# **On the Determination of Recombination Rates in Intermated Recombinant Inbred Populations**

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### ABSTRACT

The recurrent intermating of  $F_2$  individuals for some number of generations followed by several generations of inbreeding produces an intermated recombinant inbred (IRI) population. Such populations are currently being developed in the plant-breeding community because linkage associations present in an  $F_2$ population are broken down and a population of fixed inbred lines is also created. The increased levels of recombination enable higher-resolution mapping in IRI populations relative to  $F_2$  populations. Herein we derive relationships, under several limiting assumptions, for determining the expected recombination fraction in IRI populations from the crossover rate per meiosis. These relationships are applicable to situations where the inbreeding component of IRI population development is by either self-fertilization or full-sib mating. Additionally, we show that the derived equations can be solved for the crossover rate per meiosis if the recombination fraction is known for the IRI population. Thus, the observed recombination fraction in any IRI population can be expressed on an  $F_2$  basis. The implications of this work on the expansion of genetic maps in IRI populations and limits for detecting linkage between markers are also considered.

HIGH-RESOLUTION genetic maps are an impor- this population are available at the Maize Mapping Project tant resource for identifying genes associated with (http://www.maizemap.org/)]. These intermated requantitative traits, for understanding gene regulation combinant inbred (IRI) populations are randomly and expression, and for understanding allele diversity mated for some number of generations after the  $F_2$ , at the gene level. In principle these maps provide more followed by cycles of self-fertilization to produce a popupower for detecting genes of interest than has tradition- lation of homozygous lines (*e.g*., Figure 1). The developally been provided at the level of quantitative trait loci. ment of an IRI mapping population has advantages The human, animal, and plant genetics communities are over a RM mapping population in that a permanent all pursuing a number of methods for creating such maps population of fixed lines is produced. A mapping popuin an effort to progress in the above areas. The resolu-<br>tion developed by selfing from the  $F_2$  also produces a<br>tion of genetic maps is directly related to the number of population of fixed lines, but mapping resolution opportunities for recombination events between closely limited relative to an IRI population.<br>linked genes prior to mapping. In plants, the traditional Although IRI populations are being linked genes prior to mapping. In plants, the traditional Although IRI populations are being developed, no the-<br>approach has been to develop recombinant inbred lines or has been established to relate the frequency of reapproach has been to develop recombinant inbred lines ory has been established to relate the frequency of re-<br>from an  $F_2$  population derived from a single biparental combination events in IRIs to the crossover frequency from an  $F_2$  population derived from a single biparental combination events in IRIs to the crossover frequency<br>cross. It has been noted, however, that genetic linkage per meiosis (that is, the frequency expected in an  $F$ cross. It has been noted, however, that genetic linkage per meiosis (that is, the frequency expected in an  $F_2$  populations broken down more effectively by randomly mating lation). Expectations for RI populations are wel is broken down more effectively by randomly mating lation). Expectations for RI populations are well known<br>an  $F_2$  population for a small number of generations and date to the work of HALDANE and WADDINGTON an  $F_2$  population for a small number of generations and date to the work of HALDANE and WADDINGTON (LIU *et al.* 1996), and that more precise map locations of (1931). More recently, a theoretical expectation has (Liu *et al.* 1996), and that more precise map locations of (1931). More recently, a theoretical expectation has quantitative trait loci can be found in such populations been published for RM populations (Liu *et al.* 199 quantitative trait loci can be found in such populations been published for RM populations (Liu *et al.* 1996).<br>(DARVASI and SOLLER 1995). Mapping studies using IRI populations typically, and

Recently, mapping populations that incorporate impor-<br>tant aspects of both randomly mated (RM) and recombi-<br>tions to adjust recombination rates observed in IRI mantant aspects of both randomly mated (RM) and recombi-<br>nant inbred (RI) populations have been constructed in  $\frac{1}{2}$  ning populations back to an  $\frac{1}{2}$  basis (Cor et al. 2002)

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population of fixed lines, but mapping resolution is

VARVASI and SOLLER 1995).<br>
Recently, mapping populations that incorporate impor-<br>
Froneously apply equations for RI and RM populanant inbred (KI) populations have been constructed in ping populations back to an  $F_2$  basis (Coe *et al.* 2002; SHAROPOVA *et al.* 2002; details on  $F_2$  basis (Let *et al.* 2002; SHAROPOVA *et al.* 2002; details on  $F_$ LEE *et al.* 2002). In this study we derive a relationship between the observed recombination fraction, *R*, and the  $F_2$ -adjusted recombination rate,  $r$ , in an IRI mapping <sup>1</sup> Corresponding author: 7250 NW 62nd Ave., P.O. Box 552, Johnston, <sup>population. The derived relationship may be used to *Corresponding author:* 7250 NW 62nd Ave., P.O. Box 552, Johnston, population. The derived relations</sup> estimate *R* given some knowledge of *r*, and vice versa.

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### **TABLE 1**

**List of symbols**

Symbol	Meaning			
$\boldsymbol{R}$	Recombination fraction observed in a mapping population			
$\boldsymbol{r}$	Recombination fraction per meiosis ( <i>i.e.</i> , adjusted to an $F_2$ basis)			
$\boldsymbol{t}$	Number of generations of random mating			
$\boldsymbol{n}$	Number of generations of inbreeding, either via selfing or full-sib mating			
$C_n, D_n, E_n, F_n, G_n$	Proportion of genotypes in genotypic classes $C$ , $D$ , $E$ , $F$ , and $G$ after $n$ generations of inbreeding (see text for definitions)			
$C'_n, D'_n, E'_n, F'_n, G'_n$	Proportion of genotypes in each class after $t$ generations of random mating and $n$ generations of inbreeding			
$c_n, f_n$	Differences between proportions of genotypic classes after $n$ generations of inbreeding (see text for definitions)			
$c'_n, f'_n$	Differences between proportions of genotypic classes after $t$ generations of random mating and $n$ generations of inbreeding			
RI	Recombinant inbred			
RM	Randomly mated			
IRI	Intermated recombinant inbred			

### **THEORY**

An extension of HALDANE and WADDINGTON's (1931) method was used to derive the results for IRI popula-<br>tions. The method is first illustrated for self-fertilized<br>populations, however. Different values of  $\lambda$  are appropriate for<br>populations where the relationship betwee populations where the relationship between *R* and *r* is<br>well known. A list of the symbols used in this section<br>and their meaning is provided in Table 1.<br>**Selfed populations:** Consider two loci on a chromo-<br>some with two

with genotypes *AABB* and *aabb*, are crossed to produce an F<sub>1</sub> population. Following *n* generations of self-fertilization, the population contains a distribution of genotypes. We separate this distribution into the classes and thus  $f_{\infty} = 0$ . So

$$
C_n = AABB + aabb
$$
  
\n
$$
D_n = AAbb + aABB
$$
  
\n
$$
E_n = AABb + AABB + Aabb + aabb
$$
  
\n
$$
F_n = AB.ab
$$
  
\n
$$
G_n = Ab.aB
$$

where  $F_n$  and  $G_n$  represent coupled and repulsion double heterozygotic genotypes, respectively. Moreover, define the relationships

$$
c_n \equiv C_n - D_n
$$
  

$$
f_n \equiv F_n - G_n.
$$

Thus,  $c_n$  is the difference between the proportions of bomozygous individuals having parental and nonparental which is the standard formula relating the fraction of<br>tal gametes (and similarly for  $f_n$  but with regard to dou-<br>ble heterozygous genotypes). HALDANE and WAD-<br>pINGT

$$
c_i + \lambda f_i = c_j + \lambda f_j, \tag{1}
$$

$$
\lambda = \frac{1 - 2r}{2 + 4r}.\tag{2}
$$

$$
E_{\infty} = F_{\infty} = G_{\infty} = 0
$$
  

$$
C_{\infty} + D_{\infty} = 1
$$
 (3)

$$
C_n = AABB + aabb \qquad c_\infty + \lambda f_\infty = c_\infty = c_1 + \lambda f_1, \qquad (4)
$$

where the subscript 1 indicates the  $F_2$  generation *(i.e.,* after one generation of selfing). Using Haldane and Waddington's Equation 1.1 to show that  $2c_1 = f_1 = 1 - 2r$  and substituting into Equation 4 yields

$$
c_{\infty} = c_1(1 + 2\lambda) = \frac{1 - 2r}{2}(1 + 2\lambda).
$$
 (5)

Since  $c_{\infty} = C_{\infty} - D_{\infty}$ , Equations 2 and 3 can be used to solve for  $D_{\infty}$  in Equation 5. This gives

$$
D_{\infty} = \frac{2r}{1+2r},\tag{6}
$$

just described; it is simply presented to illustrate the method used for IRI populations.

**IRI populations:** Equation 6 does not hold, however,



by multiple generations of inbreeding are shown. The syn4 RM designation refers to populations that have been randomly mated populations (HALDANE and WADDINGTON 1931), the for four generations after the  $F_2$ . The term syn4 IRI refers to derivation outlined above can be repeated wi for four generations after the  $F_2$ . The term syn4 IRI refers to<br>a syn4 RM population that has subsequently undergone self-<br>value of  $\lambda$ . Doing so yields the equation fertilization or full-sib mating to create a population of inbred lines.

equations for RM populations (DARVASI and SOLLER inbred populations,  $R = 4r/(1 + 6r)$ . 1995; Liu *et al.* 1996) are inappropriate for IRI popula-<br>tions due to the generations of inbreeding present in rather than R. By knowing the recombination rate on tions due to the generations of inbreeding present in rather than *R*. By knowing the recombination rate on IRIs. The correct relationship between *R* and *r* for IRIs an *F*<sub>p</sub> basis it is possible to relate marker inform IRIs. The correct relationship between *R* and *r* for IRIs an  $F_2$  basis it is possible to relate marker information is outlined below.

two loci is expected to decrease by  $1 - r$  for each gen-<br>eration of random mating (FALCONER and MACKAY 1996; follows (and similarly for Equation 10, but not shown). OTT 1999), and thus the value of *c* decreases by  $1 - r$  Set the dummy variable  $z = 1 - r$  and substitute into for each generation of random mating after the  $F_2$  gen-<br>Equation 9. A little algebra gives eration. Defining  $c'$  to be the value of  $c$  after  $t$  generations of random mating, we have the following relationship:

$$
c_1' = c_1(1 - r)^t. \tag{7}
$$

*Because of the random mating, Equation 4 becomes* 

$$
c'_{\infty} = c'_1 + \lambda f'_1 \tag{8}
$$

$$
2c'_1 = f'_1
$$
 RESULTS  
\n
$$
c'_\in C'_\infty - D'_\infty
$$
 **Expansion of genetic maps:**  
\n
$$
C'_\infty + D'_\infty = 1
$$
 **Expansion in RI, RM, and IRI** p

$$
R = \frac{1}{2} \left( 1 - \frac{1 - 2r}{1 + 2r} (1 - r)^t \right).
$$
 (9)

crossover events in an IRI population to the crossover frequency seen at the  $F_2$  generation. Note that if  $t = 0$ , Equation 9 reduces to the equation used for purely selffertilized populations (*i.e.*, Equation 6) so the recombination fractions in RI *and* IRI populations developed via selfing can be determined with this theory. A population that was randomly mated after the  $F_2$  but had no generations of self-fertilization follows the relationship

$$
R = \frac{1}{2}(1 - (1 - 2r)(1 - r)^t)
$$

as was previously shown by Liu *et al.* (1996) and in somewhat different form by DARVASI and SOLLER (1995).

Additionally, IRI populations can be developed by full-sib mating, rather than by selfing, after undertaking FIGURE 1.—Schematic illustration of the development of an generations of random mating. Such populations also<br>IRI population. Four generations of random mating followed follow an analytic relationship between R and r. Reco nizing that  $\lambda = (1 - 6r)/(2 + 12r)$  for full-sib mating

$$
R = \frac{1}{2} \left( 1 - \frac{1 - 2r}{1 + 6r} (1 - r)^t \right).
$$
 (10)

if *t* generations of random mating have taken place Here again, note that if  $t = 0$ , Equation 10 reduces to prior to the inbreeding process (*e.g*., Figure 1). Likewise, the correct relationship for full-sib-mated recombinant

between mapping studies and hence produce dense The amount of linkage disequilibrium between any consensus genetic maps. If *R* has been measured in an follows (and similarly for Equation 10, but not shown).

$$
1 - 2R = \frac{2z - 1}{3 - 2z} \cdot z^t.
$$

*c*<sub>1</sub> Rearranging terms yields the expression

$$
2z^{t+1} - z^t + (2 - 4R)z + 3(2R - 1) = 0.
$$
 (11)

 $c' = c' + \lambda f'$  (8) Although this equation is complex, it can be solved numerically with standard methods. and, likewise,

**Expansion of genetic maps:** The degree of map expansion in RI, RM, and IRI populations depends on  $\phi$  expansion occurs as  $r \to 0$ <br>Substitution of the above relations into Equation 8 and  $\phi$  and decreases as *r* increases (HALDANE and WAD-Substitution of the above relations into Equation 8 and and decreases as *r* increases (HALDANE and WAD-<br>reduction of terms gives the equation<br> $N_{\text{DNCTON}}$  1931. I III *et al* 1996) By taking the derivative ping the derivative of terms gives the equation of the equation of the derivative of the equations showing the relationship between *R* and *r* and solving for  $r = 0$ , it can be shown that the maximum expansion factor is 2 for self-fertilized RI This relationship associates the observed fraction of populations and 4 for full-sib-mated RI populations. In

**TABLE 2 Genetic map expansion as a function of population type and marker density**

No. of marker pairs	Spacing (cM)	syn4 IRI <sup>a</sup>	syn4 <b>RM</b>	$\mathbb{R}$ <sup>a</sup>
5	20.0	206.11	188.56	142.86
10	10.0	281.30	237.56	166.67
100	1.0	385.37	293.08	196.08
200	0.5	392.59	296.52	198.02

Expansion of a genetic map with a total length of 100 cM in the F<sub>2</sub>. Different marker densities considered are shown in the first two columns, and the last three columns contain the total map length (in centimorgans) observed in three types of mapping population. Morgan's mapping function (Morgan 1928; OTT 1999) was used to convert map distances to recombination frequencies.

<sup>*a*</sup>Reported values are for IRI and RI populations developed by self-fertilization.

RM and IRI populations, however, the degree of map expansion varies as a function of the number of generations of random mating undertaken. For example, in an RM population,

$$
\frac{d}{dr}\left(\frac{1}{2}[1-(1-2r)(1-r)^{r}]\right)\Big|_{r=0}=\frac{t+2}{2}.
$$

Likewise, in IRI populations that are inbred by selfing self-fertilization. the map expansion factor is  $(t+4)/2$  and in IRI populations that are indired by full-sib mating this factor is  $(t +$ 8)/2. Sib-mated populations have greater map expan- breeding generations of the RI and IRI populations. sion than selfed populations, thus making them poten-<br>The RM and IRI populations have  $t = 4$ . Such populatially more useful for fine mapping. This greater expan- tions are referred to as syn4 RM or syn4 IRI populations, sion, however, is a consequence of the relatively slow decay respectively, in Table 2, a notation that is commonly of heterozygosity in the population; sib-mated RI popu- used in the plant-breeding literature (*e.g.*, Lee *et al.* lations are thus not preferable to IRI populations devel- 2002). The total length of the genetic maps observed oped by selfing for organisms where self-fertilization is in the final mapping population varies markedly depossible. For example, an F2 population that is randomly pending on the marker coverage (*e.g*., a sparse syn4 IRI mated for 4 generations and then self-fertilized for 6 map is almost half the size of a dense syn4 IRI map). generations will have a maximum map expansion equal Moreover, a densely mapped RI population can have to that of a full-sib-mated RI population, but with an nearly as much map expansion as a less dense syn4 IRI inbreeding coefficient greater than that of a population or RM population. Clearly, the amount of expansion that had been full-sib mated for 10 generations (Fehr expected in a genetic map depends on *both* the marker

We stress, however, that the *maximum* map expansion ered. factors apply *only* when *r* 0; marker pairs that have larger **Limits for detecting linkage between markers:** The recombination frequencies will have less map expan- extent of map expansion is dependent upon the number sion. This point appears to have been misunderstood of opportunities for crossover events during the creation in the literature (Coe *et al.* 2002; Lee *et al.* 2002). To of a mapping population. Additionally, it is difficult to illustrate the fact that map expansion varies as a function detect linkage relationships when *R* is large. Liu *et al.* of *r*, as well as the type of mapping population used, (1996) thus presented the equation consider Table 2. Data relating to four fictitious 100-cM genetic maps that differ in marker density are presented. The theoretically expected genetic map expansions for IRI, RM, and RI populations are shown, where to show the number of generations of random mating only self-fertilization mating is considered for the in- required for *R* to reach 0.4 in a RM mapping popula-



FIGURE 2.—The relationship between *r* and  $t_{R\geq 0.4}$  for RM (top) and IRI (bottom) populations that have been inbred by

1987). coverage and the type of mapping population consid-

$$
t_{R\geq 0.4} = \min\left(\text{integer} > \frac{\ln(1 - 2R) - \ln(1 - 2r)}{\ln(1 - r)}\bigg|_{R=0.4}\right) \quad (12)
$$

tion. Here  $t_{R\geq 0.4}$  is the number of generations of random SHAROPOVA *et al.* 2002) need to be revised and that the mating required for *R* to reach 0.4 and it is assumed degree of map expansion is in general larger in IRI that no linkage can be detected between markers when populations than in RM or RI populations. The rela- $R \geq 0.4$ . Equation 12 can also be used to determine the tively larger genetic map afforded by an IRI population maximum value of *r* for which linkage can be detected is a direct result of the additional meioses that occur after *t* generations of random mating. Figure 2, top, in the construction of these populations. shows this relationship and indicates that in a syn4 RM The theoretical expectations presented here, howpopulation, for example, linkage would not be detected ever, are not without limitations. Notably, we have asbetween markers with  $r \geq 0.224$ . Similarly, linkage sumed that crossover interference does not take place, would not be detected between markers with  $r \geq 0.124$  that multiple crossover events between two markers do in a syn10 RM population. It has therefore been sug- not occur during a single meiotic event, and that segregested that a first approximation of marker order be gation distortion is not a factor. Strictly speaking, the determined using an F<sub>2</sub> mapping population (LIU *et al.* theory holds only when the number of generations of 1996). Clearly the main purpose of developing random selfing or sib mating in the IRI population is sufficient

plished via selfing, the maximum value of *r* at which technically correct only for "large" population sizes. The linkage can be detected between two markers for a given results of Equations 9 and 10 will hold on average, of generation of intermating is course, but individual realizations of the breeding pro-

$$
t_{R\geq 0.4} = \min\left(\text{integer} > \frac{\ln(1 - 2R) - \ln(1 - 2r) + \ln(1 + 2r)}{\ln(1 - r)}\bigg|_{R=0.4}\right). \tag{13}
$$

Equation 13 is plotted in Figure 2, bottom, and shows LITERATURE CITED that linkage disequilibrium is broken down more rapidly COE, E., K. CONE, M. D. MCMULLEN, S.-S. CHEN, G. DAVIS *et al.*, 2002 in such IRI populations relative to RM populations. In Access to the maize genome: an integrated physical and genetic<br>a syn $A$  (syn $10$ ) IPI population for example, linkage can map. Plant Physiol. 128: 9–12. a syn4 (syn10) IRI population, for example, linkage can-<br>not be detected between markers where  $r \ge 0.186$  ( $r \ge 0.199$ –1207).<br>Narvast, A., and M. Soller, 1995 Advanced intercross lines, and the comparison of the genetic 0.109). Significantly, estimates of  $r$  from an  $F_2$  popula-<br>  $1199-1207$ .<br>
FALCONER, D. S., and T. F. C. MACKAY, 1996 *Introduction to Quantita*tion in maize (and that has subsequently been devel that converse in the C. MACKAY, 1996 *Introduction to Quantita*<br>oped into an IRI; Let *et al.* 2002) indicate that at least 8<br>of 10 chromosomes contain regions with recom of 10 chromosomes contain regions with recombination *and Technique*. MacMillan, New York.<br>
The s. S., and C. H. WADDINGTON, 1931 Inbreeding and **PLACE ISLANDER CONTAINS**, J. B. S., and C. H. WADDINGTON, 1931 Inbreeding an rates too large for detecting linkage in a syn4 IRI popula-<br>iinkage.cometics 16: 357–374.<br>Linkage.cometics 16: 357–374. N. SHAROPOVA W. D. BEAV.

We have derived a relationship between R, the ob-<br>served recombination fraction, and r, the equivalent<br>recombination fraction in an F<sub>®</sub> population. for a map-<br>Haven, T. H., 1928 The Theory of Genes. Yale University Press, recombination fraction in an  $F_2$  population, for a map-<br>  $\frac{1}{2}$  Haven, CT.<br>  $\frac{1}{2}$  Port, J., 1999 Analysis of Human Genetic Linkage, Ed. 3. Johns Hopkins ping population that has undergone some number of UTT, J., 1999 Analysis of Human Genetic Linkage, Ed. 3. Johns Hopkins<br>generations of intermating followed by generations of SHAROPOVA, N., M. D. MCMULLEN, L. SCHULTZ, S. SC either self-fertilization or full-sib mating. Our results SANCHEZ-VILLEDA *et al.*, 2002 Development and m<br>SSR markers for maize. Plant Mol. Biol. 48: 463-481. suggest that recombination estimates previously reported for IRI maize populations (*e.g.*, Lee *et al.* 2002; Communicating editor: J. B. WALSH

mating populations is to provide increased resolution to create inbreds (that is, in the limit  $n \to \infty$ ). In prac-<br>for closely linked marker pairs (e.g.,  $r < 0.1$ ). tice, however, the equations presented are still valid fice, however, the equations presented are still valid In an IRI population where the inbreeding is accom- for smaller, realistic values of *n*. Finally, the theory is cess can, and usually will, produce discrepancies from expectation when population sizes are small.

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- LEE, M., N. SHAROPOVA, W. D. BEAVIS, D. GRANT, M. KATT *et al.*, 2002 Expanding the genetic map of maize with the intermated B73  $\times$ Mo17 (*IBM*) population. Plant Mol. Biol. **48:** 453–461.
- DISCUSSION LIU, S.-C., S. P. KOWALSKI, T.-H. LAN, K. A. FELDMANN and A. H. PATERSON, 1996 Genome-wide high-resolution mapping by re-<br>current intermating using Arabidopsis thaliana as a model. Genet-
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