

# The stimulatory effects of caffeine, theophylline, lysine-theophylline and 3-isobutyl-1-methylxanthine on human sperm motility

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The potencies of caffeine, theophylline, lysine-theophylline and 3-isobutyl-1-methylxanthine (IBMX) in stimulating sperm motility have been compared, and we have found IBMX to be significantly more potent than the other three compounds, which did not exhibit significant differences in potency from each other.

**Keywords** sperm motility caffeine theophylline IBMX

## Introduction

Methylxanthines such as caffeine and theophylline increase sperm motility (Burge, 1973; De Turner *et al.*, 1978; Garbers *et al.*, 1971; Haesungcharern & Chulavatnatol, 1973; Hicks *et al.*, 1972; Levin *et al.*, 1980; Makler *et al.*, 1980; Schoenfield *et al.*, 1973). This effect has been attributed to their potent phosphodiesterase (PDE) inhibitory action and elevation of adenosine 3',5'-cyclic monophosphate (cAMP) levels within the sperm, following inhibition of cAMP-PDE (Tash & Means, 1983). We have measured the effects of theophylline and caffeine at stimulating sperm motility and compared them with IBMX and lysine-theophylline. IBMX exhibits a higher degree of PDE inhibitory activity than either caffeine or theophylline (Davis & Kuo, 1977; Glass & Moore, 1978; Helfman & Kuo, 1982) and lysine-theophylline is a more soluble theophylline preparation with a pharmacokinetic profile after oral administration almost identical to that of aminophylline in normal subjects (Johnston *et al.*, 1983).

Only samples with sperm concentrations higher than  $25 \times 10^6 \text{ ml}^{-1}$  and a percentage of progressive forward moving sperms higher than 20% were used. All drugs were dissolved in phosphate buffered saline (PBS, Oxoid Ltd) at pH 7.3. Sperm motility was measured using the method developed by Hong *et al.* (1981). Semen aliquots (100  $\mu\text{l}$ ) were mixed with 50  $\mu\text{l}$  of methylxanthine dissolved in PBS at pH 7.3. The semen-drug mixture (100  $\mu\text{l}$ ) was immediately placed onto a Nucleopore 5  $\mu\text{m}$  diameter membrane within the motility measuring apparatus and incubated for 2 h at 37°C, as previously described (Hong *et al.*, 1981). Change in sperm motility was measured from the percentage of sperm crossing the membrane compared with that of the control (2:1 semen:PBS mixture). Measurement of the effect of the methylxanthines was conducted on at least six different semen samples and in each sample the response to each methylxanthine concentration was estimated in duplicate.

## Methods

Fresh semen samples were obtained from 27 healthy volunteers aged between 18–24 years.

## Materials

All methylxanthines were obtained from Sigma Chemicals Ltd, phosphate buffered saline from Oxoid Ltd.

## Results

The responses to theophylline and caffeine were measured on the same semen samples in six cases and comparison of responses to caffeine and theophylline were made using a paired *t*-test. The responses to IBMX and lysine-theophylline were determined on different semen samples from those used to examine theophylline and caffeine and comparison of responses to IBMX and lysine theophylline with theophylline, was made, therefore, using the Mann Whitney U test. The response to IBMX differed significantly from that of theophylline ( $P < 0.004$ ), but the concentrations of caffeine, theophylline and lysine-theophylline producing half maximal increase in motility ( $EC_{50}$ ) did not differ significantly. Dose-response curves of theophylline, caffeine and IBMX are illustrated in Figure 1 and the data are in Table 1.

## Discussion

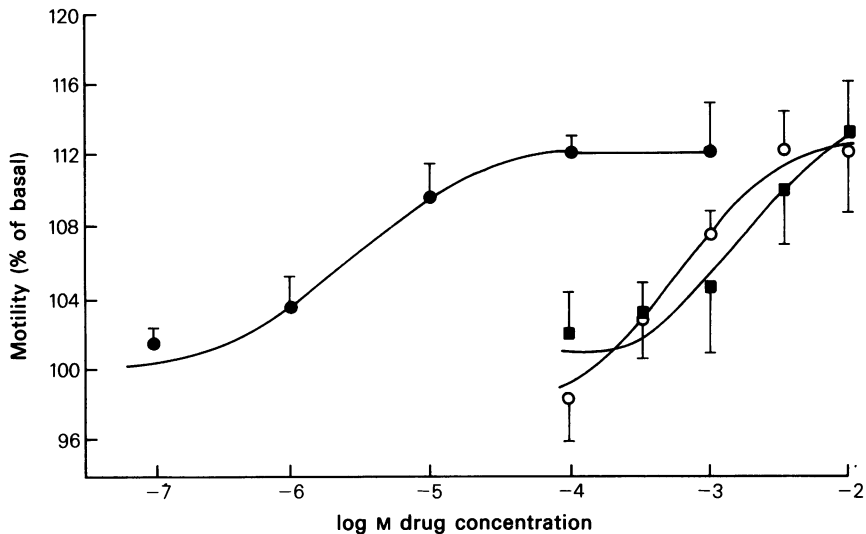
During the past decade there have been several reports indicating that methylxanthine derivatives can exert a facilitatory effect on sperm motility. Our findings are in agreement with these reports and extend the generalisation to include IBMX as a stimulator of sperm motility. The mechanism of this action of methylxanthines is not established. Some investigators have emphasised the primary involvement of phosphodiesterase inhibition and consequent elevation of basal cAMP levels as a mediator of methylxanthine induced increase in sperm motility. Direct application of dibutyryl-cAMP on to sperm produces an increase in sperm motility (Hartmann *et al.*, 1983; Hoskins *et al.*, 1975) and from the observed increase in sperm cAMP content during stimulation of motility by methylxanthines (Garbers *et al.*, 1971; 1973; Hoskins *et al.*, 1975) and other factors (reviewed by Tash & Means, 1983) it appears that a positive relationship exists between sperm cAMP content and motility stimulation with methylxanthines.

In addition to phosphodiesterase inhibition, however, the methylxanthines exhibit at least two other major pharmacological actions in various tissues, namely facilitation of  $Ca^{2+}$  mobilisation and direct antagonism of adenosine receptors (Bergstrand, 1980; Fredholm *et al.*, 1979; Rall, 1980; Svedmyr & Simonsson, 1978). The role of adenosine receptor interaction in the movement of sperm has not been investigated, but the effects of externally applied  $Ca^{2+}$  and internal  $Ca^{2+}$  have received considerable

**Table 1** Effect of caffeine, theophylline, lysine theophylline and IBMX on human sperm motility.

Caffeine (n = 9)			Theophylline (n = 6)			Lysine theophylline (n = 8)			IBMX (n = 10)		
Concentration (M)	Motility (% basal $\pm$ s.d.)	P	Concentration (M)	Motility (% basal $\pm$ s.d.)	P	Concentration (M)	Motility (% basal $\pm$ s.d.)	P	Concentration (M)	Motility (% basal $\pm$ s.d.)	P
$1.0 \times 10^{-4}$	102.2 $\pm$ 6.0		$1.0 \times 10^{-4}$	98.3 $\pm$ 6.4		$1.0 \times 10^{-4}$	99.7 $\pm$ 4.8		$1.0 \times 10^{-7}$	101.7 $\pm$ 1.6	<0.05
$3.3 \times 10^{-4}$	103.1 $\pm$ 4.5	<0.05	$3.3 \times 10^{-4}$	103.0 $\pm$ 5.6		$2.0 \times 10^{-4}$	100.8 $\pm$ 5.4		$1.0 \times 10^{-6}$	103.8 $\pm$ 4.1	<0.05
$1.0 \times 10^{-3}$	104.6 $\pm$ 11.4		$1.0 \times 10^{-3}$	107.3 $\pm$ 2.9	<0.05	$5.0 \times 10^{-4}$	102.2 $\pm$ 6.5		$1.0 \times 10^{-5}$	109.8 $\pm$ 4.1	<0.01
$3.3 \times 10^{-3}$	110.0 $\pm$ 9.6	<0.01	$3.3 \times 10^{-3}$	112.3 $\pm$ 6.9	<0.05	$1.0 \times 10^{-3}$	108.1 $\pm$ 11.6	<0.05	$1.0 \times 10^{-4}$	112.2 $\pm$ 3.2	<0.01
$1.0 \times 10^{-2}$	113.2 $\pm$ 9.3	<0.01	$1.0 \times 10^{-2}$	112.0 $\pm$ 7.8	<0.05	$2.0 \times 10^{-3}$	109.8 $\pm$ 11.3	<0.05	$1.0 \times 10^{-3}$	112.2 $\pm$ 9.5	<0.01
						$5.0 \times 10^{-3}$	116.7 $\pm$ 7.6	<0.01			
						$1.0 \times 10^{-2}$	102.1 $\pm$ 10.7				

*P* = significance level of increase above basal compared with a paired *t*-test.)



**Figure 1** Effect (mean  $\pm$  s.e. mean) of IBMX ( $\bullet$ ,  $n=10$ ), theophylline ( $\circ$ ,  $n=6$ ) and caffeine ( $\blacksquare$ ,  $n=9$ ) on human sperm motility.

attention. A correlation between the dependency of sperm on  $\text{Ca}^{2+}$  for initiation of motility and the epididymal  $\text{Ca}^{2+}$  concentration has been found (Morton *et al.*, 1978). This relationship, however, is species specific; in some species external  $\text{Ca}^{2+}$  appears to play an inhibitory role (McGrady *et al.*, 1974). Maintenance of low intracellular  $\text{Ca}^{2+}$  levels seems to be important in many species for optimum motility (Tash & Means, 1983) and the presence of  $\text{Ca}^{2+}$  pumps in the plasma membrane support this concept (Peterson *et al.*, 1979). Furthermore, stimulation of  $\text{Ca}^{2+}$  pump activity may be mediated through cAMP following the stimulation of adenylate cyclase by  $\text{Ca}^{2+}$  (Garbers & Kopf, 1980), and stimulation of the acrosome reaction.

In view of the important effects of  $\text{Ca}^{2+}$  on sperm motility it should be noted that in this study measurements were conducted with a total  $\text{Ca}^{2+}$  concentration estimated to be between  $5.0 \times 10^{-3}$  M at the start of incubation to  $2.4 \times 10^{-4}$  M during the measurement. This inference is valid if it is assumed that free  $\text{Ca}^{2+}$  equilibrates across the membrane separating the two chambers in the incubation apparatus. The free  $\text{Ca}^{2+}$  concentration, however, would be influenced by the degree of interaction between  $\text{Ca}^{2+}$  and proteins of seminal plasma. It is difficult, therefore, without accurate measurements of the free  $\text{Ca}^{2+}$  concentrations to interpret our findings in terms of involvement of methylxanthine effects on  $\text{Ca}^{2+}$  mobilisation. It is possible that the reduction of free  $\text{Ca}^{2+}$  concentration during the 2 h incubation period

assisted the effect of the methylxanthines on sperm motility.

The considerably higher potency of IBMX compared with caffeine and theophylline observed in this study, may reflect the large difference in phosphodiesterase inhibitory activity of these compounds. IBMX has been shown to be a more potent inhibitor of phosphodiesterase in many tissues. Little is known, however, of the relative potencies of these compounds to directly increase intracellular free  $\text{Ca}^{2+}$  levels and, therefore, the possibility that a proportion of the difference in capacity of these compounds to stimulate sperm motility can be attributed to effects on  $\text{Ca}^{2+}$  or adenosine activity cannot be excluded.

Caffeine has been demonstrated to enhance the fertilising ability of human sperm (Rogers 1981; Aitken *et al.*, 1983) and could, therefore, be of clinical benefit in men with reduced sperm motility. Aitken *et al.* (1983) reported, however, that increases in motility and fertilising ability of human sperm were dependent on consistent presence of caffeine and were not present in sperm washed free of stimulatory concentrations of caffeine; indicating that caffeine must be present at the point of conception to maintain enhanced fertilising ability. Caffeine has been reported to have teratogenic potential (Fujii & Nishimura, 1974; Morris & Weinstein, 1981; Nishimura & Nakal, 1960) and other methylxanthines could have teratogenic properties limiting their clinical use in improving sperm motility and fertilising ability.

Further studies with compounds having specific effects on cAMP and Ca<sup>2+</sup> activity should, however, yield a better understanding of the mechanisms governing sperm motility and thus increase our capacity to influence sperm motility in either direction for purposes of fertility control or contraception.

## References

- Aitken, R. J. (1983). Influence of caffeine on movement characteristics, fertilising capacity and ability to penetrate cervical mucus of human spermatozoa. *Reproduction and Fertility*, **67**, 19–27.
- Bergstrand, H. (1980). Phosphodiesterase inhibition and theophylline. *Eur. J. resp. Dis.*, **61**, (suppl. 109), 37–44.
- Burge, R. G. (1973). Caffeine-stimulation of ejaculated human spermatozoa. *Virology*, **1**, 371.
- Davis, C. W. & Kuo, J. F. (1978). Differential effects of nucleotides and their analogs and various agents on cyclic GMP-specific and cyclic AMP-specific phosphodiesterases purified from guinea pig lung. *Biochem. Pharmacol.*, **27**, 89–95.
- De Turner, E. A., Aparicio, N. J., Turner, D. & Schwarzstein, L. (1978). Effect of two phosphodiesterase inhibitors, cyclic adenosine 3':5'-monophosphate and a beta-blocking agent on human sperm motility. *Fertility and Sterility*, **29**, 328.
- Fredholm, B. B., Brodin, K. & Strandberg, K. (1979). On the mechanism of action of relaxation of the tracheal muscle by theophylline and other cyclic nucleotide phosphodiesterase inhibitors. *Acta Pharmac. Tox. (Kbh.)*, **45**, 336–344.
- Fujii, O. & Nishimura, H. (1974). Reduction in frequency effects of caffeine in mice by pre-treatment with propranolol. *Teratology*, **10**, 149.
- Garbers, D. L., Lust, W. D., First, N. L. & Lardy, H. A. (1971). Effects of phosphodiesterase inhibitors and cyclic nucleotides on sperm respiration and motility. *Biochemistry*, **10**, 1825–1831.
- Garbers, D. L., First, N. L. & Lardy, H. A. (1973). The stimulation of bovine epididymal sperm metabolism by cyclic nucleotide phosphodiesterase inhibitors. *Biology of Reproduction*, **8**, 589–598.
- Garbers, D. L. & Kopf, G. S. (1980). The regulation of spermatozoa by calcium cyclic nucleotides. *Advances In Cyclic Nucleotide Research*, **13**, 251–306.
- Glass, W. F. & Moore Jr, J. B. (1979). Inhibition of human lung cyclic GMP and cyclic AMP phosphotides, and pharmacological phosphodiesterase inhibitors. *Biochem. Pharmacol.*, **28**, 1107.
- Haesungcharern, A. & Chulavatnatol, M. (1973). Stimulation of human spermatozoa motility by caffeine. *Fertility and Sterility*, **24**, 662.
- Hartmann, R., Steiner, R., Hofmann, N. & Kaufmann. (1983). Human sperm motility-enhancement and inhibition measured by laser doppler spectroscopy. *Andrologia*, **15**, 120–134.
- Helfman, D. M. & Kuo, J. F. (1982). Differential effects of various phosphodiesterase inhibitors, pyrimidine and purine compounds, and inorganic phosphates on cyclic CMP, cyclic AMP and cyclic GMP phosphodiesterases. *Biochem. Pharmacol.*, **31**, 43–47.
- Hicks, J. J., Martinez-Manautou, J., Pedron, N. & Rosado, A. (1972). Metabolic changes in human spermatozoa related to capacitation. *Fertility and Sterility*, **23**, 172–179.
- Hommonai, Z. T., Paz, G., Sofer, A., Kraicer, P. F. & Harrell, A. (1976). Effect of caffeine on the motility, viability, oxygen consumption and glycolytic rate of ejaculated human normokinetic and hypokinetic spermatozoa. *Int. J. Fertility*, **21**, 163–170.
- Hong, C. Y., Chaput de Saintonge, D. M. & Turner, P. (1981). A simple method to measure drug effects on human sperm motility. *Br. J. clin. Pharmacol.*, **11**, 385–387.
- Hoskins, D. D., Hall, M. L. & Munstermann, D. (1975). Induction of motility in immature bovine spermatozoa by cyclic AMP phosphodiesterase inhibitors and seminal plasma. *Biology of Reproduction*, **131**, 168–176.
- Johnston, A., Hedges, A., Freedman, A., Aldhous, M. E., Weerasuriya, K. & Turner, P. (1983). The pharmacokinetics of lysine-theophylline, a new soluble theophylline, in human volunteers. *Postgrad. med. J.*, **59**, 86.
- Levin, R. M., Shofer, J. & Greenberg, S. H. (1980). A quantitative method for determining the effects of drugs on spermatozoal motility. *Fertility and Sterility*, **33**, 631–635.
- McGrady, A. V., Nelson, L. & Ireland, M. (1974). Ionic effects on the motility of bull and chimpanzee spermatozoa. *J. of Reproduction and Fertility*, **40**, 71–76.
- Makler, A., Makler, E., Itzkovitz, J. & Brandes, J. M. (1980). Factors affecting sperm motility. IV. Incubation of human semen with caffeine, kallikrein, and other metabolically active compounds. *Fertility and Sterility*, **33**, 624.
- Morris, M. B. & Weinstein, L. (1981). Caffeine and the foetus—is trouble brewing? *Am. J. Obstet. Gynaecol.*, **140**, 607–610.
- Morton, B. E., Sagradaca, R. & Fraser, C. (1978). Sperm motility within the mammalian epididymus: Species variation and correlation with free calcium levels in epididymal plasma. *Fertility and Sterility*, **29**, 695–698.

- Nishmura, M. B. & Nakal, K. (1960). Congenital malfunctions in offspring of mice treated with caffeine. *Proc. Soc. exp. Biol. Med.*, **104**, 140.
- Peterson, R. N., Seyler, D., Bundman, D. & Freund, M. (1979). The effect of theophylline and dibutyryl cAMP on the uptake of radioactive calcium and phosphate ions by boar and human spermatozoa. *J. Reprod. Fertil.*, **55**, 385-390.
- Rall, T. W. (1980). Central nervous system stimulants. The xanthines. In *The pharmacological basis of therapeutics*, eds. Goodman Gilman, A., Goodman, L. S. & Gilman, A., New York: Macmillan Publishing Co.
- Rogers, B. J. (1981). Factors affecting mammalian *in vitro* fertilization. In *Bioregulators of reproduction*, eds. Jagiello, G. & Vogel, H. J., pp. 459-486. New York: Academic Press.
- Schoenfeld, C., Richad, D., Amelar, S. & Dubin, L. (1973). Stimulation of ejaculated human spermatozoa by caffeine: preliminary report. *Fertility and Sterility*, **24**, 772.
- Svedmyr, N. & Simonsson, B. G. (1983). Drugs in the treatment of asthma. *Pharmacol. Ther. (B)*, **13**, 397-440.
- Tash, J. S. & Means, A. R. (1983). Cyclic adenosine 3',5' monophosphate, calcium and protein phosphorylation in flagellar motility. *Biology of Reproduction*, **28**, 75-104.

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