

Biogeochemical Processes Governing Exposure and Uptake of Organic Pollutant Compounds in Aquatic Organisms

by John W. Farrington*

This paper reviews current knowledge of biogeochemical cycles of pollutant organic chemicals in aquatic ecosystems with a focus on coastal ecosystems. There is a bias toward discussing chemical and geochemical aspects of biogeochemical cycles and an emphasis on hydrophobic organic compounds such as polynuclear aromatic hydrocarbons, polychlorinated biphenyls, and chlorinated organic compounds used as pesticides. The complexity of mixtures of pollutant organic compounds, their various modes of entering ecosystems, and their physical chemical forms are discussed. Important factors that influence bioavailability and disposition (e.g., organism-water partitioning, uptake via food, food web transfer) are reviewed. These factors include solubilities of chemicals; partitioning of chemicals between solid surfaces, colloids, and soluble phases; variable rates of sorption, desorption; and physiological status of organism. It appears that more emphasis on considering food as a source of uptake and bioaccumulation is important in benthic and epibenthic ecosystems when sediment-associated pollutants are a significant source of input to an aquatic ecosystem. Progress with mathematical models for exposure and uptake of contaminant chemicals is discussed briefly.

Introduction

Modern societies derive many benefits from mobilization of natural geochemical resources, such as oil, and from synthetic chemicals; for example, medicine, generation of usable energy, material for shelter, and chemicals for agriculture. The Organization for Economic Cooperation and Development (OECD) estimates that approximately 1000 to 1500 new substances enter daily use each year and add to the approximately 60,000 already in daily use (1), an estimate that is similar to that of the United States Council on Environmental Quality (2). Fortunately, only a small proportion of these chemicals pose threats to human health and living natural resources once they are released to the environment. This small proportion is still a large number. Butler estimated that 11,000 chemicals are manufactured in quantities that require assessment for environmental concern in that these quantities have the potential to pollute significant sectors of global ecosystems if released. In addition to these manufactured chemicals, there are chemicals of environmental concern generated by processes of treatment of industrial and domestic wastes by chlorination (4-6). The exact numbers are not important considerations in this paper. Rather, it is important that the assessment of toxicity and environmental fates and effects for these chemicals, and most important, assessment of human health risks associated with the presence of these chemicals in the environment, are substantial tasks requiring extensive knowledge of the behavior of these chemicals in the environment and their modes of toxic action. Studies of the past 15 to 20 years

have provided evidence of the widespread distribution of recalcitrant synthetic organic chemicals such as the chlorinated pesticides and polychlorinated biphenyls (PCBs) in ecosystems worldwide even in locations far removed from sources of input. Aquatic ecosystems are not an exception, as has been well documented in numerous studies and reviews (2,7,8).

The presence of a xenobiotic compound in a segment of an aquatic ecosystem does not, by itself, indicate an adverse effect. Connections must be established between levels of exposure or tissue contamination and adverse effects. The combined understanding of the inputs, fates, and effects of chemical contaminants or pollutants in the environment has been termed ecotoxicology by groups of scientists (3,9,11). This paper focuses on a few advances in biogeochemical research concerned with the movement of organic chemicals through aquatic ecosystems and conditions of exposure and routes of uptake by aquatic organisms. The focus is mainly on nonpolar, hydrophobic, medium molecular weight compounds such as PCBs, chlorinated pesticides, and aromatic compounds found in fossil fuels and their combustion products, chemicals often identified with the current concern about human health risks associated with chemical contaminants in fish and other aquatic species.

A stylized version of the biogeochemical cycle of a group of these compounds, polynuclear aromatic compounds (PAHs) is presented in Figure 1. It would be an immense task to quantitatively measure all the pathways and rates of reaction and movement for the many thousands of chemicals of concern through each of their individual biogeochemical cycles. Although individual chemical structure confers some specificity of environmental behavior and toxic action to each compound,

*Environmental Sciences Program and Urban Harbors Institute, University of Massachusetts at Boston, Harbor Campus, Boston, MA 02125.

there are principles by which we can group chemicals of similar structure and obtain some predictive capability of their biogeochemical behavior. This knowledge can then be used in concert with knowledge derived from effects studies to provide human health risk assessments and ecological risk assessments.

Physical Chemical Research

Fundamental properties related to the biogeochemical behavior of many organic chemicals of environmental concern have not been well characterized, and it is only in the past years that substantial progress has been made. For example, water solubilities are available for only relatively few compounds, especially for the medium to higher molecular weight nonpolar compounds such as PAHs, PCBs, and chlorinated pesticides. This situation is improving rapidly (12–16). The influence of temperature and salinity on solubility has been less extensively investigated (16–19) and requires much further research. Initial assessments of the influence of temperature and salinity on solubilities yield factors of 2 to 6 for medium to higher molecular

weight range PAHs (e.g., phenanthrene to benzo[*a*]pyrene) over the range of temperatures and salinities normally found in coastal waters and the open ocean (19). However, the anomalous behavior of benzo[*a*]anthracene compared to other PAH tested suggests that important knowledge has yet to be gained for key aspects of factors influencing solubilities in seawater (19).

Calculation or estimation of water solubilities from theoretical and empirical considerations is an active area of research, and some progress has been made using molecular surface area and volume and activities of organic solutes in organic phase and activity coefficients in aqueous phase (20–22). The wide range of solubilities of these essentially hydrophobic compounds and the very low solubilities for compounds in the medium to higher molecular weight range, e.g., phenanthrene and 2,2',4,4',5,5'-hexachlorobiphenyl, are illustrated by the examples in Table 1.

Compounds with low solubilities have a tendency to sorb to surfaces, and this is another of the important physical-chemical parameters needed to understand the biogeochemical behavior of organic chemicals of environmental concern. Results from several investigations during the past 10 years have demonstrated

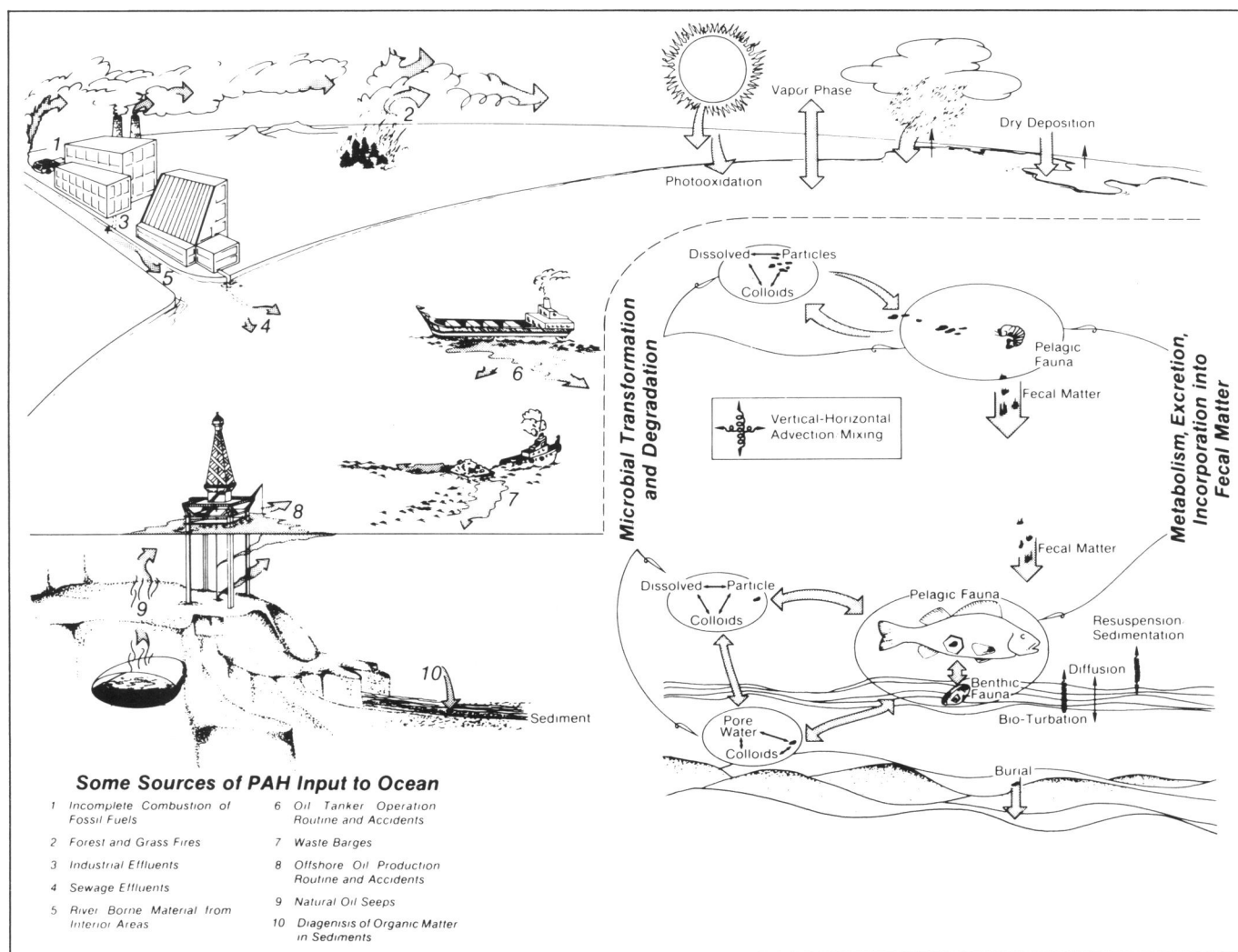


FIGURE 1. Biogeochemical cycle of PAHs in the ocean.

Table 1. Water solubilities of selected organic compounds at 25°C (20).

Compound	log S ^a
Acenaphthene	-3.86
Acrylonitrile	0.15
Aniline	-0.15
Benzene	-1.68
Benzo(a)pyrene	-6.26
Biphenyl	-3.88
1,3-Butadiene	-1.86
Carbon tetrachloride	-2.31
Chlorobenzene	-2.35
Dibenz(ah)anthracene	-6.28
<i>m</i> -Dichlorobenzidine	-3.01
3,3'-Dichlorobenzidine	-3.84
2,4'-Dichlorobiphenyl	-5.32
1,2-Dichloroethane	-1.09
7,12-Dimethylbenzanthracene	-5.71
Ethylbenzene	-2.80
Hexachlorobenzene	-5.48
2,2',4,4',5,5'-Hexachlorobiphenyl	-7.66
3-Methylanthrene	-6.54
Naphthalene	-3.04
Pentachlorobenzene	-4.85
Phenanthrene	-4.42
Styrene	-2.57
1,2,3,5-Tetrachlorobenzene	-4.33
2,2',5,5'-Tetrachloroethane	-1.75
Toluene	-2.22
1,3,5-Trichlorobenzene	-4.09
1,1-Trichloroethane	-2.00

^aS, molar solubility.

that sorption is mainly a simple partitioning of neutral, nonpolar hydrophobic compounds from the aqueous phase to a bulk non-aqueous phase (23–26). Organic constituents of natural sorbents are primarily responsible for sorption of hydrophobic organic compounds if the organic carbon content of the sorbent exceeds 0.1% (27). Thus, a partition coefficient K_p can be expressed in terms of the organic carbon content of the sorbent and in terms of a partition coefficient K_{oc} between water and a hypothetical sorbent of 100% organic carbon of a natural organic material:

$$K_p = f_{oc} \cdot K_{oc} \quad (1)$$

The partition coefficient K_{oc} can in turn be related to the octanol/water partition coefficient of a given compound by the following equation:

$$\log K_{oc} = a \log K_{ow} + b \quad (2)$$

where parameters a and b are empirically derived for different

groups of compounds. Some of the values reported for these parameters in different studies are summarized in Table 2 to illustrate the type of variation to expect.

The K_{ow} octanol/water partition coefficient of a compound has proven to be a very useful parameter in predicting the environmental behavior of hydrophobic chemicals. There are means beyond the scope of this paper to derive the relationship between K_{ow} and solubility (29–31), as reviewed in Brownawell (32) and Lyman et al. (33), among others. Thus, it is possible to determine the K_{ow} of a given compound experimentally and then calculate its solubility and sorption in aqueous systems. Handbooks with some of these properties are now becoming available (33), and on-line computer systems for environmental data of this sort are also becoming available.

The caveat that must be kept foremost in the minds of environmental scientists is that data are available for very few of the compounds of environmental concern and the concepts have been tested in relatively few of the myriad geochemical milieu found in aqueous ecosystems. For example, there have been only a few studies of compounds such as amines, carboxylic acids, and phenols (34,35). These studies demonstrate that the simple partitioning model used to describe sorption of neutral, hydrophobic molecules is of limited applicability to compounds that are partially or fully ionizable at natural waters pH values. It is possible to account for the influence of ionization and assemble predictive relationships (35–37) but relatively few types of compounds have been studied. The influence of pH and Eh on the environmental behavior of ionizable organic pollutants needs much further research because many of the present and future problems with environmental fates and effects of compounds involve considerations of polluted sediments and estuarine and coastal waters (38–40) where substantial ranges of pH and Eh are encountered (41,42).

Natural waters can be partitioned into particulate and dissolved phases by operational means of filtration, and the definitions of dissolved and particulate are operational. The partition parameters discussed in the preceding paragraphs, K_p and K_{oc} , are based on the simplest partitioning process, two phases, and this works reasonably for many soil-water systems and sediment-water systems. Three lines of evidence led Brownawell (32) and Brownawell and Farrington (43,44) to hypothesize that at least a three-phase partitioning model is needed to model the distributions of many hydrophobic compounds in interstitial waters of sediments: a) experimental evidence showing sorption of hydrophobic organic compounds to colloidal organic matter (45,46); b) high dissolved organic carbon (DOC) concentrations

Table 2. Estimation of K_{oc} based on K_{ow} : $\log K_{oc} = a \log K_{ow} + b$ (28,33).

Coefficients correlation			Number of compounds	Range of $\log K_{ow}$	Type of chemicals
a	b	r^2			
0.544	1.377	0.74	45	– 3.0–6.6	Primarily agricultural chemicals
1.00	– 0.21	1.00	10	2.1–6.3	Polycyclic aromatic hydrocarbons (PAH)
0.937	– 0.006	0.95	19	2.1–6.3	(PAH) triazines, nitroalines
1.029	– 0.19	0.91	13	0.4–6.3	Herbicides, insecticides (23)
1	– 0.317	0.98	13	1.6–6.5	heterocyclic aromatic compounds
0.72	0.49	0.95	13	2.6–4.7	Chlorinated hydrocarbons, alkylbenzenes
0.52	0.64	0.84	30	0.5–3.3	Substituted phenyl ureas and alkyl- <i>N</i> -phenyl carbamates

in the interstitial waters of marine sediments (47-51); and c) the fact that 50 to 90% of the interstitial water DOC is colloidal (48,52,53).

Field and laboratory evidence from several studies with PCBs has established that partitioning of hydrophobic compounds to colloids or very small particles must be considered in many natural water systems to account for the observed distributions of hydrophobic compounds in the dissolved and particulate fractions of water samples (43,44,54,55). Nomographs similar to those of Figure 2 (32) have been presented (46,55,56) and are useful in providing first order assessments of the physical chemical state of a given hydrophobic pollutant in coastal, estuarine, riverine, lacustrine, and interstitial waters.

The association of a significant proportion of the interstitial water hydrophobic compounds such as the PCBs with a colloid fraction has important implications for the assessment of the flux of these compounds across the sediment-water interface (32). First-principle arguments indicate that hydrophobic compound interactions with colloids will also be important in modeling the environmental behavior of compounds that enter estuarine waters across a salinity and particulate matter gradient as material is carried down river, or in the case of discharge of an industrial or municipal sewage effluent rich in organic matter content. Much of what is known about colloidal organic matter in natural waters is descriptive in nature (51,57,58). The relatively new findings and hypotheses about the importance of hydrophobic compound interactions with colloidal organic matter should add to previous observations and hypotheses concerning the importance of macromolecular organic matter and colloidal organic matter in contemporary biogeochemical cycles and stimulate new research efforts on colloids in natural water systems.

Biological Uptake, Release, Metabolism, and Distribution

Equilibrium Considerations

The concept of treating the relationship between chlorinated pesticides in aquatic organisms and their aqueous habitat as a pseudoequilibrium process was first proposed by Hamelink et al. (59). Stated in simple terms, exchange across membrane surfaces (e.g., gills) controls partitioning of a pollutant between the organism and water. The concept was shown to be consistent with data for DDT family compounds and PCBs in oceanic ecosystems (8,39) and was widely adapted for understanding of the relationship between hydrophobic pollutants such as PCBs, chlorinated pesticides, and PAHs in organisms and in waters of the organisms' habitat (3,8,33,60). Aquatic mammals and birds, being air breathing organisms, were exceptions to the concept.

Thus, direct uptake and release of organic pollutants from and to water joined food transfer and excretion as modes of input/output for pollutants in aquatic organisms and became the dominant modes in conceptual thinking and data interpretation. A procedure for assessing bioaccumulation potential under equilibrium conditions was needed, and Neeley et al. (61) were among the first to introduce the useful concept that K_{ow} could be used for this purpose. A compilation of data depicting the relationship between K_{ow} and K_b (bioaccumulation = concentration in organism/concentration in water) shows that the relationship

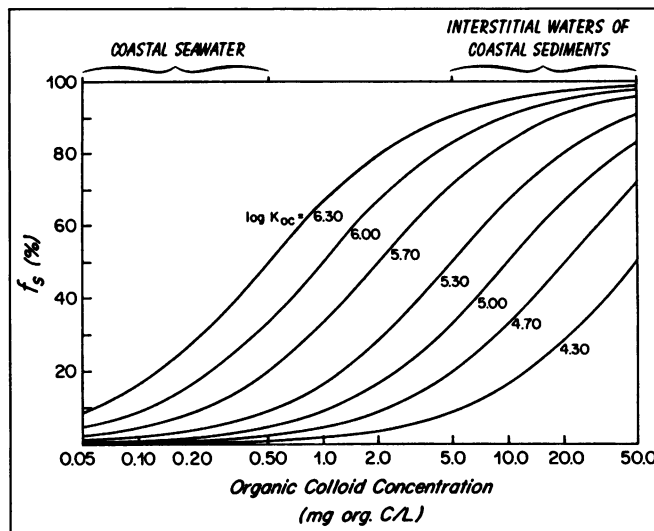


FIGURE 2. Nomograph for relationships between fraction (f_s) of a given hydrophobic compound sorbed to organic colloids, organic colloid concentration, and $\log K_{oc}$ of the compound. From Brownawell (32).

is predictive to a first approximation within a factor of 2 using Eq. (3) (62).

$$\log K_b = 1.00 \log K_{ow} - 1.32 \quad (3)$$

There are also significant departures from the predictive relationship (10,63,64). These departures could be due to non-equilibrium conditions at the time of the measurements used to calculate K_b . Eq. (3) may not apply for hydrophobic compounds with $K_b > 6$, i.e., superlipophilic compounds (62). Connell and Miller (65) submit that the equation must be modified to take into account molecular shape, e.g., planar and nonplanar PCBs. Hawker and Connell (63) have noted that little attention has been paid to the time required to attain equilibrium, with the result that some of the data in the literature may underestimate K_b . There is also the problem of the physical form of the total of the organic chemical concentration present in the water. However, the influence of dissolved organic material in the water and colloidal organic matter cannot be ignored. It may well be that only that portion of the material in the water not associated with the organic matter is available for partitioning with the organism. Experiments with the uptake of nonionic hydrophobic organic compounds such as petroleum compounds and PAHs by organisms have demonstrated reduced uptake when DOC-humiclike material is present in comparison to aqueous systems with low dissolved organic matter (DOM) concentrations (66,67). Table 3 (67) illustrates this point.

Another important physical-chemical consideration is sequestering or entrapment of an organic pollutant within a particle. Equilibrium partitioning calculations for sediment-water interactions for the hydrophobic compounds using K_{ow} s may not be applicable for sequestered or entrapped compounds. Farrington et al. (68,69) noted that comparison of PAHs for bivalve mollusks, sediments, and polychaetes in sediments in the same general locations revealed substantial differences between sediments and the biota. The causative factor was hypothesized

Table 3. Bioconcentration factors observed after 30-hr exposure of *D. magna* to 3-methylcholanthrene in the presence of different concentrations of dissolved humic material.*

DHM concentration, mg C/L	Fraction of MC bound to DHM	Observed BCF (\pm SD)	Fractional decrease in BCF due to presence of DHM
0	0	13.209 (\pm 653)	—
0.15	0.05	12.171 (\pm 194)	0.07
1.5	0.32	8.121 (\pm 161)	0.38
15.0	0.82	2.311 (\pm 127)	0.82

*See McCarthy et al. (67) for details. Abbreviations: DHM, dissolved humic material; MC, 3-methylcholanthrene; BCF, bioconcentration factor.

to be different sources and physical chemical form for the PAHs. Two types of PAHs were thought to be present: those found principally sequestered or entrapped in soot and pyrolytic source particles but extractable with normal solvent extraction procedures; and those entering aquatic ecosystems in water accommodated form or sorbed in a relatively easily partitioned form with particulate matter, e.g., petroleum from spilled oil or waste oil in effluents. These observations and hypothesis are consistent with observation concerning physical-chemical speciation of PAHs in an estuarine ecosystem (70) and the recent data on bioavailability of PAHs in a coastal sediment (71). The preceding are but a few examples illustrating that the accurate application of the equilibrium partitioning concept or other approaches to estimating bioaccumulation of a pollutant organic compound from a concentration of the compound measured or estimated for an organism's habitat must take into account the physical-chemical form of the compound.

Kinetic Considerations

The need for more accurate, predictive capabilities beyond those of equilibrium considerations seems apparent. Hawker and Connell (63) and Connell (72) have offered an insightful review of the relationship between kinetic considerations and equilibrium considerations. Eqs. (4) to (7) are from their work:

$$dC_b/dt = k_1 C_w - k_2 C_b \quad (4)$$

where C_b is concentration of pollutant in an organism; C_w is concentration of pollutant in the water phase; k_1 is a first-order uptake rate; k_2 is a first-order release rate or clearance rate from the organism; and t is time.

Water in the natural environment is a very large component relative to the organism, and thus C_w is assumed to be constant and not affected by bioaccumulation or release in this treatment of the equations. It is easily shown that at equilibrium

$$C_b/C_w = k_1/k_2 \quad (5)$$

$$\text{since } C_b/C_w = K_b, \text{ then } K_b = k_1/k_2 \quad (6)$$

and substituting into Eq. (3), it can be shown that $k_1/k_2 = 0.048 k_{ow}$. Thus, k_1 and k_2 are related to K_{ow} in the equations, and recent assessments of available data show that this is indeed the case (63). There is an inverse correlation between $\log(1/k_2)$ and $\log K_{ow}$ ($r = 0.974$) and a direct correlation between $\log(k_1)$ and $\log K_{ow}$ ($r = 0.974$) up to $\log K_{ow} = 7$. Equations of the form

$$\log K_1 = 0.337 \log K_{ow} - 0.373 \quad (7)$$

can be written and adequately describe most of the data in the literature. The fault with Eq. (7) is that it requires ever increasing k_1 with increasing K_{ow} , and it is not valid above $\log K_{ow} = 6$, i.e., for the extremely hydrophobic or superlipophilic compounds. Hawker and Connell (63) submit that there must be an upper limit to k_1 in part based on the efficiency with which a compound can be transferred across membranes, which is in turn related to compound shape and size and in part the ventilation rate of organisms which must have an upper limit for tissue such as gills.

First-order rate kinetics are useful in explaining some of the data concerning uptake and release of pollutants. An example illustrates this point. Two small no. 2 fuel oil spills occurred in the Cape God Canal 3 years apart but at the same time of the year within 1 week of each other [nearly duplicate experiments (73)]. Biological half-lives derived from Eq. (8) and (9) were determined from data exemplified by Figure 3.

$$C_t = C_0 e^{-kt} \quad (8)$$

where C_t is concentration in tissue at time t ; and C_0 = concentration in tissue at time 0.

$$t_{1/2} = \frac{0.693}{k_2} \quad (9)$$

where $t_{1/2}$ is the biological half-life.

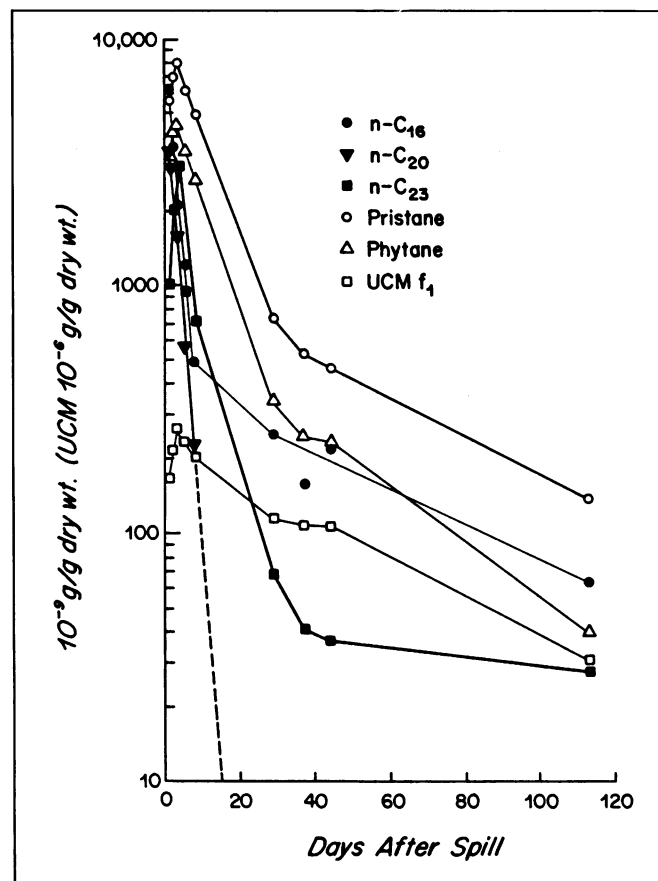


FIGURE 3. Time course of no. 2 fuel oil compounds released from *Mytilus edulis* after a contamination from a no. 2 fuel oil (73).

Table 4. Comparison of biological half-lives of no. 2 fuel oil compounds: 1978 and 1983 oil spill (73).

Compounds	$T_{1/2}$, days	
	1978 ^a Day 1-21	1983 Day 3-29
<i>n</i> -C ₁₆	0.99	8.7
Pristane	7.7	6.3
Phytane	6.9	5.8
<i>n</i> -C ₂₃	4.6	5.8
Σ C ₂ -naphthalenes	5.8	1.5
Σ C ₃ -naphthalenes	7.7	5.8
Phenanthrene	17	5.8
Σ C ₁ -phenanthrenes	9.9	6.9
Σ C ₂ -phenanthrenes	69 ^b	9.9
Unresolved complex mixture f ₁	14	17
Unresolved complex mixture f ₂	14	35

^aRecalculated for 1978 data. These are slightly different from those reported in Farrington et al. (73) because of an error in a calculator program discovered since that publication.

^bCorrelation fit of data to equation is poor.

A compilation of half-lives is given in Table 4. Review of the data for uptake and release of petroleum compounds, PAHs, and PCBs by bivalves indicates clearly that the relatively simple equations of the form of Eqs. (8) and (9) are applicable during the first 14 to 30 days. There are discernible departures from the semilog plots after 14 to 21 days in the example shown in Figure 3 depending on the compound considered. Concentrations of pollutant chemicals in the habitat and duration of exposure are known to be important considerations influencing rate of uptake and release of pollutants (60,77). For example, it has been suggested that a combination of high exposure concentrations and longer time of exposure, as would be expected in urban harbor areas, result in much slower overall release of pollutant hydrocarbons when bivalve mollusks are moved to cleaner waters.

The Stegeman and Teal (78) multiple-compartment model approach to explaining data from experiments and field observations derives from the fact that there are many tissue types in bivalves. It is not difficult to visualize this model, and it is supported by several studies (60,73,75,79). An example would be a simple three-compartment model bivalve of gills, circulatory fluid, and energy storage reserve lipids. Initial uptake across the gills is rapid, followed by slightly less rapid transfer to circulatory fluid, followed by much slower transfer to, and accumulation in, storage lipid reserves. Long-term exposure would result in accumulation of pollutant in lipid energy reserves until equilibrium or saturation of the storage capacity is attained. Transfer of the bivalve to clean water or removal of the pollutant from the water of its habitat reverses the process. Exchange of the pollutant from gill tissue to water is rapid, mobilization of the pollutant in the energy storage lipid is slower, and this accounts for an initial rapid release followed by much slower release in the longer term. Equations for assessment of multiple compartment models have been described by Mackay and Hughes (80) and involve steady-state assumptions in all but the target compartment, i.e., the compartment for which the flux of compound is of interest. Further elaboration of those models is beyond the scope of this synopsis.

It is well established that physiological status of organisms such as spawning, postspawning, prespawning, temperature of the habitat, food availability, as well as exposure concentration and duration, influence the uptake, retention, and release of

pollutant organic chemicals by bivalve mollusks (60,77,79). Metabolism of PAHs and xenobiotics such as PCBs is thought to be much less active for bivalves than for fish, crustacea, or polychaetes (40,81). However, there are reports that bivalve tissues contain enzymatic activity related to metabolism of PAHs, and perhaps similar compounds (82,83). This is consistent with data showing isomer-specific changes in relative abundance of C₂ and C₃-alkylated phenanthrenes in the latter stages of release of no. 2 fuel oil compounds by mussels contaminated by an oil spill (73).

A fallacy of the use of first-order kinetics in this situation is readily apparent if we derive Eq. (10) from Eq. (4).

$$C_b = (k_1/k_2) C_w (1 - e^{-k_2 t}) \quad (10)$$

Theoretically, equilibrium can only be attained after infinite time. It is useful to circumvent this problem and work with the concept of a close approach to equilibrium, e.g., 0.99 of value, as has been used by Hawker and Connell (63). They have then derived an equation that relates time to equilibrium to K_{ow} :

$$\log t_{eq} = 0.663 \log K_{ow} - 0.284 \quad (11)$$

This allowed them to predict that with compounds with $\log K_{ow} < 6$ attain equilibrium within 1 year. Compounds for which $\log K_{ow} > 6$ require too much time to approach equilibrium for reasonable measurements to be made. Hawker and Connell (63) also proposed a term t_s , as time to significant bioaccumulation with significant bioaccumulation being 1% of the bioaccumulation at equilibrium. They then used an equation relating t_s to $\log K_{ow}$ to calculate that compounds with a $\log K_{ow}$ of 10 are bioaccumulated significantly only after a minimum of 0.5 years. Aquatic organisms bioaccumulate a significant amount, i.e., 1% of equilibrium bioaccumulation concentration, of compounds with $\log K_{ow}$ of 13 after a minimum of 50 years. The latter time exceeds the lifetime of most aquatic organisms of concern, and the former time exceeds seasonality in temperate zones. The physiological status and other factors described previously that influence concentrations of organic chemical pollutants in an organism will change conditions of the organism or habitat well within the time required to reach significant bioaccumulation for hydrophobic pollutants with $\log K_{ow}$ of approximately 8 or 9 or greater. This limits the predictive capabilities of the kinetic equations described above.

Metabolism

Organisms other than bivalves are capable of reducing concentrations of hydrophobic organic chemical pollutants by exchange with water. In fact, some of the initial equilibrium partitioning work related to fish. In addition, fish, crustacea, and polychaetes have active enzyme systems capable of metabolizing substantial portions of bioaccumulated PAHs, some PCBs, and similar compounds (39,40,71,81). Research concerned with metabolism of organic chemical pollutants has expanded greatly in the past decade, and much detailed information is becoming available. The scope of this paper cannot provide many details. It suffices to state the obvious: concentrations in tissues and distributions among excretion of metabolites can be a function of a variety of tissues and conditions controlling enzymatic activity. These

conditions include spawning, nutritional status, conditions and duration of exposure to organic pollutants, mix of pollutants, and life cycle stage of the animal.

Exposure to xenobiotic organic compounds or PAHs has been shown to induce activity of mixed-function oxidases capable of metabolizing these xenobiotics. Extent of enzyme activity can be species-specific even for related species. Reichert et al. (84) exposed two species of deposit feeding amphipods to sediment-associated radioactively labeled benzo(a)pyrene (BaP). One species converted a higher proportion to metabolites, even though both species bioaccumulated the compound. The explicit lesson is that there are drawbacks to mathematical modeling of uptake, retention, metabolism, and release by extrapolating data from one species to another. This must be kept in mind when pragmatic approaches to the problems of a complex environment with thousands of species forces such extrapolations.

Another important example involves interactive effects of one chemical pollutant on another. Stein et al. (85) have shown that benzo(a)pyrene and PCBs interact and influence the extent of uptake and metabolism of each. Simultaneous exposure of English sole (*Parophrys vetulus*) to radioactively labeled BaP and PCBs increased concentrations of BaP-derived metabolites in the whole fish and decreased concentrations of PCBs and metabolites in some tissues and bile relative to results from separate exposure to radioactively labeled compounds in sediment-associated form.

Transfer in Food As a Source of Organic Pollutants

The water-organism partition hypothesis of Hamelink et al. (59) contributed significantly to the understanding of aquatic pollution and bioaccumulation and has prevailed for 15 years (39,40,86). This hypothesis evolved from experimental evidence in the laboratory and explained observations in the field obtained during the late 1960s and early 1970s when there were significant discharges of pollutant organic chemicals via effluents and transfer to water in aquatic ecosystems from atmospheric transport and runoff from land.

Several decades of input have resulted in accumulation of some compounds such as PAHs and PCBs and chlorinated pesticides in sediments (9,39,40,87,88). Several researchers have demonstrated that organisms living in or on polluted sediments can bioaccumulate the pollutants (71,88-91). Almost all experimental designs or field observations that were reviewed by these authors did not allow a sorting out of the relative importance of water-organism partitioning and sediment ingestion or water-organism partitioning and food ingestion in terms of contributions to bioaccumulated pollutant. In many cases, release of the pollutant from sediment by desorption was thought to cause elevated concentrations in the water followed by organism-water partitioning to achieve bioaccumulation. Oliver (92) has presented estimates (Table 5) for Lake Ontario that loadings of chlorinated hydrocarbons by desorption from sediments are in the same order of magnitude as are Niagara River inputs.

An elegant experiment by Rubenstein et al. (89) involving a multiphase experimental exposure design of a demersal fish (*Leiostomus xanthurus*) feeding on a polychaete (*Nereis virens*) with both fish and polychaete exposed to PCB-contaminated

Table 5. Comparison of contaminant loadings to Lake Ontario from bottom sediment desorption and Niagara River discharge (kg/y) (92).

Chemical ^a	Sediment desorption	Niagara River	Chemical	Sediment desorption	Niagara River
1,3-DCB	3	1600	2,4,5-TCT	4	190
1,4-DCB	11	5800	2,3,6-TCT	1	120
1,2-DCB	2	2600	PCT	6	50
1,3,4-TCB	9	140	HCBd	6	130
1,2,4-TCB	19	2400	OCS	3	10
1,2,3-TCB	2	540	PCB 18	1	70
1,2,4,5-TeCB	12	340	PCB 52	4	120
1,2,3,4-TeCB	7	760	PCB 40	1	16
QCB	7	240	PCB 101	4	90
HCB	19	120	PCB 153	4	60

^aAbbreviations: DCB, dichlorobenzene; TCB, trichlorobenzene; TeCB, tetrachlorobenzene; QCB, pentachlorobenzene; HCB, hexachlorobenzene. TCT, trichlorotoluene; PCT, pentachlorotoluene; HCBd, hexachlorobutadiene; OCS, octachlorostyrene; PCB, polychlorobiphenyl.

sediments has been completed. Rubenstein et al. conclusively demonstrated the importance of dietary input of PCBs from the polychaetes that had, in turn, been contaminated by PCBs originally in the sediment.

These results have very important implications for future efforts at modeling biogeochemical behavior, bioaccumulation, and, indeed, risk assessment for hydrophobic organic chemical pollutants in aquatic ecosystems. If a primary source of input to an aquatic ecosystem is slow release from sediments that have been polluted with inputs from various sources that have been reduced or eliminated in more recent times, then it is plausible that uptake by benthic organisms such as small bivalves, polychaetes, and crustacea followed by predation by larger organisms such as fish may be a significant source of bioaccumulated chemicals for the larger organism. In essence, food web transfer among benthic and epibenthic species may be as important as organism-water partitioning when aquatic ecosystems switch from top down (from effluent and atmospheric inputs) to bottom up (releases or transfers from polluted sediments) as regulatory controls reduce the top down sources of input. Even for aquatic ecosystems where inputs continue from effluents, the atmosphere, and runoff, there may be some ecosystems where a pseudo-steady-state approximation exists or is being approached, and food web transfer is now important for some species.

Mathematical Models of Coastal Ecosystems: From Inputs to Concentrations in Edible Tissues of Marine Organisms

It is now possible to incorporate the knowledge and predictive capabilities reviewed in earlier sections of this paper into dynamic models of entire ecosystems involving geophysical fluid dynamics, turbulence, sediment transport, and life-cycle stages of various species and arrive at some predictive capabilities with respect to contamination of edible tissues from marine organisms. O'Connor et al. (93) and Spaulding et al. (94) among others have presented such models.

One example suffices for the purpose of this paper. O'Connor et al. (95) have presented a mathematical model for the distribution and movement of the chlorinated pesticide, kepone, in the James River Estuary. Figure 4 shows the food chain portion of

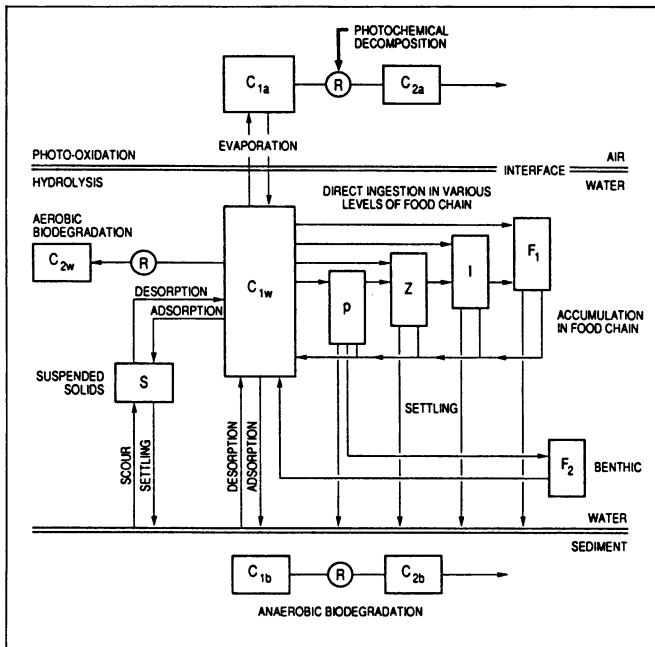


FIGURE 4. Kinetic and transfer routes of toxic chemicals in the water, bed, and food chain. From O'Connor and Piazza (9).

the model. The comparisons between the calculated kepone concentrations and the measured kepone concentrations in white perch, Atlantic croaker, and striped bass are given in Figure 5. The agreement seems reasonable to a first approximation although reasonable agreement depends on the use of the model and the degree of uncertainty that will be accepted, and this probably will vary depending on the perceived or real importance of relative risks to aquatic biota or human health. The types of mathematical models exemplified by the work of O'Connor et al. (93,95), Thomann (96), Spaulding et al. (94), and references cited therein are becoming an essential part of environmental risk assessment for issues ranging from oil spills to remedial action plans for Superfund sites in coastal estuaries (94,97).

General Discussion

There has been substantial progress during the 1970s and 1980s in all aspects of biogeochemical research related to the issues of bioavailability and disposition of toxic organic chemicals: solubility, sorption, uptake, metabolism, retention, release/excretion. Predictive equations have been derived or have evolved empirically that tie molecular structural characteristics or properties to biogeochemical behavior. Coupling of the portions of biogeochemical models that deal with transfer back to people via consumption of living aquatic resources and potential impacts on human populations, e.g., consumption of carcinogens in edible portions of fish tissues (98), presents an important and powerful tool for realistic regulation of pollution for the protection of human health.

However, caution must be exercised that the elegance and complexity of a series of coupled mathematical equations does not evoke a false sense that accurate predictive capabilities of wide-ranging applicability are a proven reality in either the scientific community or in the policy, regulation, and management communities concerned with aquatic pollution problems. Thus far, the concepts and hypotheses reviewed briefly in this paper have been tested on relatively few chemicals and relatively few biota and ecosystems and, with few exceptions, for relatively short periods of time of days to months. Reuber et al. (99), in the closing statement of a recent paper concerned with chemical equilibria and transport at the sediment-water interface, stated, "Finally, models as described here will remain of limited value until they can be applied and validated in real situations." Nevertheless, an optimistic view is in order because research is gaining on the problems, and the science has evolved from mainly a descriptive endeavor to quantitative approaches involving a mature healthy mix of theory, experimentation, field observation, research, and monitoring.

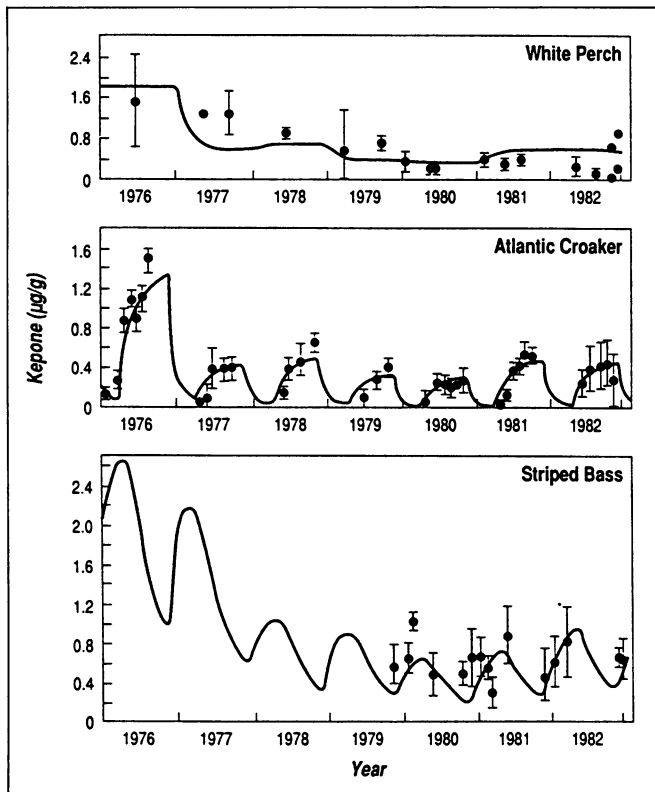


FIGURE 5. Comparison of observed and model calculated kepone concentrations in Atlantic croaker, white perch, and striped bass in the James River, Chesapeake Bay example. From O'Connor and Piazza (93).

The author thanks Judy Grassle, Judy McDowell-Capuzzo, Anne McElroy, John Teal, Bruce Brownawell, John Westall, John Stegeman, Jay Gooch, Rene Schwarzenbach, and Philip Gschwend for stimulating discussions concerned with this topic over the past several years. Paul Sherblom provided valuable comments on a draft of this paper. I gratefully acknowledge financial support from the University of Massachusetts-Boston for time to complete this review and acknowledge earlier financial support from the Woods Hole Oceanographic Institution Sea Grant Program and the Andrew W. Mellon and Richard King Mellon Foundation Grants to the Coastal Research Center of Woods Hole Oceanographic Institution for portions of the research described in this paper.

REFERENCES

- Stumm, W., Schwarzenbach, R., and Sigg, L. From environmental analytical chemistry to ecotoxicology—a plea for more concepts and less monitoring and testing. *Angew. Chem. Int. Ed. Engl.* 22: 380–389 (1983).
- CEQ. Environmental Quality 1983. 14th Annual Report of the Council on Environmental Quality, Executive Office of the President of the United States, Washington, DC, 1984.
- Butler, G. C., Ed. Principles of Ecotoxicology, SCOPE 13. John Wiley and Sons, New York, 1978.
- de Leer, E. W. B., Baggerman, T., van Schaik, P., Zuydeweg, C. W. S., and de Galan, L. Chlorination of ψ -cyanoalkanoic acids in aqueous medium. *Environ. Sci. Technol.* 20: 1218–1223 (1986).
- de Leer, E. W. B., Sininghe Damste, J. S., Erkelens, C., and de Galan, L. Identification of intermediates leading to chloroform and C-4 diacids in the chlorination of humic acid. *Environ. Sci. Technol.* 19: 512–522 (1985).
- Kringstad, K. P., de Souza, F., and Stromberg, L. M. Studies on the chlorination of chlorolignins and humic acid. *Environ. Sci. Technol.* 19: 427–431 (1985).
- Goldberg, E. The Health of the Oceans. The UNESCO Press, Paris, 1976.
- Harvey, G. R., and Steinhauer, W. Biogeochemistry PCB and DDT in the North Atlantic. In: *Environmental Biogeochemistry—Carbon, Nitrogen, Phosphorus, Sulfur and Selenium Cycles*, Vol. 1. (J. O. Nriagu, Ed.), Ann Arbor Science Publishers, Inc., Ann Arbor, MI, 1976, pp. 203–221.
- Sheehan, P. J., Miller, D. R., Butler, G. C., and Bourdeau, P., Eds. Effects of Pollutants at the Ecosystem Level, SCOPE 22. John Wiley and Sons, New York, 1984.
- Connell, D. Ecotoxicology—a new approach to understanding hazardous chemicals in the environment. *Search* 17: 27–31 (1986).
- Levin, S. A., Harwell, M. A., Kelly, J. R., and Kimball, K. D., Eds. *Ecotoxicology: Problems and Approaches*. Springer-Verlag, New York, 1989.
- Mackay, D., and Shiu, W. Y. Aqueous solubility of polynuclear aromatic hydrocarbons. *J. Chem. Eng. Data* 22(4): 399–402 (1977).
- Mackay, D., Mascarenhas, R., and Shiu, W. Y. Aqueous solubility of polychlorinated biphenyls. *Chemosphere* 9: 257–264 (1980).
- May, W. E., and Wasik, S. P. Determination of the solubility behavior of some PAH in water. *Anal. Chem.* 50: 997–1000 (1978).
- May, W. E., Wasik, S. P., and Freeman, D. Determination of the aqueous solubility of PAH by a coupled column LC technique. *Anal. Chem.* 50: 175–179 (1978).
- May, W. Aqueous solubilities of PAH. In: *Petroleum in the Marine Environment, Advances in Chemistry Series 185* (L. Petrakis and F. T. Weiss, Eds.), American Chemical Society, Washington, DC, 1980, pp. 141–192.
- Sutton, C., and Calder, J. A. Solubility of alkylbenzenes in distilled water and seawater at 25°C. *J. Chem. Eng. Data* 20: 320–322 (1975).
- Eganhouse, R. P., and Calder, J. A. The solubility of medium molecular weight aromatic hydrocarbons and the effects of hydrocarbon co-solutes and salinity. *Geochim. Cosmochim. Acta* 40: 555–561 (1976).
- Whitehouse, B. G. The effects of temperature and salinity on the aqueous solubility of polynuclear aromatic hydrocarbons. *Mar. Chem.* 14: 319–332 (1984).
- Banerjee, S. Calculation of water solubility of organic compounds with UNIFAC-derived parameters. *Environ. Sci. Technol.* 19: 369–370 (1985).
- Dunnivan, F. M., Coates, J. J., and Elzerman, A. W. Experimentally determined Henry's Law Constants for 17 polychlorinated biphenyl congeners. *Environ. Sci. Tech.* 22: 448–453 (1988).
- Andrew, A. W., Doucette, W. J., and Dickhut, R. M. Methods for estimating solubilities for hydrophobic organic compounds. In: *Sources and Fates of Aquatic Pollutants, Advances in Chemistry Series 216* (R. A. Hites and S. J. Eisenreich, Eds.), American Chemical Society, Washington, DC, 1987, pp. 3–26.
- Karickhoff, S., Brown, D. S., and Scott, T. Sorption of hydrophobic pollutants on natural sediments. *Water Res.* 13: 241–248 (1979).
- Means, J. C., Wood, S. G., Hassett, J. J., and Banwart, W. L. Sorption of polynuclear aromatic hydrocarbons by sediments and soils. *Environ. Sci. Technol.* 14: 1524–1528 (1980).
- Chiou, C. T., Peters, L. J., and Freed, V. J. A physical concept of soil-water equilibria for non-ionic organic compounds. *Science* 206: 831–832 (1979).
- Schwarzenbach, R. P., and Westall, J. Transport of nonpolar organic compounds from surface water to groundwater. Laboratory sorption studies. *Environ. Sci. Technol.* 15: 1360–1367 (1981).
- Schwarzenbach, R. P., and Westall, J. Sorption of hydrophobic trace organic compounds in groundwater systems. *Water Sci. Technol.* 17: 39–55 (1985).
- Farrington, J. W., and Westall, J. Organic chemical pollutants in the oceans and groundwater: a review of fundamental chemical properties and biogeochemistry. In: *The Role of the Oceans as a Waste Disposal Option* (G. Kullerberg, Ed.), D. Reidel Publishing Company, Boston, MA, 1986, pp. 361–425.
- Leo, A., Hansch, C., and Elkins, D. Partition coefficients and their uses. *Chem. Rev.* 71: 525–616 (1971).
- Miller, M. M., Wasik, S. P., Huang, G. L., and Mackay, D. Relationship between octanol-water partition coefficient and aqueous solubility. *Environ. Sci. Technol.* 19: 522–529 (1985).
- Chiou, C. T., Schmedding, D. W., and Manes, M. Partitioning of organic compounds in octanol-water systems. *Environ. Sci. Technol.* 16: 4–10 (1982).
- Brownawell, B. J. The Role of Colloidal Organic Matter in the Marine Geochemistry of PCBs. Ph.D. Dissertation, Woods Hole Oceanographic Institution/Massachusetts Institute of Technology Joint Program in Oceanography, Woods Hole, MA, 1986.
- Lyman, W. J., Reehl, W. F., and Rosenblatt, D. H. *Handbook of Chemical Property Estimation Methods: Environmental Behavior of Organic Compounds*. McGraw-Hill Book Co., New York, 1982.
- Means, J. C., Wood, S. G., Hassett, J. J., and Banwart, W. L. Sorption of amino- and carboxy-substituted polynuclear aromatic hydrocarbons by sediments and soils. *Environ. Sci. Technol.* 16: 93–98 (1982).
- Westall, J., Leuenberger, C., and Schwarzenbach, R. P. Influence of pH and ionic strength on the aqueous-nonaqueous distribution of chlorinated phenols. *Environ. Sci. Technol.* 19: 193–198 (1985).
- Xie, T. M., and Dyrssen, D. Simultaneous determination of partition coefficients and acidity constants of chlorinated phenols and guaiacols by gas chromatography. *Anal. Chim. Acta* 160: 21–30 (1984).
- Schellenberg, K., Leuenberger, C., and Schwarzenbach, R. P. Sorption of chlorinated phenols by sediments and aquifer materials. *Environ. Sci. Technol.* 18: 652–657 (1984).
- Baker, R. A., Ed. *Contaminants and Sediment*. Ann Arbor Science Publishers, Inc., Ann Arbor, MI, 1980.
- NRC. Polychlorinated Biphenyls. National Research Council, National Academy of Sciences, Washington, DC, 1979.
- NRC. Oil in the Sea, Inputs, Fates and Effects, National Research Council, National Academy Press, Washington, DC, 1985.
- Berner, R. A. *Early Diagenesis—A Theoretical Approach*. Princeton University Press, Princeton, NJ, 1989.
- Stumm, W., and Morgan, J. J. *Aquatic Chemistry*. Wiley-Interscience, New York, 1981.
- Brownawell, B. J., and Farrington, J. W. Partitioning of PCBs in marine sediments. In: *Marine and Estuarine Geochemistry* (A. C. Sigleo and A. Hattori, Eds.), Lewis Publishers, Chelsea, MI, 1985, pp. 97–120.
- Brownawell, B. J., and Farrington, J. W. Biogeochemistry of PCBs in interstitial waters of a coastal marine sediment. *Geochim. Cosmochim. Acta* 50: 157–169 (1986).
- Means, J. C., and Wijayaratne, R. Role of natural colloids in the transport of hydrophobic pollutants. *Science* 215: 968–970 (1982).
- Carter, C. W., and Suffet, I. H. Binding of DDT to dissolved humic material. *Environ. Sci. Technol.* 16: 735–740 (1982).
- Starikova, N. D. Vertical distribution patterns of dissolved organic carbon in seawater and interstitial solutions. *Oceanology* 10: 796–807 (1970).
- Krom, M. D., and Sholkovitz, E. R. Nature and reactions of dissolved organic matter in the interstitial waters of marine sediments. *Geochim. Cosmochim. Acta* 41: 1565–1573 (1977).
- Henrichs, S. M. Biogeochemistry of Dissolved Free Amino Acids in Marine Sediments. Ph.D. Dissertation, Woods Hole Oceanographic Institution, Massachusetts Institute of Technology Joint Program in Oceanography and Ocean Engineering, Woods Hole, MA, 1980.
- Henrichs, S. M., and Farrington, J. W. Peru upwelling region sediments near 15°S. 1. Remineralization and accumulation of organic matter. *Limnol. Oceanogr.* 29: 1–19 (1984).
- Thurman, E. M. *Organic Geochemistry of Natural Waters*. Kluwer Academic Publishers, Hingham, MA, 1985.
- Orem, W. H., and Gaudette, H. E. Organic matter in anoxic marine pore water: oxidation effects. *Org. Geochem.* 5: 175–181 (1984).
- Orem, W. H., Hatcher, P. G., Spiker, E. C., Szeverneyi, N. M., and Maciel, G. E. Dissolved organic matter in anoxic pore waters from Mangrove Lake, Bermuda. *Geochim. Cosmochim. Acta* 50: 609–618 (1986).

54. Gschwend, P. M., and Wu, S. On the constancy of sediment-water partition coefficients of hydrophobic organic pollutants. *Environ. Sci. Technol.* 19: 90-96 (1985).
55. Duinker, J. The role of small, low density particles on the partition of selected PCB congeners between water and suspended matter (North Sea area). *Neth. J. Sea Res.* 20: 229-238 (1986).
56. Baker, J. E., Capel, P. D., and Eisenreich, S. J. Influence of colloids on sediment-water partition coefficients of polychlorobiphenyl congeners in natural waters. *Environ. Sci. Technol.* 20: 1136-1143 (1986).
57. Means, J. C., and Wijayartne, R. D. Chemical characterization of estuarine colloidal organic matter: implications for adsorptive processes. *Bull. Mar. Sci.* 35: 449-461 (1984).
58. Morel, F. M. M., and Gschwend, P. M. The role of colloids in the partitioning of solutes in natural waters. In: *Aquatic Surface Chemistry—Chemical Processes at the Particle-Water Interface* (W. Stumm, Ed.), Wiley Intersciences, New York, 1987, pp. 405-422.
59. Hamelink, J. L., Waybrant, R. C., and Ball, R. C. A proposal: exchange equilibria control the degree chlorinated hydrocarbons are biologically magnified in lentic environments. *Trans. Am. Fish. Soc.* 100: 207-214 (1971).
60. NRC. The International Mussel Watch: Report of a Workshop. National Research Council, U.S. National Academy Press, Washington, DC, 1980.
61. Neeley, W. B., Branson, D. R., Blau, G. E. Partition coefficient to measure bioconcentration potential of organic chemicals in fish. *Environ. Sci. Tech.* 13: 1113-1115 (1974).
62. Mackay, D. Correlation of bioconcentration factors. *Environ. Sci. Technol.* 16(5): 274-278 (1982).
63. Hawker, D. W., and Connell, D. W. Relationships between partition coefficient, uptake rate constant, clearance rate constant, and time to equilibrium for bioaccumulation. *Chemosphere* 14: 1205-1219 (1985).
64. Farrington, J. W., Davis, A. C., Brownawell, B. J., Tripp, B. W., Clifford, C. H., and Livramento, J. B. Some aspects of the biogeochemistry of polychlorinated biphenyls in the Achushnet River Estuary. In: *Organic Marine Geochemistry*, ACS Symposium Series No. 305 (M. Sohn, Ed.), American Chemical Society, Washington, DC, 1986, pp. 174-197.
65. Connell, D. W., and Miller, G. J. *Chemistry and Ecotoxicology of Pollution*. John Wiley and Sons, New York, 1984.
66. Boehm, P. D., and Quinn, J. G. The effect of dissolved organic matter in seawater on the uptake of mixed individual hydrocarbons and No. 2 fuel oil by a marine filter feeding bivalve (*Mercenaria mercenaria*). *Mar. Biol.* 44: 227-233 (1976).
67. McCarthy, J. F., Jimenez, B. D., and Barbee, T. Effect of dissolved humic material on accumulation of polycyclic aromatic hydrocarbons: structure-activity relationships. *Aquat. Toxicol.* 7: 15-24 (1985).
68. Farrington, J. W., Goldberg, E. D., Risebrough, R. W., Martin, J. H., and Bowen, V. T. U.S. "Mussel Watch" 1976-1978: an overview of the trace metal, DDE, PCB, hydrocarbon and artificial radionuclide data. *Environ. Sci. Technol.* 17: 490-496 (1983).
69. Farrington, J. W., Teal, J. M., Tripp, B. W., Livramento, J. B., and McElroy, A. Biogeochemistry of petroleum components at the sediment-water interface. Final report prepared for Department of Energy, Bureau of Land Management, U.S. Department of the Interior, Washington, DC, March 1983.
70. Rheadman, J. W., Mantoura, R. F. C., and Rhead, M. M. The physico-chemical speciation of polycyclic aromatic hydrocarbons (PAH) in aquatic systems. *Fresenius. Z. Anal. Chem.* 319: 126-131 (1984).
71. Varanasi, U., Reichert, W. L., Stein, J. E., Brown, D. W., and Sanborn, H. R. Bioavailability and biotransformation of aromatic hydrocarbons in *Environ. Sci. Technol.* 19: 836-841 (1985).
72. Connell, D. W. Bioaccumulation behavior of persistent organic chemicals with aquatic organisms. *Rev. Environ. Contam. Toxicol.* 101:118-154 (1988).
73. Farrington, J. W., Jia, X., Clifford, C. H., Tripp, B. W., Livramento, J. B., Davis, A. C., Frew, N. M., and Johnson, C. G. No. 2 Fuel Oil Compound Retention and Release by *Mytilus edulis*. 1983 Cape Cod Canal Oil Spill. Woods Hole Oceanographic Institution Technical Report WHOI-86-8 (CRC-86-1), Woods Hole, MA, 1986.
74. Marchand, M., and Cabane, F. Hydrocarbures dans les moules et les huîtres. *Rev. Int. Oceanogr. Med.* LIX: 3-30 (1980).
75. Farrington, J. W., Davis, A. C., Frew, N. M., and Rabin, K. S. No. 2 fuel oil compounds in *Mytilus edulis*. Retention and release after an oil spill. *Mar. Biol.* 66: 15-26 (1982).
76. Pruell, R. J., Lake, J. L., Davis, W. R., and Quinn, J. G. Uptake and depuration of organic contaminants by blue mussels, *Mytilus edulis*, exposed to environmentally contaminated sediment. *Mar. Biol.* 91: 497-507 (1986).
77. Phillips, J. D. H. *Quantitative Biological Indicators: Their Use to Monitor Trace Metal and Organochlorine Pollution*. Applied Science Publishers, London, 1980.
78. Stegeman, J. J., and Teal, J. M. Accumulation, release, and retention of petroleum hydrocarbons by the oyster *Crassostrea virginica*. *Mar. Biol.* 22: 37-44 (1973).
79. Langston, W. J. Persistence of polychlorinated biphenyls in marine bivalves. *Mar. Biol.* 46: 35-40 (1978).
80. Mackay, D., and Hughes, A. I. Three parameter equation describing the uptake of organic compounds by fish. *Environ. Sci. Technol.* 18: 439-444 (1984).
81. Stegeman, J. J. Polynuclear aromatic hydrocarbons and their metabolites. In: *Polycyclic Hydrocarbons and Cancer*, Vol. 3 (H. V. Gelboin and P. O. P. Ts'O, Eds.), Academic Press, New York, 1981, pp. 1-60.
82. Stegeman, J. J. Benzo(a)pyrene oxidation and microsomal enzyme activity in the mussel (*Mytilus edulis*) and other bivalve mollusc species from the western North Atlantic. *Mar. Biol.* 89: 21-30 (1985).
83. Livingstone, D. R., Moore, M. N., Lowe, D. M., Casci, C., and Farrar, S. Responses of the cytochrome P-450 monooxygenase system to diesel oil in the common mussel *Mytilus edulis* L. and the periwinkle, *Littorina littorea* L. *Aquat. Toxicol.* 7: 79-91 (1985).
84. Reichert, W. L., Le Ederhart, B. -T., and Varanasi, U. Exposure of two species of deposit-feeding amphipods to sediment-associated (³H) benzo(a)pyrene: uptake, metabolism and covalent binding to tissue macromolecules. *Aquat. Toxicol.* 6: 45-56 (1985).
85. Stein, J. W., Hom, T., and Varanasi, U. Simultaneous exposure of English sole (*Parorhynchus vetulus*) to sediment-associated xenobiotics: Part I—Uptake and disposition of ¹⁴C-polychlorinated biphenyls and ³H-benzo(a)pyrene. *Mar. Environ. Res.* 13: 97-119 (1984).
86. Levin, S. A., Kimball, K. D., McDowell, W. H., and Kimball, S. F. New perspectives in ecotoxicology. *Environ. Manag.* 8: 375-442 (1984).
87. NRC. *Polycyclic Aromatic Hydrocarbons: Evaluation of Sources and Effects*. National Research Council, National Academy Press, Washington, DC, 1983.
88. NRC. *Testing for Effects of Chemicals on Ecosystems*. National Research Council, National Academy Press, Washington, DC, 1981.
89. Rubinstein, N., Gilliam, W. T., and Gregory, N. R. Dietary accumulations of PCBs from a contaminated sediment source by a demersal fish (*Leiostomus xanthurus*). *Aqua. Toxicol.* 5: 331-42 (1984).
90. Foster, G. D., Baksi, S. M., and Means, J. C. Bioaccumulation of trace organic contaminants from sediment by Baltic clams (*Macoma balthica*) and soft-shell clams (*Mya arenaria*). *Environ. Toxicol. Chem.* 6: 969-976 (1987).
91. McElroy, A. E., Farrington, J. W., and Teal, J. M. Bioavailability of polynuclear aromatic hydrocarbons in the aquatic environment. In: *Metabolism of Polynuclear Aromatic Hydrocarbons (PAHs) in the Environment* (U. Varanasi, Ed.), CRC Press, Boca Raton, FL, 1987, pp. 1-39.
92. Oliver, B. G. Desorption of chlorinated hydrocarbons from spiked and anthropogenically contaminated sediments. *Chemosphere* 14: 1087-1106 (1985).
93. O'Connor, D. J., and Piazza, J. C. Pharmacokinetic model for the accumulation of PCB in marine fish. In: *Ocean Processes in Marine Pollution. Biological Processes and Wastes in the Ocean*, Vol. 1 (J. M. Capuzzo and D. R. Kester, Eds.), Robert E. Kreiger Publishing Company, Malabar, FL, 1987, pp. 119-129.
94. Spaulding, M. L., Reed, M., Anderson, E., Isaji, T., Swanson, J. C., Saila, S. B., Lorda, E., and Walker, H. Oil spill fishery impact assessment model: sensitivity to spill location and timing. *Est. Coast. Shelf Sci.* 20: 41-54 (1985).
95. Connor, D. J., Connolly, J. P., and Garland, E. J. Mathematical models—fate, transport, and food chain. In: *Ecotoxicology: Problems and Approaches* (S. A. Levin, M. A. Harwell, J. R. Kelley, and K. D. Kimball, Eds.), Springer-Verlag, New York, 1989, pp. 221-243.
96. Thomann, R. V. Deterministic and Statistical Models of Chemical Fate in Aquatic Systems. In: *Ecotoxicology: Problems and Approaches* (S. A. Levin, M. A. Harwell, J. R. Kelley, and K. D. Kimball, Eds.), Springer-Verlag, New York, 1989, pp. 245-277.
97. Superfund Information Sheet, New Bedford Harbor Site, April, 1987 U.S. E.P.A. Region I, J. F. Kennedy Federal Building, Boston, MA.
98. Connor, M. S. Comparison of the carcinogenic risks from fish vs. ground-water contamination by organic compounds. *Environ. Sci. Technol.* 18: 628-631 (1984).
99. Reuber, B., Mackay, D., Paterson, S., and Stokes, P. A discussion of chemical equilibria and transport at the sediment-water interface. *Environ. Toxicol. Chem.* 6: 731-739 (1987).