# **The Signature of Positive Selection at Randomly Chosen Loci**

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## ABSTRACT

In Drosophila and humans, there are accumulating examples of loci with a significant excess of highfrequency-derived alleles or high levels of linkage disequilibrium, relative to a neutral model of a randommating population of constant size. These are features expected after a recent selective sweep. Their prevalence suggests that positive directional selection may be widespread in both species. However, as I show here, these features do not persist long after the sweep ends: The high-frequency alleles drift to fixation and no longer contribute to polymorphism, while linkage disequilibrium is broken down by recombination. As a result, loci chosen without independent evidence of recent selection are not expected to exhibit either of these features, even if they have been affected by numerous sweeps in their genealogical history. How then can we explain the patterns in the data? One possibility is population structure, with unequal sampling from different subpopulations. Alternatively, positive selection may not operate as is commonly modeled. In particular, the rate of fixation of advantageous mutations may have increased in the recent past.

CONSIDERABLE debate has focused on what pro-<br>
portion of genetic changes is favored by natural<br>
include FY (HAMBLIN *et al.* 2002), MAO-A (GILAD *et al.*<br>
and served a considerable to the served of all the served and serve and Wu 2001). Answers to these questions will help to Przeworski, unpublished results), psGBA (data from

lar genomic region, population geneticists usually se- and CROWELL 2000; M. PRZEWORSKI, unpublished requence a number of individuals at a locus and test sults), and 3 out of 19 intergenic regions (FRISSE *et al.* whether the pattern of polymorphism seen in the sam- 2001; L. Frisse and A. Di Rienzo, personal communicaple is unexpected under the standard neutral model of tion). Considered together with multilocus evidence a random-mating population of constant size. Unfortu- (*e.g.*, AQUADRO *et al.* 1994; ANDOLFATTO and PRZEWORnately, a departure from null model expectations can ski 2001; NACHMAN 2001) and an accumulating number be due to one of many causes, so it is hard to establish of individual loci that show evidence of positive selection that adaptation is responsible. In particular, an excess (reviewed in ANDOLFATTO 2001), these frequency specof rare variants may reflect a selected substitution at a trum results suggest that a large fraction of genetic closely linked site, but it may also be caused by popula- changes may be favored (Fay and Wu 2001). tion expansion or purifying selection, just to list a couple In addition, patterns of linkage disequilibrium (LD) of alternatives. For this reason, an ideal "test of neutral- depart from the expectations of the standard neutral ity" would not only have high power to detect positive model in these species. There appears to be a genomeselection, but would also focus on an aspect of the data wide excess of intralocus linkage disequilibrium in *D.* unlikely to be affected by demography or other factors. *melanogaster* and non-African populations of *D. simulans* Such a test statistic (*H*) was recently proposed by FAY (ANDOLFATTO and PRZEWORSKI 2000; J. D. WALL, P. and Wu (2000), to detect a single, recent episode of ANDOLFATTO and M. PRZEWORSKI, unpublished results)

reported for samples from *Acp26Aa* (FAy and Wu 2000), tances in humans (*e.g.*, RIEDER *et al.* 1999; TAILLON-<br>achaete (FAY and WU 2000) Attacins A and B (LAZZARO MILLER *et al.* 2000; GILAD *et al.* 2001; reviewed in PRI *achaete* (Fay and Wu 2000), Attacins A and B (Lazzaro Miller *et al.* 2000; Gilad *et al.* 2001; reviewed in Pritchand CLARK 2001), and *desat2* (TAKAHASHI *et al.* 2001) ard *ARD* and *PRZEWORSKI* 2001). It is often argued that these<br>in *Drosobbila melanogaster* and the *janA-ocn* region in *D* patterns reflect the action of positive patterns reflect the action of positive selection at or in *Drosophila melanogaster* and the *janA*-*ocn* region in *D.*

selection, as well as what types of substitutions are most  $2001$ , and several noncoding loci: a subset of olfactory likely to have been selected (ANDOLFATTO 2001; FAY receptor pseudogenes (data from GILAD *et al.* 2000; M. elucidate the genetic basis of adaptation. Martinez-Arias *et al.* 2001; M. Przeworski, unpub-To infer that positive selection has acted on a particu-<br>lished results), the intron DMD7 (data from Nachman

positive selection (OTTO 2000). and there are numerous examples of pairwise linkage<br>Since its introduction, significant H values have been disequilibrium extending over unexpectedly large dis-Since its introduction, significant *H* values have been disequilibrium extending over unexpectedly large dis-<br>exported for samples from  $Acb26Aa$  (Fay and W<sub>U</sub> 2000). Tances in humans (*e.g.*, RIEDER *et al.* 1999; TAILLON near the sampled region (*e.g.*, TAILLON-MILLER *et al.* 2000; Gilad *et al.* 2001; Parsch *et al.* 2001; other refer- $^{1}$ Address for correspondence: Max Planck Institute for Evolutionary<br> $^{1}$ Address for correspondence: Max Planck Institute for Evolutionary

If so, patterns of polymorphism in many regions will

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have been shaped by repeated episodes of positive selec- constant and chosen so that there is a small probability tion. However, as I show here, the *H* test has very low that two or more would occur simultaneously [using power to detect the effects of positive selection on a  $1 -$  Equation 6 in BRAVERMAN *et al.* (1995)—this is a randomly chosen locus. Similarly, the effect of selection slight overestimate as it ignores the effects of interferon LD is short-lived, so even neutral loci affected by ence between selected loci]. When a sweep occurs, the multiple adaptive substitutions at linked sites are un-<br>likely to show unusually high levels of allelic association. One side of the neutral locus. Selection is additive, with likely to show unusually high levels of allelic association.

*H* statistic presented in Fay and Wu (2000) is the differ- according to the infinite-sites model. ence between two estimates of the population mutation This selective sweep model is implemented as a sucrate  $\theta = 4N\mu$ , where *N* is the diploid effective popula- cession of neutral and selective phases (when there are tion size of the species and  $\mu$  the mutation rate per two alleles at a selected site). The algorithm for the generation. The two estimates are the average number neutral phase is the standard coalescent with recombiof pairwise differences in the sample,  $\pi$  (Tajima 1983) nation (*cf.* Hudson 1993). The selective sweep phase is and  $\theta_H = \sum_{i=1}^{n-1} p_i^2 / {n \choose 2}$ , where *n* is the sample size and  $p_i$  implemented as in BRAVERMAN *et al.* (1995), with the the frequency of the derived (*i.e.*, nonancestral) allele addition of intralocus recombination. During a sweep, at segregating site *i* (Fu 1995). *H* is negative when there are effectively two subpopulations at the neutral is an excess of high-frequency-derived alleles relative to locus: lineages carrying the favored allele at the is an excess of high-frequency-derived alleles relative to

(1989a): Tajima's *D* is the (approximately) normalized subpopulation can coalesce, (2) a lineage can recom-<br>difference between  $\pi$  and  $\theta_w$ , an estimate of  $\theta$  based on bine onto the same selective background, and (3) the number of segregating sites in the sample. In con-<br>trast to  $H$ ,  $D$  does not use information about ancestral and a ground. Patterns of polymorphism at the neutral locus and derived states. Negative *D* values reflect a relative are affected by events of type  $(2)$  only if the recombina-<br>averses of the alleles in a folded frequency spectrum tion breakpoint is within the neutral locus. excess of rare alleles in a folded frequency spectrum.<br>Here, both *H* and *D* are used as one-tailed tests of neu-<br>During the sweep, time changes in small increments,

of *H* to detect a model of recurrent "selective sweeps" (1 *<sup>x</sup>*(*t*))*t*, (*cf.* Kaplan *et al.* 1989; Stephan *et al.* 1992; Braverman *et al.* 1995). The model assumes a random-mating population of constant size. My implementation of this model where  $x(t)$  is the frequency of the favored allele at time follows the description in Braverman *et al.* (1995), ex- *t*, *i* is the number of lineages carrying the favored allele, cept for two features. First, I use a fixed value of the and  $j$  is the number of lineages carrying the unfavored population mutation rate, rather than a fixed number allele (BRAVERMAN *et al.* 1995), population mutation rate, rather than a fixed number of segregating sites (HUDSON 1993; WALL and HUDSON 2001). Second, I allow for recombination within the neutral locus, both during neutral and selective phases and

(see below).<br>
In the model, a neutral locus is affected by selective  $\Pr{\text{event}(3)} = [i(\rho + c)(1 - x(t)) + j(\rho + c)x(t)]\Delta t$ . sweeps that occur at some random genetic distance c,<br>where c is uniform on  $(0, M)$  and M is the maximum<br>eled deterministically, from frequency  $\varepsilon$  to  $1 - \varepsilon$ , using levels. (What is meant by genetic distance is the popula-<br>tion recombination rate between the neutral and se-<br>all t. A path x can also be found by simulating the rise tion recombination rate between the neutral and se-<br>lected locus.) M is on the order of  $4Ns$  (KAPLAN *et al.* of a selected allele forward in time, thereby allowing 1989); in this implementation,  $M = 4Ns$  (*s* is the selective for a fully stochastic treatment of the selective sweep. coefficient of the favored allele). In simulations of a Modeling the rise in frequency by binomial sampling single selective sweep, the value of  $c$  is specified, as is or a diffusion approximation does not change the qualithe time since the fixation of the beneficial allele. In tative results (results not shown). the model of repeated sweeps, the rate of sweeps is Call the sum of the probabilities of all possible events

fitnesses  $1, 1 + s, 1 + 2s$  for the three genotypes. Crossing over occurs within the neutral locus at rate  $\rho$ , where METHODS  $\rho = 4Nr$  (*r* is the crossover rate per generation). There is no gene conversion, and I assume a constant rate of **Frequency spectrum-based "tests of neutrality":** The crossing over per base pair. The neutral locus evolves

the standard neutral model.<br>This statistic is similar to one introduced by TAIIMA types of events can occur: (1) Two lineages in the same This statistic is similar to one introduced by Tajima types of events can occur: (1) Two lineages in the same<br>989a): Tajima's *D* is the (approximately) normalized subpopulation can coalesce, (2) a lineage can recom-

*trality.*  $\Delta t$ . Within  $\Delta t$ , the probabilities of the events of interest simulations of positive selection: I estimate the power are given by

$$
\Pr\{\text{event}(1)\} = \left[\frac{\binom{i}{2}}{x(t)} + \frac{\binom{i}{2}}{(1 - x(t))}\right]\Delta t
$$

$$
Pr\{\text{event}(2)\} = [\hat{ip}x(t) + j\rho(1 - x(t))] \Delta t
$$

 $\} = [i(\rho + c)(1 - x(t)) + j(\rho +$ 

distance at which a single sweep has an effect on diversity Equation 3a in STEPHAN *et al.* (1992). I set  $\varepsilon = 1/2N$ <br>levels. (What is meant by genetic distance is the popula-<br>(as do FAY and W<sub>U</sub> 2000) so  $x(t)$  is given e of a selected allele forward in time, thereby allowing

within a time interval  $S_i$ ;  $(1 - S_i)$  is approximately the to have roughly the right nominal rejection probability event, I solve  $\Pi^{y}(1 - S_i) < U$  for *y*, where *U* is a uni- (WALL and HUDSON 2001). form random variable on (0, 1) and the product is taken The *H* test relies on identification of the ancestral over successive time intervals. Which event occurs at allele. In practice, this is done with one or more outtime *y* is chosen randomly with probability Pr{event  $|t =$  groups, and the inference may be incorrect if there are *y* / *S<sub>v</sub>*.

If the event is of type 3, then with probability  $\rho/(\rho +$ *c*) the crossover event occurs within the neutral locus the extent of mutation rate variability across sites. Fay and with probability  $c/(\rho + c)$  between the selected and neutral locus. When a crossing-over event occurs within ity of an incorrect inference by assuming a constant the neutral locus, a breakpoint *b* is chosen uniformly mutation rate and the use of one outgroup, while I on [0, *L*] where *L* is the length of the neutral locus. assume a known ancestral state.

in C and based on coalescent programs kindly provided deficiency in the number of distinct haplotypes for the  $\sim$ rhudson1/). The program was error checked by com- *al.* 2001; WALL 2001; other references in ANDOLFATTO paring the output to the results in Figure 3 of Fay and 2001). To examine this aspect of the data, I consider

in Fay and Wu (2000). (For ease of comparison, note that the results in Fay and Wu are actually for a selective the sample and *S* the number of segregating sites). With sweep model with fitnesses  $1, 1 + 0.5s, 1 +$ three genotypes.) First, the 5% significance levels for *H* is 1. Under the standard neutral model, lower levels of (or *D*) are determined by simulations of the standard neutral model with no recombination. I make the latter total of  $10<sup>4</sup>$  simulations were run for each set of parameassumption for ease of comparison with Fay and Wu ters. In simulations used to examine levels of LD, cross- (2000) and because researchers have used critical values ing over occurs within the neutral locus at rate  $\rho > 0$ . of *H* established for no recombination. The neutral model is implemented for a fixed number of segregating <br>sites; *i.e.*, I generate genealogies and then place a fixed RESULTS number of segregating sites on the tree. Second, data **Selective sweeps with recombination:** Most of the thesets are generated under the alternative model for a oretical attention paid to models of positive selection given  $\theta$  value (with or without recombination). If the has focused on the "selective sweep" or "hitch hiking" value of *H* for a data set is more extreme than the model (MAYNARD SMITH and HAIGH 1974). This model significance level established for that number of segre-<br>describes the rapid increase in frequency (and ultimate gating sites under the null model, the null model is fixation in the population) of an initially rare and rejected. strongly favored allele. The effects of a selective sweep

would do in practice, when they come across a region be understood as follows: Imagine first that there is no with low diversity. Since the population mutation rate recombination and that we draw a sample of chromois unknown, one might ask to what extent the locus is somes from the present. They all bear a particular faconsistent with the neutral model and a low mutation vored mutation, A. This allele increased in frequency rate by testing if *H* is more extreme than expected for very rapidly, such that, not very long ago, there were the *observed number of segregating sites.* If no segregating only a few copies in the population. As the number of sites were found, no test would be performed. When copies of the favored allele decreases (going backward estimating power, I exclude all runs in which there are in time), coalescences between lineages ancestral to our no segregating sites. [For sake of comparison, note that sample happen faster and faster. This means that mem-Fay and Wu (2000) do not.] This procedure turns out bers of a sample from this region are much more closely

probability that no event occurs, when the probabilities for a wide range of  $\theta$  values (results not shown). The of all events are small. To calculate the time to the next same is true for *D*, as well as other tests of neutrality

> *mutations at the same site on the outgroup lineage(s).*  How likely this is depends on the mutation rate and on and Wu (2000) introduce a correction for the probabil-

Assume, as an illustration, that the selected locus is to **Linkage disequilibrium:** There are many possible the left to the neutral locus and that the lineage carries summaries of LD and none is an obvious choice. Here, the favored allele. Segments in the neutral locus right I consider two measures of linkage disequilibrium. The of *b* would then "migrate" to the subpopulation of the first is  $r^2$  (*cf.* WEIR 1996), a commonly used summary unfavored background. The number of lineages in both of the extent of allelic association between a pair of subpopulations has to be updated accordingly for those with sites. I plot the decay of  $r^2$  with distance for all polymorsegments. Other cases are treated analogously.  $\qquad \qquad$  phisms with a frequency of the minor allele  $\geq 0.1$ . A The computer code for these simulations is written relative excess of LD is sometimes characterized as a by R. Hudson (available at http://home.uchicago.edu/ observed number of segregating sites (*e.g.*, Parsch *et* Wu (2000; for which  $\rho = 0$ ). a second summary of LD: the number of haplotypes **Power tests:** The *H* and *D* tests are implemented as normalized by the number of segregating sites,  $n_{\text{Hans}}/$  $(5 + 1)$  ( $n_{\text{Haps}}$  is the number of distinct haplotypes in  $+$  *s* for the no recombination, the maximum value of  $n_{\text{Haps}}/(S+1)$ recombination result in a smaller  $E(n_{\text{Hans}}/(S+1))$ . A

This procedure is meant to mimic what researchers on the frequency spectrum of linked neutral sites can



FIGURE 1.—One possible genealogical tree for a sample of six at a neutral site linked to a selected site. The frequency of the favored allele, A, is illustrated on the graph to the left, with time on the *x*-axis. As the frequency of the favored allele decreases, the rate of coalescence increases. However, if one of the neutral lineages (shown as long dashes) recombines onto a nonfavored background (going backward in time), it may have to wait (at least) until after the original mutation from A to a (represented by the gray star), to coalesce with other lineages. Any mutation on the dotted branch will be at high frequency in the sample.

The genealogy is close to star-shaped, so, as in the case frequency-derived alleles. of population growth (Tajima 1989b), we expect an *H* **has low power to detect old sweeps:** On the basis

advantageous allele A, but (going backward in time) Instead, sweeps might be thought of as occurring at recombines onto a chromosome with the unfavored random locations and times. In this case, the power of *H* allele, a. For that lineage to coalesce with the other is much reduced. First, the power of  $H$ ,  $P(H)$ , decreases lineages still associated with A, one of two things must rapidly with the time since the fixation of the favored happen: Either it must recombine back onto an A back-<br>allele, as the high-frequency variants fix in the populaground, or we have to wait until after the original muta- tion and no longer contribute to polymorphism (Kim tion from A to a (represented by a star in Figure 1). If and STEPHAN 2000). For example, in Figure 2, if  $N =$ the latter, two lineages will be present at the beginning  $10<sup>6</sup>$ , the power is roughly equal to the nominal rejection of the sweep, as in Figure 1; their mean time to coales- probability after  $5 \times 10^5$  generations or one-eighth of cence is given by the neutral expectation, 2*N*. At the the mean time to coalescence under neutrality,  $4N(t=$ neutral site, we will obtain an unbalanced tree that looks 0.125 in Figure 2). For *D. melanogaster*, assuming 10 like Figure 1 (note that this drawing is not to scale). Any generations a year (and if  $N = 10^6$ ), this corresponds mutation on the dotted line will be at high frequency in to  $5 \times 10^4$  years. For some time after the sweep, the our sample. Thus, in the presence of recombination, power is actually  $\leq 0.05$  (see also KIM and STEPHAN selective sweeps will produce not only rare variants, but 2001): Of the variation that preexisted the sweep event, also high-frequency ones (in practice, high- and low- all the high-frequency variants have fixed (at least in frequency variants can be distinguished by using out- the sample) so that any remaining alleles are at lower groups to infer which allele is ancestral). While popula- frequency; those that arose after the sweep are young tion growth and purifying selection also predict an and therefore also at low frequency. As a result, there

related than they would be at an unlinked neutral site. excess of rare alleles, they do not predict excess high-

excess of rare variants in our sample relative to the of these insights, Fay and Wu (2000) constructed a test, standard neutral model. *H*, which focuses on the number of high-frequency-With recombination, selective sweeps can no longer derived alleles (see METHODS). They demonstrated that be treated as population size reductions (Barton 1998). the power of *H* to detect a sweep that ended at time As we go back in time, the frequency of the favored  $t = 0$  can be high. Thus, if we consider a "candidate" allele decreases, but the frequency of the unfavored locus" where there is independent evidence for the acallele increases. One way to think of this is as a subdi- tion of recent positive selection (*e.g.*, Takahashi *et al.*) vided population model, where the two populations are 2001), we can be fairly confident that a significant *H* changing size over time (Barton 1998). Consider the test is indicative of positive selection. However, this genealogy of a neutral site linked to the selected site. model is unlikely to describe the situation where re-Suppose that a lineage is currently associated with the searchers apply the *H* test to a randomly chosen locus.



simulations (see METHODS). Black lines are for an effective<br>population size  $N = 10^6$  and a selection coefficient  $s = 0.005$  if the neutral locus is very close to the sweep, or too far are more as in Figure 3 of Fay and Wu 2000) and gray lines are for away,  $P(H)$  is substantially reduced (Figure 3 in Fay and  $N = 10^4$  and  $s = 0.05$ . The sample size is 50, the population Wu 2000; results not shown).  $N = 10<sup>4</sup>$  and  $s = 0.05$ . The sample size is 50, the population mutation rate  $\theta = 5$ , and the genetic distance to the selected The power of *H* depends on *s* and *c*, not just on their locus, *c*, is chosen such that  $c/s = 0.01$ . There is no recombination Keeping *c*/s constant does

under the null model (for a given number of segregating sites).

period of time since the sweep than does *H.* These only 10% while *P*(*D*) is 58%. For the same *c*/*s* value, if results suggest that *D* might be a better test for detecting  $s = 0.05$ ,  $P(H)$  is 51% and  $P(D)$  is 62% (Figure 2).

(with  $N = 10^6$ , with other parameters as in Figure 2)

while  $P(H | \theta = 5)$  is 69%. The power of *H* also increases with larger sample size (results not shown).

Of fundamental importance in determining *P*(*H*) is the number of lineages that recombine on to the unfavored background during the sweep. As can be seen in Figure 1, for the ancestral genealogy to have long internal branches requires at least one recombination event between selected classes. How likely this is depends on the strength of selection and on the recombination rate between the selected and neutral loci (*c*). If *c* is too small, there will be no recombination events, and all lineages will coalesce during the sweep. If *c* is very large, FIGURE 2.—The power of *H* and *D* as a function of the time<br>since the fixation of the favored allele, as estimated from  $10^4$ <br>tral locus will not reflect the effects of selection. Thus,<br>if the goutpel locus is used in g

locus, c, is chosen such that  $c/s = 0.01$ . There is no recombination within the neutral locus. The powers of H (triangles) and<br>
D (diamonds) are shown as solid and dashed lines, respectively. The two lines for  $P(D)$  are es linearly on *s.* In fact, for the same *c*/*s* value, stronger are fewer high-frequency-derived alleles than expected selection (and therefore larger *c* values) will result in higher  $P(H)$ . As an illustration, if  $N = 10<sup>4</sup>$ , as might be the case for humans (Li and SADLER 1991),  $c/s = 0.01$ , The *D* test retains substantial power for a much longer and  $s = 0.005$ , then immediately after a sweep,  $P(H)$  is

selective sweeps. When selection is recent, however, the **The power of** *H* **in practice:** Researchers have assessed use of *D* and *H* is not redundant. For example, if the the significance of the *H* test with critical values estabparameters are as in Figure 2 and  $t = 0$ , the proportion lished under the assumptions of a constant population of runs where *H* is significant but *D* is not is 19% (for size and no recombination. In reality, however, there is *D* but not *H*, it is 13%). recombination within the neutral locus. In the presence **The effect of other parameters on**  $P(H)$ **: With a larger of recombination, the use of critical values for the case**  value, there is a higher probability of having a muta- of no recombination is conservative; *i.e.*, the null model tion on the dotted branch in Figure 1 and therefore is rejected  $\leq 5\%$  of the time at the 5% level. This can more power to detect the effects of a sweep. For exam-<br>ple, immediately after a sweep,  $P(H|\theta = 10)$  is 79% for different values of  $\rho$ , the population recombination for different values of  $\rho$ , the population recombination rate for the neutral locus. Even though the  $H$  test is

			$t=0$	0.02	0.05	0.10	0.25	0.50	1.00	No sweep
$N = 10^6$ , $s = 0.005$	$\rho = 0$	P(H)	0.69	0.39	0.19	0.08	0.02	0.04	0.05	0.05
		P(D)	0.65	0.65	0.62	0.45	0.12	0.05	0.05	0.05
	$p = 20$	P(H)	0.76	0.48	0.23	0.08	0.01	${<}10^{-2}$	${<}10^{-2}$	0.01
		P(D)	0.69	0.64	0.59	0.44	0.14	0.03	0.01	0.01
$N = 10^4$ , $s = 0.05$	$\rho = 0$	P(H)	0.51	0.27	0.14	0.06	0.02	0.04	0.05	0.05
		P(D)	0.62	0.66	0.62	0.44	0.11	0.05	0.05	0.05
	$\rho = 5$	P(H)	0.57	0.32	0.16	0.07	0.01	$<$ 10 <sup>-2</sup>	0.02	0.03
		P(D)	0.63	0.65	0.59	0.45	0.14	0.04	0.03	0.03

**TABLE 1 The power of** *H* **and** *D* **as a function of the time since the sweep ended**

The time *t* since the fixation of the beneficial mutation is scaled in units of 4*N* generations, where *N* is the effective population size,  $\rho$  is the population recombination rate for the neutral locus, and  $s$  is the selection coefficient of the favored allele. The sample size is 50 and the population mutation rate at the neutral locus,  $\theta$ , is 5. A total of  $10^4$  simulations were run for each set of parameters.



mosomes. The population recombination rate for the neutral locus, *ρ*, is 20. On the *x*-axis is the expected number of selective sweeps per base pair per 4*N* generations, assuming a recombi-

tion, some recombination increases the power to detect linked sites.) As can be seen in Table 1, the increase and the rate of sweeps. in power is slight, and *P*(*H*) still decreases extremely **The effect of a single sweep on LD:** As shown above, quickly with *t*.

In humans, the violation of a second assumption will lead one to overestimate the power of *H* to detect a sweep. The human population size has increased dramatically in the recent past. The effect of population growth is to increase the rate of coalescences going backward in time. For the same average diversity levels, the tree in Figure 1 would therefore have shorter internal branches than it does under a constant-size model. This will reduce the number of high-frequency-derived alleles found at neutral sites linked to a selective sweep. Thus, the finding of numerous loci with extreme *H* values is even more surprising when this aspect of human demography is taken into account.

**The power to detect sweeps at a randomly chosen locus:** Results for the recurrent selective sweep model are shown in Figure 3. There is essentially no power to detect the effects of selection using *H* and the power FIGURE 3.—The power of *H* and *D* (one-tailed) to detect does not increase with the strength of selection or the repeated selective sweeps, as estimated from 10<sup>4</sup> simulations (see METHODS). The effective population size gest that, if  $N = 10^6$ ,  $s = 0.005$ , and the sample size is sweeps per base pair per 4*N* generations, assuming a recombi-<br>nation rate of  $5 \times 10^{-9}$ /bp/generation. Dashed lines are for<br>a population mutation rate  $\theta = 5$  and solid ones are for  $\theta =$ <br> $\Gamma$ 10. The two lines for  $P(H)$  are essentially superimposed.<br>
Tor these parameters,  $P(H) > 20\%$  for a distance be-<br>
tween  $0.00035 < c/s < 0.02$  (Figure 3 in FAY and WU) 2000). If sweeps occur at a distance chosen uniformly conservative in the presence of intralocus recombina-<br>tion some recombination increases the power to detect be within the relevant range. In addition, the beneficial a sweep at a linked site. (Obviously this is true only up allele will have fixed at some random time in the past, to a point: If there is a very high level of recombination,  $t \ge 0$ , and the power of *H* decreases with increasing *t*. the neutral locus will no longer reflect selection at In contrast to *H*, the power of *D* increases with both *s*



Figure 4.—The effect of selective sweeps on the expected decay of pairwise linkage disequilibrium. The effective population size is  $N = 10^6$ , the selection coefficient  $s = 0.01$ , the population mutation rate  $\theta$  = 40, and the sample size is 50. The population recombination rate for the neutral locus,  $\rho$ , is 20 (which corresponds to 1 kb for a recombination rate of  $5 \times 10^{-9}$ /bp/generation). The genetic distance to the sweep, *c*, is chosen so that *c*/  $s = 0.005$ . The time since the fixation of the favored allele, *t*, is scaled in units of 4*N* generations. A total of  $10<sup>4</sup>$  simulations were run for each value of *t.* Only segregating sites with a minor allele frequency  $\geq 0.1$ are included.



set. Only segregating sites with a minor allele frequency  $\geq 0.1$ are included. The effective population size  $N = 10^4$ , the selec-

data, levels of linkage disequilibrium. In both Drosoph- Nonetheless, the effect of the selective sweep has all but ila and humans, numerous loci appear to exhibit unex-<br>vanished by  $t = 0.1$ , unless selection is very strong (*e.g.*, pectedly high levels of LD. In Drosophila, this is usually  $Ns = 5 \times 10^3$ . Pairwise linkage disequilibrium exhibits quantified as a paucity of haplotypes (*e.g.*, PARSCH *et al.* a similar behavior to the number of haplotypes: For 2001; further references in ANDOLFATTO 2001) or a example, in Figure 4, a sweep that ended at  $t = 0.2$  has lower than expected estimate of the population recom- $\frac{1}{2}$  an undetectable effect on  $r^2$ . For these parameters, there bination rate,  $\rho$  (ANDOLFATTO and PRZEWORSKI 2000; is still a relative excess of LD by  $t = 0.1$ ; however, this WALL 2001). In particular, in *D. melanogaster* and *D.* would be hard to discern in any one data set, because  $r^2$  $simulas$ , it appears that one estimate of  $\rho$ ,  $C_{\text{had}}$  (Hudson varies greatly from one locus to another under neutrality 1987), is systematically lower than would be expected (PRITCHARD and PRZEWORSKI 2001). from independent estimates of the mutation and recom- One implication of these results is that selection

LD extends in many regions that is unusual  $(e.g.,\mathsf{RIEDER})$ *et al.* 1999; GILAD *et al.* 2001; reviewed in PRITCHARD and Przeworski  $2001$ ). For a couple of regions,  $\rho$  has also been shown to be lower than expected for European samples (PRITCHARD and PRZEWORSKI 2001). These patterns have not yet been explained.

As is illustrated in Figures 4 and 5, a recent sweep can substantially increase levels of LD. In Figure 4, I plot the expected decay of a summary of pairwise LD,  $r^2$ , for alleles with a minor allele frequency  $\geq 0.1$ . Parameters are chosen to be plausible for *D. melanogaster.* If the beneficial allele fixed at time  $t = 0$ , there is a much slower rate of decay with distance than under the standard neutral model. Note, however, that fewer alleles satisfy the frequency cutoff after a sweep, so long sequences may be required for this pattern to be apparent in actual data. Figure 5 presents scatterplots of  $r^2$  *vs.* distance for parameters germane to humans; as can be seen, a selected substitution at a linked site increases the number of distant pairs in significant LD.

The effect of a sweep on levels of LD dissipates quickly, depending on the summary of LD used and particularly on the sensitivity of the measure to changes in allele frequencies. Consider first the effect of a single sweep on the mean number of haplotypes normalized by the number of segregating sites,  $E(n_{\text{Haps}}/(S+1))$ . As can be seen in Table 2, a neutral locus affected by a very recent sweep can exhibit a paucity of haplotypes relative to a standard neutral model (depending on the values of *s* and *c*). This suggests an increase in LD. However, the summary  $E(n_{\text{Haps}}/(S+1))$  becomes *greater* than expected under neutrality shortly after the sweep FIGURE 5.—An illustration of the effect of a selective sweep (see Table 2). This is easily understood: As the high-<br>on a neutral locus: a scatterplot of  $r^2$  for one simulated data frequency variants fix and new mutation alleles are now rare and many form new haplotypes.

are included. The effective population size  $N = 10^4$ , the selec-<br>tion coefficient  $s = 0.05$ , and the population mutation rate<br>sidered the effect of selective sweeps on allelic association coefficient  $s = 0.05$ , and the population mutation rate<br>  $\theta = 40$ . The population recombination rate for the neutral<br>
locus,  $\rho$ , is 200 (which corresponds to 1 Mb for a recombina-<br>
tion rate of 0.5 cM/Mb/generation diately adjacent to the neutral locus. The sample size is 50, corresponds to what is sometimes referred to as "haplo-<br>so points >0.0768 are in significant linkage disequilibrium by type structure" in the literature (*e.g.* so points >0.0768 are in significant linkage disequilibrium by<br>a  $\chi^2$  test (*cf.* PRITCHARD and PRZEWORSKI 2001). (A) The tratio is sharply decreased by a sweep and monotoni-<br>beneficial allele fixed at time  $t = 0$ . (B) time since the sweep. These results suggest that this tive sweep. This is also true of another feature of the statistic might be useful for detecting positive selection.

bination rates. In humans, it is the distance over which would have to be strong and recent for selective sweeps

*t* 0 0.01 0.05 0.10 0.25 0.50 1.00 No sweep  $N = 10^6$ ,  $s = 0.005$ , *s* 0.005 (0.44) (0.60) (0.82) 0.89 0.98 0.99 0.93 0.89  $N = 10^4$ ,  $s = 0.01$ , *s* 0.01 0.76 0.79 0.83 0.85 0.83 0.76 0.70 0.67  $N = 10^4$ ,  $s = 0.05$ , *s* 0.05 (0.58) 0.69 0.81 0.84 0.84 0.77 0.70 0.67  $N = 10^6$ ,  $s = 0.005^{\circ}$ , *s* 0.005*<sup>a</sup>* (0.53) (0.57) (0.81) (0.93) (1.10) (1.19) (1.23) 1.25  $N = 10^4$ ,  $s = 0.01^a$ , *s* 0.01*<sup>a</sup>* (0.70) (0.76) (0.84) (0.88) (0.90) (0.90) 0.93 0.93

The effect of a selective sweep on the mean  $n_{\text{Haps}}/(S+1)$ 

The time since the fixation of the favored allele, *t*, is scaled in units of 4*N* generations, where *N* is the effective population size.  $n_{\text{Hans}}$  is the number of distinct haplotypes and *S* is the number of segregating sites. The sample size is 50, the population mutation rate for the neutral locus,  $\theta$ , is 5, and the genetic distance to the selected locus, *c*, is chosen such that  $c/s = 0.01$  (where *s* is the selection coefficient of the favored allele). In simulations where  $N=10^6$ , the population recombination rate for the neutral locus,  $\rho$ , is 20 (corresponding to 1 kb if the recombination rate is  $5 \times 10^{-9}$ /bp/generation); where  $N = 10^4$ ,  $\rho$  is 5 (corresponding to  $\sim$ 25 kb if the recombination rate is 0.5 cM/Mb/generation). In parentheses are those entries for which  $E(n_{\text{Hans}}/n)$  $(S + 1)$ ) is less than the neutral expectation.

 $a_{\text{Haps}}/(S+1)$  is calculated excluding singleton mutations.

spots, with most of the genome experiencing very low least as modeled. rates of crossing over (*e.g.*, JEFFREYS *et al.* 2001). If so, Repeated sweeps do produce a relative excess of LD "recombination coldspots" may preserve allelic associa- when attention is restricted to intermediate frequency

increase in LD is short-lived, anonymous loci subject to  $1.05$  for  $\lambda = 10^{-5}$ , and 0.90 for  $\lambda = 5 \times 10^{-5}$  ( $\lambda$  is the repeated selective sweeps do not show a marked excess rate of sweep per base pair per 4*N* generations). Figure of LD. In fact, summaries of LD that are highly sensitive  $\overline{6}$  plots the expected decay of  $r^2$  with distance for these to the frequency spectrum, such as  $C_{\text{hud}}$  or  $E(n_{\text{Haps}}/(S +$ 1)), suggest *less* LD under this model of recurrent sweeps chosen to be plausible for *D. melanogaster.* The increase than under neutrality. *C*hud, in particular, is smaller when relative to a neutral model is slight. Note further that the sample variance in the number of pairwise differ- the rate  $\lambda = 5 \times 10^{-5}$  is probably unrealistically high. For ences is larger. Selective sweeps skew the frequency spec-  $s = 0.01$ , and assuming a fixation probability of 2*s* (*cf.*) trum toward rare alleles, leading to a smaller variance Crow and Kimura 1970, p. 426), roughly one in every

to account for the unexpectedly large distances over in pairwise differences and larger values of *C*hud (results which LD sometimes extends in humans. This said, re- not shown). Thus, repeated sweeps cannot account for cent evidence suggests that most crossing-over events in the low values of  $C_{\text{had}}$  found at most loci in both species humans may occur within narrow recombination hot- of Drosophila (ANDOLFATTO and PRZEWORSKI 2000), at

tions longer than suggested by these simulations. variants. For example, in  $10^4$  simulations,  $E(n_{\text{Haps}}/(S +$ **The effect of repeated sweeps on LD:** Because the 1)) excluding singletons is 1.24 in the absence of sweeps, two rates of sweeps, with the other parameter values



Figure 6.—The effect of repeated selective sweeps on the expected rate of decay of pairwise linkage disequilibrium. The effective population size  $N = 10^6$ , the selection coefficient *s* 0.01, the population mutation rate  $\theta = 40$ , the population recombination rate  $\rho =$ 20, and the sample size is 50. The neutral locus is affected by repeated sweeps occurring at rate  $\lambda/bp/4N$ generations (assuming a recombination rate of 5  $\times$  $10^{-9}$ /bp/generation).

### **TABLE 3**

	Panmixia	$4 Nm = 1$ (sampled 48/2)	$4 Nm = 1$ (sampled 50/0)	$4 Nm = 0.5$ (sampled 50/0)	$4 Nm = 2$ (sampled 50/0)
P(H)	0.05	0.14	0.14	0.19	0.09
P(D)	0.05	0.09	0.06	0.09	0.05

**The power of** *H* **and** *D* **to detect a symmetric two-island model**

The power of *H* and *D* is estimated from  $10^4$  simulations, as described in METHODS.  $4Nm$  is the number of migrants per deme per generation. The sample size is 50. The population mutation rate per deme is 2.5. There is no intralocus recombination.

three newly arising mutations would have to be advanta- model. In addition, population structure can produce geous to obtain this rate of selective sweeps (if the neutral high levels of LD (Li and Nei 1974; Wall 1999). mutation rate is taken to be  $2 \times 10^{-9}$ /generation/bp; McVean and Vieira 2001). Thus, for plausible parame- both Drosophila and humans. However, the purpose of ters, the decay of LD is barely less steep than under a these simulations is simply to illustrate that a demoneutral model. Randomly chosen loci are therefore not graphic model that produces trees such as Figure 1 expected to show strikingly high levels of LD, even if there more often than the standard neutral model will have have been multiple selective sweeps at linked sites. the same effect on *H* as a selective sweep. In fact, recent

**The possible effect of population structure:** If old or<br>
ted under the standard neutral model. In other words,<br>
tecurrent sweeps lead neither to high levels of LD nor<br>
to significant *H* tests, how do we interpret these island model, each of size  $N/2$ , with 0.5–2 migrants per<br>demographic explanations is that positive selection does<br>migration rate corresponds to an  $F_{ST}$  value of ~0.11–0.33<br>(HUDSON *et al.* 1992). As can be seen in Tabl Even if samples are collected from only one locality, and ANDOLFATTO (2001) have estimated the rate of  $P(H) > 5\%$ , as the samples sometimes contain individu-<br>selective sweeps needed to account for the positive cor- $P(H) \geq 5\%$ , as the samples sometimes contain individu-<br>als whose ancestors were migrants from other demes.<br>relation between diversity levels and crossing-over rates If levels of differentiation are higher (*e.g.*,  $F_{ST} = 0.33$ , observed in humans and in Drosophila, respectively. corresponding to 0.5 migrant per deme per generation The probability of overlap can be estimated from Equa-<br>in a two-island model),  $P(H)$  can be as high as 19%. If the probability of overlap can be estimated from Equathere are more than two islands, then, for approximately these rough calculations, it appears that in both species, the same  $F_{ST}$  value, the power is similar (results not selective sweeps will often occur concurrently (results shown). In general, the power of  $H$  to detect population not shown). structure increases with higher  $\theta$  or lower migration When two or more alleles are simultaneously favored, rates (results not shown). In summary, the null model interference between them might alter the patterns of can be rejected by the *H* test at substantially higher than polymorphism relative to the predictions of a single-site the nominal rejection probability when samples are model of positive selection (KIRBY and STEPHAN 1996). drawn unequally from different islands in an island However, the selected sites would have to be very close

This particular model is likely to be unrealistic for bottlenecks (results not shown) and a metapopulation model (Wakeley and Alicar 2001) can also lead to DISCUSSION high-frequency-derived alleles more often than expec-

> relation between diversity levels and crossing-over rates. tion 6 in BRAVERMAN et al. (1995). On the basis of

to one another on the chromosome for interference to with polymorphism data that a candidate locus has unhave an effect. If the locations of the selected substitu-<br>dergone a recent sweep (*e.g.*, PARSCH *et al.* 2001; TAKAtions are chosen uniformly, as in this model, this condi-<br>
Hashi *et al.* 2001). However, it has low power to detect tion is unlikely to be met. Under an alternative model, the effects of positive selection at a randomly chosen where several adaptive changes occur in a small region locus. In addition, it may not be conservative if there in short succession, interference between sweeps may is hidden population structure. Similarly, while sweeps be more likely. It is unknown whether such a scenario increase LD between intermediate frequency variants, would lead to higher levels of LD or more high-fre-<br>the effect is short-lived. Thus, randomly chosen data quency-derived alleles. Even so, the effects are likely to sets with significant *H* values and high levels of LD may be short-lived, as recombination will rapidly break down reflect demography rather than adaptation. Alternaallelic associations after the sweeps, and high-frequency tively, positive selection may not operate as it is most alleles will drift to fixation. Thus, occasional overlaps commonly modeled.

selective sweeps is constant. If, instead, there has been Andolfatto, Y. Gilad, R. Hudson, G. McVean, and J. Wall as well as an increase in the rate of genetic adantations toward D. Charlesworth and two anonymous reviewers an increase in the rate of genetic adaptations toward<br>the present, many loci may reflect recent sweeps. In the<br>case of cosmopolitan species of Drosophila, this time<br>case of cosmopolitan species of Drosophila, this time frame could reflect recent colonization of temperate habitats. Similarly, anatomically modern humans are<br>
thought to have left Africa and spread across the globe<br>
starting  $\sim$ 50 thousand vears ago, and there have been ANDOLFATTO, P., 2001 Adaptive hitchhiking effects on ge starting  $\sim$  50 thousand years ago, and there have been<br>major changes in population density over the past 10<br>knoot and MNDOLFATTO, P., 2001 Adaptive hitchhiking effects on genome vari-<br>hility. Curr. Opin. Genet. Dev. 11: kya (JONES *et al.* 1994). The emergence of modern hu-<br>mans and their spread through the world may have Drosophila. Genetics 155: 257–268. mans and their spread through the world may have Drosophila. Genetics 155: 257–268.<br>ANDOLFATTO, P., and M. PRZEWORSKI, 2001 Regions of lower recom-

Note further that the sojourn time of a selected allele *Drosophila melanogaster*. Genetics 158: 657–665.<br>
a random-mating population of constant size is  $\sim$  9 AQUADRO, C. F., D. J. BEGUN and E. C. KINDAHL, 1994 Selection in a random-mating population of constant size is  $\sim$  2<br>  $\frac{10(2N)}{s}$  and E. C. KINDAHL, 1994 Selection,<br>  $\frac{10(2N)}{s}$  (assuming that the allele was selected when<br>
first introduced), where N is the diploid effective first introduced), where *N* is the diploid effective popu-<br>lation size and *s* the selection coefficient of the favored<br>BARTON, N. H., 1998 The effect of hitch-hiking on neutral genealo-BARTON, N. H., 1998 The effect of the favored BARTON, N. H., 1998 The effect of the selection coefficient of the favored gies. Genet. Res. **72:** 123–133. allele (*cf.* STEPHAN *et al.* 1992). With the *N* values as-<br>BEGUN, D. J., and C. F. Aquadro, 1993 African and North American<br>populations of *Drosophila melanogaster* are very different at the sumed throughout and a selection coefficient of  $1\%$ , populations of *Drosophila melanog*<br>this translates into  $\approx 9 \times 10^3$  generations for humans DNA level. Nature **365:** 548–550. this translates into  $\approx 2 \times 10^3$  generations for humans<br>and  $2.9 \times 10^3$  generations for Drosophila (respectively,<br>and  $2.9 \times 10^3$  generations for Drosophila (respectively,<br>W. STEPHAN, 1995 The hitchhiking effect on th 4  $\times$  10<sup>4</sup> years assuming 20 years per generation and 300 spectrum of DNA polymorphisms. Genetics 140: 783–796.<br>
vears assuming 10 generations a year) The demo- Cavalli-Srorza, L. L., P. Manozzi and A. Piazza, 1994 The H years assuming 10 generations a year). The demo-<br>graphic assumptions behind this calculation are likely<br>to be invalid for the recent past of many cosmopolitan<br>the CROW, J. F., and M. KIMURA, 1970 An Introduction to Populat to be invalid for the recent past of many cosmopolitan Crow, J. F., and M. KIMURA, 1970 *An Introduction* Species However they suggest that if there has been an *ics Theory*. Alpha Editions, Edina, MN. species. However, they suggest that if there has been an its Theory. Alpha Editions, Edina, MN.<br>increase in the rate of sweeps in the recent past, a subset<br>of loci may reflect incomplete sweeps—ones that are<br> $\frac{155}{1200}$ of loci may reflect incomplete sweeps—ones that are FAY, J. C., and C.-I Wu, 2001 The neutral theory in the selected variant is no longer era. Curr. Opin. Genet. Dev. 11: 642–646.

disequilibrium levels. Am. J. Hum. Genet. **69:** 831–843.<br>Fu, Y.-X., 1995 Statistical properties of segregating sites. Theor. Fu, Y.A., 1995 Statistical properties of segregating sites. Theor. population structure in both *D. melanogaster* (*e.g.*, HALE GILAD, Y., D. SEGRE, K. SKORECKI, M. NACHMAN, D. LANCET *et al.*, population structure in both *D. melanogaster* (*e.g.*, HALE GILAD, Y., D. SEGRE, K. SKORECKI, M. NACHMAN, D. LANCET *et al.*, and SINGH 1991: REGIJN and AQUADRO 1993) and hu- 2000 Dichotomy of single-nucleotide polymorphi and SINGH 1991; BEGUN and AQUADRO 1993) and hu-<br>mans (e.g., CAVALLI-SFORZA et al. 1994) as well as for<br>geographic differences in selective pressures at particu-<br>geographic differences in selective pressures at particu-<br>GIL geographic differences in selective pressures at particu-<br>
lar loci (reviewed in ANDOLFATTO 2001). Departures 2001 Evidence for positive selection and population structure at lar loci (reviewed in ANDOLFATTO 2001). Departures<br>from random mating could distort the signature of selec-<br>HALE, L. R., and R. S. SINGH, 1991 A comprehensive study of genic tion relative to our expectations for a panmictic popula- variation in natural populations of *Drosophila melanogaster.* IV.

In summary, the *H* test is a useful tool to confirm HAMBLIN, M. T., E. E. THOMPSON and A. DI RIENZO, 2002 Complex

are unlikely to explain the observed patterns.<br>I thank P. Andolfatto, A. Di Rienzo, P. Donnelly, J. Fay, I. Gordo,<br>More problematic is the assumption that the rate of R. Griffiths, J. Pritchard, and J. Wall for helpful dis R. Griffiths, J. Pritchard, and J. Wall for helpful discussions and P.

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- Coincided with a burst of genetic adaptations.<br>Note further that the sojourn time of a selected allele<br>Drosophila melanogaster. Genetics 158: 657-665.
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- still ongoing or where the selected variant is no longer<br>favored.<br>An additional assumption of this sweep model that is<br>An additional assumption of this sweep model that is<br>and et al., 2001 Gene conversion and different pop may explain the contrast between polymorphism and linkage disequilibrium levels. Am. J. Hum. Genet. 69: 831-843.
	-
	-
	-
- tion, resulting in high levels of LD and, perhaps, in the Mitochondrial DNA variation and the role of history vs. selection in the genetic structure of geographic populations. Genetics 129:<br>
103-117.
	-

- Hudson, R. R., 1987 Estimating the recombination parameter of a gene, *Dmd*, in humans. Genetics **155:** 1855–1864. 250. quence data. Trends Genet. **16:** 526–529.
- HUDSON, R. R., 1993 The how and why of generating gene genealo-<br>gies, pp. 23–36 in *Mechanisms of Molecular Evolution*, edited by N. sequence variation suggest the recent action of positive selection Tаканата and A. G. CLARK. Japan Scientific Society, Tokyo.
- HUDSON, R. R., M. SLATKIN and W. P. MADDISON, 1992 Estimation 647–657.<br>
of levels of gene flow from DNA sequence data. Genetics 132: PRITCHARD, J. 583–589. in humans: models and data. Am. J. Hum. Genet. **69:** 1–14.
- JEFFREYS, A. J., L. KAUPPI and R. NEUMANN, 2001 Intensely punctate meiotic recombination in the class II region of the maior histocompatibility complex. Nat. Genet. **29:** 217–222. JONES, S., R. MARTIN and D. PILBEAM (Editors), 1994 The Cambridge
- bridge. results based on diffusion theory. Theor. Popul. Biol. **41:** 237–254.
- 
- and background selection on neutral variation. Genetics 155: 1415–1427. Tajima, F., 1983 Evolutionary relationships of DNA sequences in
- Kim, Y., and W. Stephan, 2001 Detecting a local signature of genetic finite populations. Genetics **105:** 437–460. hitchhiking along a recombining chromosome. Genetics **160:** Tajima, F., 1989a Statistical method for testing the neutral mutation
- Kirby, D. A., and W. Stephan, 1996 Multi-locus selection and the Tajima, F., 1989b The effect of change in population size on DNA structure of variation in a segment of the *white* gene of *Drosophila* polymorphism. Genetics **123:** 597–601.
- Attacin genes of *Drosophila melanogaster*. Genetics **159:** 659–671. Lt, W. H., and M. NEI, 1974 Stable linkage disequilibrium without
- epistasis in subdivided populations. Theor. Popul. Biol. **6:** 173– lation. Genetics **159:** 893–905.
- LI, W. H., and L. SADLER, 1991 Low nucleotide diversity in man.<br>Genetics 129: 513–523.
- *al.*, 2001 Sequence variability of a human pseudogene. Genome Opin. Genet. Dev. **11:** 647–651.
- MAYNARD SMITH, J., and J. HAIGH, 1974 The hitch-hiking effect of a favourable gene. Genet. Res. 23: 23-35.
- McVEAN, G. A., and J. VIEIRA, 2001 Inferring parameters of muta-<br>tion, selection and demography from patterns of synonymous WRIGHT S. 19
- NACHMAN, M. W., 2001 Single nucleotide polymorphisms and recombination rate in humans. Trends Genet. 17: 481-485. Communicating editor: D. CHARLESWORTH
- signatures of natural selection at the duffy blood group locus. NACHMAN, M. W., and S. L. CROWELL, 2000 Contrasting evolutionary Am. J. Hum. Genet. **70:** 369–383. histories of two introns of the duchenne muscular dystrophy
	- OTTO, S. P., 2000 Detecting the form of selection from DNA se-
	- sequence variation suggest the recent action of positive selection<br>in the janus-ocnus region of *Drosophila simulans*. Genetics 159:
	- PRITCHARD, J. K., and M. PRZEWORSKI, 2001 Linkage disequilibrium
	- Sequence variation in the human angiotensin converting enzyme.<br> Nat. Genet.  $22: 59-62$ .
- STEPHAN, W., T. H. E. WIEHE and M. LENZ, 1992 The effect of *Encyclopedia of Human Evolution.* Cambridge University Press, Cam- strongly selected substitutions on neutral polymorphism: analytic
- AN, N. L., R. R. Hudson and C. H. Langley, 1989 The "hitch- Taillon-Miller, P., I. Bauer-Sardina, N. L. Saccone, J. Putzel, hiking effect" revisited. Genetics 123: 887–899. T. Laitinen *et al.*, 2000 Juxtaposed regions of hiking effect" revisited. Genetics 123: 887–899. T. LAITINEN *et al.*, 2000 Juxtaposed regions of extensive and KIM, Y., and W. STEPHAN, 2000 Joint effects of genetic hitchhiking minimal linkage disequilibrium in human Xq2 minimal linkage disequilibrium in human Xq25 and Xq28. Nat. Genet.  $25: 324-328$ .
	-
	- 765–777. hypothesis by DNA polymorphism. Genetics **123:** 585–595.
		-
- TAKAHASHI, A., S. C. TSAUR, J. A. COYNE and C.-I Wu, 2001 The Lazzaro, B. P., and A. G. Clark, 2001 Evidence for recurrent paralo- nucleotide changes governing cuticular hydrocarbon variation gous gene conversion and exceptional allelic divergence in the and their evolution in *Drosophila melanogaster*. Proc. Natl. Acad. Attacin genes of *Drosophila melanogaster*. Genetics 159: 659-671. Sci. USA 98: 3920-3925.
	- WAKELEY, J., and N. ALICAR, 2001 Gene genealogies in a metapopu-
	- 183. WALL, J. D., 1999 Recombination and the power of statistical tests<br>W. H., and L. SADLER, 1991 Low nucleotide diversity in man. of neutrality. Genet. Res. 73: 65–79.
- Genetics **129:** 513–523. WALL, J. D., 2001 Insights from linked single nucleotide polymor-<br>MARTINEZ-ARIAS, R., F. CALAFELL, E. MATEU, D. COMAS, A. ANDRES et phisms: what we can learn from linkage disequilibrium. Curr. phisms: what we can learn from linkage disequilibrium. Curr.
	- Res. 11: 1071–1085.<br>
	NARD SMITH, J., and J. HAIGH, 1974 The hitch-hiking effect of the statistical tests of neutrality. Mol. Biol. Evol. 18: 1134–1135.
		- WEIR, B. S., 1996 *Genetic Data Analysis II. Sinauer Associates, Sunder-*
	- tion, selection and demography from patterns of synonymous WRIGHT, S., 1951 The genetical structure of populations. Ann. Eusite evolution in Drosophila. Genetics 157: 245–257. gen. 15: 323–354.