

# Respiratory Gas Tensions of Thoracic Duct Lymph: An Index of Gas Exchange in Splanchnic Tissues

CHARLES L. WITTE, M.D., ROY H. CLAUSS, M.D.,  
ALLAN E. DUMONT,\* M.D.

*From the Department of Surgery, New York University School of Medicine, and  
the III and IV Surgical Divisions of Bellevue Hospital, New York City*

STUDIES of respiratory gas tensions in blood have helped clarify the relationship between alveolar ventilation, cardiac output and blood flow through major organs. Ultimately, however, cellular viability depends on microcirculatory dynamics and gas exchange in tissues outside the vascular compartment. Artificial or *in vitro* techniques to measure oxygen and carbon dioxide in living tissues are limited. Excision injures tissues or impairs blood supply to cells which then utilize oxygen factitiously from the surrounding atmosphere and accumulate carbon dioxide through continued metabolism.<sup>17</sup> In 1823, Davy<sup>9</sup> estimated extracellular oxygen tension by periodic sampling of an artificial air bubble in the pleural space. With slight modification, the gas pocket is still a popular method for measuring tissue oxygen,<sup>2, 6, 10, 13, 15, 18, 22</sup> but it is not applicable when changes in tissue oxygen are rapid.

The cellular exchange of oxygen and carbon dioxide depends on the gradient of partial pressures of these dissolved gases between plasma and interstitial fluid which governs diffusion of the gases through

tissue fluid or lymph. Since interstitial fluid from the splanchnic area is transported mainly by the thoracic duct as lymph,<sup>23</sup> studies were performed in patients and experimental animals to determine whether or not gas tensions of thoracic duct lymph reflect oxygen and carbon dioxide exchange between blood and splanchnic tissues. The effects of variations in splanchnic blood flow, oxygen consumption and pH on PO<sub>2</sub> and PCO<sub>2</sub> in thoracic duct lymph were determined.

## Methods

Twenty-two mongrel dogs (10–15 Kg.) were anesthetized with intravenous pentobarbital (24 mg./Kg.). Respirations were controlled with succinylcholine and endotracheal positive pressure ventilation. In some animals supplemental oxygen was administered at constant rate and content. The thoracic duct was exposed in the right chest and cannulated with polyethylene tubing. The spleen was removed and the portal vein was cannulated via the splenic vein. The right atrium, hepatic vein and aorta were cannulated via the femoral vein, the external jugular vein and the femoral artery respectively. Splanchnic blood flow (SBF) was measured by square-wave electromagnetic flow meters on the hepatic and superior mesenteric arteries and recorded on a four-channel direct-writing Grass polygraph. Splanchnic oxygen consumption (SOC) was calculated by the Fick prin-

\* Research Career Development Award USPHS, NIH.

Submitted for publication March 14, 1967.

Supported by grant HE-09073-CV-02, USPHS and by the John A. Hartford Foundation, Inc. and the NYC Health Research Council contract 1-208.

Presented in part before the New York Surgical Society, December, 1964 and the American Physiological Society, April, 1965.

ciple: SOC = oxygen content (aorta – hepatic vein ml./L.) × splanchnic blood flow (L./min.). All samples of blood and lymph were collected anaerobically and gas content was measured with an Instrumentation Laboratory gas analyzer utilizing a Clark electrode polarograph,<sup>7, 8</sup> a PCO<sub>2</sub> Severinghaus glass electrode<sup>19</sup> and glass electrode pH meter. PO<sub>2</sub> determinations were corrected for pH and temperature, and oxygen saturation calculated according to the Severinghaus nomogram based on shifts in the oxygen dissociation curve.

Experimental conditions were varied as follows: 1) altering the oxygen content of inspired air using 6%, 10%, and 100% oxygen balanced with nitrogen; 2) changing splanchnic blood flow by induced cardiac standstill, occlusion of the hepatic and superior mesenteric arteries, and rapid hemorrhage to a blood pressure of 40 mm./Hg; 3) altering metabolic activity of splanchnic viscera with intravenous sodium cyanide and glucagon; and 4) observing the Bohr effect or shift of oxygen dissociation curve caused by alkalosis (NaHCO<sub>3</sub>) and acidosis (NH<sub>4</sub>Cl).

tion curve caused by alkalosis (NaHCO<sub>3</sub>) and acidosis (NH<sub>4</sub>Cl).

Samples of thoracic duct lymph were also obtained from 12 patients undergoing thoracic duct cannulation for diagnostic or therapeutic purposes. The thoracic duct was cannulated in the neck under local anesthesia. Seven patients had cirrhosis of the liver, two severe congestive heart failure, two had rheumatic valvular heart disease without circulatory congestion, and one had hepatic fibrosis and cholestasis with deranged liver functions. Blood samples were obtained from the femoral artery, vena cava or right atrium and at times from the portal vein. These samples and thoracic duct lymph were analyzed for PO<sub>2</sub>, PCO<sub>2</sub> and pH in six patients with cirrhosis after intravenous administration of either vasopressin or glucagon, in one patient with severe congestive failure after intravenous administration of metaraminol and in one patient with intrahepatic cholestasis and fibrosis after administration of 100% oxygen by nasal catheter.

TABLE 1. *Effect of Changes in Splanchnic Blood Flow and Oxygen Administration on PO<sub>2</sub> (mm. Hg) in Arterial and Venous Blood and Thoracic Duct Lymph. Data Expressed as the Average Maximum Deviation from Control*

Subjects	No.	Procedure	PO <sub>2</sub> Artery	PO <sub>2</sub> Right Atrium	PO <sub>2</sub> Thoracic Duct Lymph
Dogs	3	Control	83	33	51
		I.V. Pentobarbital (cardiac arrest)	+11, -12	-12	-47
Dogs	2	Control	98	38	61
		Ligation of hepatic and superior mesenteric arteries	+5, -10	-8	-53
Dogs	4	Control	110	40	66
		Hemorrhage to B.P. of 40 mm. Hg	+23, -12	+2	-42
Dogs	3	Control	75	42	50
		100% O <sub>2</sub>	+500	+25	+75
		10% O <sub>2</sub>	-30	-20	-16
		6% O <sub>2</sub>	-60	-35	-42
Patient with cholestasis and hepatic fibrosis		Control	68	22	18
		100% O <sub>2</sub> *	+108	+9	+60
Patient with severe congestive heart failure		Control*	150	40	32
		Metaraminol*	+8, -30	-18	-9

\* 100% O<sub>2</sub> via nasal catheter.

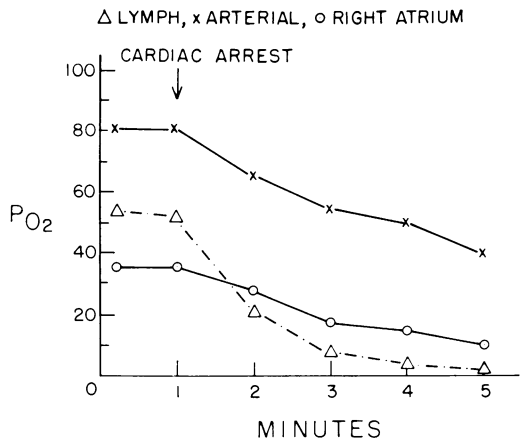


FIG. 1. Effect of induced cardiac standstill in dog on PO<sub>2</sub> in blood and thoracic duct lymph. This figure is representative of three experiments. Thoracic duct lymph PO<sub>2</sub> is normally higher than venous PO<sub>2</sub>.

**Results**

In control samples of thoracic duct lymph PO<sub>2</sub> normally ranged from 55-60 mm./Hg in patients and 45-55 mm./Hg in dogs (contrasted with an earlier report<sup>4</sup>). The peak effects on lymph PO<sub>2</sub> in 12 dogs and two patients after changes in splanchnic blood flow and oxygen administration are shown in Table 1. In five dogs, after either induced cardiac arrest or clamping the hepatic and superior mesenteric arteries, thoracic duct lymph PO<sub>2</sub> progressively decreased with only a moderate diminution in the oxygen content of central venous blood. A similar sequence occurred in four dogs during hemorrhagic shock produced by rapid bleeding to a blood pressure of 40 mm./Hg. Characteristic responses are reproduced in Figures 1, 2, 3.

Varying oxygen composition of inhaled air with 6%, 10% and 100% oxygen with balanced nitrogen for 3-4 minutes in three dogs evoked an immediate (within 20-30 seconds) change in lymph PO<sub>2</sub> which paralleled the change in arterial oxygen tension (Fig. 4). A similar change occurred after 100% oxygen was administered to the patient with hepatic fibrosis and cholestasis.

The effect of changes in pH, oxygen consumption and splanchnic blood flow on

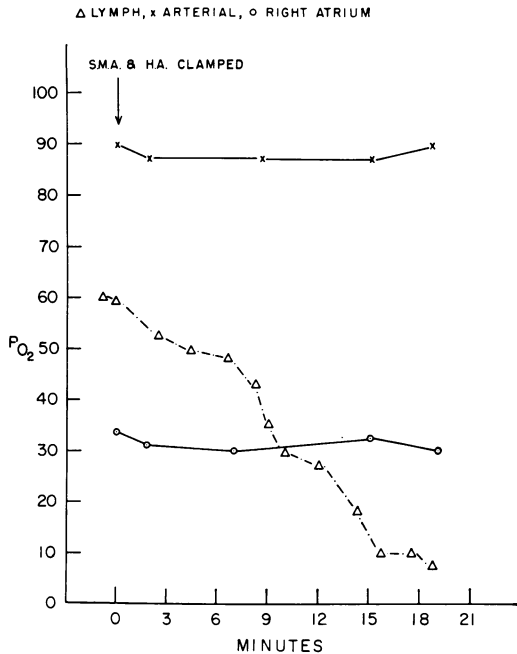


FIG. 2. Effect of ligation of superior mesenteric and hepatic arteries in dog on PO<sub>2</sub> in blood and thoracic duct lymph. This figure is representative of two experiments.

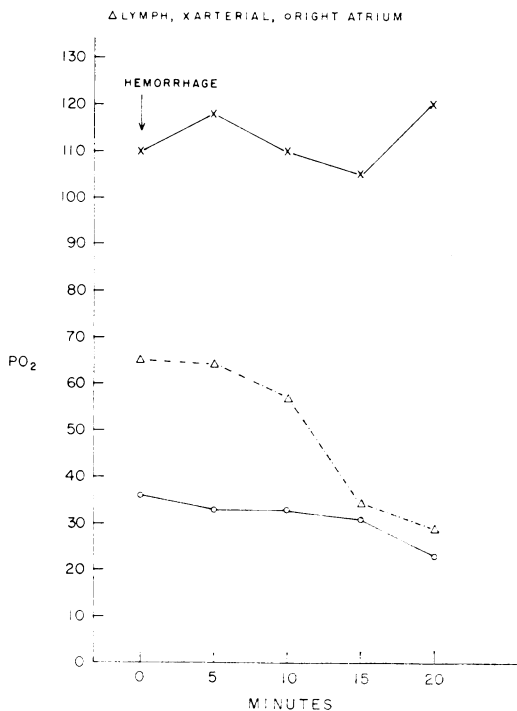


FIG. 3. Effect of rapid bleeding in dog on PO<sub>2</sub> in blood and thoracic duct lymph. This figure is representative of four experiments.

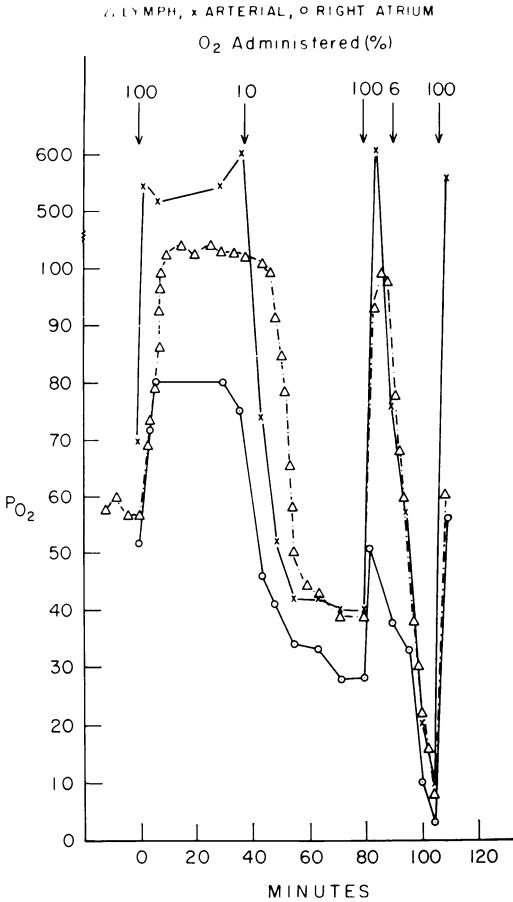


FIG. 4. Effect on  $PO_2$  in blood and thoracic duct lymph in dog after changing the oxygen content of inspired air using 6%, 10%, and 100% oxygen with balanced nitrogen. This figure is representative of three experiments.

respiratory gases in arterial and venous blood and thoracic duct lymph are shown in Tables 2 and 3. Infusion of large amounts (44 mEq.) of sodium bicarbonate transiently changed splanchnic blood flow and oxygen consumption in three dogs; yet progressive decrease in lymph  $PO_2$  was characteristic. The initial loading dose of bicarbonate caused a temporary rise in blood flow and venous  $PO_2$ , widening of the pulse pressure but lowering of the lymph  $PO_2$  (Fig. 5). Acidosis which followed 44 mEq.  $NH_4Cl$  intravenously, on the other hand, in three dogs failed to influence splanchnic gas exchange consistently despite pH values as low as 6.9. Only

transient alterations in splanchnic blood flow and oxygen consumption, blood gases and lymph gas tensions were observed (Fig. 6).

Characteristic changes in lymph gas tensions in response to variations in splanchnic blood flow and splanchnic oxygen consumption are shown in Figures 7-11. Intravenous sodium cyanide (0.5 mg./Kg.), a cytochrome enzyme inhibitor, raised lymph  $PO_2$  and regional blood flow and lowered  $PCO_2$  (Fig. 7). Intravenous glucagon (0.03 mg./Kg.) which increases oxygen consumption, raised splanchnic oxygen consumption and splanchnic blood flow but lowered lymph  $PO_2$ , pH and raised  $PCO_2$  (Fig. 8).

Extremely low values of  $PO_2$  in lymph were occasionally found in patients with

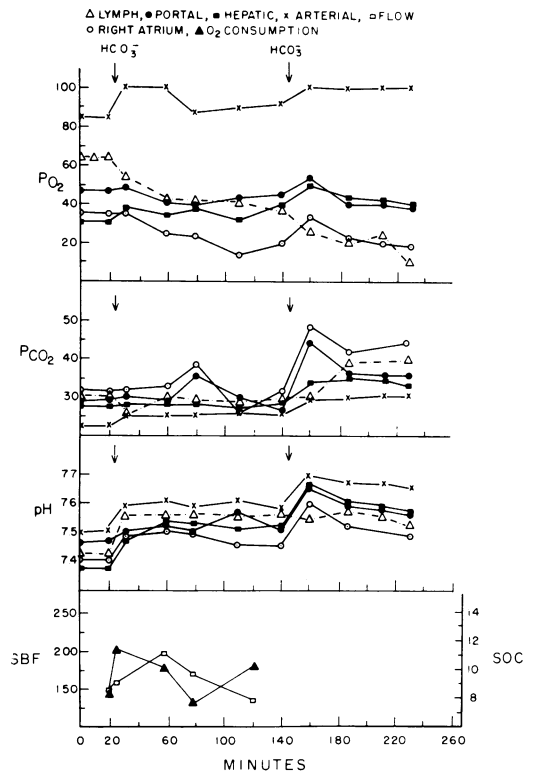


FIG. 5. Effect of sodium bicarbonate administration in dogs on splanchnic blood flow and oxygen consumption on  $PO_2$ ,  $PCO_2$  and pH in blood and thoracic duct lymph. This figure is representative of three experiments.

TABLE 2. *Effect of Changes in pH on Respiratory Gases (mm. Hg) in Arterial and Venous Blood and Thoracic Duct Lymph. Data Expressed as the Average Maximum Deviation from Control (pre-)*

Subject	No.	Agent	Sample	PO <sub>2</sub>		PCO <sub>2</sub>		pH	
				Pre	Post	Pre	Post	Pre	Post
Dogs	3	NH <sub>4</sub> Cl	Artery	100	-12	22	-2	7.42	-0.52
			Right Atrium	30	+3, -4	29	+15	7.39	-0.53
			Hepatic Vein	40	-10	25	+8	7.42	-0.53
			Portal Vein	39	+15	27	+8	7.40	-0.62
			Thoracic duct lymph	43	+12	25	+15	7.40	-0.51
Dogs	3	NaHCO <sub>3</sub>	Artery	99	+20	26	+8	7.52	+0.21
			Right Atrium	27	+7, -3	34	+16	7.41	+0.14
			Hepatic Vein	36	+8, -7	33	+13	7.50	+0.15
			Portal Vein	46	+7, -8	32	+14	7.44	+0.14
			Thoracic duct lymph	52	-33	34	+5	7.46	+0.07

advanced hepatic disease (Fig. 9) and in those with severe congestive heart failure. In five patients with cirrhosis, 20 units of vasopressin administered intravenously over 10-minute periods produced a fall in PO<sub>2</sub> and pH and a rise in PCO<sub>2</sub> in lymph (Fig. 10). In one patient with cirrhosis glucagon caused a fall in lymph PO<sub>2</sub> similar to that in experimental animals (Fig. 11).

### Discussion

Fifty years ago Starling specifically emphasized the role of lymph in tissue gas exchange: "The physical diffusion of oxygen and carbon dioxide between capillaries and cells involves not only passage across capillary endothelium and cell wall but through intervening tissue fluid or lymph. Dissociation of oxygen from hemoglobin at the arteriolar end of the capillary de-

TABLE 3. *Effect of Agents Altering Splanchnic Blood Flow and Splanchnic Oxygen Consumption on Respiratory Gases (mm. Hg) and pH in Arterial and Venous Blood and Thoracic Duct Lymph. Data Expressed as the Average Maximum Deviation from Control (pre-)*

Subjects	No.	Agent	Splanchnic Blood Flow (ml./min.)		Splanchnic O <sub>2</sub> Consumption (ml. O <sub>2</sub> /min.)		Sample	PO <sub>2</sub>		PCO <sub>2</sub>		pH	
			Pre	Post	Pre	Post		Pre	Post	Pre	Post		
Dogs	2	NaCN	165	+168	10.8	-6.2	Artery	85	+10	22	-6	7.46	+0.02, -0.03
							Right Atrium	29	+6	29	-9	7.42	+0.01, -0.04
							Hepatic	38	+17	25	+2, -6	7.44	-0.04
							Portal Vein	37	+11	25	-3	7.45	+0.03, -0.06
							T.D. lymph	48	+22	31	-17	7.45	+0.03, -0.06
Dogs	2	Glucagon	195	+380	11.6	+9.8	Artery	90	-15	27	-4	7.45	-0.04
							Right Atrium	38	-13	29	+6	7.40	-0.09
							Hepatic Vein	41	+5, -5	25	+3	7.42	+0.02, -0.06
							Portal Vein	36	+25, -5	30	+2, -5	7.40	+0.04, -0.02
							T.D. lymph	52	-23	24	+14	7.39	+0.03, -0.12
Patient with Cirrhosis	1	Glucagon				Artery	62	-3	26	+3	7.50	-0.06	
						Right Atrium	35	+2, -2	22	+8	7.48	-0.03	
						T.D. lymph	36	-18	24	+10	7.46	-0.03	
Patients with Cirrhosis	5	Vasopressin				Artery	66	+4, -7	29	+9	7.46	+0.05, -0.09	
						Right Atrium	32	-4	30	+11	7.39	+0.03, -0.09	
						Portal Vein*	40	-10	25	+2	7.29	+0.03, -0.03	
						T.D. lymph	41	-21	33	+9	7.48	+0.01, -0.12	

\* Samples obtained from umbilical vein in two patients.

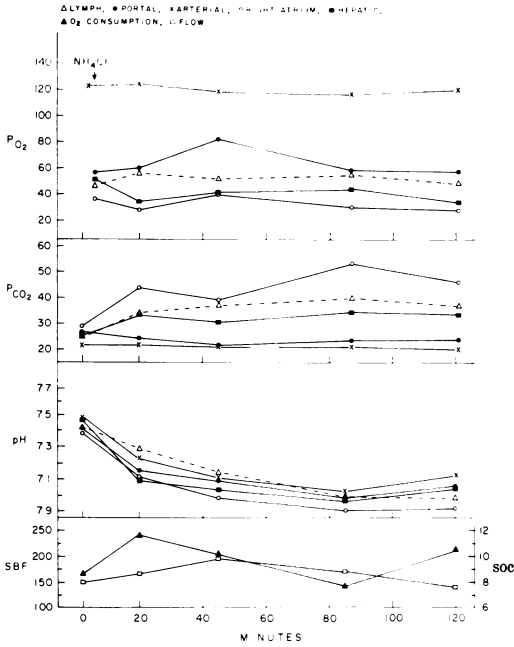


FIG. 6. Effect of ammonium chloride administration in dog on splanchnic blood flow and oxygen consumption on  $PO_2$ ,  $PCO_2$  and pH in blood and thoracic duct lymph. This figure is representative of three experiments.

depends on lowering of the partial pressure of oxygen in plasma at that point. The extent of this lowering is directly determined

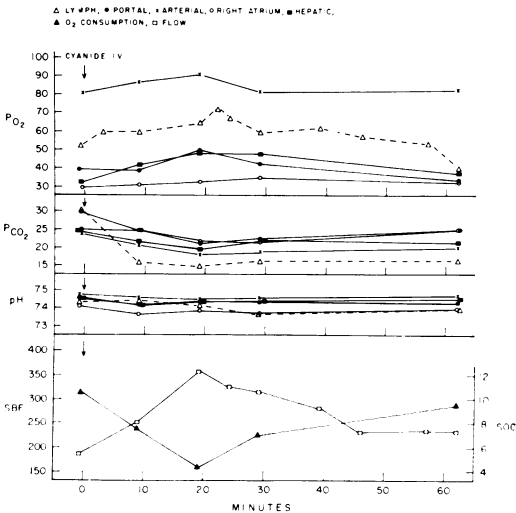


FIG. 7. Effect of sodium cyanide administration in dog on splanchnic blood flow and oxygen consumption on  $PO_2$ ,  $PCO_2$  and pH in blood and thoracic duct lymph. This figure is representative of two experiments.

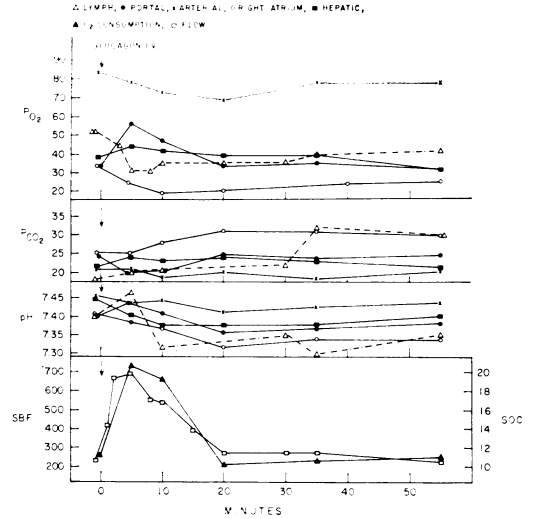


FIG. 8. Effect of glucagon administration in dog on splanchnic blood flow and oxygen consumption on  $PO_2$ ,  $PCO_2$  and pH in blood and thoracic duct lymph. This figure is representative of two experiments.

by the gradient of dissolved oxygen between capillary plasma and interstitial fluid.”<sup>21</sup>

It is generally recognized that the primary factors regulating tissue oxygen tension include arterial oxygen content, rate of capillary blood flow, arteriovenous shunts,

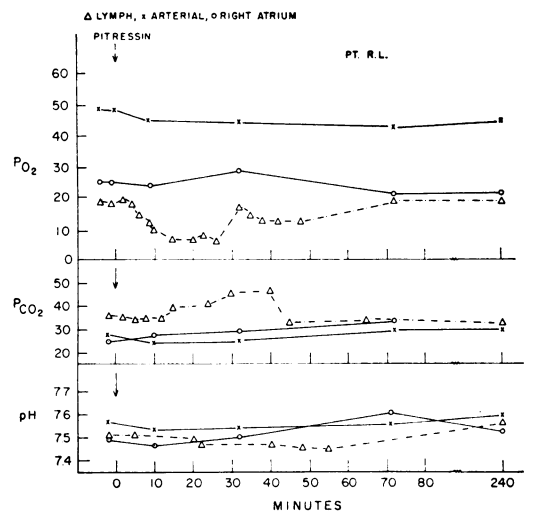


FIG. 9. Effect of vasopressin administration with advanced hepatic cirrhosis on  $PO_2$ ,  $PCO_2$  and pH in blood and thoracic duct lymph. Note low control level of lymph  $PO_2$  which rapidly approaches zero.

tissue oxygen consumption and the dissociability of hemoglobin.<sup>12, 14</sup> Increased arterial oxygen content, increased capillary blood flow, decreased oxygen consumption and a shift of the oxygen dissociation curve to the right (Bohr effect) tend to raise tissue oxygen tension. Conversely, decreased arterial oxygen content, decreased capillary blood flow, increased oxygen consumption or a shift of the oxygen dissociation curve to the left tend to lower tissue oxygen tension. Diversion of capillary blood flow through arteriovenous shunts may reduce or eliminate gas exchange at the cellular level. Each of these variables exerts its influence on tissue gas tensions by altering the plasma-interstitial fluid gas gradient.

The data presented in this study demonstrate that factors known to alter splanchnic tissue oxygen tension influence thoracic duct lymph oxygen tension in a predictable manner. Reduction of blood flow in experimental animals by induced cardiac arrest, rapid hemorrhage or interruption of major splanchnic arteries produces splanchnic tissue or lymph "hypoxia" by restricting the amount of oxygen reaching capillaries. Changes in oxygen content in in-

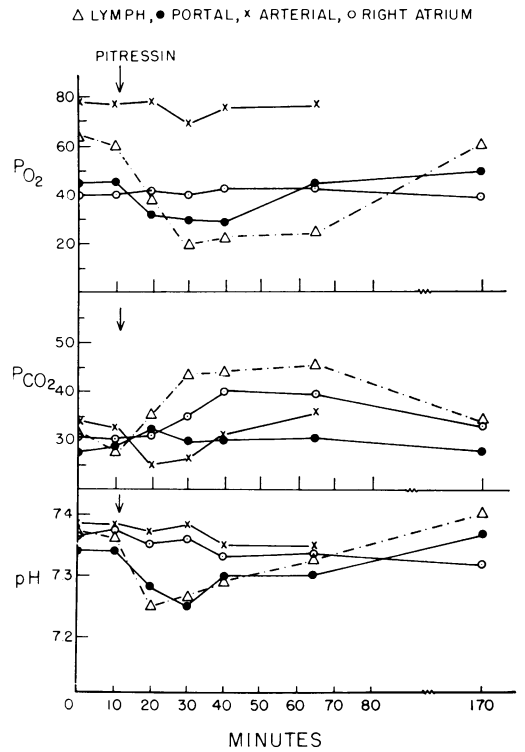
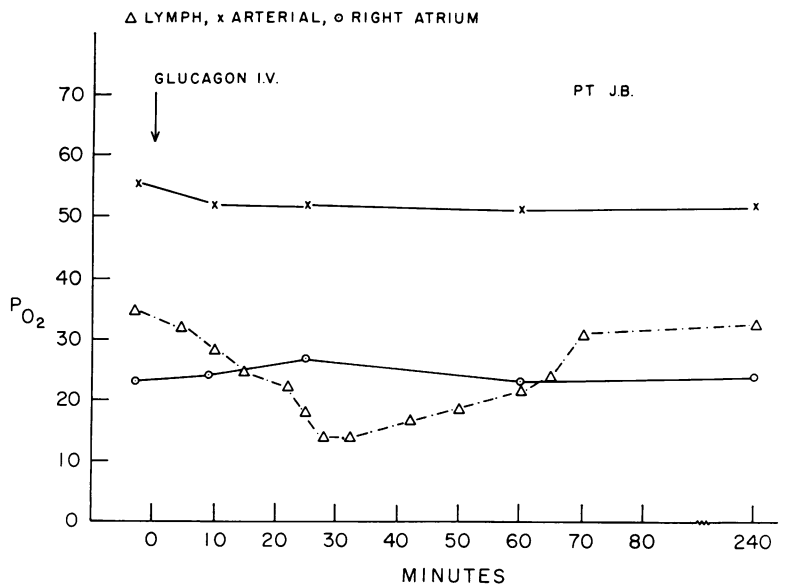


FIG. 10. Effect of vasopressin administration in patient with hepatic cirrhosis on  $PO_2$ ,  $PCO_2$  and pH in blood and thoracic duct lymph. Lymph  $PO_2$  falls below right atrial and portal vein  $PO_2$ .

haled air regulate the supply of available oxygen carried in arterial blood and there-

FIG. 11. Effect of glucagon administration in patient with hepatic cirrhosis on  $PO_2$ ,  $PCO_2$  and pH in blood and thoracic duct lymph.



by alter lymph  $PO_2$ . Administration of sodium cyanide inhibits oxygen consumption by interfering with oxidative phosphorylation which accompanied by an increase in regional blood flow from vasodilatation, raises  $PO_2$  and lowers  $PCO_2$  in lymph. Administration of sodium bicarbonate tends to lower lymph  $PO_2$  by diverting capillary blood flow through arteriovenous shunts as suggested by an increase in blood flow, venous  $PO_2$  and pulse pressure (*vide supra*), and perhaps by a shift in the oxygen dissociation curve to the left. Acidosis following intravenous ammonium chloride, however, does not lead to significant changes in blood or lymph gas tensions.

The effect of increased splanchnic oxygen consumption on splanchnic tissue gas tensions is difficult to assess because an autoregulatory rise in blood flow occurs during increased functional activity.<sup>1, 20</sup> Patients with hepatic disease are limited in their ability to compensate for increased splanchnic oxygen consumption with a rise in splanchnic blood flow.<sup>5, 16</sup> Instead, increased cellular activity is offset by a wider extraction of oxygen content (aortic-hepatic vein) a derangement which limits the availability of oxygen to splanchnic tissues. Thus, when splanchnic oxygen consumption rises a diminution in tissue  $PO_2$  is likely to occur. After administration of glucagon, a hormone capable of almost doubling the splanchnic oxygen consumption in cirrhosis,<sup>16</sup> thoracic duct lymph  $PO_2$  decreases.

Administration of vasopressin to patients with hepatic disease results in a decrease of lymph  $PO_2$  and pH and a rise in  $PCO_2$ . Metaraminol administration in congestive heart failure similarly results in a diminution in lymph  $PO_2$ . These potent vasoconstrictive drugs, in all likelihood, favor anaerobic metabolism by raising arteriolar resistance thereby reducing capillary blood flow. Moreover, in patients with severe hepatic disease or congestive heart failure,

$PO_2$  of lymph may already be as low as 14–35 mm. Hg. Although reduction of hepatic blood flow could account for a low  $PO_2$  in splanchnic interstitial fluid in these conditions, in cirrhosis there may be, in addition, a mechanical block to diffusion of oxygen secondary to parenchymal fibrosis analogous to the alveolar-capillary block in pulmonary interstitial fibrosis. When 100% oxygen was administered to a patient with marked hepatic dysfunction a prompt rise in arterial and lymph  $PO_2$  occurred. This response resembles that in the dog<sup>3</sup> and suggests that administration of oxygen to such patients may increase tissue oxygen tension toward normal levels and provide more oxygen for parenchymal cells.

### Summary

Samples of thoracic duct lymph from dogs and patients were analyzed for  $PO_2$ ,  $PCO_2$  and pH under experimental conditions that were varied by altering pH, splanchnic blood flow, splanchnic oxygen consumption and the  $PO_2$  of inspired air. Evidence was obtained which indicated that changes in gas tensions of thoracic duct lymph provide direct information concerning alterations in gas exchange in splanchnic tissues. This technic may be useful in unraveling some of the poorly understood problems of tissue oxygenation in hemorrhagic shock, cirrhosis and congestive heart failure.

### References

1. Barcroft, J.: Respiratory Function of the Blood. Cambridge, England, Cambridge University Press, 1914, p. 134.
2. Bazett, H. D. and Sribyatta, L.: The Effect of Local Changes in Temperature on Gas Tensions in the Tissues. *Amer. J. Physiol.*, **86**:565, 1928.
3. Bergofsky, E. H., Wang, M. C. H., Yamaki, T. and Jacobson, J. H.: Tissue Oxygen and Carbon Dioxide Tensions During Hyperbaric Oxygenation. *JAMA*, **189**:841, 1964.
4. Bergofsky, E. H., Jackson, H., II and Fishman, A. P.: The Use of Lymph for the Measurement of Gas Tensions in Interstitial Fluid and Tissues. *J. Clin. Invest.*, **141**: 1971, 1962.



5. Bradley, S. E., Ingelfinger, F. J., Groff, A. E. and Bradley, G. P.: Estimated Hepatic Blood Flow and Hepatic Venous Oxygen Content in Cirrhosis of the Liver. *Proc. Soc. Exper. Biol. Med.*, **67**:206, 1948.
6. Campbell, J. A.: Gas Tensions in the Tissues. *Physiol. Rev.*, **11**:1, 1931.
7. Clark, L. C., Wolf, R., Granger, D. and Taylor, Z.: Continuous Recording of Blood Oxygen Tensions by Polarography. *J. Appl. Physiol.*, **6**:189, 1958.
8. Connelly, C. M.: Methods for Measuring Tissue Oxygen Tension; Theory and Evaluation. In *Oxygen in the Animal Organism IUB Symposium Series*. Dickens, F. and Neil, E., Eds. New York, Macmillan, 1964, Vol. 31, p. 681.
9. Davy, J.: Observations on Air Found in the Pleura in a Case of Pneumatothorax; with Experiments on the Absorption of Different Kinds of Air Introduced into the Pleura. *Phil. Trans. Roy. Soc. London*, **113**:496, 1923.
10. Farhi, L. E.: Tissue Gas Tensions. In *Ann. Rev. of Physiol.* Hall, V., Ed. Palo Alto, Hanta, 1965, Vol. 27, p. 244.
11. George, W. S., Lesko, W. S. and Leevy, C. M.: Observations on the Relationship of Arterial and Hepatic Vein Oxygen Content to Splanchnic Oxygen Consumption in Man. *Gastroenterology*, **40**:555, 1961.
12. Groth, C. G., Lofstrom, B. and Werner, B.: Oxygen Tension of Thoracic Duct Lymph in Man. *Acta Chir. Scand.*, **129**:586, 1965.
13. Haggard, H. H. and Henderson, Y.: Gas Tensions of the Abdominal Cavity with Some Evidence on the Diffusion of Gases Within the Body. *J. Biol. Chem.*, **38**:71, 1919.
14. Kety, S.: Symposium American Physiologic Society. *Fed. Proc.*, **16**:666, 1957.
15. Lambertson, C. J., Stroud, M. W., Ewing, J. W. and Mack, C.: Oxygen Toxicity Effects of Oxygen Breathing at Increased Ambient Pressure Upon  $PCO_2$  of Subcutaneous Gas Depots in Man, Dogs, Rabbits and Cats. *J. Appl. Physiol.*, **6**:358, 1953.
16. Leevy, C. M., George, W., Lesko, W., Deyssine, M., Abbott, C. C. and Halligan, E. J.: Observations on Hepatic Oxygen Metabolism in Man. *JAMA*, **178**:565, 1961.
17. Montgomery, H.: Oxygen Tension of Tissues *In Vivo*. *Circulation*, **15**:646, 1957.
18. Rahn, H.: Gasometric Method for Measurement of Tissue Oxygen Tension. In *Symposium on Tissue Oxygen Tension*. Chairman, J. H. Comroe, Jr., *Fed. Proc.*, **16**:685, 1957.
19. Severinghaus, J. W. and Bradley, A. F.: Electrodes for Blood  $PO_2$  and  $PCO_2$  Determination. *J. Appl. Physiol.*, **13**:515, 1958.
20. Schmidt, C.: Cerebral Blood Supply and Cerebral Oxidative Metabolism. In *Oxygen in the Animal Organism IUB Sump*. Dicken, F. and Neil, E., Eds. New York, Macmillan, 1964, Vol. 31, p. 433.
21. Starling, E. H.: *Principles of Human Physiology*. Philadelphia, Lea and Febiger, 1915, p. 1063.
22. Taylor, H. J.: The Effect of Breathing Oxygen at Atmospheric Pressure on Tissue Oxygen and Carbon Dioxide Tensions. *J. Physiol.*, **108**:264, 1949.
23. Yoffey, J. M. and Courtice, F. C.: *Lymphatics, Lymph and Lymphoid Tissue*. Cambridge, Harvard University Press, 1956.