

Island radiation on a continental scale: Exceptional rates of plant diversification after uplift of the Andes

Colin Hughes* and Ruth Eastwood

Department of Plant Sciences, University of Oxford, South Parks Road, Oxford, OX1 3RB, United Kingdom

Edited by Peter R. Crane, Royal Botanic Gardens, Kew, Surrey, United Kingdom, and approved May 19, 2006 (received for review March 9, 2006)

Species radiations provide unique insights into evolutionary processes underlying species diversification and patterns of biodiversity. To compare plant diversification over a similar time period to the recent cichlid fish radiations, which are an order of magnitude faster than documented bird, arthropod, and plant radiations, we focus on the high-altitude flora of the Andes, which is the most species-rich of any tropical mountains. Because of the recent uplift of the northern Andes, the upland environments where much of this rich endemic flora is found have been available for colonization only since the late Pliocene or Pleistocene, 2–4 million years (Myr) ago. Using DNA sequence data we identify a monophyletic group within the genus *Lupinus* representing 81 species endemic to the Andes. The age of this clade is estimated to be 1.18–1.76 Myr, implying a diversification rate of 2.49–3.72 species per Myr. This exceeds previous estimates for plants, providing the most spectacular example of explosive plant species diversification documented to date. Furthermore, it suggests that the high cichlid diversification rates are not unique. Lack of key innovations associated with the Andean *Lupinus* clade suggests that diversification was driven by ecological opportunities afforded by the emergence of island-like habitats after Andean uplift. Data from other genera indicate that lupines are one of a set of similarly rapid Andean plant radiations, continental in scale and island-like in stimulus, suggesting that the high-elevation Andean flora provides a system that rivals other groups, including cichlids, for understanding rapid species diversification.

leguminosae | *Lupinus* | phylogeny | species diversification

Studies of rapid episodes of species diversification or species radiations have provided a continuously rich source of new insights into the evolutionary processes underlying diversification and modern patterns of biodiversity for the last 150 years (1–6). A striking feature of species radiations is the discrepancy between the very high rates of species diversification reported for cichlid fish radiations in east African rift lakes and all bird, arthropod and plant radiations (2, 7, 8). This discrepancy has been attributed in part to the recency of the fish radiations (7). Accurate measurement of peak episodes of diversification embedded within older radiations requires fully sampled and resolved phylogenies (9). In the absence of such phylogenies, measurements of diversification rates for most radiations are less precise because they average out episodes of faster and slower speciation. Furthermore, these approaches measure net diversification rates, and the effects of extinction are not assessed. If speciation is concentrated in the early phases of radiations (3, 7, 10, 11), this could explain the discrepancy between the exceptional rates of species diversification reported for very recent [<2 million years (Myr)] lacustrine fish and other documented bird, arthropod, or plant radiations, which are generally older (>5 Myr) (7). Direct comparisons with the cichlid fish diversification rates have been lacking because few comparably recent radiations have been found in other groups.

To compare plant diversification over a similarly short time period to the very recent fish radiations, we examined patterns of species diversification in the high-altitude flora of the northern Andes, which is by far the most species-rich of any tropical

mountain massif (12). It forms part of the tropical Andean biodiversity hotspot, which contains an estimated 45,000 plant species, 44% of which are endemic (13). Endemism is even higher, reaching 60%, in the high-altitude north Andean páramos (14). The cold upland habitats where much of this rich endemic flora is found today have been available for plant colonization only since the late Pliocene or early Pleistocene 2–4 Myr ago (15–18) after final uplift of the northern Andes (15). This recent uplift implies that at least some Andean plant groups must have diversified very recently and rapidly (16, 19). Despite this presumption, there are few reliable estimates of species diversification rates for Andean groups because of uncertainty about species numbers and the lack of robust, well resolved, and densely sampled phylogenies (20, 21).

The prevalence of north and south temperate genera in the high-altitude Andean flora (12) supports the idea that colonization of the emerging high Andes was comparable to colonization of a newly formed island or island archipelago (16, 22) (Fig. 1). On recently formed islands and island-like formations such as lakes and mountains, rapid diversification has been attributed to ecological opportunities afforded by the availability of new habitats and absence of competition (1, 3, 7, 8, 10, 11, 23). Conversely, on continents, where ecological opportunity is harder to demonstrate, rapid episodes of diversification have generally been associated with key morphological or physiological innovations (4, 11). Numerous north temperate plant genera, such as *Alnus*, *Draba*, *Lupinus*, *Quercus*, *Salix*, *Sambucus*, *Valeriana*, and *Viburnum*, are postulated to have arrived in the Andes from North America after uplift of the northern Andes (17, 18). The altitudinal distribution of Andean species diversity for these genera shows that diversification is restricted largely to those growing at high elevations (Fig. 2). Although the monophyly of most of these endemic Andean species flocks has not been tested, this pattern supports the idea that island-like opportunities for diversification were available above the tree line in the high-altitude grassland zone and that plants that were pre-adapted to cooler conditions were most able to exploit such opportunities (Fig. 2).

The Genus *Lupinus*

To gain insight into colonization of the recently formed high-elevation Andean habitats, we examined species diversification in the legume genus *Lupinus*, which comprises ≈ 275 species of annual herbs and herbaceous and woody perennials with an amphiatlantic distribution. The majority of *Lupinus* species occur in the New World, with two main centers of species diversity in western North America (≈ 100 species) and the Andes (≈ 85 species). Guided by previous estimates of *Lupinus*

Conflict of interest statement: No conflicts declared.

This paper was submitted directly (Track II) to the PNAS office.

Abbreviations: Myr, million years; ITS, internal transcribed spacer.

Data deposition: The sequences reported in this paper have been deposited in the GenBank database (accession nos. DQ524181–DQ524328 and DQ529744–DQ529979).

*To whom correspondence should be addressed. E-mail: colin.hughes@plants.ox.ac.uk.

© 2006 by The National Academy of Sciences of the USA

chromosome number, which differ consistently between these groups (our unpublished data).

Sequence Data and Phylogenetic Analysis. One hundred forty-eight accessions representing 98 species covering previously recognized *Lupinus* clades (24), the geographic range, and eight outgroup species representing five genera from tribe Genisteae were sampled. To test the monophyly of the Andean species, 53 accessions representing 36 species were sampled spanning the range of life forms (Fig. 3, a–j), geography, and altitudes. (Fully annotated parsimony and Bayesian trees, locality and voucher details, and GenBank accession nos. are available in supporting information, which is published on the PNAS web site.) The 5.8S subunit and flanking ITS (ITS1+ITS2) of nuclear ribosomal DNA and one copy (*LEGCYCIA*) of the rapidly evolving regulatory gene *CYCLOIDEA* were sequenced by using standard protocols. Two copies of *CYCLOIDEA*-like *LEGCYCI* genes have been characterized and sequenced in the Genistoid legumes including *Lupinus* (36, 37). These sequences were used to design complementary locus-specific primers located between the conserved TCP and R domains (38), which can be used in combination with general *LEGCYCI* primers to amplify the entire *LEGCYCIA* ORF of $\approx 1,000$ bp in two fragments. An additional pair of locus-specific primers (available upon request) was designed to amplify *LEGCYCIA* for taxa in the lowland eastern New World clade (Fig. 3, clade A), which have a deletion at the original locus-specific primer site. The *LEGCYCIA* paralogue was chosen because it is more variable than *LEGCYCIB* (37, 38) and, unlike *LEGCYCIB*, does not show allelic length variation. ITS and *LEGCYCIA* sequences from Ree *et al.* (37) were included in the analysis. New sequences have been submitted to the GenBank database (see supporting information). Sequences were aligned by using CLUSTALX (39) and manually adjusted. Indels were coded by using the simple gap-coding method in SEQSTATE (40) and included in the parsimony analysis. Parsimony analysis of the combined ITS/*LEGCYCIA* data set was conducted with NONA (41) by using 1,000 random addition sequences, tree bisection, and reconnection, holding 100 trees per replication and attempting to swap to completion (see supporting information). A reduced data set of 89 sequences representing one outgroup and one sequence per species was compiled for Bayesian analysis (42) using the GTR+G and GTR+I models (43) for the ITS and *LEGCYCIA* data partitions, respectively. This analysis involved 3,000,000 Metropolis-coupled Markov Chain Monte Carlo permutations of tree parameters and four chains heated to 0.01. The consensus

Bayesian tree was virtually congruent with the parsimony tree but slightly more resolved (see supporting information). Clade stability tests involved Bayesian posterior probabilities and parsimony bootstrap resampling.

Evolutionary Rates Analysis. The relative rate test implemented in MR BAYES rejected a molecular clock for the consensus Bayesian tree (likelihood ratio = 3313.72, df = 142, $P = 0.00$). The program R8S (44) was used to assess variation in substitution rates and incorporate this into the estimation of ages of lineages. Branching order and branch lengths from 100 Bayesian trees sampled every 10,000 generations after stationarity were analyzed to obtain means and standard deviations of ages of clades. Cross-validation and penalized likelihood analysis yielded an average smoothing parameter of 13, which was used in final dating analyses. Relative ages estimated with R8S were converted to absolute ages by fixing the age of the *Lupinus* stem node using mean (16.01 Myr) and maximum (21.16 Myr) dates for the most recent common ancestor of *Spartium* and *Lupinus* from the legume-wide analysis of *matK* sequences calibrated using 13 well studied fossils (25).

Species Diversification Rate. Species diversification rates, assuming an equal rate of random speciation Yule model, were calculated as $SR = (\ln n_1 - \ln n_0)/t$, where n_1 is the number of extant species, n_0 is the initial species diversity, here taken as 1, and t is time in Myr. Upper and lower standard deviations of age estimates were used in calculations of speciation rate.

We thank Helene Citerne for primers; Rosemary Wise for artwork; Simon Ho and Matt Lavin for advice; Matt Lavin for age estimates based on but not presented in ref. 25; Donovan Bailey, Stephan Beck, Tim Budden, Ruth Clark, Aniceto Daza, Alfonso Delgado, Chris Fagg, Rob Forrest, Martin Gardner, Greg Kenicer, Bente Klitgaard, Gwil Lewis, Helga Ochoterena, David Neill, Arely Palabral, John Pannell, Terry Pennington, Toby Pennington, Carolyn Proenca, Carlos Reynel, Mario Sousa, Salvador Talavera, John Wood, Marty Wojciechowski, and especially Chris Drummond (University of Idaho, Moscow), Richard Spellenberg (New Mexico State University, Las Cruces), and Silvia Miotto and Maria-Teresa Schifino-Wittman (Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil) for assistance with fieldwork and provision of plant samples; the authorities in Ecuador, Peru, Bolivia, and Brazil for permission to collect material; the B, CAS, CGE, E, F, FHO, G, GH, ISC, K, LPB, M, MO, MOL, MSB, NY, QCNE, UB, UC, US, USM, and USZ herbaria (Index Herbariorum acronyms) for loan of material; and Julie Hawkins, Jane Langdale, John Pannell, Mike Sanderson, and an anonymous reviewer for comments. R.E. created Figs. 1 and 3. This work was supported by the Royal Society, Biotechnology and Biological Sciences Research Council, the Genetics Society, and the Stanley Smith Horticultural Trust.

- Schluter, D. (2000) *The Ecology of Adaptive Radiation* (Oxford Univ. Press, Oxford).
- Kocher, T. D. (2004) *Nat. Rev. Genet.* **5**, 288–298.
- Gravilets, S. & Vose, A. (2005) *Proc. Natl. Acad. Sci. USA* **102**, 18040–18045.
- Kay, M. K., Reeves, P. A., Olmstead, R. G. & Schemske, D. W. (2005) *Am. J. Bot.* **92**, 1899–1910.
- Ricklefs, R. E. (2003) *Proc. Biol. Sci.* **270**, 2285–2291.
- Richardson, J. E., Pennington, R. T., Pennington, T. D. & Hollingsworth, P. M. (2001) *Science* **293**, 2242–2245.
- McCune, A. R. (1997) in *Molecular Evolution and Adaptive Radiation*, eds. Givnish, T. J. & Sytsma, K. J. (Cambridge Univ. Press, Cambridge, U.K.), pp. 585–610.
- Verheyen, E., Salzburger, W., Snoeks, J. & Meyer, A. (2003) *Science* **300**, 325–329.
- Nee, S., Mooers, A. O. & Harvey, P. H. (1994) *Proc. Natl. Acad. Sci. USA* **89**, 8322–8326.
- Baldwin, B. G. & Sanderson, M. J. (1998) *Proc. Natl. Acad. Sci. USA* **95**, 9402–9406.
- Klak, C., Reeves, G. & Hedderson, T. (2003) *Nature* **427**, 63–65.
- Smith, J. M. B. & Cleef, A. M. (1988) *J. Biogeogr.* **15**, 631–645.
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., Da Fonseca, G. A. B. & Kent, J. (2000) *Nature* **403**, 853–858.
- Luteyn, J. L. (1999) *Mem. N.Y. Bot. Gard.* **84**, 1–278.
- Gregory-Wodzicki, K. M. (2000) *Geol. Soc. Am. Bull.* **112**, 1091–1105.
- Simpson, B. B. (1975) *Paleobiology* **1**, 273–294.
- Van der Hammen, T. & Cleef, A. M. (1986) in *High Altitude Tropical Biogeography*, eds. Vuilleumier, F. & Monasterio, M. (Oxford Univ. Press, New York), pp. 153–201.
- Burnham, R. J. & Graham, A. (1999) *Ann. Mo. Bot. Gard.* **86**, 546–589.
- Monasterio, M. & Sarmiento, L. (1991) *Trends Ecol. Evol.* **6**, 387–391.
- Bell, C. D. & Donoghue, M. J. (2005) *Organisms Divers. Evol.* **5**, 147–159.
- Von Hagen, K. B. & Kadereit, J. W. (2001) *Organisms Divers. Evol.* **1**, 61–79.
- Vuilleumier, F. (1970) *Am. Nat.* **104**, 373–388.
- Böhle, U.-T., Hilger, H. H. & Martin, W. F. (1996) *Proc. Natl. Acad. Sci. USA* **93**, 11740–11745.
- Ainouche, K. & Bayer, R. (1999) *Am. J. Bot.* **86**, 590–607.
- Lavin, M. T., Herendeen, P. S. & Wojciechowski, M. F. (2005) *Syst. Biol.* **54**, 575–594.
- Richardson, J. E., Weitz, F. M., Fay, M. F., Cronk, Q. C. B., Linder, H. P., Reeves, G. & Chase, M. W. (2001) *Nature* **412**, 181–183.
- Koch, M. & Al-Shehbaz, I. A. (2002) *Ann. Mo. Bot. Gard.* **89**, 88–109.
- Wojciechowski, M. F., Sanderson, M. J. & Hu, J.-H. (1999) *Syst. Bot.* **24**, 409–437.
- Losos, J. & Schluter, D. (2000) *Nature* **408**, 847–850.

30. Hooghiemstra, H. & Van der Hammen, T. (2004) *Philos. Trans. R. Soc. London B Biol. Sci.* **359**, 173–181.
31. Janzen, D. H. (1967) *Am. Nat.* **101**, 233–249.
32. Kozak, K. H., Weisrock, D. W. & Larson, A. (2005) *Proc. R. Soc. London Biol. Sci.* **273**, 539–546.
33. Hughes, C. E., Eastwood, R. J. & Bailey, C. D. (2006) *Philos. Trans. R. Soc. London B Biol. Sci.* **361**, 211–225.
34. Kocher, T. D. (2003) *Nature* **423**, 489–491.
35. Jørgensen, P. M. & León-Yánes, S., eds. (1999) *Catalogue of the Vascular Plants of Ecuador* (Missouri Botanical Garden Press, St. Louis).
36. Citerne, H., Luo, D., Pennington, R. T., Coen, E. & Cronk, Q. C. B. (2003) *Plant Physiol.* **131**, 1042–1053.
37. Ree, R. H., Citerne, H. L., Lavin, M. T. & Cronk, Q. C. B. (2004) *Mol. Biol. Evol.* **21**, 321–331.
38. Citerne, H. L. (2006) *Edinburgh J. Bot.* **62**, 119–126.
39. Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D. G. (1997) *Nucleic Acids Res.* **25**, 4876–4882.
40. Müller, K. (2005) *Appl. Bioinformatics* **4**, 65–69.
41. Goloboff, P. (2000) *NONA: A Tree Searching Program* (Fundación e Instituto Miguel Lillo Tucumán, Argentina).
42. Huelsenbeck, J. P. & Ronquist, F. (2001) *Bioinformatics* **17**, 754–755.
43. Nylander, J. A. A. (2004) *MRMODEL TEST* (Uppsala Univ., Uppsala), Version 2.
44. Sanderson, M. J. (2004) *R8S Users Manual* (Univ. of California, Davis), Version 1.7.