>gva1b</i>	1	CDO455-RM5	4.18	0.6	5.6				4.66	0.3	4.4			NS, **
<i>gva1c</i>	1	RM283-C101				3.46	0.4	7.1						NS
<i>gva2a</i>	2	Y1-400-R208							10.2	0.5	8.5	14.00	0.6	8.3
<i>gva2b</i>	2	C920-C1769							3.33	-0.3		3.33	-0.3	2.7
<i>gva3a</i>	3	y4831-RM135	5.27	-0.8	9.7									NS
<i>gva3b</i>	3	RM55-RM203							5.44	-0.3	5.2	4.07	-0.3	4.5
<i>gva3c</i>	3	RM282-G162				3.02	-0.4	7.3	3.59	-0.4	7.9	3.13	-0.3	4.3
<i>gva5</i>	5	C624-RM26	3.14	-0.6	5.5							4.55	-0.3	3.1
<i>gva6</i>	6	R1679-C751B	5.40	-0.7	6.8									*
<i>gva7</i>	7	C615-R565	8.13	0.9	13.1				3.16	-0.3	4.3			
<i>gva8a</i>	8	S2108-C483	6.08	0.7	6.3				7.08	0.3	5.5	3.62	0.3	3.6
<i>gva8b</i>	8	R2662-RZ572										8.04	-0.5	8.2
<i>gva9</i>	9	RM205-S11302										5.33	0.4	3.0
<i>gva10</i>	10	G1084-RG257	4.24	-0.7	6.9				4.49	-0.3	3.5			
<i>gva11</i>	11	R2918-S1559B							17.66	-0.9	21.4	8.04	-0.5	6.2
<i>gpp1</i>	1	S1501-C904	18.08	-3.4	31.3	6.08	-2.6	13.3	4.23	-1.6	10.8	6.93	-1.5	15.4
<i>gpp3a</i>	3	R1925-G1318	4.53	1.4	6.6									NS, *
<i>gpp3b</i>	3	G162-y4831										3.13	1.1	7.8
<i>gpp4</i>	4	C2807-RM252	3.06	-1.2	4.6									
<i>gpp6</i>	6	C688-S1520				3.37	-5.2	10.5	2.95	-4.3	7.7			NS
<i>gpp8</i>	8	C309-S2108				5.68	-6.6	17.1	8.08	-2.6	16.4			**
<i>gpp10</i>	10	RM304-G2155							6.89	-4.5	11.8			

NS, no significance; *significance levels of $P < 0.05$; **significance levels of $P < 0.01$.

^aQTLs are named by trait abbreviations plus chromosomal number.

^bA LOD score of 3.0 was used for declaring the existence of a putative QTL, according to the method of LYNCH and WALSH (1998).

^cIn the RILs, QTL effect is the additive effect, *i.e.*, the average substitution effect of the B5 allele by the Minhui allele. In the TC populations, QTL effects were estimated by the difference between the heterozygote tester/Minhui and the heterozygote tester/B5. The heterozygosity was 17.22, 16.67, 14.44, and 17.78% for Zhenshan97B/B5, II-32B/B5, YuetaiB/B5, and Peiai64s/B5, respectively, and 19.94, 20.00, 19.44, and 18.89% for Zhenshan 97B/Minhui, II-32B/Minhui, YuetaiB/Minhui, and Peiai64s/Minhui, respectively. The genetic expectation of the QTL effect is the additive gene effect (α) when estimated from the RILs, the additive effects, and the dominance effects ($\alpha + d$) from the F₁ mean values.

^dVariation explained by each QTL.

^eFor testing the difference of QTL effect estimated among different populations.

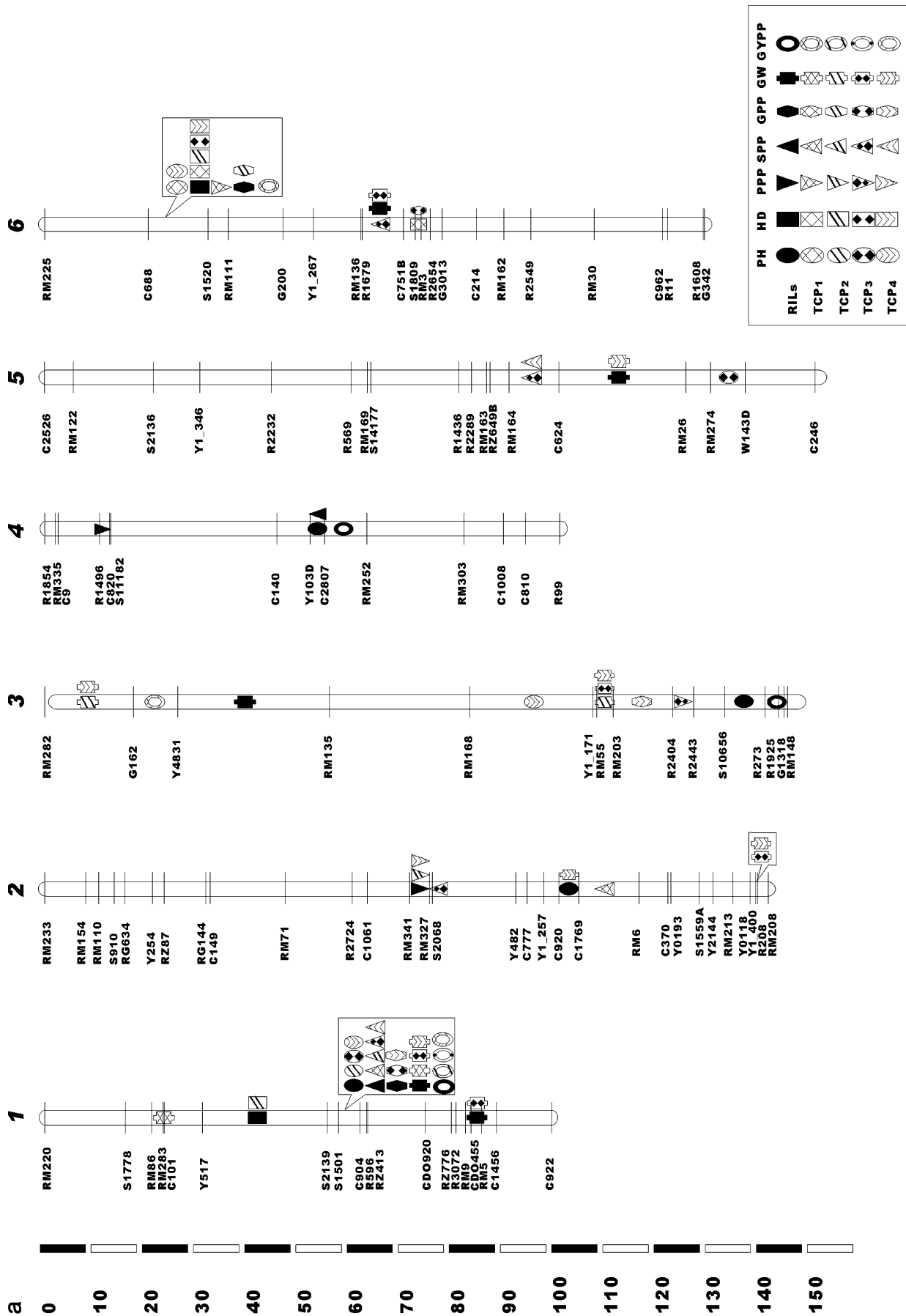


FIGURE 1.—Genomic locations of main-effect QTL for the seven investigated traits of agronomic importance in the Minghui63/B5 RILs and four testcross F₁ populations.

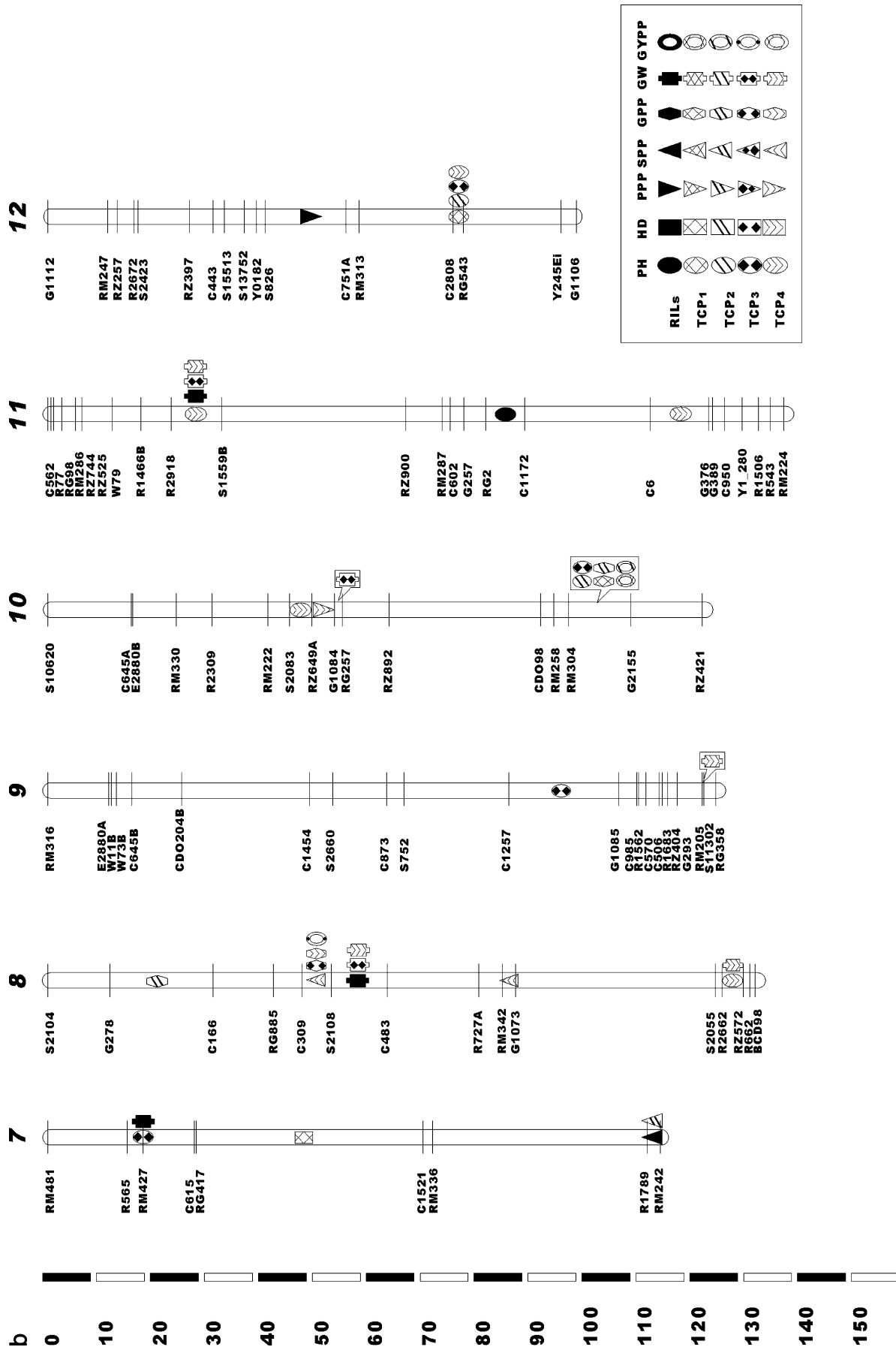


FIGURE 1.—Continued.

TABLE 5
Epistatic QTL affecting traits of agronomic importance detected in RILs and TC progenies

Trait	Population	Chromosome	Interval i	Chromosome	Interval j	LOD ^a	Variance (%)		Phenotypic effect ^d
							Locus ^b	Trait ^c	
PH	RILs	6	RM30-C962	11	C602-G257	5.25	6.1	6.1	-2.8
	TCP ₂	2	Y1257-C920	6	R11-R1608	5.48	4.9		-2.3
		2	Y0193-S1559A	10	RM304-G2155	18.2	4.0	8.9	-2.2
	TCP ₄	4	R1854-RM335	10	S2083-RZ649A	5.23	2.8	2.8	-1.6
PPP	RILs	3	y1-171-RM55	9	C985-R1562	5.58	5.9	5.9	-0.4
	TCP ₁	8	RM337-G278	10	E2880B-RM330	5.21	11.0	11.0	-1.2
	TCP ₃	2	y254-RZ87	12	RM313-C2808	5.47	13.5	13.5	0.5
	TCP ₄	2	y2144-RM213	4	RM303-C1008	5.00	6.2		-0.4
		4	C140-y103D	9	G293-RM205	5.03	5.6		0.4
	5	R2289-RM163	6	R11-R1608	5.06	6.7	18.5	0.4	
SPP	RILs	4	R1854-RM335	9	C1257-G1085	5.88	5.9		3.9
		11	R2918-S1559B	12	S13752-Y0182	6.00	6.2	12.1	4.0
	TCP ₁	2	C1769-RM6	3	R2404-R2443	6.09	3.7	3.7	3.6
	TCP ₄	2	RG144-C149	8	C309-S2108	10.2	3.3		-2.7
	8	RM342-G1073	11	R1466B-R2918	6.77	7.8	11.1	4.1	
GPP	TCP ₁	6	C688-S1520	10	RM304-G2155	17.9	14.4	14.4	-16.2
	TCP ₄	5	RZ649B-RM164	10	RM330-R2309	6.60	6.9	6.9	3.5
GW	TCP ₁	4	RM303-C1008	10	E2880B-RM330	5.00	10.1	10.1	-0.4
	TCP ₃	1	R596-RZ413	2	S910-RG634	9.38	3.2		0.2
		3	RM168-y1-171	7	C615-R565	6.20	6.7		0.3
		8	S2108-C483	11	RG543-y245EI	7.08	2.8	12.7	-0.2
	TCP ₄	2	C370-Y0193	11	G257-RG2	5.46	1.9		-0.2
	5	C624-RM26	9	S752-C1257	7.30	3.8	5.7	-0.3	
GYPP	TCP ₂	2	RG634-y254	4	y103D-C2807	5.40	5.9		3.6
		8	RM337-G278	10	RM304-G2155	8.73	5.8	10.7	-3.5
	TCP ₃	4	y103D-C2807	5	R2232-R568	5.00	5.3	5.3	-1.4
	TCP ₄	1	y517-S2139	1	C904-R596	9.81	9.2		1.8
		2	RM233-RM154	6	R1679-C751B	5.26	3.6		-1.1
	8	G1073-S2055	9	C873-S752	5.07	6.2	19.0	-1.5	

^a A LOD score of 5.0 was used for declaring the existence of putative pairs of epistatic QTL, according to the method of LYNCH and WALSH (1998).

^b Variance (%) for locus is the proportion of variance explained by the component epistasis.

^c Variance (%) for trait is the total variance of this trait explained by all epistatic QTL detected in a population.

^d Phenotypic effect is the effect arising from interactions between alleles at the loci i and j, as defined by MATHER and JINKS (1982).

alleles segregate at each polymorphic locus. In rare cases, TC progenies and immortalized F₂ populations have been used (LI *et al.* 2001; HUA *et al.* 2002). In studies seeking common QTL in inbred lines and their hybrid progenies, backcross populations are usually employed. In the study reported in this article, we introduced a variant of design III (COMSTOCK and ROBISON 1948), which produced four TC progenies by mating three maintaining lines for different types of cytoplasmic male sterility and one PTGMS line with the RILs. With such populations, we could compare the QTL across the RILs and the four TC populations with a set of common male parents (RILs). Our design was expected to increase the scope to identify QTL, and even multiple alleles at

QTL loci, because of the inclusion of four testers as parents of the mapping populations. This was particularly true since the four testers were genetically divergent and unrelated in pedigree to the RILs. As shown by the data presented here, we were able to detect a total of 67 main-effect QTL (or 9.6 QTL/trait) and 29 epistatic QTL pairs (or 4.1 pair/trait) in the populations.

Another feature of this experiment is the selection of improved modern rice lines as the testers for TCs and the parents for the RILs. Minghui63, a parental variety of the RILs, is a restorer line of the most popular hybrid rice variety, Shanyou 63. The total planting area of Shanyou63 has exceeded 67 million hectares (ha) in China and other Asian countries. Minghui63 is also a restorer line

for many other hybrid rice varieties, with a combined total planting area of 130 million ha. The high general combining ability and the genetic basis of Minghui63 are attractive to rice breeders. Three maintaining lines used in this experiment are among the best in China. Zhenshan97B is the maintainer of Zhenshan97A, which is the female parent of Shanyou63 and a number of other hybrid varieties with a total planting area of 108 million ha. In addition, the respective CMS lines of II-32B and YuetaiB, which have different types of cytoplasmic male sterility, are widely employed in rice breeding programs and the areas planted with them are rapidly growing. Another tester, Peiai64S, is a PTGMS rice line that is the parental line of the first super hybrid rice in China. Utilizing Peiai64s, many hybrid rice combinations have been released for rice production. For field trials, we used TC progenies derived directly from crossing the breeding lines (RILs) with maintaining lines for different types of CMS and PTGMS lines, closely resembling practices in modern rice breeding strategies. Data on QTL and the genetic features associated with them, detected using improved modern rice varieties, are expected to be more attractive to breeders than data obtained using other types of material since such results are likely to be highly applicable to contemporary rice breeding programs.

Comparison of QTL mapped in RILs and their TC populations in rice: In this study, F_1 performance was related to the performance of the parental lines according to both the phenotypic correlation analysis (Table 2) and the QTL mapping (Table 4). In all, 67 distinct main-effect QTL were identified for seven traits in the RILs and four TC populations. Of these, 27 were identified in the RIL population. For each trait, the QTL that had the strongest effect in the RILs (giving a total of 9 such QTL) were also detected in two, three, or four TC populations. Six main-effect QTL in RILs were identified in one TC populations. Another 12 main-effect QTL detected in RILs were not identified in any TC populations. In all instances of common main-effect QTL across the RILs and multiple TC progenies, the direction of parental contribution was the same. Several features of the distribution of main-effect QTL in the RILs and TC progenies for the seven traits under study can be noted. First, only the main-effect QTL with the largest effects in the RILs for each trait were detected in two or more TC populations, and except for GW only one such QTL was identified for the other six traits considered. Second, some main-effect QTL that were detected in RILs were identified in one TC population, and more than half of the main-effect QTL detected in RILs were identified in TC populations. Third, effects of the common main-effect QTL among the RILs and four TC progenies may vary in the magnitude of their substitution effects, but do not change in parental contribution. Fourth, both Minghui 63 and B5 contributed to increased trait values across the RILs and four TC populations for the seven traits evaluated.

A number of main-effect QTL were detected either in TC populations or in RILs. When comparing QTL mapped in RILs and their TC populations, we have to take into account the fact that TC progenies are likely to show only half of the difference in performance attributable to any specific marker compared to the difference between the testers and the RILs. In RILs, a QTL is identified when the additive effect between lines homozygous for the allele from the parents is significant. TC progenies carry only one allele from RILs in combination with another allele from the tester. A QTL is detected when the substitution effect of replacing an allele from RILs with the allele from the tester is significant. Possible interactions between the parental allele with the tester allele also have to be considered when comparing different types of progeny. The discrepancies between QTL mapped in RILs and their TC populations could be caused by genetic effects and are easy to explain by considering the genetic components of the RILs and TC populations. Main-effect QTL effects detected in the TC populations represent the differential intralocus interactions between the parental alleles from the RILs and those from the respective testers—due to the segregation of alleles with additive, partial recessiveness, intermediate gene action, or dominance—or the difference between the two heterozygous loci (Minghui63/tester-B5/tester). If a tester carries an allele that is fully dominant over the alleles carried by RILs, the corresponding QTL will not be detected in RILs but may be detected in the TC progenies. Overdominance of the RIL alleles over tester alleles can also lead to divergent results among TC progenies unless the four testers carry the same allele. The parental (Minghui 63 and B5) alleles in QTL associated with a single tester presumably have specific dominance interactions with the respective testers that do not occur with the other testers. Another possible explanation for the differences between QTL mapped in RILs and TC populations is the presence of epistatic effects. Twenty-nine pairs of distinct epistatic QTL were identified for six traits, and no common epistatic QTL were identified for the same trait across the RILs and the four TC progenies in this study. Hence, epistasis is very likely a major cause for the inconsistencies of QTL detection across RILs and their TC populations and is specific to the cross combination.

Cluster distribution of the main-effect QTL: In this study, 67 distinct main-effect QTL distributed among 12 chromosomes were identified for seven traits across the RILs and four TC populations. A very interesting feature is the highly concentrated distribution of the QTL in a few chromosomal regions and the existence of QTL hot spots (Figure 1). This is particularly true for the region around the S1501-C904 locus on chromosome 1 and the C688-S1520 locus on chromosome 6, where QTL for several traits were detected in the RILs and the four TC populations. Similar concentrated distributions of QTL

have also been observed in previous studies (XIONG *et al.* 1999; LI *et al.* 2000). Particular attention should be given to such QTL hot spots in future studies of gene cloning and functional genomics.

Positional convergence of main-effect QTL in rice:

Traits of agronomic importance, including those in this study, are useful characters for QTL analysis in rice. A QTL associated with grain weight or length has been reported in the centromere region of rice chromosome 3 in at least 10 different inter- and intraspecific populations of independent studies, suggesting that a homologous gene determining seed weight or size may be associated with domestication and subsequent selection (LI *et al.* 2004). For the seven QTL with strongest main effect (*i.e.*, *ph1*, *ppp2*, *spp1*, *gpp1*, *gw1*, *gypp1*, and *hd6a*), we explored the publicly available QTL database (<http://www.gramene.org>) to search their alignment QTL identified in the same chromosome region in previous studies. For *ph1*, the same QTL associated with plant height has been reported in six populations: CNHZAU Zh97/Ming63 RI (CUI *et al.* 2002), Cornell 9024/LH422 RI (XIAO *et al.* 1996), Cornell Jef/Oruf BC (THOMSON *et al.* 2003), IRRI Mor/CO39 (HUANG *et al.* 1996), IRRI IR64/Azu DH (VENUPRASAD *et al.* 2002), and IRRI Lem/Teq RI QTL (MEI *et al.* 2003). In the vicinity of *ph1*, two genes associated with plant height have been identified and isolated: one is the gibberellin biosynthetic gene *OsGA3ox2* (ITOY *et al.* 2001) and the other is the brassinosteroid biosynthetic gene *D2* (HONG *et al.* 2003). The same QTL of *hd6a* has been identified in four populations, including CNHZAU Zh97/Ming63 RI (YU *et al.* 2002), IRRI IR64/Azu DH (LI *et al.* 2003), JRGP Nip/Kas F₂ (LIN *et al.* 1998), and NIAS Kosh/Kas BIL (YAMAMOTO *et al.* 2001). One allele of the gene (*Hd3a*) has been cloned, which encodes a protein closely related to Arabidopsis FT (KOJIMA *et al.* 2002). For five yield-related QTL (*i.e.*, *ppp2*, *spp1*, *gpp1*, *gw1*, and *gypp1*), the same QTL have been reported in the corresponding chromosome regions in 5, 9, 3, 3, and 4 populations, respectively, among total 13 populations: CNHZAU Zh97/Ming63 RI (CUI *et al.* 2002; HUA *et al.* 2002, 2003), CNHZAU Zhe97/Wuy2 (JIANG *et al.* 2004), CNRRI Tes/CB (ZHUANG *et al.* 1997), CNRRI Zh97B/Mil46 RI (ZHUANG *et al.* 2001, 2002), Cornell 9024/LH422 RI (XIAO *et al.* 1996), Cornell IR64/IRG105 (SEPTININGSIH *et al.* 2003), Cornell Jef/Oruf BC (THOMSON *et al.* 2003), HNAES MIL23/Aki RI (YAGI *et al.* 2001), IGCAS ZYQ8/JX17 F₂ (XU *et al.* 1995), IRRI Lem/Teq RI (MEI *et al.* 2003), IRRI Mil23/Aki RI (NAGATA *et al.* 2002; KOBAYASHI *et al.* 2003), JRGP Nip/Kas F₂ (YAMAYA *et al.* 2002), and NIAS Kosh/Kas near isogenic lines (OBARA *et al.* 2004). Recently, the gene *Gn1a*, which produces more grains per panicle near the QTL *gpp1* and encodes a cytokinin oxidase, has been cloned (ASHIKARI *et al.* 2005). The results show that these main-effect QTL have been identified in a much wider range of populations and under different environ-

TABLE 6

Phenotypic variance (%) for seven traits explained by main-effect QTL and epistatic QTL detected in RILs and TC progenies

Trait	Loci type	RILs	TCP ₁	TCP ₂	TCP ₃	TCP ₄
PH	Main-effect QTL	50.1	37.1	53.0	40.2	55.7
	Epistatic QTL	6.1	—	8.9	—	2.8
	Total	56.2	37.1	61.9	40.2	58.5
HD	Main-effect QTL	61.9	67.1	71.6	21.5	36.5
	Epistatic QTL	—	—	—	—	—
PPP	Main-effect QTL	28.6	14.1	11.6	10.3	30.9
	Epistatic QTL	5.9	11.0	—	13.5	18.5
	Total	34.5	25.1	11.6	23.8	32.4
SPP	Main-effect QTL	47.5	39.9	30.5	54.0	40.7
	Epistatic QTL	12.1	3.7	—	—	11.1
	Total	59.6	43.6	30.5	54.0	51.8
GPP	Main-effect QTL	49.0	21.5	29.2	38.2	49.0
	Epistatic QTL	—	14.4	—	—	6.9
	Total	49.0	35.9	29.2	38.2	55.9
GW	Main-effect QTL	70.1	20.2	15.2	59.1	48.4
	Epistatic QTL	—	10.1	—	12.7	5.7
	Total	70.1	30.3	15.2	71.8	54.1
GYPP	Main-effect QTL	42.5	27.6	37.4	22.6	23.2
	Epistatic QTL	—	—	10.7	5.3	19.0
	Total	42.5	27.6	48.1	27.9	19.0

—, indicates that no epistatic QTL was identified in this population for the trait evaluated.

ments in independent studies in rice. Correspondence in the location of QTL in different taxa suggests that some of the underlying genes are identical (PATERSON *et al.* 1995). On the basis of the positional convergence of QTL across different populations, the structure and function of the underlying genes might be conserved across different varieties in the rice gene pool.

The data listed in Table 4 and Table 6 indicate that the genes underlying the main-effect QTL, with large effects, might determine the phenotype of the traits studied in RILs and TC hybrid populations. The effect of these QTL is more evident in modern improved varieties than in traditional varieties, indicating that the loci have been the targets of selection associated with breeding practice. The main-effect QTL detected in RILs are expressed in heterozygous F₁ as showed in this report, which is an important part of the genetic basis of heterosis. The QTL are expressed in different genetic backgrounds and environments, making them valuable targets for gene manipulation and also for application in rice breeding. By comparing QTL mapped in RILs and their hybrid populations in rice, and with a publicly available QTL data reservoir, specific candidate loci will be identified to address fundamental problems in rice

improvement. Recently, a fine mapping of a grain-weight QTL has been constructed (Li *et al.* 2004). Such information should be valuable for positional cloning genes underlying QTL and for marker-aided selection of QTL in rice breeding programs. As correspondence in the location of QTL exists in different taxa of grasses (PATERSON *et al.* 1995), main-effect QTL identified in rice will be useful in other cereal crops.

Implications for genetic improvement and marker-aided breeding of hybrid rice: In our study, the detected main-effect QTL had distinct effects on both the RILs and the TC hybrids. The results indicate that the contributions of the inbred line on F₁ performance were quite stable across unrelated testers. The common main-effect QTL showing effects on multiple TC progenies may be associated with the general combining ability in hybrid breeding. On the other hand, a number of epistatic QTL were detected. Compared with the main-effect QTL, the effects of the epistatic QTL were relatively weak and present in some, but not all, crosses (Table 6). These cross-specific epistatic QTL are likely to be related to the special combining ability in hybrid breeding. Our findings have several implications for contemporary hybrid rice breeding. First, the elite varieties can be selected for use as restorers, as long as they carry fertility-restoring genes, and may contain a relatively high number of main-effect QTL that give large contributions to yields. Second, when a restorer line that carries the common main-effect QTL is bred, it can be used in a number of cross combinations. The direction of the contribution from the main-effect QTL of the restorer will be consistent in the cross combinations, according to our results. Third, to further improve the hybrid performance, the selection of the genetic background of the restorer line and the maintaining line should be seriously considered in rice breeding, since the epistatic QTL are cross combination dependent.

Since most quantitative traits of interest to plant breeders are considered to have poor heritabilities, plant breeders are more interested in the phenotypic variability among progenies of crosses between the inbred lines than in the lines *per se*. Breeders of hybrid crops need to improve the performance of the traits in the inbred lines and evaluate the progenies of test-crosses with unrelated testers. A crucial question in hybrid rice breeding is whether the QTL in inbred lines are stable across different testers. The identification of QTL in RILs that affect the performance of F₁ populations and the accurate estimation of their genetic effects, including epistasis, is essential for efficient hybrid breeding of crops. Common QTL exhibiting effect across RILs and the F₁ population in this study have also been detected in the large number of previous studies in rice. With the development of dense linkage maps based on molecular markers, marker-aided transfer and selection of the common QTL to improve hybrid productivity is expected to be possible. The common

main-effect QTL for each trait with the largest effects in both the RILs and different TC populations, and the chromosome regions harboring multiple main-effect QTL for different traits such as those between S1501 and C904 on chromosome 1 and between C688 and S1520 on chromosome 6, will be valuable targets in marker-aided selection for efficient hybrid rice breeding and functional genomic studies.

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