

Cathepsin S inhibitor prevents autoantigen presentation and autoimmunity

Kaoru Saegusa,¹ Naozumi Ishimaru,¹ Kumiko Yanagi,¹ Rieko Arakaki,¹ Kouichi Ogawa,¹ Ichiro Saito,¹ Nobuhiko Katunuma,² and Yoshio Hayashi¹

¹Department of Pathology, Tokushima University School of Dentistry, Tokushima, Japan

²Institute for Health Science, Tokushima Bunri University, Yamashirocho, Tokushima, Japan

The cysteine endoprotease cathepsin S mediates degradation of the MHC class II invariant chain Ii in human and mouse antigen-presenting cells. Studies described here examine the functional significance of cathepsin S inhibition on autoantigen presentation and organ-specific autoimmune diseases in a murine model for Sjögren syndrome. Specific inhibitor of cathepsin S (Clik60) *in vitro* markedly impaired presentation of an organ-specific autoantigen, 120-kDa α -fodrin, by interfering with MHC class II-peptide binding. Autoantigen-specific T cell responses were significantly and dose-dependently inhibited by incubation with Clik60, but not with inhibitors of cathepsin B or L. Clik60 treatment of mouse salivary gland cells selectively inhibited autopeptide-bound class II molecules. Moreover, the treatment with Clik60 *in vivo* profoundly blocked lymphocytic infiltration into the salivary and lacrimal glands, abrogated a rise in serum autoantibody production, and led to recovery from autoimmune manifestations. Thus, inhibition of cathepsin S *in vivo* alters autoantigen presentation and development of organ-specific autoimmunity. These data identify selective inhibition of cysteine protease cathepsin S as a potential therapeutic strategy for autoimmune disease processes.

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Introduction

Sjögren syndrome (SS) is an autoimmune disorder characterized by lymphocytic infiltrates and destruction of the salivary and lacrimal glands, and systemic production of autoantibodies to the ribonucleoprotein particles SS-A/Ro and SS-B/La (1–3). We have investigated an animal model for SS in *NFS/sld* mutant mice thymectomized 3 days after birth (3d-Tx) (4–9). All 3d-Tx *NFS/sld* mice develop autoimmune lesions in the salivary and lacrimal glands, starting at 3 weeks of age, and the disease mediated by CD4⁺ T cells is chronic and progressive (4, 5). Previously, we reported a 120-kDa α -fodrin autoantigen in the salivary gland tissues from SS model mice and identified autoantigen-specific T cell responses associated with Th1 cytokine production of IL-2 and IFN- γ (10). However, the role of antigen-presenting cells (APCs) in organ-specific T cell activation in this model has not yet been analyzed.

MHC class II molecules encounter and bind antigenic peptides as class II-peptide complexes on the cell sur-

face of APCs for recognition by CD4⁺ T cells (11–13). The molecular mechanisms leading to formation of class II-peptide complexes and presentation of antigen on the cell surface begin with synthesis of class II $\alpha\beta$ heterodimers in the endoplasmic reticulum. These class II $\alpha\beta$ heterodimers associate early during biosynthesis with a type II membrane protein, the invariant chain (Ii) (14, 15). Inhibition of Ii degradation in B lymphoblastoid cells and murine spleen cells induces accumulation of class II-associated Ii fragments and inhibition of class II-peptide formation (16–19). Selective inhibition of the proteases responsible for both these degradative processes is a potential mechanism for modulating the immune response. Several lysosomal proteases have been implicated in the processing of Ii and antigenic peptides. Cathepsin B, the most abundant lysosomal cysteine protease, has been tied to Ii degradation using purified class II-Ii complexes (20). Cathepsin L, a potent cysteine-class endoprotease, is specifically inhibited by a fragment of the alternatively spliced Ii form p41 (21). Cathepsin S containing potent endoproteolytic activity is highly expressed in the spleen and professional APCs and other class II-positive cells and is inducible by IFN- γ (22, 23). In mouse splenocytes, inhibition of cathepsin S also induces buildup of Ii breakdown products and attenuation of class II-peptide association, although the extent of this effect appears to be haplotype-dependent (24). We have developed specific inhibitors of cathepsin B (CA074), cathepsin L (Clik148), and cathepsin S (Clik60), *in vivo* as well as *in vitro* (25–27). Matsunaga et al. first reported that CA074 suppresses immune responses (28), suggesting that cysteine proteases in

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Address correspondence to: Yoshio Hayashi, Department of Pathology, Tokushima University School of Dentistry, 3 Kuramotocho, Tokushima 770-8504, Japan. Phone: 81-88-633-7327; Fax: 81-88-633-7327; E-mail: hayashi@dent.tokushima-u.ac.jp.

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Nonstandard abbreviations used: Sjögren syndrome (SS); thymectomized 3 days after birth (3d-Tx); nonthymectomized (non-Tx); antigen-presenting cell (APC); invariant chain (Ii); lymph node cell (LNC); ovalbumin (OVA); concanabalin A (ConA); mouse salivary gland (MSG); propidium iodide (PI).

lysosomes play an important role in the functional differentiation of MHC class II-restricted CD4⁺ T cells. However, it is uncertain whether the inhibition of cathepsins B, L, and S blocks generation of the antigenic peptide on the development of autoimmune diseases. To address this important issue, antigen processing and presentation after specific inhibition of cathepsins were examined in a murine model for SS.

Studies presented here suggest that cathepsin S plays an important role in processing of class II-Ii in autoantigen-presenting cells to generate class II molecules competent for binding antigenic peptide, and that inhibition of cathepsin S has important functional consequences in modulating the autoimmune response.

Methods

Mice. Female NFS/N-strain mice carrying the mutant gene *sld* (29) were reared in our specific pathogen-free mouse colony, and given food and water ad libitum. Thymectomy was performed on day 3 after birth (3d-Tx) in NFS/*sld* mice. Nonthymectomized (non-Tx) NFS/*sld* mice and C56BL/6 mice purchased from Charles River Japan Inc. (Atsugi, Japan) were used as controls.

Measurement of endogenous cathepsin activities. Salivary glands, regional lymph nodes, and spleens from 3d-Tx NFS/*sld* SS model, non-Tx NFS/*sld*, and control C57BL/6 mice were used to assay for cathepsin B, L, and S activity. Lysosomes were isolated for the assay by gentle homogenization of samples using a Teflon homogenizer (Microtec Co. Ltd., Funabashi, Japan) pestle in 0.25 M cold sucrose. The suspension was centrifuged at 3,500 *g* for 10 minutes at 4°C. The supernatant was centrifuged at 25,000 *g* for 20 minutes at 4°C. The resulting pellet was resuspended with 50 mM acetate buffer (pH 5.0). The suspension fluid was frozen and thawed three times to disrupt lysosomal membranes. After three cycles of freezing and thawing, the fluid was centrifuged and the supernatant was used as a mitochondria and lysosome fraction. Cathepsin activities were assayed with Z-Arg-Arg-methyl coumarylamide (Peptide Institute, Osaka, Japan) as substrate at pH 5.0 for cathepsin B, with Z-Phe-Arg-methyl coumarylamide for cathepsin L, and using the method described by Inubushi et al. (30) for cathepsin S. The reaction was initiated by addition of substrate (10 μM final concentration) after preincubation with the test compound for 3 minutes at 37°C. The fluorescence of the liberated 7-amino-4-methylcoumarin was measured in a fluorescence spectrophotometer (Hitachi Science Systems Ltd., Hitachinaka, Japan). Emission at 460 nm was measured with excitation at 370 nm.

Cathepsin inhibitors. Specific inhibitors for cathepsin B (CA074), cathepsin L (Clik148), and cathepsin S (Clik60) have been developed with the help of computer-graphic modeling based on the stereo-structure as described previously (25–27). The common fragment, *N*-(*L*-trans-carbamoyloxyrane-2-carbonyl)-phenylalanine-dimethylamide, is required for specific inhibition of cathepsin L. Seven novel inhibitors of the cathepsin L inhibitor

Katunuma (Clik) specifically inhibited cathepsin L at a concentration of 10⁻⁷ M in vitro, while almost no inhibition of cathepsins B, C, S, and K was observed. Four of the Clik inhibitors are stable and showed highly selective inhibition for hepatic cathepsin L in vivo. One of the Clik inhibitors contains an aldehyde group and specifically inhibits cathepsin S at 10⁻⁷ M in vitro (27).

In vivo treatment with cathepsin inhibitors. We examined the in vivo therapeutic effects of cathepsin S inhibitor (Clik60), cathepsin B inhibitor (CA074), and cathepsin L inhibitor (Clik148) in a murine model for SS. Each inhibitor (Clik60, CA074, and Clik148), dissolved in PBS, was administered intraperitoneally into model mice at doses of 0.1 mg/mouse/day from 4 weeks to 7 weeks (*n* = 10 for each), and then analyzed at 8 weeks, compared with untreated SS model mice (*n* = 7).

Histology. All organs were removed from the mice, fixed with 4% phosphate-buffered formaldehyde (pH 7.2), and prepared for histologic examination. The sections were stained with hematoxylin and eosin. Histological grading of the inflammatory lesions was done according to the method proposed by White and Casarett (31). These slides were scored by three independent, pathologists in a blinded manner.

Proliferation assay. Single-cell suspensions of spleen cells or regional lymph node cells (LNCs) from 3d-Tx, non-Tx NFS/*sld*, and C57BL/6 mice were cultured in 96-well flat-bottom microtiter plates (Nalge Nunc Intl. Co., Rochester, New York, USA) in RPMI-1640 containing 10% FCS, penicillin/streptomycin, and β-mercaptoethanol. For proliferation assay, a total of 5 × 10⁵ cells per well were cultured for 72 hours under stimulation of recombinant α-fodrin protein (JS-1, 10 μg/ml) (10), ovalbumin (OVA) (10 μg/ml), and concanabalin A (ConA) (5 μg/ml), and pulsed with 1 μCi/well of [³H]thymidine (NEN Life Science Products Inc., Boston, Massachusetts, USA) during the final 20 hours of the culture. [³H]thymidine incorporation was evaluated using an automated β liquid scintillation counter. We further examined the in vitro preventive effects of cathepsin inhibitors (10⁻⁷ to 10⁻⁴ M CA074, Clik148, and Clik60) for antigen-specific proliferative T cell responses. T cell purification was done using CD4 mAb-bounded immunomagnetic beads (Dyna, Oslo, Norway).

Flow cytometric analysis. Spleen cells and regional LNCs from 3d-Tx and non-Tx NFS/*sld* mice were analyzed by flow cytometry. Single-cell suspensions were stained with antibodies conjugated to phycoerythrin (anti-CD4; Cedarlane Laboratories Ltd., Hornby, Ontario, Canada) and FITC (anti-CD8 from Cedarlane Laboratories Ltd.; anti-CD44, anti-CD45RB, anti-Mel-14, and anti-I-A^b from Pharmingen, San Diego, California, USA), and analyzed with an EPICS (Beckman Coulter, Miami, Florida, USA). Apoptotic or necrotic cells were detected by flow cytometry with an EPICS (Beckman Coulter) using the Annexin V-FITC Apoptosis Detection Kit (Genzyme Pharmaceuticals Co. Ltd., Cambridge, Massachusetts, USA). Viable cells (1 × 10⁵) were

gated according to size and scatter to eliminate dead cells and debris from analysis.

Measurement of fluid secretion. Detection of tear and saliva volume of the treated and untreated SS animal model of NFS/*sld* mice was done according to a method as described (32). Five mice in each group were analyzed.

Measurement of cytokine production. Cytokine production from spleen cells of 3d-Tx and non-Tx NFS/*sld* mice was tested by two-step sandwich ELISA using a mouse IL-2, IL-4, and IFN- γ kit (Genzyme Pharmaceuticals). In brief, culture supernatants from spleen cells activated with immobilized anti-CD3 mAb (10 μ g/ml) (Cedarlane Laboratories Ltd.) for 3 days were added to microtiter plates precoated with anti-IL-2, -IL-4, and -IFN- γ capture antibody and incubated overnight at 4°C. After addition of biotinylated detecting antibody and incubation at room temperature for 45 minutes, avidin-peroxidase was added and incubated at room temperature for 30 minutes. Plates were washed extensively with 1% Tween in PBS between each step. Finally, 2,2'-azino-di-3-ethylbenzothiazoline sulfonate (ABTS) substrate containing H₂O₂ was added and the colorimetric reaction was read at an absorbance of 450 nm using an automatic microplate reader (BioRad Laboratories Inc., Hercules, California, USA). The concentrations (in pg/ml) of IL-2, IL-4, and IFN- γ were calculated according to the standard curves produced by various concentrations of recombinant cytokines.

Primary culture of mouse salivary gland cells. Mouse salivary gland (MSG) epithelial cells were prepared as previously described (33). Briefly, MSGs were minced into 1-mm² pieces, washed with HBSS without Ca²⁺ and Mg²⁺, and placed in a 60-mm dish containing HBSS with 0.76 μ g/ml EDTA, 4.9 μ g/ml L-ascorbic acid, and 4.9 μ g/ml reduced glutathione. Fragments were washed with DMEM/STI and placed in a mixture of collagenase (type I, 75 U/ml) and hyaluronidase (type IV, 500 U/ml) dissolved in DMEM/F12 containing 10% FBS. The first digest suspension was passed through sterile 100- μ m nylon mesh filter and were redigested for 30 minutes by the same digestion procedure, and then the digest suspension was passed through a 100- μ m nylon mesh filter. Adherent cells, after culture in MEM containing 10% FBS for 24 hours at 37°C, were isolated as salivary gland epithelial cells. We confirmed that the cells over 95% were positively stained with anti-keratin polyclonal antibody.

Detection of serum autoantibodies against 120-kDa α -fodrin. Serum autoantibody production against 120-kDa α -fodrin was analyzed by immunoblotting and ELISA as described previously (6, 7, 10).

Confocal immunofluorescence analysis. IFN- γ -stimulated and nonstimulated MSG cells were fixed with 1% paraformaldehyde and were incubated with mAb to I-A^b molecule (PharMingen) and FITC-labeled AFN₃₀₃₋₃₁₈ peptide (described below). In vitro effects of cathepsin inhibitors were examined by the preincubation with each inhibitor (10⁻⁵ M) for 6–24 hours. The

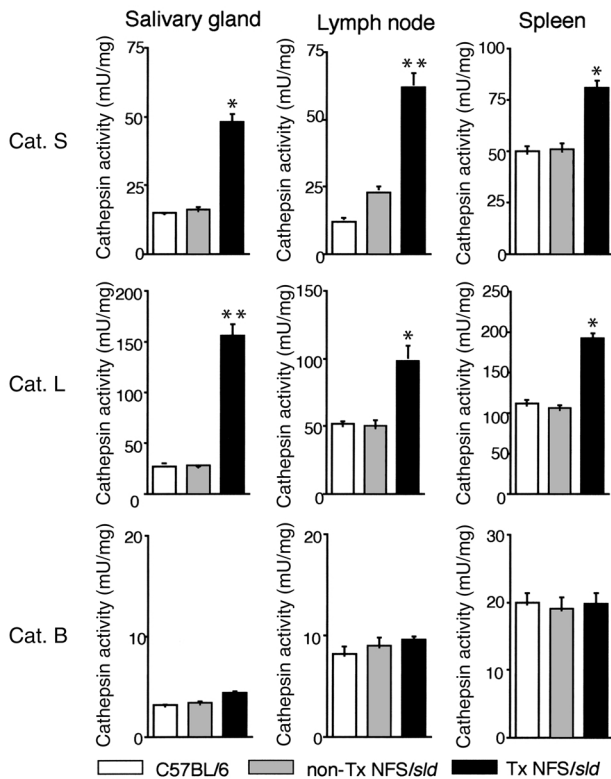
labeled second antibody was Texas red-conjugated goat anti-mouse IgG (Molecular Probes Inc., Eugene, Oregon, USA). For microscopy, a Leica TCS-NT laser scanning microscope (Leica Microsystems Nussloch GmbH, Nussloch, Germany) was used.

Results

Endogenous activities of cathepsins B, L, and S in tissue samples. We first investigated endogenous activities of cathepsin S, cathepsin L, and cathepsin B in tissue samples of the salivary glands, regional lymph nodes, and spleens from SS model mice and controls. As shown in Figure 1, the activities of both cathepsin S and cathepsin L were significantly higher in the salivary glands, lymph nodes, and spleens from model mice than those of controls. No difference in activities of cathepsin B was found in any organ from these mice. Significantly inhibitory effects of cathepsin S inhibitor (Clik60), cathepsin L inhibitor (Clik148), and cathepsin B inhibitor (CA074) were observed in each organ in a dose-dependent manner. Moreover, we confirmed that cathepsin S activity in each organ was clearly inhibited by Clik60, but not by Clik148 and CA074 (data not shown).

Inhibitory effect of a specific inhibitor of cathepsin S (Clik60) on proliferative T cell response to autoantigen. To address the role of autoantigen-reactive T cells, we examined the proliferative T cell responses in the LNCs and spleen cells from model mice and controls. We found that the LNCs and spleen cells in SS model mice, but not in non-Tx NFS/*sld* and C57BL/6 control mice, at 8 weeks of age showed a significant increase in autoantigen-specific (JS-1-specific) T cell proliferation (Figure 2a; data of the spleen cells not shown). No significant differences were observed in the proliferative response stimulated with OVA (10 μ g/ml) and ConA (5 μ g/ml) among these mice. We examined the inhibitory effects of cathepsin inhibitors against autoantigen-specific T cell responses in vitro. In regional LNCs, but not in spleens, from model mice, a significantly inhibitory effect of Clik60 was observed in a dose-dependent manner (Figure 2b). In contrast, no inhibitory effects of Clik148 and CA074 were found. Annexin V/propidium iodide (PI) flow cytometric analysis revealed a small proportion of early apoptotic cells (annexin V-positive, PI-negative) and necrotic cells (annexin V-positive, PI-positive), indicating that Clik60 is not toxic and favors T cell survival in the LNCs (Figure 2c). Indeed, the Clik60 inhibitors at 10⁻⁷ to 10⁻⁴ M concentrations do not block T cell proliferation to ConA (Figure 2d). These findings indicated that the dose of Clik60, 10⁻⁵ to 10⁻⁴ M, was sufficient to inactivate the autoantigen-specific T cell responses in vitro.

In vivo administration of cathepsin inhibitors. We next examined the in vivo therapeutic effects of Clik60, Clik148, and CA074 in a murine model for SS. Interestingly, the treatment with intraperitoneal injection of Clik60 (0.1 mg/mouse/day) was effective in preventing the development of autoimmune lesions in the lacrimal ($P < 0.01$), parotid ($P < 0.05$), and submandibular



($P < 0.05$) glands of the SS model mice, but not in those of groups injected with CA074 and Klik148 (Figure 3a). Representative histological features in the salivary and lacrimal glands are shown in Figure 3b. Moreover, the average saliva and tear volumes of Klik60-treated model mice were significantly higher than those of the untreated SS model mice (Figure 3c). The activation markers (CD44^{high}, CD45RB^{low}, Mel-14^{low}) were clearly downregulated in LNCs gated on CD4 from Klik60-treated model mice (Figure 4a). In addition, autoantigen-specific (JS-1-specific) T cell response was significantly inhibited in LNCs from Klik60-treated mice (Figure 4b). Culture supernatants from anti-CD3 mAb-stimulated splenic T cells obtained from Klik60-treated mice, but not those obtained from non-Tx

Figure 2

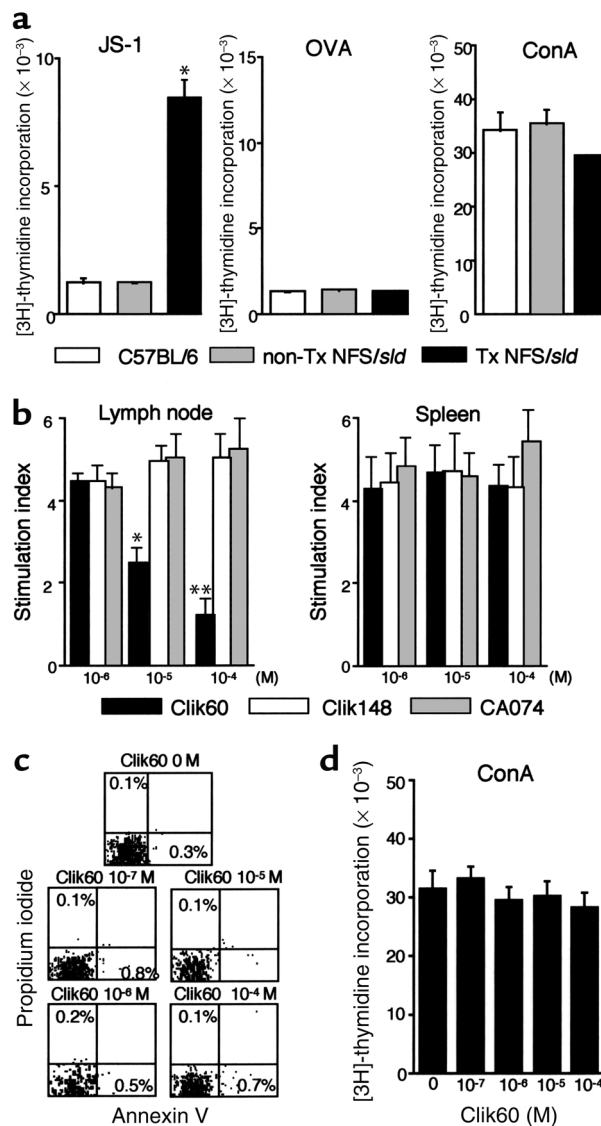
(a) Detection of proliferative T cell response of LNCs from SS model mice to organ-specific autoantigen (JS-1) ($*P < 0.001$, Student's *t* test). No differences were found in OVA (10 $\mu\text{g/ml}$) and ConA (5 $\mu\text{g/ml}$) responsiveness. (b) In vitro preventive effect of proliferative T cell response of LNCs, but not spleen cells, to JS-1 by Klik60 in a dose-dependent manner. No inhibitory effects of Klik148 and CA074 were found. Data are expressed as stimulation indices \pm SEM. Three experiments from each group were performed at 8 weeks of age, and the mean values of index were statistically significant at $*P < 0.05$ and $**P < 0.01$ (Student's *t* test). (c) Annexin V/propidium iodide (PI) flow cytometric analysis revealed a small proportion of early apoptotic cells (annexin V-positive, PI-negative) and necrotic cells (annexin V-positive, PI-positive). (d) The inhibitors at 10^{-7} to 10^{-4} M concentrations of Klik60 do not block T cell proliferation to Con A (5 $\mu\text{g/ml}$).

Figure 1

Endogenous cathepsin (Cat.) activities in tissue samples from SS model mice and controls. The activities of both cathepsin S and cathepsin L were significantly higher in the salivary glands, regional lymph nodes, and spleens from model mice than in those from controls ($*P < 0.05$, $**P < 0.01$, Student's *t* test). Cathepsin activities were assayed as described in Methods.

NFS/*sld* mice, contained high levels of IL-4, but low levels of IL-2 and IFN- γ , by ELISA (Figure 5, a and b). Moreover, serum autoantibody production against 120-kDa α -fodrin was exclusively inhibited in Klik60-treated mice, but not in other groups (Figure 5c). A dilution curve, obtained by ELISA, illustrates that each case is capable of reacting with antigen (Figure 5d). These results strongly suggest that Klik60 plays an important role in preventing autoantigen presentation to generate class II molecules competent for binding antigenic peptide, resulting in inhibition of autoimmunity.

Possible role of MSG cells in autoantigen presentation. It has been well known that nonlymphoid cells that



express MHC class II molecules provoke autoimmune responses in combination with costimulation (34, 35). To determine whether specific inhibition of cathepsin S activity can functionally alter antigen presentation, primary cultured MSG cells, regional LNCs, and spleen cells from model mice and non-Tx control mice were compared in terms of their capacity to express class II molecule, and to react with organ-specific autoantigen as determined by flow cytometry and proliferation assay. Among these cells, a large proportion of class II-expressing (I-A^q-expressing) cells was observed on MSG cells compared with spleen cells and LNCs from SS model mice (Figure 6a). Moreover, class II molecule can be stably induced by IFN- γ stimulation on MSG cells from non-Tx NFS/*sld* control mice by flow cytometry (Figure 6b). Indeed, purified CD4⁺ T cells from model mice are capable of responding to IFN- γ -stimulated MSG cells, but not with nonstimulated MSG cells (Figure 6c). Autoantigen-stimulated (JS-1-stimulated) T cell response of LNCs using MSG cells was significantly inhibited by incubation with Clk60, but not incubation with Clk148 and CA074 (Figure 6d). These results suggest that MSG cells may function, at least in part, as autoantigen-presenting cells in the development of murine SS, and that inhibition of cathepsin S prevents both autoantigen presentation in the salivary gland cells and autoantigen-specific T cell proliferation.

Inhibition of cathepsin S prevents class II-peptide association. We confirmed that activity of cathepsin S in MSG cells was clearly inhibited by Clk60 (10⁻⁵ M) (Figure 7a). Surface expression of class II molecule induced by IFN- γ stimulation in MSG cells from non-Tx NFS/*sld* mice was clearly inhibited by incubation with Clk60, but not incubation with Clk148 and CA074, as determined by flow cytometry (Figure 7b). MHC class II molecules are synthesized with their peptide-binding site blocked by Ii, and they acquire the capacity to bind antigens only after Ii has been degraded in the compartments. Cathepsin S is known to play a pivotal role in proteolytic degradation of the Ii as a prerequisite for peptide binding to class II molecules (36). Recently, we have identified the pathogenic T cell epitope peptide (AFN₃₀₃₋₃₁₈) on α -fodrin autoantigen involved in autoimmune responses in this model (unpublished data). We observed colocalization of FITC-labeled AFN₃₀₃₋₃₁₈ peptide and class II (I-A^q) molecule in the IFN- γ -stimulated MSG cells (Figure 7c), suggesting that MSG cells may play a role in presenting autopeptide in the autoimmune responses. Moreover, confocal immunofluorescence analysis confirmed that inhibition of cathepsin S prevents the surface expression of peptide-class II complex formation, while inhibition of cathepsin L and cathepsin B did not effect expression of peptide-bound class II molecules (Figure 7c).

Discussion

Therapeutic strategies for modulating the immune response in autoimmune disorders have centered on

altering generation of the effector molecules that mediate the inflammatory reactions, e.g., corticosteroids, cyclosporin, and inhibitors of cytokines (37, 38). Our experiments demonstrate that selective inhibition of cysteine proteases is an additional potential strategy for modulating the autoantigen-derived immune response in class II-restricted disease processes. To our knowledge, this manuscript presents the first evidence that selective inhibition of cysteine protease cathepsin S *in vivo* has important functional consequences for MHC class II-dependent immune responses in autoimmune disease.

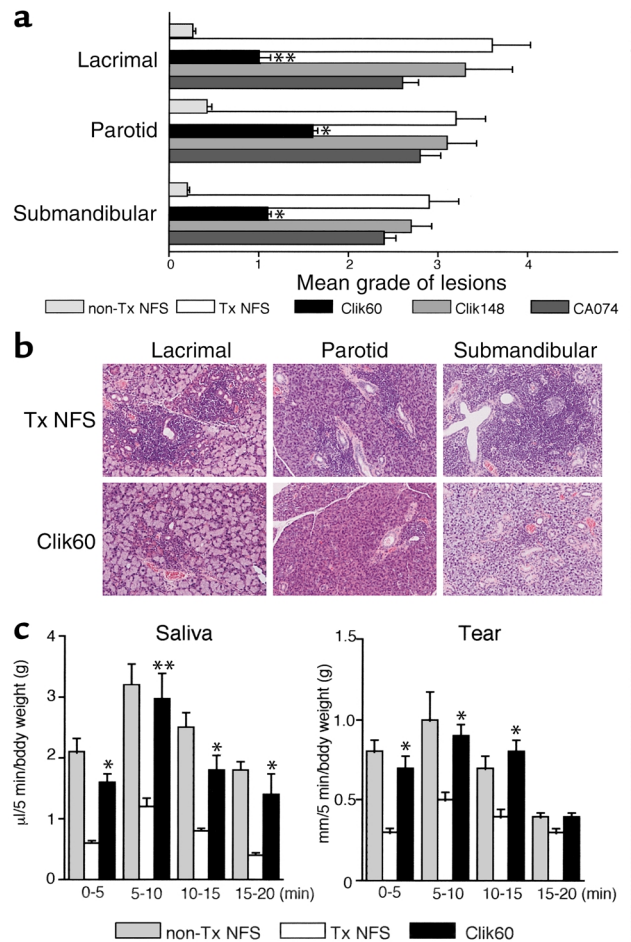


Figure 3

(a) Effects of *in vivo* administration of intraperitoneal injection of cathepsin inhibitors (Clk60, Clk148, and CA074). Each inhibitor was administered into SS model mice ($n = 10$ for each) at doses of 0.1 mg/mouse/day from 4 weeks to 7 weeks, and the results were analyzed at 8 weeks compared with those in untreated SS model mice ($n = 7$). Treatment with intraperitoneal injection of Clk60, but not treatment with Clk 148 and CA074, was effective in preventing the development of autoimmune lesions in the lacrimal, parotid, and submandibular glands of the SS model mice ($*P < 0.01$, $**P < 0.005$, Student's *t* test). (b) Representative histologic features in the salivary and lacrimal glands showing preventive effects in Clk60-treated mice at 8 weeks of age. (c) The average saliva and tear volume of the SS model mice treated with Clk60 was significantly higher than that of control mice. Results are expressed as mean \pm SEM in five mice examined per group ($*P < 0.05$, $**P < 0.005$, Student's *t* test).

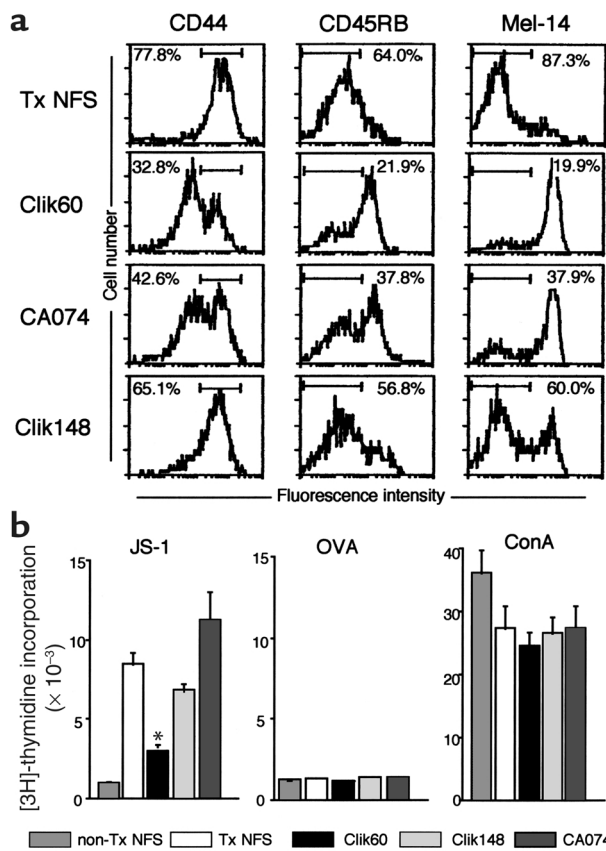


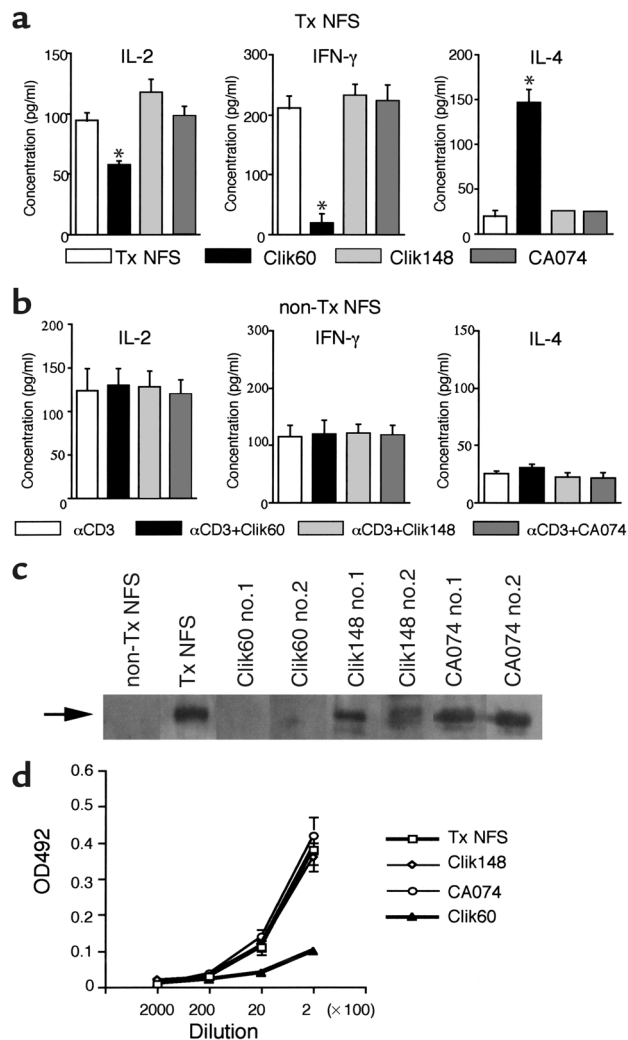
Figure 4 (a) Upregulation of the activation markers (CD44^{high}, CD45RB^{low}, Mel-14^{low}) in LNCs gated on CD4 from Clik60-treated mice, compared with control mice. (b) SS model mice treated with Clik60 showed a significant decrease of autoantigen-specific (JS-1-specific) T cell proliferation in LNCs, compared with controls ($P < 0.01$, Student's t test). Data are expressed as cpm per culture in triplicate.

cells was observed on MSG epithelial cells from SS model mice, and MHC class II molecule can be stably induced by IFN- γ stimulation on MSG cells from syngeneic control mice. Moreover, autoantigen-stimulated (JS-1-stimulated) proliferative T cell response using MSG cells was clearly inhibited by the incubation with cathepsin S inhibitor (Clik60). In light of these results, it is possible that the salivary gland epithelial cells may function, at least in part, as autoantigen-presenting cells in the development of murine SS, and that inhibition of cathepsin S prevents autoantigen presentation and subsequent peptide binding by class II molecules. MHC class II molecules bind a diverse array of peptides derived from the endocytic pathway and present them to CD4⁺ T cells. APCs have a pool of active class II mol-

We reported previously a cleavage product of 120-kDa α -fodrin as an important autoantigen in the pathogenesis of SS in both an animal model and humans (10). The observation that thyrocytes express MHC class II molecules in Graves thyroiditis led Bottazzo and coworkers (39) to theorize that nonlymphoid cells that express MHC class II molecules provoke autoimmune responses by presenting autoantigens. Salivary gland epithelial cells from SS patients have been found to express MHC class II molecules in a large proportion of biopsy samples, and the number of positive cells generally increases with the number of lymphocytes infiltrating the gland (40–42). In the present experiments, a large proportion of class II-expressing (I-A^q-expressing)

Figure 5

(a) Downregulation of Th1 cytokine production in SS model mice treated with Clik60. Culture supernatants from anti-CD3 mAb-stimulated splenic T cells from Clik60-treated model mice (8 weeks old) contained a high level of IL-4, but low levels of IL-2 and IFN- γ , as measured by ELISA ($*P < 0.005$, $**P < 0.001$, Student's t test). (b) No significant cytokine production in culture supernatants from anti-CD3 mAb-stimulated splenic T cells from non-Tx NFS/*sld* mice (8 weeks old). (c) Production of serum autoantibodies to 120-kDa α -fodrin was diminished in two mice (no. 1 and no. 2) treated with Clik60, as shown by immunoblotting. (d) A dilution curve illustrates that each case is capable of reacting with antigen. The same dilution of sera was examined in all cases by ELISA. OD492, optical density of 492 nm.



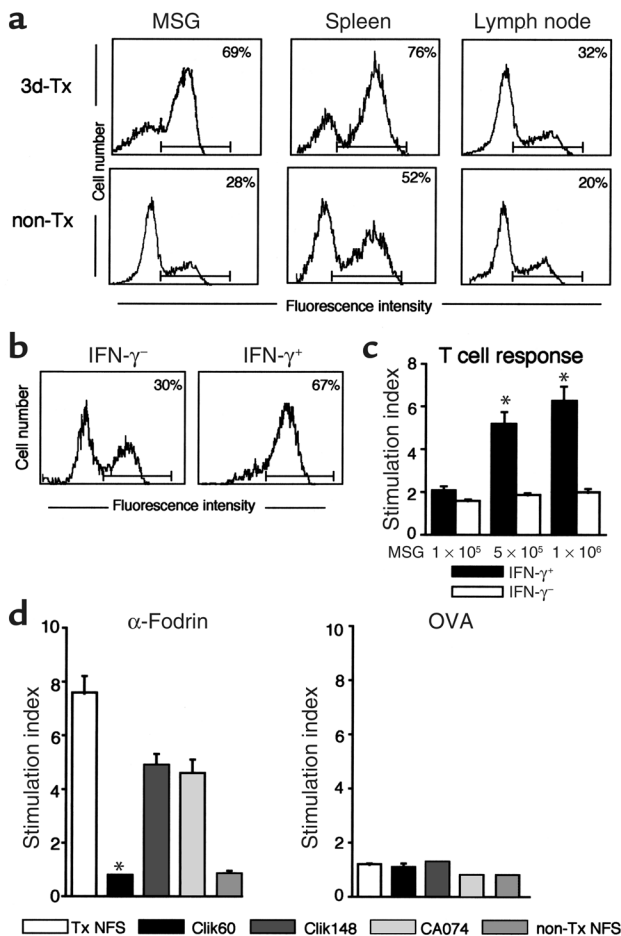


Figure 6

(a) A large proportion of class II-expressing (I-A^d-expressing) cells observed on MSG cells from SS model mice compared with those on spleen and LNCs, as determined by flow cytometry. (b) MHC class II molecule induced by IFN- γ (100 U/ml) stimulation in MSG cells from non-Tx NFS/*sld* control mice by flow cytometry. (c) Significant proliferative responses of purified CD4⁺ T cells (2×10^5) from model mice with IFN- γ -stimulated MSG cells (5×10^5 and 1×10^6 , * $P < 0.01$, Student's *t* test), but not from those with non-stimulated MSG cells. (d) Autoantigen-stimulated (JS-1-stimulated) proliferative T cell response of LNCs using MSG cells from SS model mice was significantly inhibited by the incubation with Clik60, but not the incubation with Clik148 and CA074 (* $P < 0.01$, Student's *t* test). Data are expressed as stimulation indices \pm SEM. Three experiments from each group were performed.

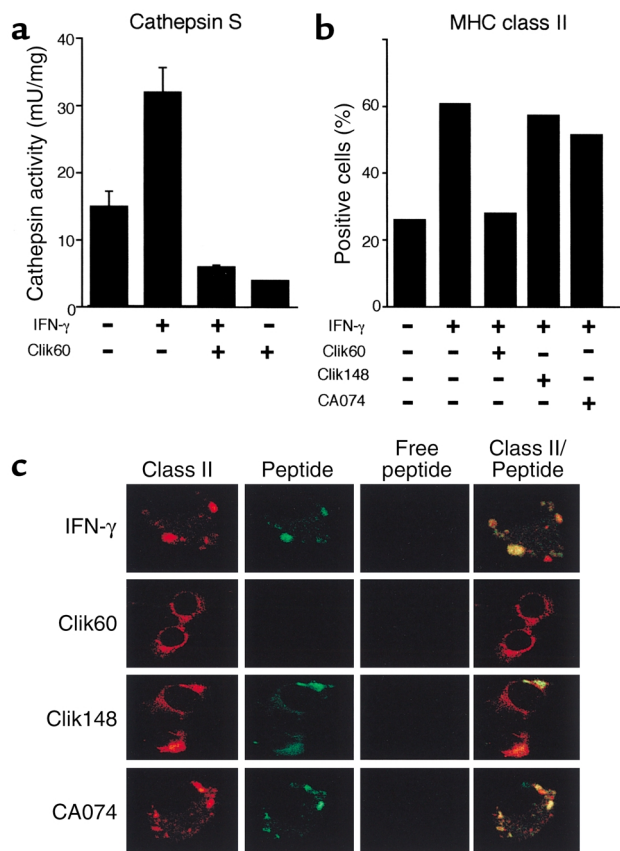
ecules on their surface that can quickly load peptides from the extracellular milieu for T cell presentation (43). The kinetics of this T cell response may correlate with the kinetics of peptide binding to the surface class II MHC molecules (44–46).

Recent studies using cathepsin-deficient mice have shown that cathepsin D and cathepsin B are unnecessary for MHC class II presentation (47, 48), whereas cathepsin S and cathepsin L are important for antigen presentation by discrete populations of cells (49–52). Active cathepsin S was detected in B cells, dendritic cells, and peritoneal macrophages, where it was shown to be involved in the late stages of Ii degradation (50–52). By contrast, cathepsin L was detected only in macrophages and cortical thymic epithelial cells, where a defect resulted in severely impaired CD4⁺ T cell selec-

tion (49). In addition to cathepsin S and cathepsin L, cathepsin F has been recently implicated in Ii degradation and antigen presentation in alveolar macrophages (53). Thus, it would appear that different APCs utilize distinct cathepsins to mediate late-stage Ii degradation and to regulate MHC class II presentation. Moreover, cathepsin S-deficient mice showed diminished susceptibility to collagen-induced arthritis, suggesting a potential therapeutic target for regulation of immune responsiveness (50). In this study, the treatment with cathepsin S inhibitor was effective in preventing the development of autoimmune lesions in the salivary and lacrimal glands of the SS model mice. Almost entire remissions of SS were induced in the salivary and

Figure 7

(a) Detection of cathepsin S activity in MSG cells and its inhibition by Clik60 (10^{-5} M). (b) Surface expression of class II molecule induced by IFN- γ stimulation on MSG cells was clearly inhibited by the incubation with Clik60, but not the incubation with CA074 and Clik148, as determined by flow cytometry. (c) Colocalization of FITC-labeled AFN₃₀₃₋₃₁₈ peptide and class II (I-A^d) molecule in the IFN- γ -stimulated MSG cells. Prevention of the surface expression of peptide–class II complex formation was confirmed by the inhibition of Clik60, but not by inhibition of Clik148 and CA074.



lacrimal glands by the Clik60 treatment. In addition, Clik60-treated mice showed a significant downregulation of autoantigen-specific (JS-1-specific) T cell response and Th1 cytokine expressions. Although we could not exclude the possibility that the Clik60 treatment in vivo inhibits only cathepsin S and that other enzymes could be involved, these results indicate that cathepsin S inhibitor plays an important role in preventing autoantigen presentation which is followed by inhibition of autoimmunity.

Ubiquitous expression and involvement in the terminal degradation of proteins that intersect the endocytic pathway were previously perceived to be the hallmarks of these proteinases. However, recent evidence has demonstrated that several cathepsins are expressed in a tissue-specific fashion and that partial proteolysis of specific biological targets is a key function of cathepsins in antigen processing. Analysis of cathepsin L-deficient mice revealed a profound defect in Ii degradation in thymic cortical epithelial cells but not in bone marrow-derived APCs. We have recently identified the pathogenic T cell epitope peptide (AFN₃₀₃₋₃₁₈) on α -fodrin autoantigen involved in organ-specific autoimmune responses (unpublished data). We confirmed here that MSG cells may play an important role for presenting autopeptide in the autoimmune responses, and that inhibition of cathepsin S prevents the surface expression of peptide-class II complex formation, while inhibition of cathepsin L and cathepsin B did not. The differential expression of proteinases by distinct APCs may affect the types of peptides that are presented to T cells and thereby the immune responses that are ultimately generated.

Taken together, our experiments support the hypothesis that cathepsin S, previously shown to be important in Ii processing in vitro, regulates MHC class II function and subsequent autoimmune responses in vivo. Thus, selective inhibition of cysteine protease cathepsin S may have important therapeutic potential in modulating class II-restricted autoimmune processes.

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