

Genotype, phenotype: upstairs, downstairs in the family of cardiomyopathies

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I loved Upstairs, Downstairs . . . you identify with the downstairs people while vicariously enjoying the life of the upstairs people.

—Alistair Cooke, Host of *Masterpiece Theatre*

Upstairs, Downstairs was beloved by a generation of Americans who became entranced by this classic story of an upstairs Edwardian family and its downstairs household. The series chronicled the daily repartee and tangled relationships between the upstairs and downstairs contingents, a tale laced with all of the ingredients of human complexity. Despite their diverse perspectives and backgrounds, the lives at 165 Eaton Place became entwined by their core values, conserved traits, and the shared challenges of the day. In terms of human biology, *Upstairs, Downstairs* was a classic saga of the interplay between genes and environment.

In the family of human cardiomyopathies, another complex story is unfolding, this time around the divergent backgrounds and perspectives of clinical cardiology and molecular genetics. In this cardiological version of *Upstairs, Downstairs*, the theme is *Genotype, Phenotype*, and the initial storyline revolves around the precept that

the primary determinant of the clinical phenotype is the molecular genotype. The clinical viewpoint has been underpinned by noninvasive analyses that can quantitatively assess differences in chamber volume, wall thickness, hypertrophy, systolic versus diastolic dysfunction, and outflow tract obstruction, leading to three clinical subtypes: hypertrophic cardiomyopathy (HCM), dilated cardiomyopathy (DCM), and restrictive cardiomyopathy (RCM). The molecular viewpoint has been driven by: 1) the discovery of diverse cardiomyopathic genotypes,

resulting in a detailed examination of the differential phenotypic effects of mutations in a myriad of sarcomeric and cytoskeletal genes, 2) cataloguing the effects of missense mutations by the severity of the charge change within a given disease gene, 3) evaluating differences between haploinsufficiency and missense mutations, and 4) correlating these diverse disease genotypes with differences in the severity, time of onset, and diversity of the cardiomyopathy phenotype. The theme of this early script is that there may be a specific set of molecular pathways

Table 1
Molecular defects linked to human cardiomyopathies

Genomic defects	Human defects		
	HCM	DCM	RCM
Sarcomere			
Myosin heavy chain	Missense (17–19)	Missense (20)	
Myosin essential light chain	Missense (21)		
Myosin regulatory light chain	Missense (21)		
Cardiac actin	Missense (22)	Missense (3)	
Troponin-T	Missense/deletion (19, 23)	Deletion (20)	
Troponin-I	Missense (7)		Missense (6)
α -Tropomyosin	Missense (19, 23)	Missense (24)	
Myosin binding protein-C	Missense/deletion (19, 25)		
Titin/titin-related protein			
Titin	Missense (26)	Missense/deletion (27, 28)	
Telethonin (T-cap)	Missense (14)		
Z-disk-associated proteins			
MLP	Missense (14)		
Sarcolemma cytoskeleton			
Dystrophin	Deletion (29–31)		
β -Sarcoglycan	Deletion/duplication (32)		
δ -Sarcoglycan	Missense (33)		
α -Dystrobrevin	Missense (34)		
Metavinculin	Deletion (35, 36)		
Intermediate filaments			
Desmin	Missense (37, 38)		
Lamin A/C	Missense (39)		

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Nonstandard abbreviations used: hypertrophic cardiomyopathy (HCM); dilated cardiomyopathy (DCM); restrictive cardiomyopathy (RCM); troponin I (TNNI3).

Table 2

In vivo and in vitro cardiac physiological phenotyping in mouse models of cardiac diseases

Anatomical and contractile phenotyping**Cardiac imaging**

Transthoracic echocardiography (40)

Transesophageal echocardiography (41)

Magnetic resonance imaging (42, 43)

LV pressure analysis (44)

LV pressure-volume loop analysis (45)

PA/RV pressure analysis (46)

LV and RV angiography (47, 48)

Aortography (49)

Isolated intact muscle contractility studies

Isolated working heart system (50)

Single cell contractility

Video-edge detection (10, 51)

Laser diffraction (52)

In vivo determinations of calcium signaling

Chemical or genetically engineered fluorescent probes (53, 54)

Electrophysiological phenotyping**In vivo electrophysiological analysis**

Surface electrocardiography (55, 56)

Telemetric electrocardiography (57, 58)

Transesophageal cardiac pacing (59)

Open-chest in vivo EP study (58, 60)

Close-chest endocardial EP study (60, 61)

Single cell electrophysiological analysis

Action potential duration (58, 62, 63)

Patch-clamp analysis (58, 62)

Measurements of intracellular calcium concentrations (10, 11, 51)

Assessment of excitation-contraction coupling (64)

Biomechanical stress assays**In vivo mechanical loading**

Thoracic aortic banding (65)

Abdominal aortic banding (66)

PA banding (47)

Arteriovenous fistula (40)

Coronary artery ligation (67)

In vitro mechanical loading

Isolated perfused heart system (50, 68)

Papillary muscle stretch (14)

Cultured neonatal cardiomyocyte stretch (14, 69)

LV, left ventricle; RV, right ventricle; PA, pulmonary artery; EP, electrophysiology.

that account for the distinct cardiac phenotypes of the three major forms of cardiomyopathy. Over a decade ago, the Seidmans made the initial, important discovery that mutations in the β -myosin heavy chain can cause HCM (for review, see ref. 1). Subsequent work by this group and others expanded this concept to include other sarcomeric gene mutations, all of which were linked with the HCM subset of cardiomyopathy. At the same time, studies in genetically engineered mice

began to uncover a role for cytoskeletal defects in DCM via studies of mice that harbor a mutation in the cardiac Z disk protein MLP (2), and links between other cytoskeletal proteins and familial forms of human DCM were subsequently established (3) (for review, see refs. 4, 5). In short, the early, neat storyline was that sarcomeric mutations always lead to HCM, while cytoskeletal mutations result in DCM, thereby reflecting specific defects in the hardwiring within cardiac muscle

cells that govern these two distinct phenotypes. In short, the phenotypic diversity of familial cardiomyopathies appeared to be primarily driven by the disease genotype. From the perspective of molecular cardiologists, the hunt was on for specific signaling pathways that might differentially connect these genetic lesions with DCM versus HCM. However, recent experimental and clinical studies suggest a more complex genotype-phenotype relationship of cardiomyopathies (Table 1). While there is little doubt that the disease genotype is a critical determinant of the clinical phenotype, perhaps the major protagonists and antagonists of this story have yet to enter the stage.

Multiple cardiomyopathy phenotypes from identical sarcomeric genotypes: *TNNI3* mutations lead to HCM and RCM

In the current issue of the *JCI* (6) Mogenson et al. reinforce this notion by clearly documenting that a single mutation within the *tropoin I* (*TNNI3*) gene can lead to either HCM or RCM within the same family. In a series of patients with RCM, a number of independent *TNNI3* mutations were uncovered, again suggesting that *TNNI3* mutations cannot only lead to HCM, as previously described (7), but also RCM. Previous studies have shown that sarcomeric mutations in the tropomyosin, troponin T, titin, and β -myosin heavy chain gene can lead to either DCM or HCM (Table 1). Taken together, it appears that mutations in a given sarcomeric gene can lead to a spectrum of cardiomyopathic phenotypes, often overlapping between the clinical subsets of DCM, HCM, and RCM. Although there is little doubt that the disease genotype plays a critical role in initiating the cardiomyopathic process, the ultimate clinical phenotype undoubtedly represents the integrated effect of multiple interacting factors. This view is supported by a host of circumstantial evidence, including the poor penetrance of many cardiomyopathic genotypes, the influence of hemodynamic stress (pressure, volume, etc.) on disease progression (8), the secondary effects of the loss of cardiac myocyte survival and subsequent replacement fibrosis (9), strong modifying effects of calcium cycling (10, 11)

and calcium signaling (12), and clear evidence of genetic background effects in gene-targeted mouse models of cardiomyopathy (13).

Resolving cardiomyopathy phenotypes and disease pathways with refined physiological technology: the MLP story

Part of the difficulty in attempting to define the molecular pathways that link the myriad number of sarcomeric and cytoskeletal mutations with specific forms of cardiomyopathy stems from our relatively primitive understanding of the precise physiological phenotype of these and other cardiac muscle diseases. Given the vast diversity of human cardiomyopathic genotypes, as well as the inherent difficulty in assessing the physiological function of cardiac muscle cells from a large number of distinct patients and their families, it is likely that many of the major insights will arise at the interface of mouse models and human disease. For example, recent studies have identified a missense mutation in the *muscle LIM domain protein (MLP)* gene that is associated with human DCM that has arisen as a result of a founder effect in a Northern European population (14). Parallel studies in MLP-deficient mice indicate that MLP plays a critical role as part of a Z disc-tethonin-titin complex that is an essential component of the cardiac muscle stretch sensor (14). The human MLP mutation disrupts this complex, indicating that the pathway that links the disease genotype with the cardiomyopathy phenotype is related to a primary defect in the cardiac muscle stretch sensor pathway (14). Accordingly, it may become possible to develop a more functional approach to the classification of human cardiomyopathies on the basis of a detailed phenotypic analysis of mouse model systems. It will become critical to continue to develop new, high-resolution, high-throughput technology to resolve cardiac muscle physiological phenotypes at the whole organ, intact muscle, and single cell level.

As noted in Table 2, an arsenal of physiological technology has already been developed by multiple laboratories, which can now be coupled with the growing power of mouse genetics and well-characterized gene-targeted model

systems. Background effects in various mouse strains have been clearly observed and attempts to map and clone these modifiers are ongoing, a task made easier by the mouse genome project. The generation of hypomorphic alleles that correspond to known human cardiomyopathy genotypes could be especially informative. Conditional mutagenesis will be valuable in assessing whether the onset of cardiomyopathy in the postnatal setting actually reflects subtle, developmental effects of the sarcomeric and cytoskeletal gene mutations on chamber morphogenesis or function and a fleet of CRE recombinase mouse lines have now been well validated to restrict the mutations to specific cardiovascular lineages, i.e., atrial, ventricular, atrioventricular nodal, epicardial, endothelial, and neural crest (for review, see ref. 15). Genetic complementation via germline gene modification or Adeno-associated virus-mediated transcoronary gene transfer should be informative (10, 16), capitalizing on candidate genes uncovered in surrogate systems, i.e., in vitro cardiac myocyte assays, zebrafish, fly, and mouse models.

Phenotype, Genotype

Of course, without parallel advances in functional cardiac phenotyping, attempts to use these models to dissect specific molecular pathways for this multifaceted disease are likely to remain superficial. Ironically, the next episodes of the continuing story on the family of cardiomyopathies may rely more on innovative strategies for precision phenotyping, as opposed to simply expanding the number of genetically engineered mouse model systems per se. It is highly likely that a re-analysis of existing gene targeted mouse models of cardiomyopathy with more refined phenotyping will uncover unsuspected physiological mechanisms of direct relevance to cardiac muscle diseases. High-throughput patch clamp arrays, in vivo expression of calcium reporter genes in intracellular micro-domains of living cardiac muscle cells, novel strategies to monitor conduction system function, and new technology to assay cardiac muscle stretch sensor and effector pathways are likely to be featured. Consult your local listing for the sequel to the cardiomyopathy story, *Phenotype, Genotype*.

1. Seidman, J.G., and Seidman, C. 2001. The genetic basis for cardiomyopathy: from mutation identification to mechanistic paradigms. *Cell*. **104**:557-567.
2. Arber, S., et al. 1997. MLP-deficient mice exhibit a disruption of cardiac cytoarchitectural organization, dilated cardiomyopathy, and heart failure. *Cell*. **88**:393-403.
3. Olson, T.M., Michels, V.V., Thibodeau, S.N., Tai, Y.S., and Keating, M.T. 1998. Actin mutations in dilated cardiomyopathy, a heritable form of heart failure. *Science*. **280**:750-752.
4. Chien, K.R. 2000. Genomic circuits and the integrative biology of cardiac diseases. *Nature*. **407**:227-232.
5. Hoshijima, M., and Chien, K.R. 2002. Mixed signals in heart failure: cancer rules. *J. Clin. Invest.* **109**:849-855. doi:10.1172/JCI200215380.
6. Mogenson, J., et al. 2003. Idiopathic restrictive cardiomyopathy is part of the clinical expression of cardiac troponin I mutations. *J. Clin. Invest.* **111**:209-216. doi:10.1172/JCI200316336.
7. Kimura, A., et al. 1997. Mutations in the cardiac troponin I gene associated with hypertrophic cardiomyopathy. *Nat. Genet.* **16**:379-382.
8. Chien, K.R. 1999. Stress pathways and heart failure. *Cell*. **98**:555-558.
9. Hirota, H., et al. 1999. Loss of a gp130 cardiac muscle cell survival pathway is a critical event in the onset of heart failure during biomechanical stress. *Cell*. **97**:189-198.
10. Minamisawa, S., et al. 1999. Chronic phospholamban-sarcoplasmic reticulum calcium ATPase interaction is the critical calcium cycling defect in dilated cardiomyopathy. *Cell*. **99**:313-322.
11. Semsarian, C., et al. 2002. The L-type calcium channel inhibitor diltiazem prevents cardiomyopathy in a mouse model. *J. Clin. Invest.* **109**:1013-1020. doi:10.1172/JCI200214677.
12. Zhang, C.L., et al. 2002. Class II histone deacetylases act as signal-responsive repressors of cardiac hypertrophy. *Cell*. **110**:479-488.
13. Suzuki, M., Carlson, K.M., Marchuk, D.A., and Rockman, H.A. 2002. Genetic modifier loci affecting survival and cardiac function in murine dilated cardiomyopathy. *Circulation*. **105**:1824-1829.
14. Knoll, R., et al. 2002. The cardiac mechanical stretch sensor machinery involves a Z disc complex that is defective in a subset of human dilated cardiomyopathy. *Cell*. **111**:943-956.
15. Ruiz-Lozano, P., and Chien, K.R. 2003. Reconstructing the heart. *Nat. Genet.* **33**:8-9.
16. Hoshijima, M., et al. 2002. Chronic suppression of heart-failure progression by a pseudophosphorylated mutant of phospholamban via in vivo cardiac rAAV gene delivery. *Nat. Med.* **8**:864-871.
17. Tanigawa, G., et al. 1990. A molecular basis for familial hypertrophic cardiomyopathy: an alpha/beta cardiac myosin heavy chain hybrid gene. *Cell*. **62**:991-998.
18. Geisterfer-Lowrance, A.A., et al. 1990. A molecular basis for familial hypertrophic cardiomyopathy: a beta cardiac myosin heavy chain gene missense mutation. *Cell*. **62**:999-1006.
19. Bonne, G., Carrier, L., Richard, P., Hainque, B., and Schwartz, K. 1998. Familial hypertrophic cardiomyopathy: from mutations to functional defects. *Circ. Res.* **83**:580-593.
20. Kamisago, M., et al. 2000. Mutations in sarcomere protein genes as a cause of dilated cardiomyopathy. *N. Engl. J. Med.* **343**:1688-1696.
21. Poetter, K., et al. 1996. Mutations in either the essential or regulatory light chains of myosin are associated with a rare myopathy in human heart and skeletal muscle. *Nat. Genet.* **13**:63-69.
22. Mogensen, J., et al. 1999. Alpha-cardiac actin is a novel disease gene in familial hypertrophic cardiomyopathy. *J. Clin. Invest.* **103**:R39-R43.
23. Thierfelder, L., et al. 1994. Alpha-tropomyosin and cardiac troponin T mutations cause familial hypertrophic cardiomyopathy: a disease of the sarcomere. *Cell*. **77**:701-712.

24. Olson, T.M., Kishimoto, N.Y., Whitby, F.G., and Michels, V.V. 2001. Mutations that alter the surface charge of alpha-tropomyosin are associated with dilated cardiomyopathy. *J. Mol. Cell. Cardiol.* **33**:723-732.
25. Bonne, G., et al. 1995. Cardiac myosin binding protein-C gene splice acceptor site mutation is associated with familial hypertrophic cardiomyopathy. *Nat. Genet.* **11**:438-440.
26. Satoh, M., et al. 1999. Structural analysis of the titin gene in hypertrophic cardiomyopathy: identification of a novel disease gene. *Biochem. Biophys. Res. Commun.* **262**:411-417.
27. Gerull, B., et al. 2002. Mutations of TTN, encoding the giant muscle filament titin, cause familial dilated cardiomyopathy. *Nat. Genet.* **30**:201-204.
28. Itoh-Satoh, M., et al. 2002. Titin mutations as the molecular basis for dilated cardiomyopathy. *Biochem. Biophys. Res. Commun.* **291**:385-393.
29. Towbin, J.A., et al. 1993. X-linked dilated cardiomyopathy. Molecular genetic evidence of linkage to the Duchenne muscular dystrophy (dystrophin) gene at the Xp21 locus. *Circulation.* **87**:1854-1865.
30. Muntoni, F., et al. 1993. Brief report: deletion of the dystrophin muscle-promoter region associated with X-linked dilated cardiomyopathy. *N. Engl. J. Med.* **329**:921-925.
31. Melacini, P., et al. 1996. Myocardial involvement is very frequent among patients affected with subclinical Becker's muscular dystrophy. *Circulation.* **94**:3168-3175.
32. Barresi, R., et al. 2000. Disruption of heart sarcoglycan complex and severe cardiomyopathy caused by beta sarcoglycan mutations. *J. Med. Genet.* **37**:102-107.
33. Tsubata, S., et al. 2000. Mutations in the human delta-sarcoglycan gene in familial and sporadic dilated cardiomyopathy. *J. Clin. Invest.* **106**:655-662.
34. Ichida, F., et al. 2001. Novel gene mutations in patients with left ventricular noncompaction or Barth syndrome. *Circulation.* **103**:1256-1263.
35. Maeda, M., Holder, E., Lowes, B., Valent, S., and Bies, R.D. 1997. Dilated cardiomyopathy associated with deficiency of the cytoskeletal protein metavinculin. *Circulation.* **95**:17-20.
36. Olson, T.M., et al. 2002. Metavinculin mutations alter actin interaction in dilated cardiomyopathy. *Circulation.* **105**:431-437.
37. Goldfarb, L.G., et al. 1998. Missense mutations in desmin associated with familial cardiac and skeletal myopathy. *Nat. Genet.* **19**:402-403.
38. Li, D., et al. 1999. Desmin mutation responsible for idiopathic dilated cardiomyopathy. *Circulation.* **100**:461-464.
39. Fatkin, D., et al. 1999. Missense mutations in the rod domain of the lamin A/C gene as causes of dilated cardiomyopathy and conduction-system disease. *N. Engl. J. Med.* **341**:1715-1724.
40. Tanaka, N., et al. 1996. Transthoracic echocardiography in models of cardiac disease in the mouse. *Circulation.* **94**:1109-1117.
41. Scherrer-Crosbie, M., et al. 1998. Determination of right ventricular structure and function in normoxic and hypoxic mice: a transeptophageal echocardiographic study. *Circulation.* **98**:1015-1021.
42. Kubota, T., et al. 1997. Dilated cardiomyopathy in transgenic mice with cardiac-specific overexpression of tumor necrosis factor-alpha. *Circ. Res.* **81**:627-635.
43. Williams, S.P., et al. 2001. Dobutamine stress cine-MRI of cardiac function in the hearts of adult cardiomyocyte-specific VEGF knockout mice. *J. Magn. Reson. Imaging.* **14**:374-382.
44. Milano, C.A., et al. 1994. Enhanced myocardial function in transgenic mice overexpressing the beta 2-adrenergic receptor. *Science.* **264**:582-586.
45. McConnell, B.K., et al. 1999. Dilated cardiomyopathy in homozygous myosin-binding protein-C mutant mice. *J. Clin. Invest.* **104**:1235-1244.
46. Zhao, Y.Y., et al. 2002. Defects in caveolin-1 cause dilated cardiomyopathy and pulmonary hypertension in knockout mice. *Proc. Natl. Acad. Sci. USA.* **99**:11375-11380.
47. Rockman, H.A., et al. 1994. Molecular and physiological alterations in murine ventricular dysfunction. *Proc. Natl. Acad. Sci. USA.* **91**:2694-2698.
48. Pashmforoush, M., et al. 2001. Adult mice deficient in actinin-associated LIM-domain protein reveal a developmental pathway for right ventricular cardiomyopathy. *Nat. Med.* **7**:591-597.
49. Nakamura, T., et al. 2002. Fibulin-5/DANCE is essential for elastogenesis in vivo. *Nature.* **415**:171-175.
50. Geisterfer-Lowrance, A.A., et al. 1996. A mouse model of familial hypertrophic cardiomyopathy. *Science.* **272**:731-734.
51. Zhou, Y.Y., et al. 2000. Culture and adenoviral infection of adult mouse cardiac myocytes: methods for cellular genetic physiology. *Am. J. Physiol. Heart Circ. Physiol.* **279**:H429-H436.
52. Wussling, M., Schenk, W., and Nilius, B. 1987. A study of dynamic properties in isolated myocardial cells by the laser diffraction method. *J. Mol. Cell Cardiol.* **19**:897-907.
53. Wier, W.G., Balke, C.W., Michael, J.A., and Mauban, J.R. 2000. A custom confocal and two-photon digital laser scanning microscope. *Am. J. Physiol. Heart Circ. Physiol.* **278**:H2150-H2156.
54. Zhang, J., Campbell, R.E., Ting, A.Y., and Tsien, R.Y. 2002. Creating new fluorescent probes for cell biology. *Nat. Rev. Mol. Cell Biol.* **3**:906-918.
55. Berul, C.I., Aronovitz, M.J., Wang, P.J., and Mendelsohn, M.E. 1996. In vivo cardiac electrophysiology studies in the mouse. *Circulation.* **94**:2641-2648.
56. Sah, V.P., et al. 1999. Cardiac-specific overexpression of RhoA results in sinus and atrioventricular nodal dysfunction and contractile failure. *J. Clin. Invest.* **103**:1627-1634.
57. Kramer, K., et al. 1993. Use of telemetry to record electrocardiogram and heart rate in freely moving mice. *J. Pharmacol. Toxicol. Methods.* **30**:209-215.
58. Nguyen-Tran, V.T., et al. 2000. A novel genetic pathway for sudden cardiac death via defects in the transition between ventricular and conduction system cell lineages. *Cell.* **102**:671-682.
59. Hagendorff, A., Schumacher, B., Kirchhoff, S., Luderitz, B., and Willecke, K. 1999. Conduction disturbances and increased atrial vulnerability in Connexin40-deficient mice analyzed by transeptophageal stimulation. *Circulation.* **99**:1508-1515.
60. Berul, C.I., et al. 1997. Electrophysiological abnormalities and arrhythmias in alpha MHC mutant familial hypertrophic cardiomyopathy mice. *J. Clin. Invest.* **99**:570-576.
61. Berul, C.I., et al. 1998. Familial hypertrophic cardiomyopathy mice display gender differences in electrophysiological abnormalities. *J. Interv. Card. Electrophysiol.* **2**:7-14.
62. Kuo, H.C., et al. 2001. A defect in the Kv channel-interacting protein 2 (KChIP2) gene leads to a complete loss of I(to) and confers susceptibility to ventricular tachycardia. *Cell.* **107**:801-813.
63. Nuss, H.B., and Marban, E. 1994. Electrophysiological properties of neonatal mouse cardiac myocytes in primary culture. *J. Physiol.* **479**:265-279.
64. Esposito, G., et al. 2000. Cellular and functional defects in a mouse model of heart failure. *Am. J. Physiol. Heart Circ. Physiol.* **279**:H3101-H3112.
65. Rockman, H.A., et al. 1991. Segregation of atrial-specific and inducible expression of an atrial natriuretic factor transgene in an in vivo murine model of cardiac hypertrophy. *Proc. Natl. Acad. Sci. USA.* **88**:8277-8281.
66. Shimoyama, M., et al. 1999. Calcineurin plays a critical role in pressure overload-induced cardiac hypertrophy. *Circulation.* **100**:2449-2454.
67. Michael, L.H., et al. 1995. Myocardial ischemia and reperfusion: a murine model. *Am. J. Physiol.* **269**:H2147-H2154.
68. Grupp, I.L., Subramaniam, A., Hewett, T.E., Robbins, J., and Grupp, G. 1993. Comparison of normal, hypodynamic, and hyperdynamic mouse hearts using isolated work-performing heart preparations. *Am. J. Physiol.* **265**:H1401-H1410.
69. Kudoh, S., et al. 1998. Mechanical stretch induces hypertrophic responses in cardiac myocytes of angiotensin II type 1a receptor knockout mice. *J. Biol. Chem.* **273**:24037-24043.