Single- and Multiple-Trait Mapping Analysis of Linked Quantitative Trait Loci: Some Asymptotic Analytical Approximations

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ABSTRACT

Estimating the resolution power of mapping analysis of linked quantitative trait loci (QTL) remains a difficult problem, which has been previously addressed mainly by Monte Carlo simulations. The analytical method of evaluation of the expected LOD developed in this article spreads the "deterministic sampling" approach for the case of two linked QTL for single- and two-trait analysis. Several complicated questions are addressed through this evaluation: the dependence of QTL detection power on the QTL effects, residual correlation between the traits, and the effect of epistatic interaction between the QTL for one or both traits on expected LOD (ELOD), etc. Although this method gives only an asymptotic estimation of ELOD, it allows one to get an approximate assessment of a broad spectrum of mapping situations. A good correspondence was found between the ELODs predicted by the model and LOD values averaged over Monte Carlo simulations.

MANY efforts have been devoted to increasing the a complicated problem arising when the considered
efficiency of marker analysis of quantitative traits,
including interval analysis (Landan and Batatain 1999).
 $\frac{7}{2}$ and including interval analysis (Lander and Botstein 1989; Zeng 1995; Korol *et al.* 1998). If one tries to fit a single-Knott and Haley 1992), selective sampling (Lebowitz locus model to such a case, a ghost QTL can be detected *et al.* 1987; Darvasi and Soller 1992, 1994; Weller in an interval that has no effect on the trait (Knott *et al.* 1997), replicated progeny testing (Soller and and Haley 1992; Martinez and Curnow 1992; Beckmann 1990), and sequential experimentation Wright and Kong 1997). Especially difficult are situa-
(Boehnke and Moll 1989; Motro and Soller 1993). tions with *trans* effects of linked QTL (Knott and (Boehnke and Moll 1989; Motro and Soller 1993). Recently, a general method to improve the efficiency Haley 1992; Luo and Kearsey 1992). That *trans*-associ-
of quantitative trait loci (QTL) mapping was proposed ation of QTL could be a common phenomenon even of quantitative trait loci (QTL) mapping was proposed ation of QTL could be a common phenomenon even by taking into account simultaneous segregation at in interspecific crosses has been demonstrated by many genomic segments that affect the trait in question DeVicente and Tanksley (1993) in tomato: they found many genomic segments that affect the trait in question DeVicente and Tanksley (1993) in tomato: they found
(Jansen and Stam 1994: Zeng 1994). A situation in that up to 36% of the detected QTL had alleles with (Jansen and Stam 1994; Zeng 1994). A situation in that up to 36% of the detected QTL had alleles with
which one QTL (or a chromosome segment) affects effects opposite to the direction expected from the pawhich one QTL (or a chromosome segment) affects effects opposite to several traits simultaneously can also be considered to rental differences. several traits simultaneously can also be considered to rental differences.

result in increased power (Korol *et al.* 1987, 1994, 1995. The usual way of dealing with several linked QTL is result in increased power (Korol *et al.* 1987, 1994, 1995, The usual way of dealing with several linked QTL is 1998; Jiang and Zeng 1995; Ronin *et al.* 1995; Zeng multiple regression analysis or mixture model analysis 1997). Such an analysis may be important in marker-
assisted breeding strategies, dissecting heterosis as a for segregation of QTL of the same chromosome (Janassisted breeding strategies, dissecting heterosis as a for segregation of QTL of the same chromosome (Jan-
multilocus multitrait phenomenon, developing opti-
sen and Stam 1994; Zeng 1994). The third possibility multilocus multitrait phenomenon, developing opti-
mized programs for evaluation and bioconservation of is to construct two- to three-interval mixture models, mized programs for evaluation and bioconservation of is to construct two- to three-interval mixture models,
genetic resources, and revealing genetic architecture of although this approach is rather cumbersome and genetic resources, and revealing genetic architecture of although this approach is rather cumbersome and
fitness systems in natural populations, etc. Multiple-trait are needs intensive calculations. Employing Monte Carlo fitness systems in natural populations, etc. Multiple-trait needs intensive calculations. Employing Monte Carlo mapping analysis proved to be very useful within the simulations with mixture models, we demonstrated re-
framework of the selective genotyping design (Weller cently the advantage of multiple trait analysis in detecframework of the selective genotyping design (Weller

The multiple-trait approach may help in coping with

et al. 1997; Ronin *et al.* 1998). The *et al.* 1998). The *et al.* 1998). The multiple-trait approach may help in coping with goal of this article is to elaborate an analytical model enabling us to evaluate in a general form the expected LOD values in cases of two linked QTL. Such a model Corresponding author: A. B. Korol, Institute of Evolution, University

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Cation one can consider the possibility of cation one can consider the possibility of calculating

the minimum sample size needed to detect linked QTL with certain effects on either of the correlated quantitative traits or to prove the existence of epistasis for any of the traits. Likewise, the proposed analysis allows us to predict situations where a ghost QTL will be detected using interval analysis and to evaluate the minimum marker density needed to prevent such a possibility for given effects of the linked QTL. Recently, a similar technique, referred to as "deterministic sampling," was applied to single-QTL situations in single-trait analysis, with the expected LOD values calculated numerically (Mackinnon and Weller 1995; Mackinnon *et al.* 1996; Wright and Kong 1997). Our major target here is
analytical and numerical deterministic sampling for twoanalytical and numerical deterministic sampling for *two*
 trait analysis with linked QTL. We first treat the case of

a range-trait analysis and then generalize the results for

the two-trait analysis. The consideratio

SINGLE-TRAIT ANALYSIS

- The major target of our analysis is analytical and numerical deterministic sampling with linked QTL. There fore, analytical expression of ELOD should be obtained where in case that allows us to compare H_2 (two linked QTL) and H_1 (single QTL) for any set of parameter values.

Single-QTL models: Let a trait *x* be dependent on two linked loci Q_1/q_1 and Q_2/q_2 and let the trait values in the four QTL groups *Q*1*Q*1*Q*2*Q*2, *Q*1*Q*1*q*2*q*2, *q*1*q*1*Q*2*Q*2, and $q_1q_1q_2q_2$ of a mapping population have normal densities $f_{11}(x)$, $f_{12}(x)$, $f_{21}(x)$, and $f_{22}(x)$ with (unknown) means μ_{11}^* , μ_{12}^* , μ_{21}^* , and μ_{22}^* and standard deviations σ_{11}^* , σ_{12}^* , σ_{21}^{*} , and σ_{22}^{*} , respectively. Usually, the mapping procedure is started with the assumption of one QTL in the chromosome and then one can try to apply some versions of single-marker or interval analysis. Reduced test power, biased parameter estimates, and detection of ghost factors may result from this simplification, as demonstrated by simulation studies (Knott and Haley
1992; Luo and Kearsey 1992; Martinez and Curnow
1992; Korol *et al.* 1998). This question is treated here alternative groups can be represented as analytically for both single-marker and single-interval analysis (see also Wright and Kong 1997).

Consider a random sample of individuals genotyped for marker loci from the chromosome that carries the two QTL. With a dense molecular map, one analyzes consequently a series of markers with five different locations relative to the linked QTL: (2)

in the alternative marker groups for an arbitrary marker will be

$$
h_{MM}(x) = h_1 = \alpha_1 f_{11}(x) + \alpha_2 f_{12}(x) + \beta_2 f_{21}(x) + \beta_1 f_{22}(x),
$$

\n
$$
h_{mm}(x) = h_2 = \beta_1 f_{11}(x) + \beta_2 f_{12}(x) + \alpha_2 f_{21}(x) + \alpha_1 f_{22}(x),
$$

(a) $\alpha_1 = (1 - r_1)(1 - r), \alpha_2 = (1 - r_1)r,$ $\beta_1 = r_1(1 - r), \qquad \beta_2 = r_1r,$ (b) $\alpha_1 = 1 - r$, $\alpha_2 = r$, $\beta_1 = 0,$ $\beta_2 = 0,$ (c) $\alpha_1 = (1 - r_1)(1 - r_2), \quad \alpha_2 = (1 - r_1)r_2,$ $\beta_1 = r_1 r_2,$ $\beta_2 = r_1(1 - r_2),$ (d) $\alpha_1 = 1 - r, \qquad \alpha_2 = 0,$ $\beta_1 = 0,$ $\beta_2 = r,$ (e) $\alpha_1 = (1 - r)(1 - r_2), \quad \alpha_2 = rr_2,$ $B_1 = (1 - r)r_2,$ $B_2 = r(1 - r_2).$ (1)

$$
\begin{aligned}\n\tilde{\mu}_1 &= \alpha_1 \mu_{11}^* + \alpha_2 \mu_{12}^* + \beta_2 \mu_{21}^* + \beta_1 \mu_{22}^*, \\
\tilde{\mu}_2 &= \beta_1 \mu_{11}^* + \beta_2 \mu_{12}^* + \alpha_2 \mu_{21}^* + \alpha_1 \mu_{22}^*, \\
\tilde{\sigma}_1^2 &= \alpha_1 \sigma_{11}^{*2} + \alpha_2 \sigma_{12}^{*2} + \beta_2 \sigma_{21}^{*2} + \beta_1 \sigma_{22}^{*2} + G(\alpha_1, \alpha_2, \beta_1, \beta_2), \\
\tilde{\sigma}_2^2 &= \beta_1 \sigma_{11}^{*2} + \beta_2 \sigma_{12}^{*2} + \alpha_2 \sigma_{21}^{*2} + \alpha_1 \sigma_{22}^{*2} + G(\beta_1, \beta_2, \alpha_1, \alpha_2),\n\end{aligned}
$$

TABLE 1

Asymptotic estimates of parameters and max ELODs (mELODs) of a single-QTL model when applied to the case of two linked QTL $(r = 0.25)$ for coupling and repulsion phases

Phase	d_1^*	d_2^*	d_1	d_2	σ_1	σ_{2g}		$mELOD_{11}$ $mELOD_{12}$	mELOD ₂
\mathcal{C}		0.5	0.75	0.75			10.11	10.11	
R	0.5	-0.5	0.25	-0.25	0.83	0.83	1.22	1.22	3.84
\mathcal{C}		0.3	0.86				13.19		
\mathbb{R}	0.7	-0.3	0.55		0.81		5.92		1.41

The results of application of single marker sliding are presented. It appears that the ELOD function may have two local maxima (mELOD₁₁ and mELOD₁₂) coinciding with the true positions of the QTL (see also Hyne and Kearsey 1995; Wright and Kong 1997), resulting in two sets of parameter estimates. For comparison, we also provide the results of two-QTL sliding, $mELOD_2$ for the hypothesis H_2 (two linked QTL) *vs.* H_1 (single QTL), assuming the sample size $n = 250$. C and R stand for coupling and repulsion phases, respectively; d *, d_2^* , d_1 , and d_2 , are the true and estimated values of the effects of the first and second QTL, respectively; σ_1 and σ_2 are the estimates of the residual standard deviations (the true value was $\sigma^* = 0.8$). Note that the same values of mELOD₂ are presented for the two linkage phases, C and R (for further details see the two-trait analysis section).

$$
G(a, b, c, d) = a(c(\mu_{11}^* - \mu_{22}^*) + d(\mu_{11}^* - \mu_{21}^*))^2
$$

+ $b(c(\mu_{12}^* - \mu_{22}^*) + d(\mu_{12}^* - \mu_{21}^*))^2$ QTI
+ $d(a(\mu_{21}^* - \mu_{11}^*) + b(\mu_{21}^* - \mu_{12}^*))^2$ in t
+ $c(a(\mu_{22}^* - \mu_{11}^*) + b(\mu_{22}^* - \mu_{12}^*))^2$ for
+ $a b(\mu_{11}^* - \mu_{12}^*)^2(1 + c + d)$
+ $c d(\mu_{22}^* - \mu_{21}^*)^2(1 + a + b)$. and

Assuming that our trial marker is tightly linked to (or coincides with) the putative (single) QTL and that the trait distributions in alternative groups are normal, we can calculate the regression of the log-likelihood as a function of parameter set $\theta = (\mu_1, \mu_2, \sigma_1, \sigma_2)$: where

$$
U_1(\theta) = \mathbf{E} \sum_{i=1}^{n_1} \log \{ (\sigma_1 \sqrt{2\pi})^{-1} \exp[-(x_i - \mu_1)^2 / (2\sigma_1^2)] \} \hat{\mu} = \frac{1}{2}
$$

+
$$
\mathbf{E} \sum_{i=n_1+1}^{n_1} \log \{ (\sigma_2 \sqrt{2\pi})^{-1} \exp[-(x_i - \mu_2)^2 / (2\sigma_2^2)] \}, \hat{\sigma}^2 = \frac{1}{2}
$$

where **E** stands for expectations. To obtain the asymp-
totic estimates of parameter values, one can calculate $\hat{\sigma}^2$ and one can obtain the expression for the maximum
max $U_1(\theta)$:

$$
\max_{\theta} U_1(\theta) = U_1(\tilde{\mu}_1, \tilde{\mu}_2, \tilde{\sigma}_1, \tilde{\sigma}_2)
$$

= -0.5n(1 + log(2\pi\tilde{\sigma}_1\tilde{\sigma}_2)) = V_1(\tilde{\sigma}_1, \tilde{\sigma}_2),

trial marker with respect to Q_1/q_1 and Q_2/q_2 loci. For Clearly, $V_1(\cdot) - V_0$ reaches a local maximum when the any trial marker position *l* the conditional max of U_1 marker coincides with one of the QTL. One can easily (e) is $V_1(\tilde{\sigma}_1(I),\tilde{\sigma}_2(I))$. We now maximize $V_1(\tilde{\sigma}_1(I),\sigma_2(I))$ see from the presented illustrations that the possibility with respect to marker position *l.* To save space we skip of finding an indication of the existence of two QTL the details and provide here only the final results. Our by revealing two local maxima depends on linkage phase

where **a** single-QTL mapping model to a situation with two a situation with two a situation with two a situation with two a situation α linked QTL. In particular, we compare the expected LOD scores corresponding to the hypotheses $H₁$, "one + $\mathit{b}(c(\mu_{12}^* - \mu_{22}^*) + d(\mu_{12}^* - \mu_{21}^*))^2$ QTL in the considered chromosome" and H₀, "no QTL in the chromosome." Then, we have consequently

+
$$
c(a(\mu_{22}^* - \mu_{11}^*) + b(\mu_{22}^* - \mu_{12}^*))^2
$$
 for $H_1 \to \max_j V_1(\tilde{\sigma}_1(I), \tilde{\sigma}_2(I)) = V_1$; for $H_0 \to U_0(\mu, \sigma)$
+ $a b(\mu_{11}^* - \mu_{12}^*)^2(1 + c + d)$
$$
= \mathbf{E} \log \prod_{i=1}^n (\sigma \sqrt{2\pi})^{-1} \exp(-(x_i - \mu)^2/(2\sigma^2)),
$$

$$
V_0 = \max_{\mu,\sigma} U_0(\mu,\,\sigma)
$$

=
$$
\mathbf{E} \log \prod_{i=1}^n (\hat{\sigma} \sqrt{2\pi})^{-1} \exp(-(x_i - \hat{\mu})^2/(2\hat{\sigma}^2)),
$$

$$
\hat{\mu} = \frac{1}{2} \{ (1 - r)\mu_{11}^* + \frac{1}{2} r \mu_{12}^* + \frac{1}{2} r \mu_{21}^* + \frac{1}{2} (1 - r) \mu_{22}^* \}
$$
\n
$$
+ \mathbf{E} \sum_{i=n_1+1}^n \log \{ (\sigma_2 \sqrt{2\pi})^{-1} \exp[-(x_i - \mu_2)^2 / (2\sigma_2^2)] \}, \qquad \hat{\sigma}^2 = \frac{1}{2} (1 - r) \sigma_{11}^{*2} + \frac{1}{2} r \sigma_{12}^{*2} + \frac{1}{2} r \sigma_{21}^{*2} + \frac{1}{2} r \sigma_{21}^{*2} + \frac{1}{2} (1 - r) \sigma_{22}^{*2}
$$
\n
$$
+ \mathbf{G} (\frac{1}{2} (1 - r), \frac{1}{2} r, \frac{1}{2} (1 - r), \frac{1}{2} r).
$$

 $\hat{\sigma}^2$) and one can obtain the expression for the maximum expected LOD value, max ELOD = $V_1 - V_0$.

These results allow us to evaluate the consequences of model misspecification. The behavior of the score $ELOD = V_1(\cdot) - V_0$ as a function of trial marker position where $\tilde{\sigma}_1$ and $\tilde{\sigma}_2$ are as defined in (2). and the parameters characterizing the effect of Q_1/q_1 Clearly, $\tilde{\sigma}_1$ and $\tilde{\sigma}_2$ depend on the position *l* of the and Q_2/q_2 are represented in Table 1 and Figure 1. task was to analyze the consequences of application of (coupling or repulsion), distance between the QTL,

and magnitudes of the QTL effects and their ratio (see Figure 1 and Table 1).

Two linked QTL: *ELOD for testing H₂ <i>vs. H₁*: As before, consider a situation when the target trait *x* depends on the two linked loci Q_1/q_1 and Q_2/q_2 with normal trait densities $f_{11}(x)$, $f_{12}(x)$, $f_{21}(x)$, $f_{22}(x)$ in the QTL groups $Q_1Q_2Q_2$, $Q_1Q_1q_2q_2$, $q_1q_1Q_2Q_2$, and $q_1q_1q_2q_2$ of the dihaploid mapping population characterized by unknown means μ_{11}^* , μ_{12}^* , μ_{21}^* , and μ_{22}^* , and (residual) standard deviations σ_{11}^* , σ_{12}^* , μ_{21}^* , and σ_{22}^* . Employment of two markers instead of one allows us to take into account both QTL. Several basic situations of marker loci positioning relative to the QTL could be considered:

Clearly, other possible situations are equivalent to these four, up to a replacement of parameters. In the foregoing single-marker sliding, we had two discrepancies between the model specification and the real situa tion: (i) only one QTL was assumed, and (ii) the trial marker was treated as if its position coincides with that of the putative QTL. Now the model is improved, because the first assumption is removed. Therefore, we can consider a process of sliding with a pair of markers along the chromosome as a tool to locate the pair of QTL. Such a procedure is equivalent to two-interval mapping analysis $\begin{array}{ccc} + & r_1(1-r_2)(1-r_3). \end{array}$ (3) (3)

$$
h_{11}(x) = \alpha_1 f_{11}(x) + \alpha_2 f_{12}(x) + \beta_2 f_{21}(x) + \beta_1 f_{22}(x)
$$

= $H(\alpha_1, \alpha_2, \beta_1, \beta_2),$

$$
h_{12}(x) = H(\tau_1, \tau_2, \delta_1, \delta_2),
$$

$$
h_{21}(x) = H(\sigma_1, \delta_2, \tau_1, \tau_2),
$$

$$
h_{22}(x) = H(\beta_1, \beta_2, \alpha_1, \alpha_2).
$$

(a)
$$
\alpha_1 = (1 - r_1)(1 - r_2)(1 - r_3)/s_{\alpha\beta}
$$
,
\n $\alpha_2 = (1 - r_1)(1 - r_2)r_3/s_{\alpha\beta}$,
\n $\beta_1 = r_1r_2r_3/s_{\alpha\beta}$,
\n $\beta_2 = r_1r_2(1 - r_3)/s_{\alpha\beta}$,
\n $s_{\alpha\beta} = r_1r_2 + (1 - r_1)(1 - r_2)$;
\n $\tau_1 = (1 - r_1)r_2r_3/s_{\tau\delta}$,
\n $\tau_2 = (1 - r_1)r_2(1 - r_3)/s_{\tau\delta}$,
\n $\delta_1 = r_1(1 - r_2)(1 - r_3)/s_{\tau\delta}$,
\n $\delta_2 = r_1(1 - r_2)r_3/s_{\tau\delta}$,
\n $s_{\tau\delta} = (1 - r_1)r_2 + r_1(1 - r_2)$;

(b)
$$
\alpha_1 = (1 - r_1)(1 - r_3), \quad \alpha_2 = (1 - r_1)r_3,
$$

\n $\beta_1 = r_1r_3, \qquad \beta_2 = r_1(1 - r_3),$
\n $\tau_1 = (1 - r_1)r_3, \qquad \tau_2 = (1 - r_1)(1 - r_3),$
\n $\delta_1 = r_1(1 - r_3), \qquad \delta_2 = r_1r_3;$

(c)
$$
\alpha_1 = (1 - r_1)(1 - r_2), \quad \alpha_2 = r_1 r_2,
$$

\n $\beta_1 = (1 - r_1) r_2, \quad \beta_2 = r_1 (1 - r_2),$
\n $\tau_1 = (1 - r_1)(1 - r_2), \quad \tau_2 = r_1 r_2,$
\n $\delta_1 = (1 - r_1) r_2, \quad \delta_2 = r_1 (1 - r_2);$

(d)
$$
\alpha_1 = (1 - r_1)(1 - r_2)(1 - r_3)/s_{\alpha\beta}
$$
,
\n $\alpha_2 = (1 - r_1)r_2r_3/s_{\alpha\beta}$,
\n $\beta_1 = r_1(1 - r_2)r_3/s_{\alpha\beta}$,
\n $\beta_2 = r_1r_2(1 - r_3)/s_{\alpha\beta}$,
\n $s_{\alpha\beta} = (1 - r_1)(1 - r_2)(1 - r_3)$
\n $+ (1 - r_1)r_2r_3 + r_1r_2(1 - r_3)$
\n $+ r_1(1 - r_2)r_3$,
\n $\tau_1 = (1 - r_1)(1 - r_2)r_3/s_{\tau\delta}$,
\n $\tau_2 = (1 - r_1)r_2(1 - r_3)/s_{\tau\delta}$,
\n $\delta_1 = r_1(1 - r_2)(1 - r_3)/s_{\tau\delta}$,
\n $\delta_2 = r_1r_2r_3/s_{\tau\delta}$,
\n $s_{\tau\delta} = (1 - r_1)(1 - r_2)r_3$
\n $+ (1 - r_1)r_2(1 - r_3) + r_1r_2r_3$
\n $+ r_1(1 - r_2)(1 - r_3)$. (3)

Jansen 1993; Korol *et al.* 1998) with vanishing lengths of For any pair of markers, one can assume that they coin-
the trial intervals. Because of the foregoing assumptions, the true expected densities of the trait distri the procedure described in the previous section. Consequently, one can calculate, for the current pair of markers, the expected LOD assuming two linked QTL (H_2 hypothesis), which can be compared to the expected log-likelihood obtained under the assumption of one
QTL (H_1) . We found that given independent variance $h_{22}(x) = H(\beta_1, \beta_2, \alpha_1, \alpha_2),$

with corresponding mixture parameters

with corresponding mixture parameters

effects of the linked QTL, the maximum of ELOD over

possible locations (l_1, l_2) of the trial pairs of mark

in the single-QTL model when applied to situations with two $a_2^* = 0.5$; **A**, $d_1^* = 0.6$ and $d_2^* = 0.4$; \bullet , $d_1^* = 0.7$ and d_2^*

values in the QTL mapping analysis in the case of two

inc. Certainly, the achievable maximum is higher in

linked QTL. However, these results are essentially as-

ymptotic and may be biased at small samples. Therefore,

pling phase; (b) repulsion phase. design depends on many factors characterizing the un-

ference (Haldane mapping function) was assumed. For each sample, we employed two subsets of markers, using the information on intervals 12 and 48. Table 2 shows the behavior of the average LOD values and the discrepancy between the estimated and simulated QTL positions as dependent on sample size and number of markers. The main conclusion from the simulations is that the proposed method can indeed serve as a basis to get an approximate prediction of the expected LOD for interval mapping of two linked QTL (compare the average max LODs with max ELODs).

It follows from the presented results that the difference between predicted max ELOD and the averaged Figure 1.—Behavior of ELOD for a single-marker sliding over simulations max LOD in repulsion phase is smaller
the single-QTL model when applied to situations with two than that in coupling phase, in spite of the fact that linked QTL, as a function of the QTL positions and effects. Theory predicts the same value for the two phases. In CP and RP, coupling and repulsion phases. The effects of the both cases the experimental LODs are smaller th CP and RP, coupling and repulsion phases. The effects of the both cases the experimental LODs are smaller than QTL are denoted by the following: \blacksquare , $d_i^* = 0.5$ and the predicted ones: *i.e.* for the same combination QTL are denoted by the following: $d_i^* = 0.5$ and the predicted ones; *i.e.*, for the same combinations of $d_i^* = 0.5$; \blacktriangle , $d_i^* = 0.6$ and $d_i^* = 0.4$; \blacktriangle , $d_i^* = 0.7$ and $d_i^* = 0.7$ and $d_i^* = 0.7$ and $d_i^* = 0.7$ Q1L are denoted by the following: $d_2^* = 0.3$ and the predicted ones; *i.e.*, for the same combinations of $d_2^* = 0.5$; \triangle , $d_1^* = 0.6$ and $d_2^* = 0.4$; \triangle , $d_1^* = 0.7$ and $d_2^* = 0.7$ and $d_2^* = 0.7$ and $d_2^$ sion phase. A simple explanation can be proposed for this effect. The simulated procedure includes analyses cide with those of the QTL (the proof is available from
authors upon request). The surface ELOD =
ELOD(I_1, I_2) represented in Figure 2 manifests an impor-
tant asymptotic property of interval QTL mapping with
the H₁ tant asymptotic property of interval QTL mapping with Haley and Knott 1992; Korol *et al.* 1998): (i) *fixed* vanishing interval length and increasing sample size: a
faster-than-linear growth of the criterion when ap-
proaching the true position of the QTL (the second
lated positions (which would not necessarily be true proaching the true position of the QTL (the second
derivatives are positive). Note that the same max ELOD
is predicted for coupling and repulsion phases.
Comparison of the analytical and simulation results: The
foregoing m

that we employed Monte Carlo simulations. Chromo-
somes with two linked QTL were modeled for two population sizes ($n = 500$ and 2000). No crossing-over inter-
for designing mapping experiments. For example, using the obtained expression of max ELOD, we can get an estimate of the *minimum sample size* needed to discriminate between H_1 and H_2 , when H_2 is true (*i.e.*, when we have a pair of linked QTL with some effects d_1 and d_2), with a certain preset test power. This is based on the fact that the expected LOD value is distributed as noncentral chi-square with degrees of freedom equal to the difference in the number of parameters specifying the alternatives $(H_2 \text{ and } H_1)$ (Wald 1943). This tool enables us to compare different practical situations with respect to the foregoing prediction of the minimum sample size (see Lander and Botstein 1989). The usefulness of Figure 2.—The ELOD surface for the alternative H_2 (two such an option is especially obvious for mapping of linked QTL, where the efficiency of the experimental linked QTL, where the efficiency of the experimental

known "configuration" of the problem: the distance of the putative QTL, their relative effects on trait mean value and variance, linkage phase (coupling *vs*. repulsion), and presence or absence of epistatic interaction, etc. We now consider two examples to illustrate the possibilities of the proposed analysis: the dependence of ELOD for H_2 *vs.* H_1 on epistasis and the detectability of epistasis provided H_2 is already proved.

The effect of epistatic interaction on ELOD for QTL detection: In the example on epistatic interactions the trait values in the four QTL classes were modeled as:

$$
\mu_{11} = \mu_0 + 0.5(d_1 + d_2) + \varepsilon,
$$

\n
$$
\mu_{21} = \mu_0 - 0.5(d_1 - d_2) - \varepsilon,
$$

\n
$$
\mu_{12} = \mu_0 + 0.5(d_1 - d_2) - \varepsilon,
$$

\n
$$
\mu_{22} = \mu_0 - 0.5(d_1 + d_2) + \varepsilon,
$$

where ε is the epistatic effect. It is of high practical importance to predict the expected power of detection of epistatic interactions within the framework of QTL of the effects and positions of the involved QTL. Clearly, Korol *et al.* 1994; Fu and Ritland 1996). This problem such a fitting. Then, when fitting H₂ for data with epistamodel of two-QTL data may depend on the peculiarities of two linked QTL as compared to one QTL). Two

Figure 3.—The behavior of the maximum ELOD (H₂ *vs.*) H_1) as a function of the effects of the QTL (*d*) and the level of epistasis (ε) . Here *d* is the effect of the first QTL and we assumed that the effect of the second QTL also varies, in such a manner that the sum of the absolute values of both effects is equal to 1. The numerical values of the parameters were $n = 250$, $r = 25\%$, and $\sigma = 0.8$.

mapping analysis (Haley and Knott 1992; Eaves 1994; if epistasis is present it also may affect the results of is addressed in the next section. Here we consider first sis, one can either adopt or ignore epistasis, which may the question of how the presence of epistasis may affect affect the final result. Figure 3 shows the behavior of the power of QTL detection. The fitted single-QTL max ELOD for testing H_2 *vs.* H_1 (testing for the presence

TABLE 2

Comparison of the average LOD values (over 200 Monte Carlo runs) with the asymptotic values of max ELOD for comparing the hypotheses H_2 (two linked QTL in the chromosome) *vs*. $H₁$ (one QTL in the choromosome)

		Phase	Number of intervals						
					12		48		
			12 48		Average estimated positions of the QTL				
\boldsymbol{n}	max ELOD		Average LOD value		L_1	L ₂	L_{1}	L ₂	
500	7.68	C	5.26 (1.46)	5.88 (1.49)	14.37 (3.36)	49.66 (3.22)	14.10 (2.38)	49.47 (2.52)	
		R	6.64 (2.17)	7.15 (2.25)	14.80 (3.64)	49.48 (3.50)	14.29 (2.60)	49.49 (2.66)	
2000	30.72	C	23.46 (3.55)	26.84 (3.61)	14.29 (1.54)	49.15 (1.12)	14.29 (0.56)	49.02 (0.62)	
		R	27.33 (4.64)	29.07 (4.73)	14.72 (1.59)	49.12 (1.18)	14.41 (0.70)	49.19 (0.73)	

Monte Carlo simulations (200 runs) were conducted for doubled haploid mapping populations with $n =$ 500 and 2000 genotypes. A single chromosome was modeled with two QTL (*Q*1/*q*¹ and *Q*2/*q*² of equal additive effects $(d_1 = d_2 = 0.5)$ in coupling (C) and repulsion (R) phases; the residual standard deviations were all 0.8. Chromosomes with 49 equidistant markers with a recombination rate of 0.02 between adjacent markers were simulated assuming no interference (hence Haldane mapping function). The simulated positions of the QTL coincided with markers 8 and 25. For the considered two marker densities (*i.e.*, with 12 and 48 intervals), correspondingly 13 and 49 markers were used in two-interval analysis. Thus, the simulated positions of the QTL were $L_1 = 14.29$ cM and $L_2 = 48.98$ cM. For both LOD values and estimated positions of the QTL, we provide the averages (first rows) and standard deviations (second rows in parentheses). The hypothesis H_2 (two linked QTL) was compared to H_1 (a single QTL) by fitting the H_1 model with no presumed constraints on the position of the putative single QTL; correspondingly, the two-QTL model was fitted by scanning over all possible pairs of intervals (see Halley and Knott 1992; Korol *et al.* 1998).

TABLE 3

		QTL with no epistasis), and H_0 (no QTL in the chromosome)						
\boldsymbol{n}		H_2 ($\epsilon \neq 0$) $VS. H_0$	H_2 ($\varepsilon = 0$) \mathbf{v} s. \mathbf{H}_0	H_2 (ε \neq 0) vs. H ₂ ($\varepsilon = 0$)	Power ($\alpha = 5\%$) H ₂ (ε \neq 0) <i>vs.</i> H ₂ (ε = 0)			
250		16.25 14.71	15.06 13.74	1.19 0.97	55%			
500	$s - t$.s	1.54 30.48 29.42	1.32 28.45 27.48	0.22 2.02 1.94	76%			
	$s - t$	1.06	0.97	0.08				

Comparison of the average LOD values (over 100 Monte Carlo runs) with the asymptotic values of max ELOD for comparing the hypotheses H₂ (two linked QTL with epistasis), H₂ (two linked $\bf QTL$ with no epistasis), and \bf{H}_0 (no $\bf QTL$ in the chromosome)

A situation with two linked epistatically interacting QTL was considered for coupling phase. The positions, the additive effects, and the residual variances of the QTL are as described in Table 2. The epistatic effect was $\varepsilon = 0.125$. Here *s* denotes the average LOD resulting from simulation experiments whereas *t* denotes the predicted max ELOD for the compared alternatives H₂ ($\varepsilon \neq 0$) *vs.* H₂ ($\varepsilon = 0$) (d.f. = 1).

0 vs. H₂ under $\varepsilon = 0$: In the foregoing section we could somes affecting either of the traits, and by pleiotropy see how epistasis affects the expected LOD values when and linkage of genes from the chromosome under consingle-QTL and two-QTL models are applied to the anal- sideration. ysis. The proposed tool allows us also to predict the As in single-trait analysis, to analyze two-QTL situaexpected LOD for the situation when one wants to con- tions we calculate max ELOD for the alternative hypothtrast two versions of the hypothesis H_2 (two linked QTL eses: H_2 *vs.* H_1 . This means that we need to develop in the chromosome): H_2 ($\varepsilon = 0$), *i.e.*, additive effects of bivariate analogues of the foregoing single-QTL and the QTL, and H₂ ($\varepsilon \neq 0$), *i.e.*, assuming epistasis. Testing two-QTL models based on single- and two-marker slidfor epistasis (coadaptation) and evaluating the magni- ing procedures. Hence, the goal of the first part of this tude of epistasis have recently become an important section is to obtain the regression of the log-likelihood component of QTL mapping analysis (Doebley *et al.* function assuming that only one QTL resides in the 1995; Li *et al.* 1997). This meaningful subject has a long chromosome that in fact carries two linked QTL. Let history in both evolutionary genetics (Dobzhansky the traits *x* and *y* be dependent on two loci, Q_1/q_1 and 1970; Wright 1977), theories of recombination and *Q*2/*q*2, residing in the marked chromosome and let the sex evolution (Barton 1995; Otto and Feldman bivariate trait distributions in the four QTL groups
1997), and agricultural genetics (Yu *et al.* 1997). How- $Q_1Q_1Q_2Q_2$, $Q_1Q_1q_2q_3$, $q_1q_1Q_2Q_2$, and $q_1q_1q_2q_3$ 1997), and agricultural genetics (Yu *et al.* 1997). How $Q_1Q_1Q_2Q_2$, $Q_1Q_1q_2q_2$, $q_1q_1Q_2Q_2$, and $q_1q_1q_2q_2$ of dihaploid ever, only with QTL mapping can epistatic effects be mapping population be normal de ever, only with QTL mapping can epistatic effects be mapping population be normal densities $f_{11}(x, y)$, $f_{12}(x, z)$ objectively detected and evaluated. Each of the forego-
 \dot{y} , $f_{11}(x, y)$, and $f_{22}(x, y)$ with unkno ing alternative versions of $H₂$, without and with epistasis, can be compared to H_0 (no QTL in the chromosome), using the proposed approximation. The difference between the resulting max ELODs will give us max ELOD for the presence of epistasis. An example presented in for the presence of epistasis. An example presented in apping procedure is started with the assumption of Table 3 illustrates the closeness between the predicted one QTL in the chromosome, and then one could try to Table 3 illustrates the closeness between the predicted one QTL in the chromosome, and then one could try to

et al. 1998), joint analysis of correlated quantitative traits two-trait analysis.

conclusions can be derived from our analysis: (i) the may increase the mapping resolution in situations with effect of epistasis on detection power is symmetric with linked QTL, *i.e.*, when H_2 (two linked QTL) and H_1 respect to the sign of ε, and (ii) epistasis may have either (one QTL) are compared. The higher the residual cora positive or negative effect on max ELOD of the test relation the better the expected LOD. In two-trait analyof H_1 *vs.* H_0 (not shown) and always a positive effect sis, the residual correlation between the traits in the when testing H_2 *vs.* H_1 (Figure 3; see also Eaves 1994). QTL groups may be caused by nongenetic mechanisms, *The detectability of epistasis (comparison of* H_2 *under* $\varepsilon \neq$ pleiotropy, or linkage of genes from other chromo-

 y , $f_{21}(x, y)$, and $f_{22}(x, y)$ with unknown vectors of means $\{\mu_{11x}^*\} = \{\mu_{11x}^*, \mu_{12x}^*, \mu_{21x}^*, \mu_{22x}^*\}$ and $\{\mu_y^*\} = \{\mu_{11y}^*, \mu_{12y}^*, \mu_{21y}^*\}$ $\{\sigma_{11x}^* \} = \{\sigma_{11x}^* \sigma_{12x}^* \}$ $\{x_{2x}^*\}\$ and $\{\sigma_y^*\} = \{\sigma_{11y}^*\,\,\sigma_{12y}^*\,\,\sigma_{21y}^*\,\,\sigma_{22y}^*\}$, and correla- $_{11}^{*}$, ρ_{12}^{*} , ρ_{21}^{*} , ρ_{22}^{*}), respectively. Usually, the LOD values and the average LODs obtained in direct apply some versions of single-marker or interval analysis. Monte Carlo simulations. Reduced test power, biased parameter estimates, and detection of ghost factors may result from this simplifi-TWO-TRAIT ANALYSIS cation, as demonstrated by many simulation studies.
The foregoing analytical treatment of this problem de-As was shown in our previous simulation study (Korol veloped for a single-trait analysis is expanded now to

terized for traits *x* and *y* and a set of marker loci from looks like the chromosome in question. For an arbitrary marker, we take into account the same five situations (a–e) as those considered above for the single-trait analysis. *Then the true expected densities of the bivariate trait* distribution in the alternative marker groups for an arbitrary scanning marker will be $N(\cdot)$ is a bivariate normal density, $\{\mu_x\} = \{\mu_{11x},$

$$
h_{MM}(x, y) = h_1 = \alpha_1 f_{11}(x, y) + \alpha_2 f_{12}(x, y)
$$

+ $\beta_2 f_{21}(x, y) + \beta_1 f_{22}(x, y),$

$$
h_{mm}(x, y) = h_2 = \beta_1 f_{11}(x, y) + \beta_2 f_{12}(x, y)
$$

+ $\alpha_2 f_{21}(x, y) + \alpha_1 f_{22}(x, y),$

where α_i and β_i are as defined in (1). To proceed with the analysis we need to make the following note.

Consider an arbitrary bivariate distribution with finite central moments (up to the fourth). Then the maximum of log-likelihood per individual for the Gaussian model will converge in probability to the maximum of the regression of the log-likelihood function per individ- ⁵ *^V*²*xy*(*˜r*, {s˜ *^x*}, {s˜*y*}, {rxy}) ⁵ *^V*²*xy*(·), ual. Assume that the trial marker is exactly at the same position as our putative QTL and the trait distributions where the components of vector $(\{\tilde{\mu}_x\},\{\tilde{\sigma}_y\},\{\tilde{\rho}_x\},\{\tilde{\rho}_x\})$ in the alternative groups *MM* and *mm* are bivariate nor-
mals. The regression of the log-likelihood will take the us to obtain the following results. mals. The regression of the log-likelihood will take the form Assume independent variance effects of the linked

$$
U_{1xy}(\theta) = \mathbf{E} \log(\prod_{j=1}^{n_1} N(x_{1j}, \mu_{1j}, \mu_{1j}, \sigma_{1j}, \sigma_{1j}, \rho_1) \times \prod_{\substack{\sigma_{21}^* \\ \sigma_{21}^* \\ j=n_1+1}}^{n_1} N(x_{2j}, \mu_{2j}, \mu_{2j}, \sigma_{2j}, \sigma_{2j}, \rho_2)), \qquad \text{tion} \qquad \text{group}
$$

$$
\max_{\theta} U_{1xy}(\theta) = -n(1 + \frac{1}{2}\log(4\pi^2 \tilde{\sigma}_{1x} \tilde{\sigma}_{2x} \tilde{\sigma}_{1y} \tilde{\sigma}_{2y} \times \sqrt{(1 - \tilde{\rho}_1^2)(1 - \tilde{\rho}_2^2)}) \n= V_{1xy}(\tilde{\sigma}_{1x} \tilde{\sigma}_{2x} \tilde{\sigma}_{1y} \tilde{\sigma}_{2y} \tilde{\rho}_{1z} \tilde{\rho}_{2}),
$$

Consider now a process of scanning with a pair of $d_{1y}/d_{2y} = c$, then the maximum of $V_{2xy}(\cdot)$ over possible markers along the chromosome. Because of the forego-
locations of the trial pairs of markers in a sufficiently markers along the chromosome. Because of the forego-clocations of the trial pairs of markers in a sufficiently
ing assumptions, the true expected densities of the trait small neighborhood of the QTL is attained exactly whe ing assumptions, the true expected densities of the trait small neighborhood of the QTL is attained exactly when
distribution in four alternative marker groups for an strucker locations coincide with those of the QTL (the distribution in four alternative marker groups for an these locations coincide with those of the QTL (the arbitrary pair of trial markers can be written as the proof is available from the authors upon request).

$$
h_{11}(x, y) = \alpha_1 f_{11}(x, y) + \alpha_2 f_{12}(x, y) + \beta_2 f_{21}(x, y) + \beta_1 f_{22}(x, y) = H(\alpha_1, \alpha_2, \beta_1, \beta_2), h_{12}(x, y) = H(\tau_1, \tau_2, \delta_1, \delta_2), \quad h_{21}(x, y) = H(\delta_1, \delta_2, \tau_1, \tau_2), h_{22}(x, y) = H(\beta_1, \beta_2, \alpha_1, \alpha_2),
$$

2) are as defined in (3). For any pair of markers, one the effect of the second QTL (Q_2/q_2) and on the residual can assume that they coincide with (or are closely linked correlation (ρ) between the quantitative traits. We are to) the corresponding QTL. If so, the parameter values interested here in testing H_2 *vs.* H_1 , assuming additive characterizing these QTL are easily derived by em- effects of the two QTL. Two situations are considered: ploying maximization of the regression of the log-likeli- in the first, each trait depends on only one of the linked

Consider a random sample of individuals each charac- hood function. The regression of the log-likelihood now

$$
U_{2xy}(\{\mu_x\}, \{\sigma_x\}, \{\mu_y\}, \{\sigma_y\}, \{\rho_{xy}\}) = U_{2xy}(\cdot)
$$

= **E** log $(\prod_{j,k=1}^{2} \prod_{jjk=1}^{njk} N(x_{ijk}, \mu_{jkx}, \mu_{jky}, \sigma_{jkx}, \sigma_{jky}, \rho_{jk}))$

 $\mu_{12x}, \mu_{21x}, \mu_{22x}, \{\mu_y\} = (\mu_{11y}, \mu_{12y}, \mu_{21y}, \mu_{22y}), \{\sigma_x\} = (\sigma_{11x},$ $\sigma_{12x}, \sigma_{21x}, \sigma_{22x}), \{\sigma_y\} = (\sigma_{11y}, \sigma_{12y}, \sigma_{21y}, \sigma_{22y}), \{\rho_{xy}\} = (\rho_{11}, \rho_{12},$ ρ_{21} , ρ_{22}), and *n_{kl}* are the frequencies of the four marker classes for the current pairs of markers, *n*₁₁ + *n*₁₂ + *n*₂₁ + $n_{22} = n$. Then,

+
$$
\alpha_2 f_{21}(x, y) + \alpha_1 f_{22}(x, y)
$$
,
\nas defined in (1). To proceed with
\n1 to make the following note.
\n- $\alpha_1 f_{22}(x, y) = U_{2xy}(\{\tilde{\mu}_x\}, {\tilde{\sigma}_y\}, {\tilde{\mu}_y\}, {\tilde{\sigma}_y\}, {\tilde{\rho}_x\})$
\n $= -n(1 + \log 2\pi)$
\n- $\nu_2((1 - \tilde{r})\log (\tilde{\sigma}_{11x}\tilde{\sigma}_{11y}\tilde{\sigma}_{22x}\tilde{\sigma}_{22y})$
\n- $\nu_2((1 - \tilde{r})\log (\tilde{\sigma}_{11x}\tilde{\sigma}_{11y}\tilde{\sigma}_{22x}\tilde{\sigma}_{22y})$
\n- $\sqrt{(1 - \tilde{\rho}_{11}^2)(1 - \tilde{\rho}_{22}^2)})$
\n- $\sqrt{1 - \tilde{\rho}_{11}^2 + \tilde{\rho}_{22}^2}$
\n- $\sqrt{1 - \tilde{\rho}_{11}^2 + \tilde{\rho}_{22}^2}$
\n- $\sqrt{1 - \tilde{\rho}_{12}^2 + \tilde{\rho}_{22}^2}$
\n- $\sqrt{1 - \tilde{\rho}_{12}^2 + \tilde{\rho}_{22}^2}$
\n- $\sqrt{1 - \tilde{\rho}_{12}^2 + \tilde{\rho}_{22}^2}$
\n- $\tilde{\rho}_{21}^2$
\n- $\tilde{\rho}_{22}^2$
\n- $\tilde{\rho}_{21}^2$
\n- $\tilde{\$

QTL for each of the traits (*i.e.*, $\sigma_{11x}^* = \sigma_x^*$, $\sigma_{12x}^* =$ α_{x}^{*} , $\sigma_{z1x}^{*} = b_{x}\sigma_{x}^{*}$, $\sigma_{z2x}^{*} = a_{x}b_{x}\sigma_{x}^{*}$, $\sigma_{11y}^{*} = \sigma_{y}^{*}$, $\sigma_{12y}^{*} = a_{y}\sigma_{y}^{*}$, $\sigma_{21y}^* = b_j \sigma_y^*$, $\sigma_{22y}^* = a_j b_j \sigma_{y^*}^*$ and equal residual correlation between the target traits in all of the four QTL *groups (i.e.*, $\rho_{ij}^* = \rho^*$; *i, j* = 1.2). Then, if one of the where $\theta = (\mu_{1x} \mu_{2x} \mu_{1y} \mu_{2y} \sigma_{1x} \sigma_{2x} \sigma_{1y} \sigma_{2y} \rho_{1}, \rho_{2})$. Then, traits, x, depends additively on both linked QTL (Q_1 / Q_2) , whereas the correlated trait y is independu dent of the considered QTL, then the global maximum of $V_{2x}(.)$ over all possible locations of the trial pair of markers in the chromosome is attained exactly when $V_{1,y}(\tilde{\sigma}_{1x}, \tilde{\sigma}_{2x}, \tilde{\sigma}_{1y}, \tilde{\sigma}_{2y}, \tilde{\rho}_1, \tilde{\rho}_2)$, these locations coincide with those of the QTL. Like-
Where $\tilde{\sigma}_{1y}$ ($u = x$ or y) and $\tilde{\rho}_i$ ($i = 1, 2$) are some functions only one of the traits or where $\tilde{\sigma}_{i\mu}$ ($u = x$ or y) and $\tilde{\rho}_i$ ($i = 1,2$) are some functions only one of the traits or (ii) has a pleiotropic effect on both traits x and v but in such a manner that $d_v/d_v =$ the main parameters.
Consider now a process of scanning with a pair of $d_x/d_x = c$, then the maximum of $V_{2x}(t)$ over possible proof is available from the authors upon request).

 $h_{11}(x, y) = \alpha_1 f_{11}(x, y) + \alpha_2 f_{12}(x, y) + \beta_2 f_{21}(x, y)$
 $h_{12}(x, y) = H(\tau_1, \tau_2, \delta_1, \delta_2),$
 $h_{22}(x, y) = H(\beta_1, \beta_2, \alpha_1, \alpha_2),$
 $h_{22}(x, y) = H(\beta_1, \beta_2, \alpha_1, \alpha_2),$
 $h_{23}(x, y) = H(\beta_1, \beta_2, \alpha_1, \alpha_2),$
 $h_{31}(x, y) = H(\beta_1, \beta_2, \alpha_1$ where the mixture parameters α_i , β_i , τ_i , and δ_i (*i* = 1, (say Q_i/q_i) and consider how max ELOD depends on

QTL (Figure 4a), whereas in the second case both QTL affect the first trait and one of the QTL affects the 5b) demonstrates a situation in which the QTL interact second trait (Figure 4b). One can conclude that the epistatically for both traits. Clearly, the provided examples second trait (Figure 4b). One can conclude that the epistatically for both traits. Clearly, the provided examples detection power increases with the residual correlation are not more than illustrations of the possibilities detection power increases with the residual correlation between the analyzed traits. An additional conclusion proposed analytical tool. Each of the questions discussed
is that the power increases with the effect of Q_2/q_2 up in these illustrations can be dealt with in necess is that the power increases with the effect of Q_2/q_2 up to some "saturation" point. In the first situation the saturation is reached when the effect of Q_2/q_2 becomes conclusion equal to that of Q_1/q_1 (see Figure 4a). The only difference in the second situation is that the saturation point Resolution power of mapping analysis of linked QTL depends on ρ : the larger abs(ρ) the earlier (at lower remains a difficult problem, which was previously adeffects of Q_2/q_2) the saturation (see Figure 4b). For dressed mainly in terms of Monte Carlo simulations. the second situation, let us consider the complication This has restricted the possibilities of detailed evaluation caused by epistasis. Namely, we allow for epistatic inter- and comparison of different mapping situations and action between the QTL with respect to the trait *x.* As experimental designs. The proposed analytical method in the foregoing example on single-trait analysis, it is of evaluating the expected LOD generalizes for the case interesting here to evaluate how epistasis affects the of two linked QTL the corresponding estimates derived expected detection power. Figure 5a demonstrates that by Lander and Botstein (1989), Mackinnon and Welepistasis may be helpful in discriminating between H₂ ler (1995), and Mackinnon *et al.* (1996) (referred to (two linked QTLs) and H_1 (only a single QTL in the as "deterministic sampling"). Our model allows us to chromosome). This effect is manifested for both positive analyze situations with variance effect and epistatic interand negative residual correlations, but the sign of ρ is action between the putative QTL. We developed here important in determining the details of the behavior of also a two-locus analogue of our previous analytical premax ELOD as a function of epistasis and the effect of *Q*2/ dictor (see Korol *et al.* 1995) of the expected LOD for *q*₂ on trait *y* (not shown). The second example (Figure two-trait mapping analysis. And again, many compli-

Figure 5.—The behavior of the maximum ELOD $(H_2 \text{ vs.})$ Figure 4.—The behavior of the maximum ELOD (H₂ *vs.* H₁) in the two-trait model with two linked QTL as a function H₁) in the two-trait model with two linked QTL as a function of the parameters. In (a), the first QTL H₁) in the two-trait model with two linked QTL as a function of the parameters. In (a), the first QTL affects only the trait of the residual correlation and the effect of one of the QTL $x (d_{1x} = 0.35)$, whereas the sec on one of the two traits (*y*). (a) A situation with the first $d_{2x} = 0.15$, and d_{2y} varies, as shown in the figure. The residual QTL affecting the trait x ($d_{1x} = 0.25$), whereas the second correlation was $\rho = -0$ x ($d_{1x} = 0.35$), whereas the second QTL affects both traits
 $(d_{2x} = 0.25; d_{2y}$ varies, as shown in the figure). (a and b) $r =$
 25% , $\sigma = 0.5$, and $n = 250$.

The residual correlation was $\rho = -0.5$. (a and b) $r =$ The residual correlation was $\rho = -0.5$. (a and b) $r = 25\%$, $\sigma = 0.5$, and $n = 250$.

cated questions can be addressed, like dependence of liang, C., and Z.-B. Zeng, 1995 Multiple trait analysis and genetic
the QTL detection power on residual correlation be-
tween the traits, accounting of epistatic interac tween the QTL for one or both traits, and the influence
of variance effect for one or both traits on ELOD. Allaction and the influence
though this method gives only an asymptotic estimation
though this method gives only an though this method gives only an asymptotic estimation ous analysis of a set of c and c and c marker c marker c mark of ELOD, it allows one to get an approximate assessment
of a broad spectrum of specific mapping situations.
Clearly, any asymptotic effect found by the proposed
Clearly, any asymptotic effect found by the proposed
Korol, A Clearly, any asymptotic effect found by the proposed Korol, A. B., Y. I. Ronin and V. M. Kirzhner, 1995 Interval mapping

tool can (and should) be shocked by Monte Carlo simu of quantitative trait loci employing correlated of quantitative trait loci employees. The checked by Monte Carlo simu-
lations for given sample sizes. Our comparisons made Genetics **140:** 1137–1147. for a series of situations indeed show a good correspon-
dance hattuation and indeed show a good correspon-
evidence from barley. Heredity **80**: 273-284. dence between the predicted max ELODs and LOD
values averaged over Monte Carlo runs. An important underlying quantitative traits using RFLP linkage maps. Genetics values averaged over Monte Carlo runs. An important underlying quantitative traits underlying quantitative maps. The considered class. point is that our results prove (for the considered class **121:** 185–199.

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