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IgE- and IgE+Ag-mediated mast cell migration in an autocrine/paracrine fashion

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Abstract

Mast cells are the major effector cells for immediate hypersensitivity and chronic allergic reactions. These cells accumulate in mucosal tissues of allergic reactions, where immunoglobulin E (IgE) is produced locally. Here we provide evidence that, in addition to antigen that can attract IgE-bound mast cells, the type of IgE molecules that efficiently activate mast cells can promote the migration of mast cells in the absence of antigen. IgE and IgE+Ag-mediated migration involves an autocrine/paracrine secretion of soluble factors including adenosine, leukotriene B₄, and several chemokines. Their secretion depends on 2 tyrosine kinases, Lyn and Syk, and they are agonists of G-protein-coupled receptors and signal through phosphatidylinositol 3-kinase γ , leading to mast cell migration. In mouse experiments, naive mast cells are attracted to IgE, and IgE-sensitized mast cells are attracted to antigen. Therefore, IgE and antigen are implicated in mast cell accumulation at allergic tissue sites with local high IgE levels.

Introduction

Mast cells are the major effector cell type in immunoglobulin E (IgE)-mediated immediate hypersensitivity, chronic allergic diseases, and the defense against certain parasites and bacteria. Traditionally, it is thought that mast cells bound to antigen-specific IgE via the high-affinity receptor (Fc ϵ ;RI) encounter multivalent antigen (the stimulation mode hereafter termed IgE+Ag), and then IgE-bound receptors are aggregated, leading to cellular activation.¹ Activated mast cells secrete preformed and newly synthesized proinflammatory mediators, such as histamine, proteases, lipids, cytokines, and chemokines.

The mouse Fc ϵ ;RI consists of an IgE-binding α subunit, a β subunit, and 2 molecules of signal-generating γ subunit.² Fc ϵ ;RI aggregation leads to phosphorylation of the immunoreceptor tyrosine-based activation motifs (ITAMs) of the β and γ subunits by Lyn, and recruitment of Syk to the tyrosine-phosphorylated ITAMs of the γ subunit results in the activation of this protein-tyrosine kinase (PTK).³ Activated Lyn and Syk as well as Fyn eventually lead to the activation of multiple signaling pathways, including phosphatidylinositol 3-kinase (PI3K),

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phospholipase C- γ /Ca²⁺/protein kinase C (PKC), and mitogen-activated protein (MAP) kinases.⁴

Cell–extracellular matrix interactions mediated by integrins play a critical role in multiple cellular functions including cell adhesion and migration. Activation of mast cells by Fc ϵ RI aggregation and stem cell factor (SCF) induces adhesion to fibronectin (FN)⁵ predominantly via integrin α 5 β 1.^{6–8} Upon Fc ϵ RI aggregation and SCF stimulation, FN-adherent cells exhibit stronger effector functions such as histamine release and cytokine production than nonadherent cells.⁷

We and others recently demonstrated that monomeric IgE can promote mast cell survival.^{9, 10} This observation, together with earlier studies showing that IgE in the absence of antigen can increase the surface expression of Fc ϵ RI,^{11–13} has transformed the traditional view of IgE–mast cell binding as a “sensitization” step prior to receptor aggregation with antigen or other crosslinking reagents into a new one that monomeric IgE can induce survival and “activation” of mast cells.¹⁴ We also found that IgE molecules display heterogeneity in that different IgE molecules induce varied levels of activation; at one extreme end of the spectrum, some IgE molecules, termed highly cytokinergic (HC) IgEs, induce the production and secretion of various cytokines and other activation events including degranulation, whereas other IgE molecules, termed poorly cytokinergic (PC), do so very inefficiently.¹⁵

Mast cells accumulate at local inflammatory mucosal tissues, as seen in allergic rhinitis and asthma. Interestingly, class switch recombination and somatic hypermutation of the immunoglobulin gene and eventually IgE synthesis and secretion occur at such inflammatory mucosae.^{16–21} In an allergic individual, local IgE production persists for a long period in the absence of allergen.²⁰ A variety of biologic agents, including growth factors (eg, SCF), chemokines (MCP-1/CCL2, MIP-1 α /CCL3, RANTES/CCL5, eotaxin/CCL11, SDF-1 α /CXCL12, etc, refer to “Chemokine/chemokine receptor nomenclature”²² for the nomenclature of chemokines), and adenosine nucleotides, are known to attract rodent mast cells.^{23–27} Consistent with effects of the chemokines on mast cell migration, mast cells express appropriate receptors including CCR1–5, CXCR1–2, and CXCR4.^{28–32} Antigen can also cause the migration of IgE-sensitized mast cells, which can be suppressed by inhibitors of Rho-kinase/ROCK and p38.³³ In this study, we have found that, in addition to IgE+Ag, HC IgEs can attract mast cells. HC IgE- and IgE+Ag-induced mast cell migration involves autocrine/paracrine secretion of soluble factors. The initial phase of migration leading to the release of such factors depends on Lyn and Syk, and the following phase downstream of G-protein–coupled receptor (GPCR) stimulation by these soluble factors requires PI3K γ . In vivo mouse experiments also suggest that mast cells can be attracted to HC IgEs as well as antigen. Therefore, this study provides a novel mechanism for mast cell accumulation to allergic inflammatory sites.

Materials and methods

Antibodies and other reagents

IgE antibodies used were described previously.¹⁵ Antitrinitrophenyl (TNP) IgG and antibodies against integrins were purchased from BD Biosciences Pharmingen (San Diego, CA); MIP-1 α , MCP-1, and RANTES were from R&D Systems, Minneapolis, MN; fibronectin, vitronectin, laminin, collagen type IV, dinitrophenyl (DNP)–lysine, adenosine, histamine, serotonin, and MRS 1523 were from Sigma, St Louis, MO; PP2, piceatannol, wortmannin, LY294002, PD98059, SP600125, Gö-6976, PKI, KT5720, Y-27632, cytochalasin D, EGTA [ethylene glycol-bis(beta-aminoethyl ether)-N,N,N',N'-tetraacetic acid], and leukotriene B₄ (LTB₄) were from EMD Biosciences (San Diego, CA); N-oleoyl dopamine, U-75302, and LY2552833 were from Cayman Chemical (Ann Arbor, MI). SCF and ER-27319 were gifts

from Kirin Brewery (Tokyo, Japan) and Eisai (Tokyo, Japan), respectively. Terreic acid was described previously.³⁴

Cells and stimulation

Bone marrow cells from wild-type and mutant mice were cultured in interleukin 3 (IL-3)-containing medium for 4 to 6 weeks to generate mast cells (BMMCs) with more than 95% purity (c-Kit⁺ Fc ϵ RI⁺ by flow cytometry). The following mutant mice were used: *Fc ϵ RI α ^{-/-}*, *lyn^{-/-}*, *fyn^{-/-}*, *hck^{-/-}*, *syk^{-/-}*, *blk^{-/-}*, *PKC β ^{-/-}*, *PKC ϵ ^{-/-}*, *PKC θ ^{-/-}*, and *PI3K γ ^{-/-}*. For IgE+Ag stimulation, BMMCs were sensitized by overnight incubation with 0.5 μ g/mL H1- ϵ -DNP-206 (206) IgE. BMMCs washed twice with buffer were used for migration assays. Animal studies were approved by the institutional review board of La Jolla Institute for Allergy and Immunology, San Diego, CA.

In vitro migration assay

BMMC migration for 8 hours was assayed using 24-well Transwell chambers (Corning, Corning, NY) separated by 5- μ m polycarbonate filters, the lower surface of which had been coated overnight with 20 μ g/mL fibronectin and followed by blocking with 4% bovine serum albumin (BSA) in phosphate-buffered saline (PBS) at 37°C for 1 hour, unless otherwise described. Under standard conditions, upper wells contained 10⁶ cells/well in 0.2 mL medium consisting of RPMI/1% BSA/20 mM HEPES [N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid] (pH 7.4) and lower wells contain 0.6 mL medium. Cells migrated into lower wells were counted using a hemocytometer. For inhibition with antibodies, BMMCs were first preincubated with neutralizing antibodies (20 μ g/mL-50 μ g/mL) for 30 minutes, and then placed in upper wells. Both upper and lower wells contained neutralizing antibodies.

In some experiments, BMMCs were prestained with 10 μ M 5- (and 6-) carboxyfluorescein diacetate, succinimidyl ester (CFSE; Molecular Probes, Eugene, OR) in 0.1% BSA/PBS at 37°C for 10 minutes before loading onto upper wells. Cells migrated into lower wells were quantified using flow cytometry.

In vivo mast cell accumulation

Gauze strips (~7 layers, 10 \times 15 mm²) spotted with 50 μ L of 10 μ g/mL SPE-7 IgE or PBS were applied to the shaved back skin of naive NC/Nga mice for 24 hours. This portion of the back skin was occluded with Tegaderm Transparent Dressing (3M, St. Paul, MN) and BAND-AID (Johnson & Johnson, New Brunswick, NJ). Mice were killed and skin regions were prepared to stain mast cells with toluidine blue. Mast cell numbers were counted under a microscope. In another type of in vivo experiment, mice were first treated similarly with 100 μ L of 10 μ g/mL anti-DNP (206) IgE or PBS into the back skin, and 1 day later the same area was applied with 50 μ L of 100 ng/mL DNP-human serum albumin (HSA) or PBS for 24 hours. For mast cell quantification, dorsal skin samples were fixed in 10% formaldehyde, paraffin embedded, and cut into 6- μ m sections. Deparaffinized sections were stained with toluidine blue (pH 4.0) and analyzed by light microscopy. Cells between epithelium and panniculus carnosus were counted at a magnification of \times 400. Images were acquired with a Nikon Optiphot photomicroscope (Nikon, Tokyo, Japan) with a Fluor 40 \times /1.30 NA or Plan 10 \times /0.25 NA objective, using a DVC-1310 camera (DVC Company, West Austin, TX) and DVC C-View v2.2 software. Immersion oil type FF (Cargille Laboratories Inc, Cedar Grove, NJ) was used for a magnification at 400 \times .

Data supplements

Four figures are provided as supplemental materials, which are available on the Blood website (see the Supplemental Figures link at the top of the online article): effects of IgE/antigen and

IgG/antigen complexes on mast cell migration (Figure S1), IgE- and IgE+Ag-induced migration of mouse mast cell lines (Figure S2), requirements of fibronectin, vitronectin, or laminin (Figure S3), and autocrine/paracrine mechanism of mast cell migration (Figure S4).

Results

HC IgEs can induce mast cell migration

Given the ability of IgEs to induce mast cell activation,¹⁵ we examined whether IgEs can promote the migration of mouse BMMCs in the absence of antigen. As shown in Figure 1A, 3 HC IgEs in a lower well induced vigorous migration of BMMCs from an FN-coated upper well after 8 hours of incubation in a transwell assay, whereas typical PC IgEs such as H1- ϵ -DNP-206 (206) and IgE-3 did not. Dose-response experiments (Figure 1B) indicated that the extent of migration induced by a typical HC IgE (SPE-7) increases in a dose-dependent manner up to 30 μ g/mL. A significant migration was observed at 1 μ g/mL IgE, a concentration reachable in a subset of atopic conditions.^{35,36} Kinetic experiments (Figure 1C) showed that HC IgE-induced migration is smaller in magnitude than that induced by SCF, and plateaus within 10 to 15 hours. Consistent with previous studies,^{33,37} migration with similar levels and kinetics was also observed with BMMCs in an upper well that had been sensitized with anti-DNP IgE (206) and moved toward antigen, DNP-HSA (Figure 1A-C).

HC IgE-induced migration signals through the Fc ϵ RI and integrins

We next confirmed that IgE-induced migration is mediated through Fc ϵ RI. BMMCs from Fc ϵ RI $\alpha^{-/-}$ mice³⁸ failed to migrate in response to HC IgEs or IgE+Ag, whereas the mutant cells were attracted to SCF as vigorously as wild-type cells (Figure 1D). Consistent with the notion that these IgE-dependent migrations are triggered by Fc ϵ RI aggregation, these migrations were inhibited by monovalent hapten, DNP-lysine (Table 1). Further, BMMC migration was induced by incubation with PC IgE plus DNP-HSA, conditions that mimic acute allergic situations in that antigens are present around IgE-bound mast cells and are different from IgE+Ag in that unbound IgE molecules were removed (IgE+Ag) or not (PC IgE plus DNP-HSA) (Figure S1A). By contrast, BMMC migration was not induced by IgG, IgM (J.K. and T.K., unpublished data, February 2004), or anti-TNP IgG plus TNP-BSA (Figure S1B). Efficient HC IgE-dependent migration was seen with BMMCs and mouse mast cell lines (MC/9 and MCP-5) (Figure S2) incubated in upper wells coated with FN, vitronectin, or laminin (Figure S3). However, collagen or BSA coating did not promote BMMC migration efficiently. These results implicated the involvement of integrins in HC IgE- and IgE+Ag-mediated mast cell migration. This was confirmed by the strong inhibition of migration with a neutralizing antibody against integrin β 1. β 2 and β 7 integrins may play a minor, if any, role in IgE-dependent mast cell migration (Figure 1E).

Comparison between HC IgE- and chemokine-induced migration

Several chemokines are known to attract mast cells.^{26,39,40} Remarkably, migration induced by 10 μ g/mL SPE-7 IgE was more pronounced than that induced by mast cell-attracting chemokines, RANTES, MCP-1, and MIP-1 α (Figure 2A). When used in combinations, HC IgEs did not synergize to induce mast cell migration with SCF or any of the tested chemokines, and HC IgEs determined the extent of migration (Figure 2B-C), suggesting a hierarchically dominant role for HC IgEs in complex situations where various chemoattractants are present. Signaling hierarchy was well known among several chemoattractants of neutrophils.⁴¹ An alternative possibility is that the mediators released by HC IgE-activated mast cells (for example, proteases) inactivate SCF, chemokines, or their receptors. Unlike SCF or the chemokines, which induce the movement of mast cells toward the increasing gradient of chemoattractant concentrations (chemotaxis), checkerboard analysis indicated that HC IgEs induce a mixture of directional and nondirectional (chemokinesis) movements (Figure 2D-E).

Signaling requirements for HC IgE- and IgE+Ag-induced migration

Fcε;RI stimulation by IgE+Ag triggers activation of nonreceptor PTKs of Src, Syk, and Tec families and several signaling pathways.^{3,4,14} Pharmacologic inhibition suggested that HC IgE- and IgE+Ag-induced migration requires Src and Syk kinases, but not Tec kinases (Table 1). Consistent with these data, IgE-dependent migration was abrogated in *syk*^{-/-} cells (Figure 3B), but was not affected by Btk deficiency (Figure 3C). Among the tested Src PTKs, Lyn and Fyn played major and minor roles, respectively, in IgE- and IgE+Ag-dependent migration, whereas Hck played no part in this function (Figure 3A). These pharmacologic experiments (Table 1) and previous genetic and biochemical experiments^{10,15} indicate that signaling events induced by HC IgEs are almost the same as those induced by IgE+Ag. Downstream of these kinases, PI3K and some MAP kinases (p38 and JNK) appeared essential from pharmacologic experiments. A general inhibitor (Ro31-8425) of PKC, an inhibitor (Gö-6976) of Ca²⁺-dependent PKC isoforms and a Ca²⁺ chelator (EGTA) all inhibited IgE-dependent migration whereas inhibitors of protein kinase A or Rho-kinase did not. Consistent with these results, deficiency of PKCβ, PKCε, or PKCθ modestly reduced IgE-dependent migration (Figure 3D).

Involvement of autocrine/paracrine soluble factors, GPCRs, and PI3Kγ in HC IgE- and IgE+Ag-induced mast cell migration

To test potential involvement of autocrine/paracrine secretion in migration, we examined whether *Fcε;RIα*^{-/-} mast cells that cannot directly respond to IgE can migrate in the presence of wild-type cells. A significant proportion of CFSE-labeled *Fcε;RIα*^{-/-} mast cells mixed at a ratio of 1:1 with nonlabeled wild-type cells in upper wells migrated to lower wells in response to HC IgE and IgE+Ag (Figure S4). Further, *Fcε;RIα*^{-/-} cells in upper wells were attracted to lower wells that contained wild-type cells in the presence of HC IgE or anti-DNP IgE-sensitized wild-type cells in the presence of DNP-HSA (Figure 4A). More directly, supernatants of wild-type mast cells cultured with HC IgE or IgE+Ag for 0.5, 1, 2, or 6 hours could attract *Fcε;RIα*^{-/-} mast cells with similar efficiency (Figure 4B). These results strongly indicate that *Fcε;RIα*^{-/-} cells alone or in the mixed cultures with wild-type cells migrated to lower wells that contained HC IgE- or IgE+Ag-induced soluble factors secreted from wild-type cells.

A variety of chemicals and peptides are released from IgE+Ag- and HC IgE-stimulated mast cells within 30 minutes and exert their functions on multiple cell types including mast cells themselves.^{10,15,42} For example, histamine (and serotonin) mediates increased vascular permeability in passive cutaneous anaphylaxis, and LTB₄ and sphingosine 1-phosphate induce the migration of various leukocytes.⁴³⁻⁴⁶ Several chemokines such as MCP-1, MIP-1α, MIP-1β/CCL4, and MIP-2 are produced by BMDCs stimulated weakly by IgE+Ag,⁴⁷ and MCP-1 secretion can be induced by fast dissociating antigens and characterized as an exception of the kinetic proofreading regimen.⁴⁸ Like mast cell-produced chemokines such as MCP-1 and MIP-1α (Figure 2), adenosine and LTB₄ vigorously induced migration of BMDCs whereas histamine, serotonin, or sphingosine 1-phosphate did not (Figure 4C and J.K., T.K., and T.K., unpublished data, June 2004). Adenosine and LTB₄ as well as chemokines are ligands of GPCRs. Consistent with this, migration of *Fcε;RIα*^{-/-} BMDCs to supernatants of wild-type mast cells that had been cultured with HC IgE or IgE+Ag was blocked by approximately 60% by pertussis toxin, indicating that some soluble factors use Gαi-coupled receptors (Figure 4D). The role of adenosine in the migration was confirmed by the inhibition with adenosine deaminase that converts adenosine to inosine and an adenosine A₃ receptor inhibitor, MRS 1523 (Figure 4D). The involvement of LTB₄ was indicated by the inhibition with a 5-lipoxygenase inhibitor (N-oleoyl dopamine) and inhibitors of LTB₄ receptors BLT₁ (U-75 302) and BLT₂ (LY2 552 833) (Figure 4E). Parenthetically, these inhibitors of LTB₄ synthesis or receptors did not affect SCF-induced migration (T.K. and T.K., unpublished data, June 2004). Furthermore, roles of mast cell-produced chemokines in the migration were shown by the

inhibition by neutralizing antibodies. Antibodies to MIP-1 α , MIP-1 β , and RANTES individually inhibited weakly (10%-25%) but in a combination inhibited HC IgE- and IgE+Ag-induced mast cell migration more strongly (~50%; Figure 4F). Adenosine, LTB₄, and chemokines are all agonists of GPCRs and signal through PI3K γ .⁴⁹ Strikingly, HC IgE- and IgE+Ag-induced mast cell migration was drastically reduced in PI3K γ -deficient cells (Figure 5A). Therefore, it appears that IgE- and IgE+Ag-dependent migration results from the signals originated from Fc ϵ RI aggregation that lead to a rapid release of multiple GPCR agonists including adenosine, LTB₄, and several chemokines. Thus, activation of the GPCRs and their downstream signaling molecule, PI3K γ , is crucial for mast cell migration.

We next examined which step, Fc ϵ RI stimulation-induced release of GPCR agonists or downstream signaling from GPCRs, is dependent on Lyn and Syk. For this purpose, we tested whether 5-(and -6)-carboxyfluorescein diacetate, succinimidyl ester (CFSE)-labeled Fc ϵ RI $\alpha^{-/-}$ BMMCs in upper wells migrate to lower wells containing wild-type, *lyn*^{-/-}, or *syk*^{-/-} BMMCs in the presence of SPE-7 IgE or SCF. In the presence of SPE-7 IgE, wild-type BMMCs attracted Fc ϵ RI $\alpha^{-/-}$ cells vigorously, but the migration of the latter cells was abrogated toward SPE-7 IgE-incubated *lyn*^{-/-} and *syk*^{-/-} cells, although Fc ϵ RI $\alpha^{-/-}$ cells migrated vigorously to SCF irrespective of the genotype of mast cells present in the lower wells (Figure 5B). Similar results were obtained when CFSE-labeled Fc ϵ RI $\alpha^{-/-}$ cells in upper wells were cultured with lower wells containing 206 IgE-sensitized mutant cells in the presence of antigen or supernatants of 206 IgE-sensitized mutant cells that had been cultured in the presence of antigen (J.K. and T.K., unpublished data, January 2004). Therefore, these results indicate that Lyn and Syk are required for HC IgE- and IgE+Ag-induced release of soluble factors.

We next examined whether Lyn and Syk are required for signaling downstream of GPCRs as well. For this purpose, we tested whether wild-type, *lyn*^{-/-}, and *syk*^{[unk]^{-/-} BMMCs in upper wells are attracted to lower wells containing 206 IgE-sensitized wild-type cells in the presence of antigen or supernatants of 206 IgE-sensitized wild-type cells cultured in the presence of antigen. The mutant cells migrated as vigorously as wild-type cells under these conditions (Figure 5C and J.K. and T.K., unpublished data, December 2003). Furthermore, the mutant cells were also attracted to adenosine, LTB₄, MCP-1, and MIP-1 α as efficiently as wild-type cells (J.K. and T.K., unpublished data, January 2004), consistent with our previous study showing that Syk is not required for mast cell migratory responses to GPCR stimuli.⁵⁰ Therefore, we conclude that Lyn and Syk are required for the initial phase of mast cell activation leading to the release of GPCR agonists, but not for the late phase of GPCR activation and further downstream events, in HC IgE- and IgE+Ag-induced migration.}

In vivo mouse models for IgE- and IgE+Ag-induced mast cell accumulation

To evaluate the pathophysiologic relevance of our in vitro observations on IgE- and IgE/Ag-induced mast cell migration, we performed in vivo mouse experiments. First, we tested whether epicutaneous application of IgE on the back skin of naive mice can attract mast cells. Microscopic analysis of toluidine blue-stained samples showed that the local density of mast cells in SPE-7 IgE-applied areas was significantly increased after 24 hours compared with that in PBS-applied areas, whereas a modest increase was observed in 206 IgE-treated areas (Figure 6A-B). Most mast cells were localized in perivascular regions of the dermis. SPE-7 IgE-applied areas contained approximately 10% mast cells that had degranulated whereas such degranulating mast cells were very rare in PBS- or 206 IgE-applied areas. Another type of in vivo experiment was designed to test whether antigen can induce mast cell migration in vivo. Mice were epicutaneously treated with anti-DNP (206) IgE or PBS into the back skin, and 1 day later the same area was applied with DNP-HSA or PBS. Microscopic analysis showed that mast cell numbers in IgE/antigen-applied areas are much higher after 24 hours of treatment

than those in PBS/PBS-, PBS/antigen-, or IgE/PBS-applied areas (Figure 6C). Similar to the effects of SPE-7 IgE on mast cell accumulation, antigen-treated areas had degranulating mast cells. Although these *in vivo* experiments do not reveal the mechanism of mast cell accumulation, the perivascular location of these cells suggests that mast cells or their precursors were recruited from the circulation rather than from nearby tissues. Alternatively, mast cells might have proliferated *in situ*, although this is less likely because of the short assay time and the low capacity of mast cells to proliferate in response to HC IgE and IgE+Ag.

Discussion

This study provides evidence that HC IgEs in addition to IgE+Ag can promote the migration of mast cells. This migration is mediated mainly through integrin $\beta 1$ and is more potent than some chemokines, and involves 2 phases: an early phase of Lyn- and Syk-dependent release of multiple soluble factors including adenosine, LTB₄, and chemokines, and a later phase of PI3K γ -dependent signaling following the activation of GPCRs by these factors. These *in vitro* observations are consistent with *in vivo* mouse experiments in which mast cell accumulation was induced by epicutaneous application of HC IgE and IgE+Ag.

The *in vivo* results suggest that HC IgE and IgE+Ag can induce accumulation of mast cells without prior inflammation. However, it is conceivable that, in allergic individuals, some inflammatory reactions such as the infiltration of helper T cells have occurred when IgE synthesis in B cells takes place at mucosal sites in the nasal cavity and lung in response to antigen exposure.¹⁶⁻²¹ Given the vast variety of proinflammatory mediators secreted from activated mast cells,⁵¹ IgE- and IgE+Ag-induced mast cell accumulation would amplify inflammatory reactions by recruiting other cells such as T cells, eosinophils, monocytes, and neutrophils. For instance, histamine plays an important role in the pathogenesis of atopic asthma by enhancing the secretion of Th2 cytokines and inhibiting the production of Th1 cytokines.⁵² LTB₄ recruits T cells and myeloid cells,⁴³⁻⁴⁵ and mast cell-produced cytokines and chemokines can recruit T cells, eosinophils, monocytes, and neutrophils. CC chemokine transcripts coding for I-309/CCL1, MIP-1 α , MIP-1 β , and MCP-3/CCL7 are among the most dramatically enhanced ones in IgE+Ag-stimulated mast cells.⁵³ The ability of HC IgEs to attract mast cells suggests that this amplification of inflammation can last as long as local IgE synthesis continues, even after the elimination of antigen. Overall, IgEs in the absence as well as presence of allergen are implicated in mast cell accumulation at allergic tissue sites with local high IgE levels.

This and previous studies^{10,15} have shown that intracellular signaling events induced by HC IgEs and IgE+Ag are very similar, if not identical: Src and Syk family PTKs are activated, intracellular Ca²⁺ concentrations are increased, and several serine/threonine kinases such as MAP kinases, PKCs, and Akt are also activated. These signaling events induced by Fc ϵ ;RI aggregation by either HC IgEs or IgE+Ag (Table 1 and Kitaura et al¹⁵) result in a variety of biologic outcomes such as degranulation, histamine synthesis, leukotrienes release, receptor internalization, cytokine production, migration, and survival.^{9,10,15,54} Consistent with the similarities in signaling between HC IgE- and IgE+Ag-stimulated cells, migrations induced by these 2 modes of Fc ϵ ;RI stimulation require identical signaling molecules such as Lyn, Syk, and PI3K γ . Importantly, both migrations use soluble autocrine factors as a part of the migratory mechanism. Among numerous chemical and peptide agents rapidly secreted from activated mast cells, adenosine, LTB₄, and several chemokines were identified as the mediators for mast cell migration. Unlike RBL-2H3 rat mast cells,⁵⁵ sphingosine 1-phosphate did not substantially induce the migration of BMMC (Figure 4C). As discussed in the preceding paragraph, these mediators influence an *in vivo* inflammatory process by recruiting not only mast cells but also other types of inflammatory cells. Assuming that PI3K γ is involved only in signal transduction of GPCRs,⁴⁹ this autocrine/paracrine mechanism seems essential for

mast cell migration. Thus, only residual levels of migration were observed with *PI3K* $\gamma^{-/-}$ mast cells (Figure 5A). Notably, pertussis toxin inhibits HC IgE-, IgE+Ag-, and adenosine-induced migrations by approximately 60%. This indicates that these migrations are mediated mainly via *G α i*. However, other *G α* proteins or G protein-independent signals may also be involved; for example, integrins might supply additional signals for migration. In line with this possibility, *Lyn* is required for FN-mediated migration in RBL-2H3 cells.⁵⁶ In any event, this study has extended the previous findings that soluble factors including adenosine are used to amplify Ca^{2+} and degranulation responses in IgE+Ag-stimulated mast cells.^{49,55} *LTB $_4$* or chemokines do not induce degranulation. It will be interesting to investigate whether other biologic responses of mast cell activation depend on an autocrine/paracrine mechanism and what molecules mediate such responses. Our knowledge on this mechanism will expand our choices of anti-inflammatory drugs beyond currently used antihistamine and antileukotriene drugs that target this process.

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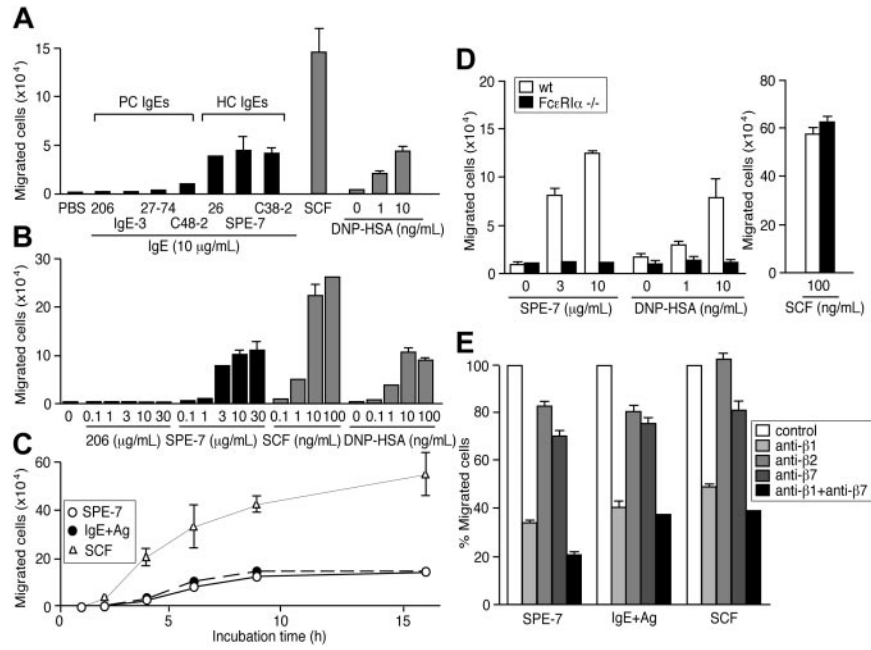


Figure 1. HC IgEs and IgE+Ag induce the migration of BMMCs through FcεRI and β1 integrin. (A) HC, but not PC, IgEs induce BMMC migration efficiently in an 8-hour transwell assay. BMMCs, sensitized overnight with anti-DNP IgE (206) and washed, were also attracted to a lower well containing DNP-HSA. For comparison, SCF (100 ng/ml) was also tested in the same assay. Also shown are dose-response (B) and time-course (C) experiments using 10 μg/mL 206 IgE, 10 μg/mL SPE-7 IgE, 10 ng/mL DNP-HAS, and 100 ng/mL SCF. (D) Migration assays were performed with *FcεRIα*^{-/-} and control BMMCs. (E) Migration of wild-type BMMCs was inhibited by neutralizing antibodies against integrins. Experiments were performed in triplicate. Representative data (± SD) are shown out of 2 or more experiments.

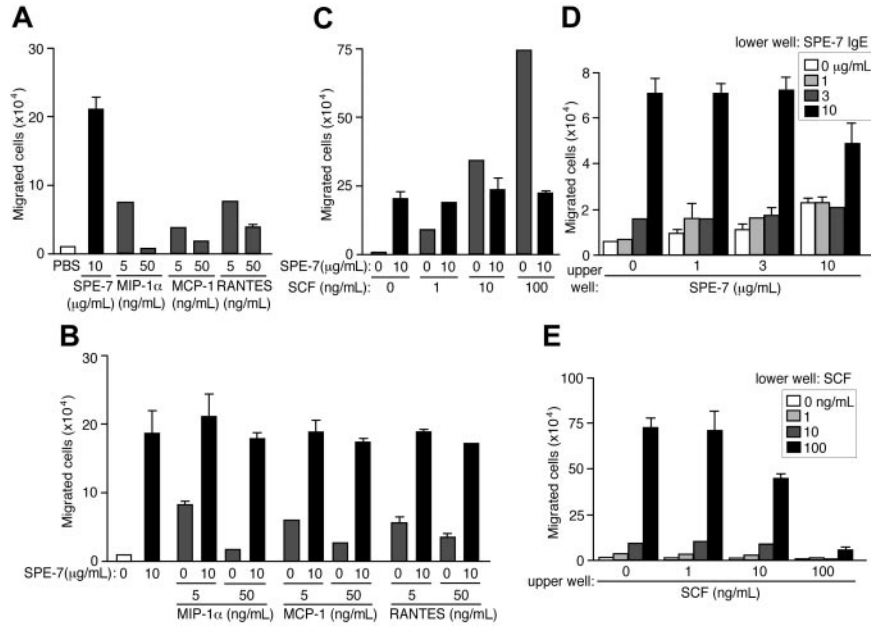


Figure 2. Comparison between HC IgE- and chemokine-induced migration of BMDCs and checkerboard analysis of HC IgE-induced mast cell migration. (A) SPE-7 IgE-induced migration of BMDCs was compared with those induced by mast cell-attracting chemokines, MIP-1α, MCP-1, and RANTES. (B-C) BMDC migration was induced by SPE-7 IgE in combination with chemokines or SCF. (D) BMDCs in upper wells containing various concentrations of SPE-7 IgE were incubated with various concentrations of SPE-7 IgE in lower wells. (E) BMDCs in upper wells containing various concentrations of SCF were incubated with various concentrations of SCF in lower wells. Values (mean ± SD) are shown out of 2 independent experiments.

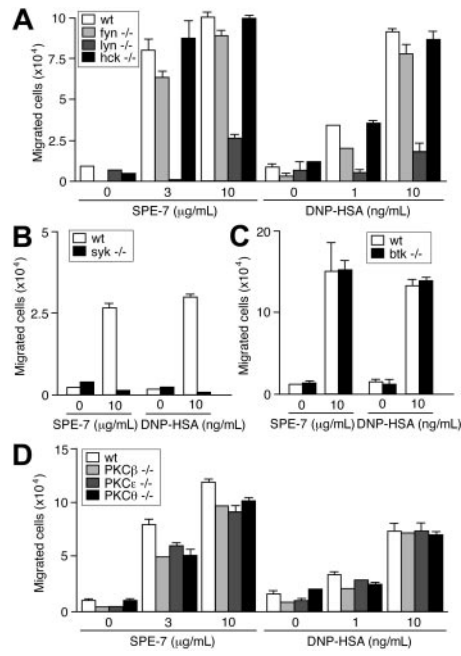


Figure 3. Effects of Src, Syk, Btk, and PKC deficiencies on IgE-induced migration. IgE- and IgE +Ag-induced migration assays were performed using Src family PTKdeficient (A), Syk-deficient (B), Btk-deficient (C), and PKC-deficient (D) BMMCs. Values (mean \pm SD) are shown out of 2 independent experiments.

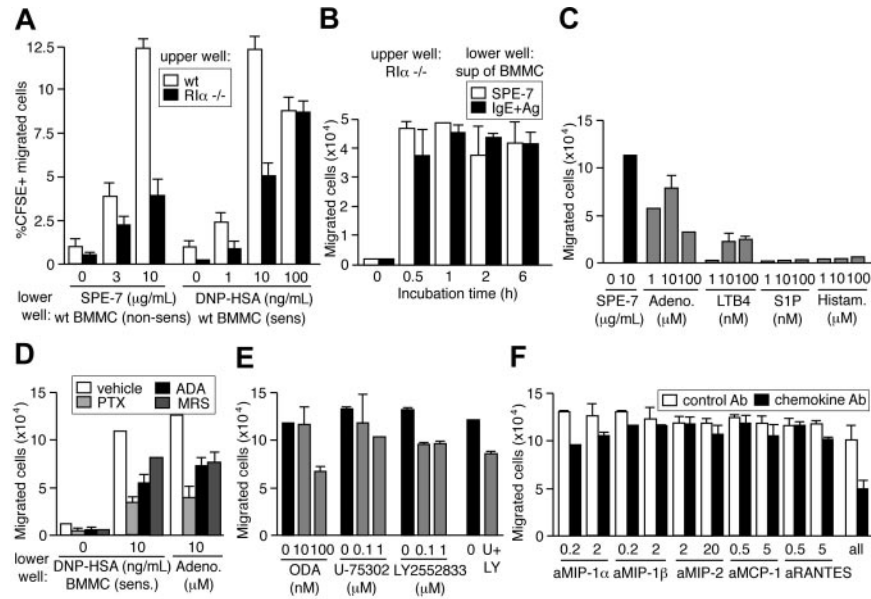


Figure 4. Involvement of adenosine, LTB₄, and chemokines in an autocrine/paracrine mechanism of IgE and IgE+Ag-induced mast cell migration. (A) CFSE-labeled wild-type and *FcεR1α*^{-/-} BMMCs in upper wells were attracted to lower wells containing wild-type cells and SPE-7 or 206 IgE-sensitized wild-type cells and antigen. (B) Supernatants of wild-type BMMCs incubated with SPE-7 for the indicated periods attracted *FcεR1α*^{-/-} cells. Supernatants of PC IgE (206)-sensitized wild-type cells incubated with antigen for the indicated periods also attracted *FcεR1α*^{-/-} cells. (C) Migration was induced by adenosine and LTB₄, but not by sphingosine 1-phosphate (S1P) or histamine. (D) Pertussis toxin (PTX), adenosine deaminase (ADA), and adenosine A₃ receptor inhibitor MRS 1523 inhibit the migration of CFSE-labeled wild-type BMMCs from upper wells to lower wells containing IgE-sensitized wild-type cells and antigen or adenosine. (E) 5-Lipoxygenase inhibitor (N-oleoyl dopamine [ODA]), BLT₁ receptor inhibitor (U-75302), and BLT₂ inhibitor (LY2552833) inhibit the migration of CFSE-labeled wild-type BMMCs from upper wells to lower wells containing IgE-sensitized wild-type cells and antigen. U-75302 and LY2552833 at 1 μM each were used in a combination (U+LY) as well. (F) Neutralizing antibodies to several chemokines inhibit the migration of CFSE-labeled wild-type BMMCs from upper wells to lower wells containing IgE-sensitized wild-type cells and antigen. All the antibodies at the higher concentrations each were used in a combination (all) as well. Mean values ± SD are shown out of 2 independent experiments.

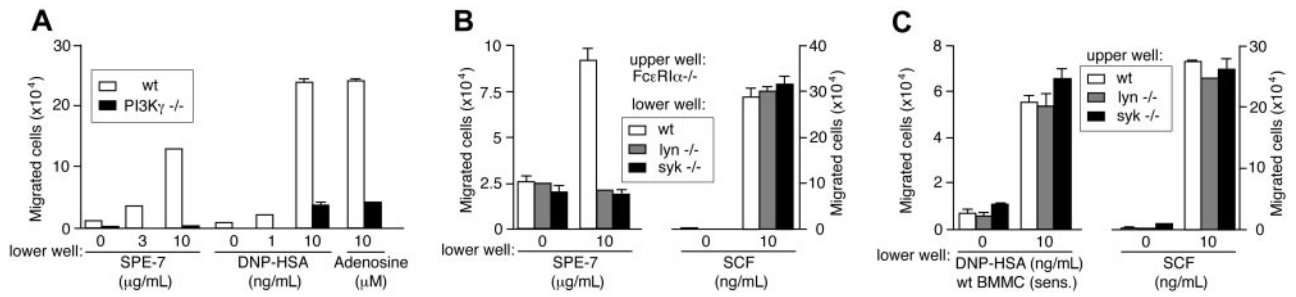


Figure 5.
IgE- and IgE+Ag-induced mast cell migration can be divided into the early phase of Lyn/Syk-dependent release of soluble factors and the later PI3K γ -dependent phase. (A) PI3K γ ^{-/-} BMMCs were defective in migration in responses to SPE-7 IgE, IgE+Ag, or adenosine. (B) Fc ϵ RI α ^{-/-} cells in upper wells were incubated with lower wells containing wild-type, lyn^{-/-}, or syk^{-/-} cells in the presence of SPE-7 or SCF. (C) Wild-type, lyn^{-/-}, or syk^{-/-} cells in upper wells were incubated with lower wells containing 206 IgE-sensitized wild-type cells in the presence of DNP-HSA or SCF. Mean values \pm SD are shown from 2 (A) or 3 (B,C) independent experiments.

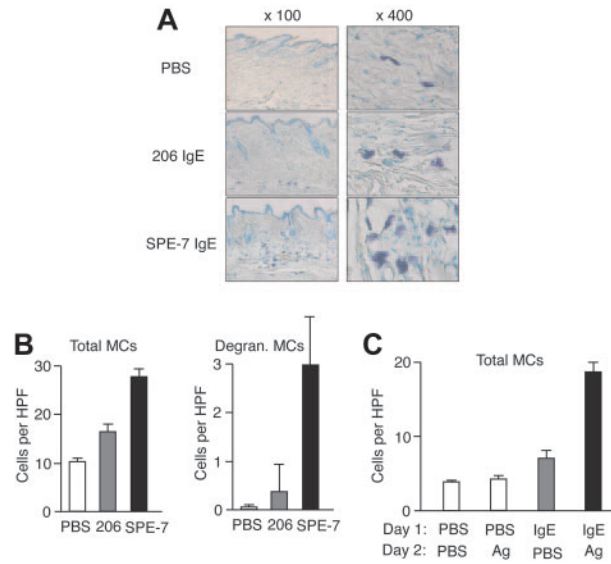


Figure 6. **In vivo models for IgE- and IgE+Ag-induced mast cell accumulation.** (A) Gauze strips spotted with 10 μ g/mL 206 or SPE-7 IgE or PBS were applied to the shaved back skin of naive NC/Nga mice for 24 hours. The mice were killed and skin samples were prepared to stain mast cells. Representative photomicrographs are shown. (B) Total and degranulating mast cell numbers per high-power field were counted. (C) NC/Nga mice were first treated with anti-DNP (206) IgE or PBS on the back skin, and 1 day later the same area was applied with gauze strips spotted with DNP-HSA or PBS for 24 hours. Total mast cell numbers per high-power field were counted. Data (mean values \pm SD) are shown representative of 3 similar experiments.

Table 1

Inhibition of BMDC migration induced by SPE-7 IgE, IgE + Ag, or SCF by various pharmacologic inhibitors

Target	Inhibitor	SPE-7 IgE	IgE+Ag	SCF
Src PTKs	PP2	2 μ M	10 μ M	0.5 μ M
Syk	Piceatannol	100 μ M	120 μ M	> 200 μ M
	ER-27319	20 μ M	20 μ M	> 100 μ M
Btk	Terreic acid	> 20 μ M	> 20 μ M	> 20 μ M
PI3K	Wortmannin	5 nM	40 nM	30 nM
	LY294002	7 μ M	10 μ M	> 50 μ M
MEK	PD98059	> 50 μ M	> 50 μ M	> 50 μ M
JNK	SP600125	30 μ M	40 μ M	40 μ M
p38	SB203380	5 μ M	2 μ M	20 μ M
PKC	Ro31-8425	4 μ M	8 μ M	20 μ M
cPKC	Goö-6976	2 μ M	0.5 μ M	2 μ M
PKA	PKI	> 100 nM	> 100 nM	> 100 nM
	KT5720	> 100 nM	> 100 nM	> 100 nM
Rho-K	Y-27632	> 10 μ M*	> 10 μ M*	> 10 μ M*
F-actin	Cytochalasin D	0.02 μ M	0.02 μ M	0.08 μ M
Ca ²⁺	EGTA	1 mM	1 mM	2 mM
IgV	DNP-lysine	0.4 μ M	0.4 μ M	> 100 μ M

BMDCs were preincubated with pharmacologic reagents for 30 minutes before being placed in upper wells at the time of addition of 10 μ g/mL SPE-7 IgE or 100 ng/mL SCF to lower wells. For IgE+Ag stimulation, BMDCs were sensitized by overnight incubation with 0.5 μ g/mL of 206 IgE. BMDCs washed with buffer were preincubated with inhibitors for 30 minutes before being placed in upper wells at the time of addition of 10 ng/mL DNP-HSA. Concentrations for 50% inhibition (IC₅₀) are indicated.

* Although BMDCs were insensitive to Y-27632, MC/9 mouse mast cells were sensitive with IC₅₀s of 5 μ M to 10 μ M for SPE-7-, IgE + Ag-, and SCF-induced migration.