Origin and evolution of circadian clock genes in prokaryotes

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Regulation of physiological functions with approximate daily periodicity, or circadian rhythms, is a characteristic feature of eukaryotes. Until recently, cyanobacteria were the only prokaryotes reported to possess circadian rhythmicity. It is controlled by a cluster of three genes: *kaiA***,** *kaiB***, and** *kaiC***. Using sequence data of 70 complete prokaryotic genomes from the various public depositories, we show here that the** *kai* **genes and their homologs have quite a different evolutionary history and occur in Archaea and Proteobacteria as well. Among the three genes,** *kaiC* **is evolutionarily the oldest, and** *kaiA* **is the youngest and likely evolved only in cyanobacteria. Our data suggest that the prokaryotic circadian pacemakers have evolved in parallel with the geological history of the earth, and that natural selection, multiple lateral transfers, and gene duplications and losses have been the major factors shaping their evolution.**

^tircadian clock genes are a vital and essential feature of eukaryotes (1). Cyanobacteria were the first prokaryotes reported to have the circadian clock regulated by a cluster of three genes: *kaiA*, *kaiB*, and *kaiC* (2). Cyanobacteria are among the oldest organisms on the earth, and they are among the most successful in terms of ecological plasticity and adaptability (3). In adaptation strategy of cyanobacteria, circadian clock genes are of particular importance, because they underlie fundamental physiological processes such as the regulation of nitrogen fixation, cell division, and photosynthesis (4). The clock genes are ubiquitous in cyanobacteria (5). In most cyanobacteria, these genes were reported as a single copy (2, 5), although some of them (*kaiB*; ref. 6) or even a whole cluster (7) may be duplicated. They were shown to operate as a single unit and to follow a feedback model of regulation: *kaiA* positively affects *kaiBC* promoter, whereas overexpression of *kaiC* represses it (2, 8). Among these genes, $kai\bar{C}$ is a crucial component of clock precession in cyanobacteria (9–11).

Although the *kai* genes are under comprehensive study with regard to the mechanism of action, their evolution has yet to be resolved completely. The *kaiC* gene has a double-domain structure, and each of the domains has an ATP/GTP-binding site, or Walker's motif (2, 11). Based on its structure and sequence homology, the *kaiC* genes were classified as a family related to the *RecA* gene family of ATP-dependent recombinases (12). In addition to the *kaiC* genes with the typical double-domain structure, there are many single-domain homologous genes in Archaea and Proteobacteria. It was assumed that an ancestral single-domain *kaiC* gene was horizontally transferred from Bacteria to Archaea and then the double-domain *kaiC* evolved through duplication and subsequent fusion in Archaea (12). Although the evolution of the *kaiC* genes has been hypothesized, no data or hypotheses are available regarding the evolution of two other circadian clock genes, *kaiA* and *kaiB*. The evidence about the key role of *kaiC* in cyanobacterial clock regulation (9, 11), along with its homology to archaeal *RecA* genes, suggests that this gene is evolutionarily the oldest among the three.

In this study we reconstruct the origin and evolutionary patterns of the circadian clock genes in prokaryotes. Using available sequences from the public databases, we performed extensive phylogenetic analysis of the *kai* genes. The results suggest that the three prokaryotic circadian pacemakers have quite different evolutionary histories, and two of them, *kaiA* and *kaiB*, originated in cyanobacteria. The three-gene *kaiABC* cluster itself evolved $\approx 1,000$ Mya.

Materials and Methods

DNA and Protein Sequences. The annotated and homolog sequences of the *kai* genes were retrieved from GenBank and public databases of the Department of Energy Joint Genome Institute (www.jgi.doe.gov/JGLmicrobial/html/index.html) by using the gapped BLASTP and PSI-BLAST (13) and the respective amino acid sequences of *Anabaena* sp. strain PCC 7120 (Gen-Bank accession no. AP003591) as queries. Among the *kaiC* homologs from the other Prokaryota, which were about half the length of the *kaiC* genes from Cyanobacteria, only those matching both domains of the *kaiC* genes were chosen for further analysis. Multiple protein sequence alignments were constructed by using CLUSTALW (14) and manually adjusted based on structural considerations. Alignments of the nucleotide sequences were modified manually according to the respective amino acid alignments.

For the comparative phylogenetic analysis, we used sequences of 16S rRNA gene from the respective or closely related strains. This gene is a common marker for evolutionary studies of cyanobacteria (15, 16). The sequences obtained from the public databases were aligned with CLUSTALW (14) and adjusted by visual inspection. The list of used sequences is given in Table 1, which is published as supporting information on the PNAS web site, www.pnas.org.

Sequence Polymorphism Analysis. Due to saturation, synonymous substitutions were not computed. For the *kaiB* genes, we estimated the number of nonsynonymous differences (*K*a). To determine the DNA substitution model corresponding to our data, we used MODELTEST 3.0 software (17). It seemed that Kimura's two-parameter model (18) with gamma distribution and a transition/transversions ratio equal to 0.56 fit our data best. Therefore, nonsynonymous nucleotide substitutions in the *kaiB* genes were calculated by using the modified Nei–Gojobori method (19) with Jukes–Cantor correction for multiple substitutions at the same site and the above transition/transversions ratio. The MEGA 2.1 software (20) was used for the computations of K_a .

Phylogenetic Analysis. The alternative topologies of the phylogenetic trees were obtained initially by using maximum-parsimony (21) and neighbor-joining algorithms (22) and then evaluated with maximum-likelihood (ML) method (23) as implemented in TREE-PUZZLE 5.0 software (24) to find the best topology and test a molecular clock hypothesis. The ML approach with a local

Abbreviations: Mya, million years ago; ML, maximum likelihood.

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clock (25) realized in PAML software (26) then was applied to estimate the lengths of the branches. Those for the 16S rRNA tree were computed based on the Tamura–Nei model of substitutions (27), and those for the *kai* genes were estimated by using respective amino acid sequences and based on the WAG matrix of substitutions (28). The statistical support for the nodes of the trees was obtained by the bootstrapping procedure with 1,000 replications.

To detect lateral transfers, the congruency of the obtained phylogenetic trees of species (16S rRNA tree) and the *kai* genes was estimated by using Templeton's test (29) and the winningsites test as implemented in PAUP* 4.0b10 (30). Most of the taxa having multiple representatives in the *kaiC* data set (e.g., *Chloroflexus* 1 and 2; Table 1) did not define monophyletic clusters in the *kaiC* tree. Because there is only one respective representative in the 16S rRNA tree (for example, *Chloroflexus*), it might generate the taxonomic ambiguity. To avoid it, we used all possible combinations of the trees (96 in total) in the analysis.

Results

Occurrence of kai Genes in Prokaryotes. A BLAST search in the available completed prokaryotic genomes revealed homologs of the *kaiC* genes in both domains of prokaryotes, Archaea and Bacteria. In Archaea, these genes were found in the species of almost all well defined major taxa (except *Methanopyri* and *Thermoplasmata*). In Bacteria, the *kaiC* genes were not so ubiquitous; they were found only in four major taxa: Proteobacteria, Thermotogae, Chloroflexi, and Cyanobacteria. No such genes were found in the other bacteria. Importantly, *kaiC* homologs of Chloroflexi and Proteobacteria were the long (double-domain) versions of the gene, similar to those of Cyanobacteria, whereas most of the *kaiC* homologs of Archaea were the short (single-domain) versions (Table 1). However, within each domain, they are characteristic only to some taxa. Their occurrence in prokaryotes sometimes lacks predictability. For example, among Methanococci, *kaiC* homologs appear in *Methanococcales* (*Methanococcus jannaschii*) but not in *Methanosarcinales* (*Methanosarcina barkeri*). In Proteobacteria, such irregularity is observed even within the same orders: For example, among *Xanthomonadales*, the *kaiC* gene occurs in *Xanthomonas campestris* but not in *Xylella fastidiosa*. Cyanobacteria are the only prokaryotic kingdom in which all species appear to possess the *kaiC* genes.

Structure and Sequence Polymorphism of kai Genes in Prokaryota. The *kaiC* genes and their homologs in prokaryotes can be divided into two major groups by their length. The longer genes usually have two ATP/GTP -binding domains and have approximately twice the length than that of the shorter genes. The shorter homologs usually match both domains but always have higher similarity to the first domain of the longer *kaiC* genes. Proteobacteria, Chloroflexi, and Cyanobacteria possessed only the double-domain genes, but in Archaea both double- and single-domain homologs occurred.

Similar to the *kaiC* genes, the *kaiB* and *kaiA* genes are variable in length. In Archaea, Chloroflexi, Proteobacteria, and unicellular cyanobacteria $kaiB$ genes are \approx 300 bp long, whereas in filamentous cyanobacteria much longer (up to 858 bp) copies also occur. The *kaiA* genes were found only in cyanobacteria and varied in length; the genes from unicellular cyanobacteria (*Synechococcus* and *Synechocystis*) were 1.5–2 times longer (852–900 bp) as compared with those from filamentous cyanobacteria (*Anabaena* and *Nostoc*). These genes seem to be the least conserved among the *kai* genes; even in closely related species such as *Synechococcus* sp. PCC7942 and *Thermosynechococcus elongatus* their amino acid sequences share only 43.6% identity. However, in contrast to the respective *kaiC* genes, shorter homologs of the *kaiA*and *kaiB*genes match only one segment of their longer versions, which is located closer to the

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3' terminus. It suggests that the latter have likely not evolved through duplication.

In some species, the *kaiB* and/or *kaiC* genes occur in multiple copies. These genes may form a cluster or be scattered throughout the genome. In species with a cluster, a single copy usually exists, except *Synechocystis* sp. PCC6803, which has two copies of the cluster.

Phylogeny of kaiC Genes. The resulting consensus tree clearly shows separation of the *kaiC* genes into three main clades, which essentially correspond to Archaea, Proteobacteria, and Cyanobacteria in the topology of the 16S rRNA tree (Fig. 1). All these branches have significant statistical support. The topology of the *kaiC* tree indicates that the genes of Cyanobacteria and Proteobacteria are monophyletic, whereas *kaiC* homologs of Archaea are polyphyletic. However, this polyphyly has low bootstrap support. Both Templeton's test (29) and the winningsites test strongly rejected the null hypothesis of 16S rRNA and *kaiC* tree compatibility ($P < 0.0001$) with either the 16S sequence data or the *kaiC* sequence data and with all 96 alternative trees. These results evidently suggest multiple lateral transfers of the *kaiC* genes both within and between the three major clades of the tree (Fig. 1). For example, there were the transfers from Cyanobacteria to Proteobacteria (*Rhodospirillum* and *Rhodobacter*), Chloroflexi (Chloroflexus 1), and Archaea (Methanobacterium). Some of the transfers within the major clades are quite recent (for example, from *Xenococcus* to *Calothrix*, which have identical *kaiC* sequences, or from *Scytonema* to *Gloeobacter*).

In some species, the *kaiC* genes are present in two or three (*Synechocystis*) copies, and based on the topology of the trees these copies may have quite different evolutionary histories. Both copies of *Mesorhizobium* most likely have evolved through recent duplication, whereas evolution of the twin copies in the other taxa (*Pyrobaculum*, *Sulfolobus*, *Archaeoglobus*, *Agrobacterium*, *Chloroflexus*, and *Synechocystis*) is more complex and probably involves lateral transfers. For example, two *kaiC* genes of *Chloroflexus* are orthologous, and among them only Chloroflexus 2 has evolved along with the species, whereas the other was likely transferred from cyanobacteria (Fig. 1).

Phylogeny of kaiA and kaiB Genes. Screening prokaryotic genomes revealed *kaiB* genes in all available cyanobacteria similar to the *kaiC* genes. However, in contrast to the *kaiC* genes, *kaiB* occurred only in a few proteobacteria and one archaeon, *Methanobacterium thermoautotrophicum* (Table 1). None of the applied methods of phylogenetic reconstruction gave a fully resolved tree for the *kaiB* genes. The best unrooted tree is shown in Fig. 2*A*. This tree clearly indicates that the *kaiB* genes are separated into three main clades, which notably differ by the amount of accumulated nucleotide substitutions. The genes of the clade B1 have $K_a = 0.092 \pm 0.018$, which is significantly lower than in the clades B2 (K_a = 0.478 \pm 0.056) and B3 (K_a = 0.693 \pm 0.068). Clade B1 comprises the genes only from cyanobacteria, whereas the two other clades include *kaiB* genes from Archaea and Proteobacteria as well. Interestingly, the *kaiB* genes in clade B3, except *Methanobacterium*, are not from the cluster, as the genes in clade B1 and B2 are, but are scattered throughout the genome.

Only a few nodes in clades B2 and B3 of the *kaiB* tree have significant bootstrap support (Fig. 2*A*). However, a phylogenetic tree literally becomes completely resolved when inferred from the whole *kaiBC* clusters (Fig. 2*B*). Notably, its topology follows that of the cyanobacterial subtree of the *kaiC* genes (Fig. 1). This fact suggests that, after formation of the cluster, the *kaiB* and *kaiC* genes evolve as a unit rather than independently. Furthermore, comparison of the trees of the *kai* genes in Figs. 1 and 2 makes it reasonable to assume that the *kaiB* genes had originated

Fig. 1. ML congruent phylogenetic trees of 16S rRNA genes and the *kaiC* homologs of prokaryotes. Designations of the genes are given in Table 1. Species of the major taxa are indicated with different colors. Bootstrap values <50% are not shown.

in cyanobacteria and then were transferred laterally to the other prokaryotes.

Screening available prokaryotic genomes revealed the *kaiA* genes only in cyanobacteria. Among the available complete cyanobacterial genomes, the *kaiA* genes were not found in *Prochlorococcus marinus*. When available, the *kaiA* genes always occur in a single copy. Because of the limited number of *kaiA* genes available, it was impossible to perform their comprehensive phylogenetic analysis.

Dating Events in the Evolution of kai Genes. In our estimation of the dates of main events in the evolution of *kai* genes, we proceeded from the assumption that *kaiC* genes are evolutionarily the oldest among the prokaryotic circadian oscillators. Indeed, only *kaiC* and its homologs occur in the prokaryotes other than cyanobacteria. Based on fossil evidence, the last common ancestor of Eubacteria and Archaea is estimated as existing $\approx 3,800$ Mya, and the oldest known cyanobacteria morphologically indistinguishable from existing *Oscillatoria* were documented in $\approx 3,500$ million-year-old rocks (31). Given that all known *kaiC* genes of cyanobacteria are monophyletic and have a double-domain structure, we thus can suppose that duplication and subsequent fusion of their single-domain predecessors took place in the period between these dates, i.e., during \approx 300 million years (Fig. 3).

Fig. 2. (*A*) The best unrooted ML tree of the *kaiB* genes. (*B*) ML phylogenetic tree of the *kaiBC* cluster of prokaryotes. For color designations of the taxa see Fig. 1. Bootstrap values $<$ 50% are not shown.

Fig. 3. Timeline of major events in evolution of the cyanobacterial circadian clock genes based on ML estimates. The time scale is not proportional. p*kaiC*, predecessor of the *kaiC* gene; sd*kaiC*, single-domain *kaiC*; dd*kaiC*, doubledomain *kaiC*.

The other major events in the evolution of the *kai* cluster were the appearance of the *kaiB* gene and its fusion with the *kaiC* to form the *kaiBC* cluster. Almost identical topologies of the cyanobacterial subtree of the *kaiC* genes and *kaiBC* tree (Figs. 1 and 2*B*) suggest that these events likely occurred in the period corresponding to the time between nodes 1 and 2 in the *kaiC* tree.

Because the ML test rejected the global clock hypothesis for all *kai* genes, we applied the local clock model (25) to date their evolution. According to these estimates, the appearance of *kaiB* genes and the formation of the *kaiBC* cluster occurred between \approx 3,500 and 2,320 Mya.

Because of the small number of available *kaiA* genes, dating their evolution is difficult. Definitely, they are evolutionarily the youngest among the three prokaryotic circadian pacemakers, because they occur only in some higher cyanobacteria (Table 1). Importantly, of the two closely related unicellular cyanobacteria, *Synechococcus* sp. PCC7942 and *P. marinus* (Fig. 1, 16S rRNA tree), only the former possesses the *kaiA* gene. Furthermore, referring to the cyanobacterial *kaiC* subtree, the *kaiA* genes were found only in genomes of the species, which belong to the clades that are evolutionarily younger than *Prochlorococcus*. It allows us to suppose that these genes originated in cyanobacteria after speciation of *Synechococcus* and *Prochlorococcus* and thus to estimate the time of origin. Remarkably, ML estimation of this time using both 16S rRNA and *kaiC* sequences gives essentially the same result, $1,051 \pm 1,16.9$ and 944 \pm 92.9 Mya, respectively.

Discussion

The results of the phylogenetic analysis suggest that the three genes of the *kai* cluster have quite different evolutionary histories. The *kaiC* gene is the oldest among the three. The three-gene cluster, as it was described originally (2), is likely characteristic only to cyanobacteria and evolved gradually through the consequent stepwise addition of the *kaiB* and *kaiA* genes to the *kaiC* gene. Moreover, the *kaiA* and *kaiB* genes originated in cyanobacteria after their separation from the other prokaryotes. The occurrence of the *kaiB* genes in some species of the other prokaryotic domains (Fig. 2) is a result of lateral transfers from cyanobacteria. Importantly, our data suggest no lateral transfers of any *kai* genes to cyanobacteria from the other prokaryotes. In light of these findings, the hypothesis about the origin of cyanobacterial *kaiC* genes through the duplication and fusion of the single-domain ancestor in Archaea and their lateral transfer to Cyanobacteria (12) is inconsistent.

Evolution of kai Genes and Geological History of the Earth. Circadian pacemakers underlie basic physiological processes and thus may influence the expression of many genes. In cyanobacteria, the clock genes were shown to be involved in the regulation of nitrogen fixation (32), cell division (33), and other metabolic processes (4, 34). Furthermore, reproductive fitness of cyanobacteria increases when the endogenous clock and the temporal environmental cycle are strictly synchronized (35).

An analysis of the *kai* gene phylogenies in prokaryotes brings us to the three key assumptions: (*i*) circadian pacemakers are involved in the regulation of photosynthesis, (*ii*) double-domain structure of the *kaiC* gene is essential for the circadian oscillation, and (*iii*) circadian systems of prokaryotes may not necessarily include all three *kai* genes. These suppositions are supported by the fact that only photosynthetic prokaryotes have either the double-domain *kaiC* gene or the *kaiBC* cluster. The only exception is for a few Euryarchaeota (Table 1), which are not photosynthetic.

Comprehensive study of the *kaiABC* cluster expression in *Synechococcus* sp. PCC7942 showed that all three *kai* genes are essential for circadian rhythmicity, and inactivation of any of them completely abolishes it (2). The *kaiA* genes do not occur in the most primitive cyanobacteria and photosynthetic proteobacteria. Does that mean that these prokaryotes do not possess circadian rhythmicity? Apparently, it does not. Indeed, the circadian system with all three *kai* genes was described, thus far, only for *Synechococcus* sp. PCC7942 (2). Most likely, it is characteristic for the most evolutionarily advanced cyanobacteria, which have all these genes. However, it does not necessarily mean that other circadian systems, without *kaiA*, are impossible. In fact, in the system with the three *kai* genes, *kaiA* acts solely as a regulator of *kaiBC* expression via binding to two *kaiA*binding domains in the *kaiC* gene (11). In the simpler systems, without *kaiA*, other yet-unknown gene(s) may play the role. Indeed, even the three-gene *kai* system includes other, either homologous to *kai* or not, genes essential for the circadian regulation (36, 37). Such genes may be elements of the simpler systems ensuring their functionality. Further studies of such circadian systems are needed to verify this hypothesis.

In the evolution of *kai* genes as a cluster controlling circadian rhythmicity, a few major events may be defined. The first was the duplication of a single-domain ancestor of *kaiC* and further fusion of the resulting genes to form the double-domain gene. It occurred ≈3,800–3,500 Mya (Fig. 3).

Another major event was the origin of the *kaiB* gene and the formation of the *kaiBC* cluster. It happened between \approx 3,500 and 2,320 Mya (Fig. 3). According to the theory of atmosphere evolution (38), the time of \approx 2,500–2,000 Mya corresponds to the period when a reducing geochemical environment, which had existed since the early history of the earth, was replaced by an oxidizing environment produced by cyanobacteria. In this period, cyanobacteria were not truly oxygen-evolving but were consuming nitrogen and producing oxygen, which was poisonous for the then-dominating methano- and sulfobacteria. Initially, oxygen was bound to $\bar{F}e^{2+}$ released from the Earth's mantle because of volcanic activity. Approximately 2,500 Mya the volcanic activity subsided, resulting in a depletion of Fe^{2+} resources \approx 2,000 Mya (39). Further, when becoming oxygenproducing, cyanobacteria out-competed early-evolved photosynthetic bacteria, because biosynthesis of bacteriochlorophyll would have been inhibited by molecular oxygen (40).

Because of their influence on the wide variety of vital metabolic cycles, circadian clock genes should be an essential part of adaptive mechanisms. They are particularly important for photosynthetic organisms (e.g., cyanobacteria and purple bacteria),

the vital functions of which are based on photochemically dependent processes. Temporal optimization of intracellular mechanisms to the day/night conditions thus should increase fitness of these prokaryotes and confer an evolutionary advantage. Origin of the circadian gene cluster therefore could be one of the key events in the evolution of cyanobacteria, which ensured their domination in the Earth's ecosystems over almost 2.5 billion years known as ''the age of Cyanobacteria'' (41).

Key Mechanisms of Adaptive Evolution of the kai Genes. In addition to the above considerations, there are a number of other evidences supporting an adaptive character of the evolution of the *kai* genes. Duplication of the *kaiC* predecessor and formation of the double-domain *kaiC* gene in the earliest stages of its evolution (Fig. 3) was likely among those adaptive characters, because gene duplication was shown to be an exceptionally efficient mechanism for rapid evolutionary advance (42–44) including the *kai* genes (7). Duplicated genes may further evolve in four ways: (*i*) one of the copies keeps the original function, whereas another may accumulate deleterious mutations and become nonfunctional (nonfunctionalization); (*ii*) one copy acquires a novel, advantageous function, and another copy still maintains the original function (neofunctionalization); (*iii*) duplicated paralogs share original functions, and both new copies are less efficient compared with the ancestral gene (subfunctionalization); or (*iv*) one of the new copies performs with the original efficiency, whereas another accumulates advantageous mutations and acquires greater efficiency under specific conditions (superfunctionalization or reinforcement) (7, 45–49).

All these scenarios may have taken place in the evolution of the *kai* genes. The evident cases of nonfunctionalization and superfunctionalization were described elsewhere (7). Neofunctionalization and subfunctionalization also might occur in evolution of the *kaiB* and *kaiC* genes of some species. For example, *Synechocystis* sp. PCC6803 has three copies of the *kaiB* and *kaiC* genes each, six in total (Table 1). Four of these copies form two clusters (one of them with the *kaiA* gene, which presents itself in a single copy), and the remaining two are scattered throughout the genome. The *kaiABC* cluster was shown to regulate circadian oscillation (2) and was supposed to be the most advanced circadian system in prokaryotes. Another cluster comprises two genes, *kaiB* and *kaiC*, and may have either different (neofunctionalization) or similar but probably less efficient (subfunctionalization) function. And finally, the same ways of evolution may be considered for the scattered *kaiB* and *kaiC* genes.

Another mechanism greatly affecting evolution of the *kai* genes is lateral transfers. From the phylogenetic trees of the *kai* genes in Figs. 1 and 2, the numerous lateral transfers of the *kaiB*

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and *kaiC* genes or *kaiBC* cluster can be detected. Remarkably, all transfers of the cluster were from evolutionarily more advanced (Cyanobacteria) to more primitive prokaryotes (Chloroflexi, Proteobacteria, and Archaea) (Fig. 2*B*). In some species such as *Chloroflexus aurantiacus*, the laterally transferred and presumably more advanced system (*kaiBC* cluster) coexists with the original less-efficient one (comprising the *kaiB* and *kaiC* genes not joined in the cluster), whereas in the others such as *Methanobacterium thermoautotrophicum* the original system (supposedly consisting of the single-domain *kaiC* gene) had been replaced in the course of evolution by the laterally transferred one represented by the *kaiBC* cluster. It is hard to believe that the transferred genes conferring lower fitness under the given conditions could replace the homologous original genes and/or persist for such a long evolutionary period.

Recently, we reported that under acute long-term environmental stress, evolution of the *kai* genes in cyanobacteria is governed mainly by various types of natural selection (7). In the reported case, one of the main stress factors was UV irradiation. During the long period of the geological history of the Earth, the natural environment of prokaryotes was harsh, and UV irradiation was among the major factors most dramatically influencing evolution of cyanobacteria (50). Thus, the effect of UV and other stress factors on the *kai* genes in this period was probably somewhat similar to that observed in the ''Evolution Canyon'' model including a high mutation rate and frequent gene duplications (7) and eventually was controlled by natural selection.

During the long period of prokaryotic evolution, since the earliest documented time (31, 51) until \approx 500 Mya, the environment has undergone radical changes in the levels of oxygen and UV irradiation. Many prokaryotes occupy stressful habitats at present. They have evolved through increasing their fitness to survive as well as adapting to the changing environment. The origin and evolution of circadian clock genes was one of the most remarkable instruments of such adaptation involving a vast variety of known evolutionary mechanisms.

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