

Microbial Infections, Immunomodulation, and Drugs of Abuse

Herman Friedman,* Catherine Newton, and Thomas W. Klein

*Department of Medical Microbiology and Immunology, College of Medicine,
University of South Florida, Tampa, Florida 33612*

INTRODUCTION AND HISTORICAL BACKGROUND	209
OPIATE EFFECTS ON IMMUNITY AND SUSCEPTIBILITY TO INFECTION	209
MARIJUANA-INDUCED ENHANCEMENT OF SUSCEPTIBILITY TO INFECTION	211
COCAINE AND INFECTIONS	212
NICOTINE EFFECTS ON RESISTANCE TO INFECTIONS	212
ALCOHOL MODULATION OF RESISTANCE TO INFECTION	213
DISCUSSION	214
ACKNOWLEDGMENTS	214
REFERENCES	214

INTRODUCTION AND HISTORICAL BACKGROUND

The recreational use of legal and illegal drugs of abuse in this country and abroad has aroused serious concerns about the consequences of these drugs on immunity. Marijuana, cocaine, heroin, and other opiates are widely used illegal drugs. There have been numerous clinical reports on the association between infectious diseases and use of illegal drugs. In addition, legal substances such as alcohol and tobacco have been linked to excessive and addictive use and have been correlated with major health problems. Heavy smokers and/or alcoholics are often hospitalized with infectious diseases. Experimental studies using drugs of abuse support the clinical observations that these substances are associated with immunomodulation.

Studies concerning the effects of addictive drugs on immunity became even more urgent with the onset of the worldwide epidemic of AIDS. AIDS is caused by human immunodeficiency virus (HIV) and results in a collapse of the immune system, making an individual highly susceptible to opportunistic microorganisms (77, 130, 150). Drugs of abuse have been suggested as possible cofactors, resulting in a more rapid progression of disease (58, 63, 69, 212). Approximately one-third of all AIDS patients in the United States are intravenous drug users (IVDUs), and contaminated needles or equipment often spreads HIV (57). AIDS patients also often use other drugs such as marijuana, alcohol, and nicotine, which some investigators think are immunosuppressive (13, 59, 106, 216). Thus, there is concern that abused drugs are serving as cofactors in AIDS progression and in alteration of susceptibility to other infectious diseases (63, 73, 124, 161, 164).

OPIATE EFFECTS ON IMMUNITY AND SUSCEPTIBILITY TO INFECTION

Opiates compose a collection of drugs derived from the poppy *Papaver somniferum* (199) which include opium, morphine, and heroin. An excellent review of the historical use of

opiates has been written by Risdahl et al. (190). It is clear from this review that opiates have had a great impact throughout history on mankind both from use and from the wars over the control of opium. Opium was derived from the Greek word meaning “of sap” or “juice”, because the drug is obtained from the juice of the poppy plant. Relics from the Stone Age, pre-dating recorded history, show widespread poppy cultivation. Over time, the addictive nature of opium was also recognized. First morphine (in the early 1800s) and then codeine, heroin, and other opium alkaloids (in the late 1800s) were synthesized from opium with claims of being the cure of opium addiction (91).

During the late 1800s and early 1900s, many medical practitioners began to recognize infections as serious complications of opiate addiction (28, 105, 165, 260). The list of infections associated with opiates continued to increase during the 1900s (89, 90). Also, experimental evidence began to accumulate during this time demonstrating the detrimental effects of opiates on immunity in humans and animals (90, 115, 116, 123, 187). For example, Cantacuzene demonstrated in the late 1890s that morphine-treated guinea pigs had altered phagocytosis and leukocyte trafficking (36). Studies of drug addicts in the early 1970s further demonstrated a connection between drug use and infectious diseases (117, 185). It is now recognized that IVDUs face many complications from the use of opiates (243). Pulmonary infections, caused by *Mycobacterium*, *Staphylococcus*, *Streptococcus*, *Haemophilus*, and other bacteria, are among the most common diagnoses of opiate abusers (190). Other serious diseases caused by microbial pathogens in IVDUs are AIDS (HIV), endocarditis (*Staphylococcus*, *Enterococcus*, *Pseudomonas*, *Klebsiella*, *Serratia*, and *Candida*), abscesses and cellulitis (*Staphylococcus*, *Streptococcus*, *Haemophilus*, *Enterobacter*, *Pseudomonas*, *Klebsiella*, *Clostridium*, *Candida*, and others), hepatitis A, B, and C (hepatitis A, B, and C viruses), sexually transmitted diseases, and skeletal infections (*Staphylococcus* and *Pseudomonas*) (77, 188, 190).

A large percentage of infections among IVDUs are related to the methods of injection and life-style practices, which increase their exposure to microbial pathogens (57). Recent reports continue to discuss the problem of contaminated heroin or drug paraphernalia and infections (15, 47). Numerous in-

* Corresponding author. Mailing address: Department of Medical Microbiology and Immunology, College of Medicine, University of South Florida, Tampa, FL 33612. Phone: (813) 974-3281. Fax: (813) 974-4151. E-mail: hfriedma@hsc.usf.edu.

TABLE 1. Effects of opiates on immune functions in vivo and in vitro

Mode of administration and model	Immune function ^a	Receptor involvement ^a	Reference(s)
In vivo			
Rodents	↓ Phagocytosis	+	168, 191, 240
	↓ Antibody production	+	32, 120, 172, 184
	↓ Mitogen-stimulated proliferation	+	18, 19
	↓ NK/CTL activity	+	18, 27, 207, 258
	↓ Cytokine production	+	27, 125, 194
	↑ Serum IL-6 levels	+	83, 84
	↑ LPS-induced sepsis	NE	195
	↑ IL-12	+	173
	↓ DTH	+	31
Humans	↓ Phagocytosis	NE	239
	↓ NK activity	NE	265
	↓ ADCC	NE	265
Monkeys	↓ Chemotaxis	NE	121
	↓ PMN killing	NE	121
Pigs	↓ DTH	NE	148, 189
In vitro			
Mice	↓ Phagocytosis	+	191, 235
	↓ Cell proliferation	+	18, 19, 203
	↓ Induction of antibody	+	60, 72, 237
	↓ Cytokine production	+	4, 22, 98, 238
Humans	↓ Chemotaxis	+	71
	↓ Superoxide	NE	176
	↓ ↑ Cytokine production	+	38, 39, 179
	↓ ↑ Chemokine production	+	237, 259
	↑ Phagocytosis	+	178

^a ↑, increase; ↓, decrease; NE, not examined; CTL, cytotoxic T lymphocyte; LPS, lipopolysaccharide; DTH, delayed-type hypersensitivity; ADCC, antibody-dependent cellular cytotoxicity; PMN, polymorphonuclear leukocyte.

investigators, however, have proposed that increased exposure is not the only factor that enhances microbial infections. They hypothesize that opiates cause immunosuppression and therefore serve as cofactors for microbial infections. These studies have been previously reviewed (59, 139). They have been expanded on by the advent of the AIDS epidemic in the 1980s (30, 176). AIDS and its corresponding decreased host immunity were, and still are, principal players in opportunistic infectious diseases in IVDUs (63, 76, 124, 150). Several studies support that intravenous use of opiates influences the outcome of HIV infection (16, 58, 63). Heroin addicts have been observed to have an increased risk of acquiring HIV (16, 227), and at one time half of IVDUs from certain areas of the United States were infected with HIV (103). Moreover, mortality rates from infectious diseases among HIV-infected IVDUs decrease when drug use was discontinued, and this abatement correlated with a decrease in the rate of progression to AIDS (256). Therefore, a correlation between the use of opiates, increased susceptibility to infection, and depressed immunity does indeed exist. Whether this correlation is due, however, to increased exposure to infectious pathogens through risky behaviors, to immunosuppressive effects of opiates, or to a combination of these two is uncertain at this time.

Experimental studies to investigate the effect of opioids on

immune responses and on microbial pathogens have extended the earlier clinical correlative observations and animal studies (Tables 1 and 2). Several good reviews of these studies have been previously published (59, 139, 188, 190). Several opiate receptors have been identified on cells of the nervous system, with μ -, κ -, and δ -receptors and their subtypes being the most predominant and being referred to as classical receptors (2). The classical receptors are G-protein coupled seven-transmembrane receptors (186). Opiates have been linked to modulations of host resistance to bacterial, protozoan, viral, and fungal infections, using animal models, cell lines, and primary cells. Opiates appear to affect the immune response directly through opioid receptors on immune cells and indirectly via the receptors on neuronal cells. The μ -, κ -, and δ -opioid receptors as well as nonclassical opioid-like receptors have been demonstrated on immune cells, suggesting possible mechanisms for the direct actions of opiates on immune cells (139). In vitro studies of immune cells have demonstrated receptor-mediated reduced phagocytosis (235), chemotaxis (71), and cytokine and chemokine production (4, 22, 39, 179).

While opiates directly modulate host immunity, their effects on physiological function of nonspecific host mechanisms are thought to also alter immune responses and play an important role in increased susceptibility to infection. These effects are proposed to act through the central nervous system (CNS) and the hypothalamus-pituitary-adrenal (HPA) axis. Opiates are known to alter the release of HPA hormones (corticotrophin-releasing hormone and adrenocorticotrophic hormone) (5), which, in turn, alter glucocorticoids (cortisol and corticosterone), the end-effectors of the HPA axis. The glucocorticoids play an important role in decreasing and regulating cellular immune responses (29). Studies have shown that morphine treatments suppress immune parameters in mice through the HPA axis (31, 64, 183, 200, 201). In addition to these corticoids, immunosuppression via the autonomic nervous system has been observed (144, 257). Shavit et al. observed that natural killer (NK) activity in rats was suppressed following morphine injection into the lateral ventricle of the brain via opioid receptors (206, 207). The central opioid pathways were involved in immunosuppression of lymphocyte proliferation (78, 82, 126). Receptor-mediated increase in the production of transforming growth factor β , an immunosuppressive cytokine, is another possible indirect method by which opiates suppress immunity (38). Thus, it appears that immunosuppression occurs through direct and indirect mechanism involving receptors on immune cells and the CNS.

TABLE 2. Opiates and microbial pathogens

Pathogen	Host species	Effect ^a	Reference(s)
<i>Salmonella enterica</i> serovar Typhimurium	Mice	↑ Gut colonization	127
<i>Toxoplasma gondii</i>	Mice	↑ Mortality	40
Endogenous ^b	Mice	↑ Sepsis	81
HSV-1	Mice	↑ Infection	169
FLV	Mice	↑ Mortality	224, 242
<i>Candida albicans</i>	Mice	↑ Mortality	240
Swine herpesvirus plus <i>Pasteurella multocida</i>	Pigs	↑ Pneumonia	189

^a ↑, increase; ↓, decrease.

^b Retrovirus.

TABLE 3. Effects of THC and other cannabinoids on immune functions in vivo and in vitro

Mode of administration and model	Immune function ^a	Reference(s)
In vivo		
Humans	↓ Lymphoproliferation	154
	↓ Antimicrobial activity	14
	↓ Cytokine production	14
Mice	↓ ↑ Serum Ig levels	155
	↓ Cellular immunity	112, 163, 214
	↓ Antimicrobial activity	8, 34, 147, 222
	↓ Humoral immunity	10, 102, 198
	↑ Apoptosis	140
	↓ ↑ Cell signaling	50, 241
In vitro		
Humans	↓ Lymphocyte proliferation	153, 221
	↓ NK cell activity	219, 220
	↓ Neutrophil antifungal activity	55
	↑ ↓ Cytokine production	223, 255
Mice	↓ Lymphocyte proliferation	113, 140, 181, 182
	↓ NK cell activity	104, 110, 133
	↓ Antibody formation	9, 107
	↓ ↑ IL-2 system receptors	46, 79, 270, 271
	↓ ↑ Cytokine production	14, 108, 162
	↑ Apoptosis	140, 269
	↓ ↑ Cell signaling	46, 62, 79

^a ↑, increase; ↓, decrease; ↑ ↓, both increase and decrease.

MARIJUANA-INDUCED ENHANCEMENT OF SUSCEPTIBILITY TO INFECTION

Marijuana is the common name for *Cannabis sativa*, a plant that has long been known for its “medicinal” and recreational properties and for its fiber (hemp). Chemical extracts of marijuana contain over 400 compounds and more than 60 cannabinoids. Cannabinoids, especially the major psychoactive component Δ⁹-tetrahydrocannabinol (THC), exert immunomodulatory effects that alter normal functions of T and B lymphocytes, NK cells, and macrophages in human and animals. These modulations have been observed during both in vivo and in vitro cannabinoid treatment (Table 3). The molecular and cellular mechanisms for these effects are not fully defined; however, it appears that receptor as well as nonreceptor mechanisms are involved (106). Like opiate receptors, cannabinoid receptors (CBRs) are G-protein-coupled seven-transmembrane receptors of which two types have been identified, CB₁ and CB₂ (85, 86, 108, 134, 151, 174). CB₁ receptors are associated with the brain and certain peripheral tissues and are responsible for behavioral effect of THC, while CB₂ receptors are located in the periphery, especially on immune cells (86, 108, 174). The discovery of CBR has led to the identification of a class of endogenous compounds that bind to these receptors, called endocannabinoids, although the majority of the compounds are eicosanoids (53, 138, 142, 261). The broad spectrum of action of THC on immune functions is thought to result in decreased host resistance to bacterial and viral infections as observed in various experimental animal models (Table 4).

Studies in the early 1970s using human peripheral blood

mononuclear cells (PBMCs) from marijuana smokers showed a tendency for heavy use to result in suppression of lymphocyte proliferation in culture as well as alterations in PBMCs immune cell subsets (154). Serum immunoglobulin (Ig) levels were also modulated by marijuana use, with IgG protein levels decreasing and IgE protein levels increasing. Animal studies started in earnest with the isolation and synthesis of THC by Mechoulam et al. (143), when it became possible to inject THC into animals or treat human and animal cells in vitro. Various groups have observed both in vivo and in vitro that THC suppresses immune functions (Table 3). These functions vary from lymphocyte proliferation and antibody production to cytotoxic activity (Table 3). Other studies have demonstrated that THC enhances certain functions (Table 3). B-cell proliferation increased in the presence of THC at nanomolar concentrations (51), and the production of the chemokines MIP1α and interleukin-8 (IL-8); increased at micromolar concentrations (223). The latter group also observed decreases in the levels of other cytokine after THC treatment (223). Therefore, the data that have accumulated over the past three decades indicate that THC and cannabinoids are immunomodulatory.

One of the important risk factors of marijuana use is its suppression of host resistance to infections (99). This aspect has been studied in both humans and animals, and the results have suggested that cannabinoids have a moderating effect on various infection paradigms and that at least some of the effects involve CBRs (Table 4) (33, 112). A correlation between marijuana smoking and herpesvirus infection was observed to increase the risk of mortality in HIV positive marijuana smokers (211). Furthermore, alveolar macrophages from marijuana smokers were found to be deficient in several functional properties including phagocytosis and bactericidal activity (14). Experimental animal studies have also suggested that THC treatment causes increased susceptibility to various infectious agents (Table 4). Disease progression and mortality in different animal models were increased on infection with herpes simplex virus (HSV) and Friend leukemia virus (FLV) (35, 147, 222) and with bacterial pathogens such as *Listeria*, *Treponema*, and *Staphylococcus* (87, 149, 170).

These studies leave major gaps in our understanding of the cellular and molecular mechanisms mediating these effects on immunity and resistance. Immune cells have been demonstrated to express CBR; therefore, it is likely that at least a portion of the cannabinoid-induced modulations of the immune cells are directly mediated via their own CBRs (79, 119). The host immunity, however, involves many cell types, both

TABLE 4. Effects of cannabinoids on resistance to infections

Infectious agent	Host	Effect ^a	Reference(s)
HSV	Mice	↑ Mortality	147, 149
<i>Listeria</i>	Mice	↑ Mortality	149
HSV	Humans	↑ Recurrence	100
HSV	Guinea pigs	↑ Infection	35
FLV + HSV	Mice	↑ Mortality	222
<i>Staphylococcus</i>	Rats	↓ Lung infection	87
<i>Treponema pallidum</i>	Rabbits	↑ Progression	170
<i>Legionella</i>	Mice	↑ Mortality	111, 163
<i>Staphylococcus</i>	Rats	↓ Macrophage activity	
HIV	Humans	↑ Risk of mortality	211

^a ↑, increase; ↓, decrease.

TABLE 5. Effects of cocaine on immune functions

Mode of administration	Immune function ^a	Reference(s)
In vivo		
Humans	↓ Antimicrobial activity ↑ ↓ Cytokine production	14 14
Rodents	↓ Lymphoproliferation ↓ Antibody formation ↓ DTH ↓ ↑ Humoral immunity ↑ ↓ Cytokine production	20, 21, 171 166, 253 253 11 250
In vitro		
Humans	↑ HIV replication ↓ Lymphocyte proliferation ↓ Cytokine production	12, 177 23, 49, 109 131, 132
Rodents	↓ Cytokine production ↓ Lymphocyte proliferation ↓ NK activity	250, 254, 262 262 262

^a ↑, increase; ↓, decrease; ↑ ↓, both increase and decrease; DTH, delayed-type hypersensitivity.

immune and nonimmune, as well as chemical factors such as cytokines and chemokines and hormones of the HPA axis. Thus, there are numerous cellular and molecular mechanisms where THC could be exerting its effects as demonstrated by our *Legionella pneumophila* infection studies. In our studies, THC pretreatment of mice infected with THC affects both innate immunity and the development of the adaptive (cell-mediated) immune response. Initially, we reported that mice receiving a THC injection 1 day before and 1 day after a sublethal *L. pneumophila* infection died of septic shock resulting from a detrimental production of high levels of proinflammatory cytokines (111). Induction of tumor necrosis factor alpha (TNF- α) has since been confirmed, most recently in CB₂-transfected HL60 cells stimulated with the cannabinoid agonist CP55,940 (52). We further observed that a single injection of THC 18 h prior to infection inhibited the development of Th1 immunity in mice (163), which involved both CB₁ and CB₂ receptors and suppression of Th1 development by inhibiting the production of gamma interferon and IL-12 and reducing the amount of IL-12R β 2 mRNA (112). This THC-induced shift away from a Th1 response has also been observed in other models involving THC treatment and tumor immunity (268), endotoxemic mice (215), and NK cell activity (133). These studies suggest that cannabinoids have the ability to bias the developing immune response from Th1 (cell-mediated) toward Th2 (antibody-mediated) immunity. Interestingly, Th shifts have also been observed toward Th2 following treatment with morphine (193) and toward Th1 following treatment with norepinephrine (229). It is possible, therefore, that drugs used either recreationally or therapeutically might enhance or suppress infections by modulating Th activity in the host. Clearly, the full extent of these findings needs clarification.

COCAINE AND INFECTIONS

Cocaine is derived from the coca plant, *Erthroxylon coca*. Being an alkaloid, cocaine is water soluble and readily absorbed through

TABLE 6. Effects of cocaine on resistance to infections

Infectious agent	Host	Effect ^a	Reference
LP-BM5 retrovirus	Mice	↑ Cryptosporidiosis	48
HIV	SCID mice implanted with human PBMCs	↑ HIV-infected PBL ↑ Virus load ↓ CD4/CD8 ratio	192 192 192

^a ↑, increase; ↓, decrease; PBL, peripheral blood leukocytes.

mucous membranes of the body. It appears to function at least partially through the sigma₁ (σ_1) receptor, a protein first proposed to be involved with morphine binding (122, 135, 137, 205, 228). The σ_1 receptors are distributed throughout the brain and periphery of the body (137), similar to the classical opiate and cannabinoid receptors.

A limited number of in vivo and in vitro studies have been done to examine cocaine-induced modulation of immune responses (Tables 5) and infections (Table 6). Much of the work with cocaine and infections has been centered on HIV and progression to AIDS (13). Epidemiologic studies on IVDUs and AIDS link abuse of cocaine, even more than other drugs, to increased incidence of HIV seroprevalence and progression of AIDS (7, 37, 42, 56). Cocaine increases HIV infection of human PBMCs in vitro (12, 177). Roth et al., using a model of human PBMCs implanted into severe combined immunodeficient (SCID) mice, demonstrated that cocaine treatments resulted in increased numbers of HIV-infected PBMCs and viral load, as well as a decreased CD4/CD8 ratio (192). These immunomodulations may be through receptors located in the periphery, as was demonstrated with cocaine-induced suppression of mitogen-stimulated lymphoproliferation (171). Therefore, while studies of the immunological impact of cocaine have started, there is much more research in this area to be done.

NICOTINE EFFECTS ON RESISTANCE TO INFECTIONS

Cigarette smoking is linked to community-acquired pneumonia and is considered one of the risk factors for respiratory

TABLE 7. Effects of nicotine and cigarette smoke on host resistance mechanisms in vivo and in vitro

Mode of administration	Immune function ^a	Reference(s)
In vivo		
Rats	↓ Antibody-forming cells ↓ Intracellular Ca ²⁺ stores ↓ Lymphocyte proliferation ↑ T-cell anergy ↓ Antimicrobial activity	66, 67 101 218 67, 218 217
In vitro		
Humans	↓ NK activity ↓ Cytokine production	156 167
Mice	↓ Splenocyte proliferation ↓ ↑ Cytokine production ↓ Antimicrobial activity	74, 75 74, 75, 136 136

^a ↑, increase; ↓, decrease; ↑ ↓, both increase and decrease.

TABLE 8. Effects of alcohol on immune functions

Mode of administration	Immune function ^a	Reference(s)
In vivo		
Mice	↓ ↑ Cytokine production	3, 94, 210, 246, 248, 249, 251
	↓ NK activity	141, 249
	↓ IgA and IgG production	249
	↑ Apoptosis	61
	↓ DTH	245, 246
Rats	↓ TNF-α	114, 158
	↓ Serum chemokines	266
	↓ TNF-α processing	267
	↓ Alveolar nitric oxide	70, 114
	↓ Chemokines production by Kupffer cells	17
Humans	↓ Monocyte stimulated proliferation	232
In vitro		
Mice	↓ Macrophage killing	26
	↑ Bactericidal capacity	263
	↓ Cytokine production	41
Humans	↓ Macrophage killing	26
	↓ ↑ Cytokine production	25, 68, 231, 233, 234, 244
	↓ TNF-α receptor	25
	↓ NF-κB activation	129
	↓ T-cell proliferation	232
Rhesus macaques	↓ TNF-α	226

^a ↑, increase; ↓, decrease; ↑ ↓, both increase and decrease; DTH, delayed-type hypersensitivity.

infections (6, 89, 196). Cigarette smoke is composed of two components, the vapor phase and the particulate phase. The immunosuppressive effects of smoke and nicotine occur in the particulate portion, thus suggesting that the nicotine is at least partially responsible for the inhibitory effects on the immune responses (Table 7) (216, 217). Nicotine is a small organic alkaloid synthesized by tobacco plants and is recognized as the addictive component of cigarettes. While its lipophilic nature allows small amounts to cross directly through cell membranes, the primary biological effects are proving to be receptor mediated. Nicotine is an agonist for nicotinic acetylcholine receptors (nAChRs), which are present on cells of the CNS as well as other cells throughout the body including immune cells (80, 118). The neural nAChRs are upregulated in smokers (118, 264). Rapid progression of nicotine from cigarette smoke in the lungs to the brain increases dopamine transmission within the brain in the shell of the nucleus accumbens, a region essential for reward processing that has been associated with addictive properties of other drugs including opiates, alcohol, and THC (180, 236).

Nicotine appears to affect the immune system through nAChRs on cells in CNS and on immune cells similar to opiates and cannabinoids. Nicotine induces glucocorticoids, through the HPA axis, that modify the immune system (29, 80, 216) as well as directly affecting immune cells (74, 75, 136, 167, 217). Nicotine was demonstrated to enhance the growth of *L. pneumophila* and cause a corresponding inhibition of IL-6,

TNF-α, and IL-12 in a murine alveolar macrophage cell line through nAChRs (136). The substance also affects murine splenocyte production of Th1- and Th2-associated cytokines in a differential manner (74, 75). Chronic nicotine treatment of rats induces T-cell anergy, depletes intracellular IP3-sensitive Ca²⁺ stores, and inhibits the antibody-forming cell response and lymphocyte proliferation, which may prevent the animals from developing a protective immune response to microbial pathogens (66, 67, 101). In vitro treatments of PBMCs by nicotine and other extracts from cigarette were observed to inhibit cytokine production (167). Rodents exposed to cigarette smoke in inhalation chambers have increased susceptibilities to infections when challenge with aerosolized bacteria or viruses (217). Smoking among HIV-positive individuals has also been linked to increased numbers of infections (164, 213).

Thus, it is important to determine how nicotine, a legal addictive drug, increases or alters susceptibility to infectious diseases. Studies of this nature have begun with the apparent linking of the effects to nAChRs. However, much more information is needed to ascertain the nature and mechanism whereby nicotine influences the immune response and thus affects host resistance to infectious diseases.

ALCOHOL MODULATION OF RESISTANCE TO INFECTION

Alcohol abuse causes widespread health problems including decreased liver functions and increased incidences of infectious diseases (45). Alcoholics have long been recognized to be particularly susceptible to infections and to be at a greater risk of community-acquired pneumonias (1, 43, 88, 128, 157, 160, 196, 225). Moderate alcohol use (one beer, or one glass of wine, or one mixed drink per day), which may be beneficial to the immune system (159, 230), is not covered here.

Alcohol, unlike the previous drugs of abuse discussed, does not appear to involve receptor mediation. Studies during the last decade have demonstrated that alcohol has multiple effects on the host immune responsiveness to microbial pathogens (Tables 8 and 9). These effects are characterized by depletion of circulating lymphocyte populations and altered lymphoid organ architecture and immune functions (95, 152, 209, 230). In addition, alcohol suppresses the production of cytokines important in antimicrobial immunity, such as TNF-α secreted by mononuclear cells in vitro and/or in vivo, including alveolar macrophages from rats (114, 160) and rhesus macaques (226). The suppression of TNF-α is posttranscriptional and involves

TABLE 9. Effects of alcohol on resistance to infections

Infectious agent	Host	Effect ^a	Reference(s)
<i>Listeria</i>	Mice	↓ Infection	4, 95, 197
<i>Salmonella</i>	Mice	↓ Infection	95, 208
<i>Streptococcus</i>	Mice	↓ Infection	204
<i>Mycobacterium</i>	Mice	↑ Disease	24, 26
<i>Mycobacterium</i> -BCG	Rats	↓ Bactericidal	145
LP-BM5 retrovirus	Mice	↑ Disease	248, 251
LP-BM5 retrovirus	Mice	↑ <i>Cryptosporidium parvum</i> infection	3
LP-BM5 retrovirus	Mice	↑ Coxsackievirus-induced myocarditis	202

^a ↑, increase; ↓, decrease.

TNF- α -converting enzyme-mediated processing of TNF- α (114, 267). Furthermore, in human monocytes, alcohol inhibits lipopolysaccharide LPS-induced activation of NF- κ B, a transcription factor for inflammatory cytokines (129). Of particular interest is the observation that alcohol use also decreases Th1 cytokine levels and responses (68, 230, 232, 246, 247) and increases Th2 cytokine levels (68, 248, 251), similar to the effect observed with THC (112, 163, 268) and morphine (193). Indeed, Peterson et al. reported that IL-12 therapy could attenuate the suppressed cell-mediated immunity in ethanol-consuming mice (175).

Rodents given alcohol orally show modulation in immune cell functions (Table 8) and infections (Table 9). Bermudez and Young showed that ethanol augments the intracellular survival of *Mycobacterium avium* complex and impairs macrophage responses to cytokines (26). Ethanol treatment increases the growth of *L. pneumophila* in nonpermissive macrophage cultures (263). Furthermore, studies by Jerrells and colleagues showed that immune cells from mice given alcohol and infected with intracellular bacteria (*Salmonella* or *Listeria*) had increased susceptibility to the bacteria (94–97, 197, 208).

Alcohol has also been connected to viral infections. The HSV-2 incidence is increased in women who abuse alcohol (44). Experimental studies indicate that alcohol inhibits Th1 responses generated to LP-BM5, a retrovirus that causes a murine AIDS-like syndrome (3, 248, 251, 252). Whether alcohol serves as a cofactor in AIDS is uncertain (54, 146). However, alcohol does exacerbate opportunistic infections in murine AIDS-like syndrome (3, 202) and opportunistic infections are correlated with the progression of AIDS. Hepatitis C virus infection has also been linked to chronic liver disease in alcoholics (92). Animal studies indicate that alcohol enhances liver damage by activating CD8 cells (93) and increasing apoptosis (65). Thus, the animal models and clinical studies imply that alcohol abuse is detrimental to the host and causes increase susceptibility to disease from microbial pathogens.

DISCUSSION

In recent years there have been more studies concerning the relationship between the use of addictive drugs of abuse and the increased incidence of susceptibility to infectious diseases, including AIDS. These studies have shown that drugs of abuse, including marijuana, cocaine, opiates, alcohol, and nicotine, alter not only neuropsychological and pathophysiological responses of individuals but also immune functions. Such studies support the earlier correlative observations that the use of these drugs is associated with enhanced susceptibility to infectious diseases.

The mechanisms by which abused drugs increase susceptibility to infections in humans as well as experimental animals have begun to be delineated. From the studies reviewed here, it appears that all five classes of drugs affect the immune system through both indirect and direct mechanisms. One indirect method is drug-induced stimulation of the HPA axis, which results in glucocorticoid production and regulation of the immune system. In addition, these abused substances have direct actions on immune cells that seem to be receptor mediated for all of the drugs except alcohol. Studies of receptor-mediated effects on immunity and infection have been per-

formed in detail with opiates and to a lesser degree with cannabinoids and have just been started with nicotine. Another common mechanism of action among the five classes of these abused drugs is their effect on Th1/Th2 responses, either by inhibition of Th1- or elevation of Th2-associated cytokines.

The correlation between IVDUs and HIV infections has led many investigators to propose that the immunomodulation mediated by drugs is a major factor contributing to the progression of AIDS in IVDUs. While it is impossible to determine the cause-and-effect relationships from epidemiological studies, there is growing consensus among investigators of drugs of abuse that drug-induced immunomodulation is involved. Studies of immunosuppression by drugs of abuse are supporting increased susceptibility to opportunistic infectious pathogens by alteration of the immune response. However, there is still convincing evidence that the social practices connected with drug abuse also contribute to increase exposure to infectious pathogens. In the end, it logically seems that it will be a combination of increased exposure and drug-induced immunomodulation that contributes to increase susceptibility to infectious pathogens. A concerted enterprise, however, is essential to determine the mechanisms by which drugs compromise immune responses in general and in concert with immunosuppressive viruses.

ACKNOWLEDGMENTS

We acknowledge with gratitude the invaluable contributions of Susan Pross and Yoshimasa Yamamoto, Department of Medical Microbiology and Immunology, University of South Florida College of Medicine, to studies of the effects of alcohol and nicotine on immunity.

Studies in our laboratories were supported by research grants from the U.S. Public Health Service (DA03646, DA10683, AI45169, and DA07245).

REFERENCES

- Adams, H. G., and C. Jordan. 1984. Infections in the alcoholic. *Med. Clin. North Am.* **68**:179–200.
- Adler, M. W., E. B. Geller, T. J. Rogers, E. E. Henderson, and T. K. Eisenstein. 1993. Opioids, receptors, and immunity. *Adv. Exp. Med. Biol.* **335**:13–20.
- Alak, J. L., M. Shahbazian, D. S. Huang, Y. Wang, H. Darban, E. M. Jenkins, and R. R. Watson. 1993. Alcohol and murine acquired immunodeficiency syndrome suppression of resistance to *Cryptosporidium parvum* infection during modulation of cytokine production. *Alcohol Clin. Exp. Res.* **17**:539–544.
- Alicea, C., S. Belkowski, T. K. Eisenstein, M. W. Adler, and T. J. Rogers. 1996. Inhibition of primary murine macrophage cytokine production *in vitro* following treatment with the kappa-opioid agonist U50,488H. *J. Neuroimmunol.* **64**:83–90.
- Allolio, B., H. M. Schulte, U. Deuss, D. Kallabis, E. Hamel, and W. Winkelmann. 1987. Effect of oral morphine and naloxone on pituitary-adrenal response in man induced by human corticotropin-releasing hormone. *Acta Endocrinol. (Copenhagen)* **114**:509–514.
- Almirall, J., I. Bolibar, X. Balanzo, and C. A. Gonzalez. 1999. Risk factors for community-acquired pneumonia in adults: a population-based case-control study. *Eur. Respir. J.* **13**:349–355.
- Anthony, J. C., D. Vlahov, K. E. Nelson, S. Cohn, J. Astemborski, and L. Solomon. 1991. New evidence on intravenous cocaine use and the risk of infection with human immunodeficiency virus type 1. *Am. J. Epidemiol.* **134**:1175–1189.
- Ashfaq, M. K., E. S. Watson, and H. N. Elshohly. 1987. The effect of subacute marijuana smoke inhalation on experimentally induced dermonecrosis by *S. aureus* infection. *Immunopharmacol. Immunotoxicol.* **9**:319–331.
- Baczynsky, W. O. T., and A. M. Zimmerman. 1983. Effects of Δ^9 -tetrahydrocannabinol, cannabinal and cannabidiol on the immune system in mice. II. *In vitro* investigation using cultured mouse splenocytes. *Pharmacologist* **26**:12–19.
- Baczynsky, W. O. T., and A. M. Zimmerman. 1983. Effects of Δ^9 -tetrahydrocannabinol, cannabinal, and cannabidiol on the immune system in mice. I. *In vivo* investigation of the primary and secondary immune response. *Pharmacologist* **26**:1–11.

11. **Bagasra, O., and L. Forman.** 1989. Functional analysis of lymphocytes subpopulations in experimental cocaine abuse. I. Dose-dependent activation of lymphocyte subsets. *Clin. Exp. Immunol.* **77**:289–293.
12. **Bagasra, O., and R. J. Pomerantz.** 1993. Human immunodeficiency virus type 1 replication in peripheral blood mononuclear cells in the presence of cocaine. *J. Infect. Dis.* **168**:1157–1164.
13. **Baldwin, G. C., M. D. Roth, and D. P. Tashkin.** 1998. Acute and chronic effects of cocaine on the immune system and the possible link to AIDS. *J. Neuroimmunol.* **83**:133–138.
14. **Baldwin, G. C., D. P. Tashkin, D. M. Buckley, A. N. Park, S. M. Dubinett, and M. D. Roth.** 1997. Marijuana and cocaine impair alveolar macrophage function and cytokine production. *Am. J. Respir. Crit. Care Med.* **156**:1606–1613.
15. **Bangsberg, D. R., J. I. Rosen, T. Aragon, A. Campbell, L. Weir, and F. Perdreau-Remington.** 2002. Clostridial myonecrosis cluster among injection drug users: a molecular epidemiology investigation. *Arch. Intern. Med.* **162**:517–522.
16. **Battjes, R. J., C. G. Leukefeld, R. W. Pickens, and H. W. Haverkos.** 1988. The acquired immunodeficiency syndrome and intravenous drug abuse. *Bull. Narc.* **40**:21–34.
17. **Bautista, A. P.** 2001. Acute alcohol intoxication and endotoxemia desensitize HIV-1 gp120-induced CC-chemokine production by Kupffer cells. *Life Sci.* **68**:1939–1949.
18. **Bayer, B. M., S. Daussin, M. Hernandez, and L. Irvin.** 1990. Morphine inhibition of lymphocyte activity is mediated by an opioid dependent mechanism. *Neuropharmacology* **29**:369–374.
19. **Bayer, B. M., M. R. Gastonguay, and M. C. Hernandez.** 1992. Distinction between the *in vitro* and *in vivo* inhibitory effects of morphine on lymphocyte proliferation based on agonist sensitivity and naltrexone reversibility. *Immunopharmacology* **23**:117–124.
20. **Bayer, B. M., M. C. Hernandez, and X. Z. Ding.** 1996. Tolerance and cross tolerance to the suppressive effects of cocaine and morphine on lymphocyte proliferation. *Pharmacol. Biochem. Behav.* **53**:227–234.
21. **Bayer, B. M., S. E. Mulrone, M. C. Hernandez, and X. Z. Ding.** 1995. Acute infusions of cocaine result in time- and dose-dependent effects on lymphocyte responses and corticosterone secretion in rats. *Immunopharmacology* **29**:19–28.
22. **Belkowski, S. M., C. Alicea, T. K. Eisenstein, M. W. Adler, and T. J. Rogers.** 1995. Inhibition of interleukin-1 and tumor necrosis factor- α synthesis following treatment of macrophages with the kappa opioid agonist U50,488H. *J. Pharmacol. Exp. Ther.* **273**:1491–1496.
23. **Berkeley, M. B., S. Daussin, M. C. Hernandez, and B. M. Bayer.** 1994. *In vitro* effects of cocaine, lidocaine and monoamine uptake inhibitors on lymphocyte proliferative responses. *Immunopharmacol. Immunotoxicol.* **16**:165–178.
24. **Bermudez, L. E., M. Petrofsky, P. Kolonoski, and L. S. Young.** 1992. An animal model of *Mycobacterium avium* complex disseminated infection after colonization of the intestinal tract. *J. Infect. Dis.* **165**:75–79.
25. **Bermudez, L. E., M. Wu, J. Martinelli, and L. S. Young.** 1991. Ethanol affects release of TNF and GM-CSF and membrane expression of TNF receptors by human macrophages. *Lymphokine Cytokine Res.* **10**:413–419.
26. **Bermudez, L. E., and L. S. Young.** 1991. Ethanol augments intracellular survival of *Mycobacterium avium* complex and impairs macrophage responses to cytokines. *J. Infect. Dis.* **163**:1286–1292.
27. **Bhargava, H. N., P. T. Thomas, S. Thorat, and R. V. House.** 1994. Effects of morphine tolerance and abstinence on cellular immune function. *Brain Res.* **642**:1–10.
28. **Biggam, A. G.** 1929. Malignant malaria associated with the administration of heroin intravenously. *Trans. R. Soc. Trop. Med. Hyg.* **23**:147.
29. **Boumpas, D. T., G. P. Chrousos, R. L. Wilder, T. R. Cupps, and J. E. Balow.** 1993. Glucocorticoid therapy for immune-mediated diseases: basic and clinical correlates. *Ann. Intern. Med.* **119**:1198–1208.
30. **Bryant, H. U., E. W. Bernton, and J. W. Holaday.** 1987. Immunosuppressive effects of chronic morphine treatment in mice. *Life Sci.* **41**:1731–1738.
31. **Bryant, H. U., and R. E. Roubesh.** 1990. Suppressive effects of morphine pellet implants on *in vivo* parameters of immune function. *J. Pharmacol. Exp. Ther.* **255**:410–414.
32. **Bussiere, J. L., M. W. Adler, T. J. Rogers, and T. K. Eisenstein.** 1992. Differential effects of morphine and naltrexone on the antibody response in various mouse strains. *Immunopharmacol. Immunotoxicol.* **14**:657–673.
33. **Cabral, G., and D. Dove Pettit.** 1998. Drugs and immunity: cannabinoids and their role in decreased resistance to infectious diseases. *J. Neuroimmunol.* **83**:116–123.
34. **Cabral, G. A., P. J. McNerney, and E. M. Mishkin.** 1986. Delta-9-tetrahydrocannabinol enhances release of herpes simplex virus type 2. *J. Gen. Virol.* **67**:2017–2022.
35. **Cabral, G. A., E. M. Mishkin, F. Marciano-Cabral, P. Coleman, L. Harris, and A. E. Munson.** 1986. Effect of Δ^9 -tetrahydrocannabinol on herpes simplex virus type 2 vaginal infection in the guinea pig. *Proc. Soc. Exp. Biol. Med.* **182**:181–186.
36. **Cantacuzene, J.** 1898. Nouvelles recherches sur le mode de destruction des vibrions dans l'organisme. *Ann. Inst. Pasteur* **12**:273.
37. **Chaisson, R. E., P. Bacchetti, D. Osmond, B. Brodie, M. A. Sande, and A. R. Moss.** 1989. Cocaine use and HIV infection in intravenous drug users in San Francisco. *JAMA* **261**:561–565.
38. **Chao, C. C., S. Hu, T. W. Molitor, Y. Zhou, M. P. Murtaugh, M. Tsang, and P. K. Peterson.** 1992. Morphine potentiates transforming growth factor-beta release from human peripheral blood mononuclear cell cultures. *J. Pharmacol. Exp. Ther.* **262**:19–24.
39. **Chao, C. C., T. W. Molitor, K. Close, S. Hu, and P. K. Peterson.** 1993. Morphine inhibits the release of tumor necrosis factor in human peripheral blood mononuclear cell cultures. *Int. J. Immunopharmacol.* **15**:447–453.
40. **Chao, C. C., B. M. Sharp, C. Pomeroy, G. A. Filice, and P. K. Peterson.** 1990. Lethality of morphine in mice infected with *Toxoplasma gondii*. *J. Pharmacol. Exp. Ther.* **252**:605–609.
41. **Chen, G. J., D. S. Huang, B. Watzl, and R. R. Watson.** 1993. Ethanol modulation of tumor necrosis factor and gamma interferon production by murine splenocytes and macrophages. *Life Sci.* **52**:1319–1326.
42. **Chiasson, M. A., R. L. Stoneburner, D. S. Hildebrandt, W. E. Ewing, E. E. Telzak, and H. W. Jaffe.** 1991. Heterosexual transmission of HIV-1 associated with the use of smokable freebase cocaine (crack). *Aids* **5**:1121–1126.
43. **Chomet, B., and B. M. Gach.** 1967. Lobar pneumonia and alcoholism: an analysis of thirty-seven cases. *Am. J. Med. Sci.* **253**:300–304.
44. **Cook, R. L., N. K. Pollock, A. K. Rao, and D. B. Clark.** 2002. Increased prevalence of herpes simplex virus type 2 among adolescent women with alcohol use disorders. *J. Adolesc. Health* **30**:169–174.
45. **Cook, R. T.** 1998. Alcohol abuse, alcoholism, and damage to the immune system—a review. *Alcohol Clin. Exp. Res.* **22**:1927–1942.
46. **Daaka, Y., W. Zhu, H. Friedman, and T. W. Klein.** 1997. Induction of IL-2 receptor α gene by Δ^9 -tetrahydrocannabinol is mediated by nuclear factor κ B and CB1 cannabinoid receptor. *DNA Cell Biol.* **16**:301–309.
47. **Dancer, S. J., D. McNair, P. Finn, and A. B. Kolsto.** 2002. *Bacillus cereus* cellulitis from contaminated heroin. *J. Med. Microbiol.* **51**:278–281.
48. **Darban, H., R. R. Watson, J. Alak, and N. Thomas.** 1993. Cocaine facilitation of cryptosporidiosis by murine AIDS in male and female C57/BL/6 mice. *Adv. Exp. Med. Biol.* **335**:143–151.
49. **Delafuente, J. C., and C. L. DeVane.** 1991. Immunologic effects of cocaine and related alkaloids. *Immunopharmacol. Immunotoxicol.* **13**:11–23.
50. **Derkinderen, P., C. Ledent, M. Parmentier, and J. A. Girault.** 2001. Cannabinoids activate p38 mitogen-activated protein kinases through CB1 receptors in hippocampus. *J. Neurochem.* **77**:957–960.
51. **Derocq, J., M. Segui, J. Marchand, G. LeFur, and P. Casellas.** 1995. Cannabinoids enhance human B-cell growth at low nanomolar concentrations. *FEBS Lett.* **369**:177–182.
52. **Derocq, J. M., O. Jbilo, M. Bouaboula, M. Segui, C. Clere, and P. Casellas.** 2000. Genomic and functional changes induced by the activation of the peripheral cannabinoid receptor CB2 in the promyelocytic cells HL-60. Possible involvement of the CB2 receptor in cell differentiation. *J. Biol. Chem.* **275**:15621–15628.
53. **Devane, W. A., L. Hanus, A. Breuer, R. G. Pertwee, L. A. Stevenson, G. Griffin, D. Gibson, A. Mandelbaum, A. Etinger, and R. Mechoulam.** 1992. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science* **258**:1946–1949.
54. **Dingle, G. A., and T. P. Oei.** 1997. Is alcohol a cofactor of HIV and AIDS? Evidence from immunological and behavioral studies. *Psychol. Bull.* **122**:56–71.
55. **Djeu, J. Y., M. Wang, and H. Friedman.** 1991. Adverse effect of Δ^9 -tetrahydrocannabinol on human neutrophil function. *Adv. Exp. Med. Biol.* **288**:57–62.
56. **Doherty, M. C., R. S. Garfein, E. Monterroso, D. Brown, and D. Vlahov.** 2000. Correlates of HIV infection among young adult short-term injection drug users. *Aids* **14**:717–726.
57. **Donahoe, R. M.** 1990. Drug abuse and AIDS: causes for the connection. *NIDA Res. Monogr.* **96**:181–191.
58. **Donahoe, R. M., and A. Falek.** 1988. Neuroimmunomodulation by opiates and other drugs of abuse: relationship to HIV infection and AIDS. *Adv. Biochem. Psychopharmacol.* **44**:145–158.
59. **Eisenstein, T. K., M. E. Hilburger, and D. M. P. Lawrence.** 1996. Immunomodulation by morphine and other opioids, p. 103–120. *In* H. Friedman, T. W. Klein, and S. Specter (ed.), *Drugs of abuse, immunity, and infections*. CRC Press, Inc., Boca Raton, Fla.
60. **Eisenstein, T. K., J. J. Meissler, Jr., T. J. Rogers, E. B. Geller, and M. W. Adler.** 1995. Mouse strain differences in immunosuppression by opioids *in vitro*. *J. Pharmacol. Exp. Ther.* **275**:1484–1489.
61. **Ewald, S. J., and H. Shao.** 1993. Ethanol increases apoptotic cell death of thymocytes *in vitro*. *Alcohol Clin. Exp. Res.* **17**:359–365.
62. **Faubert, B. L., and N. E. Kaminski.** 2000. AP-1 activity is negatively regulated by cannabinol through inhibition of its protein components, c-fos and c-jun. *J. Leukoc. Biol.* **67**:259–266.
63. **Friedman, H.** 1996. Drugs of abuse as possible co-factors in AIDS progression: summary of panel discussion. *Adv. Exp. Med. Biol.* **402**:225–228.
64. **Fuchs, B. A., and S. B. Pruett.** 1993. Morphine induces apoptosis in murine thymocytes *in vivo* but not *in vitro*: involvement of both opiate and glucocorticoid receptors. *J. Pharmacol. Exp. Ther.* **266**:417–423.

65. Gao, B. 2002. Interaction of alcohol and hepatitis viral proteins. implication in synergistic effect of alcohol drinking and viral hepatitis on liver injury. *Alcohol* **27**:69–72.
66. Geng, Y., S. M. Savage, L. J. Johnson, J. Seagrave, and M. L. Sopori. 1995. Effects of nicotine on the immune response. I. Chronic exposure to nicotine impairs antigen receptor-mediated signal transduction in lymphocytes. *Toxicol. Appl. Pharmacol.* **135**:268–278.
67. Geng, Y., S. M. Savage, S. Razani-Boroujerdi, and M. L. Sopori. 1996. Effects of nicotine on the immune response. II. Chronic nicotine treatment induces T cell anergy. *J. Immunol.* **156**:2384–2390.
68. Girouard, L., P. Mandrekar, D. Catalano, and G. Szabo. 1998. Regulation of monocyte interleukin-12 production by acute alcohol: a role for inhibition by interleukin-10. *Alcohol Clin. Exp. Res.* **22**:211–216.
69. Goedert, J. J. 1984. Recreational drugs: relationship to AIDS. *Ann. N. Y. Acad. Sci.* **437**:192–199.
70. Greenberg, S., J. Xie, J. Kolls, S. Nelson, P. Didier, and C. Mason. 1995. Ethanol suppresses *Mycobacterium tuberculosis*-induced mRNA for nitric oxide synthase in alveolar macrophages, *in vivo*. *Alcohol Clin. Exp. Res.* **19**:394–401.
71. Grimm, M. C., A. Ben-Baruch, D. D. Taub, O. M. Howard, J. H. Resau, J. M. Wang, H. Ali, R. Richardson, R. Snyderman, and J. J. Oppenheim. 1998. Opiates transactivate chemokine receptors: delta and mu opiate receptor-mediated heterologous desensitization. *J. Exp. Med.* **188**:317–325.
72. Guan, L., R. Townsend, T. K. Eisenstein, M. W. Adler, and T. J. Rogers. 1994. Both T cells and macrophages are targets of kappa-opioid-induced immunosuppression. *Brain Behav. Immun.* **8**:229–240.
73. Gurwitz, D., and Y. Kloog. 1998. Do endogenous cannabinoids contribute to HIV-mediated immune failure? *Mol. Med. Today* **4**:196–200.
74. Hakki, A., N. Hallquist, H. Friedman, and S. Pross. 2000. Differential impact of nicotine on cellular proliferation and cytokine production by LPS-stimulated murine splenocytes. *Int. J. Immunopharmacol.* **22**:403–410.
75. Hallquist, N., A. Hakki, L. Wecker, H. Friedman, and S. Pross. 2000. Differential effects of nicotine and aging on splenocyte proliferation and the production of Th1- versus Th2-type cytokines. *Proc. Soc. Exp. Biol. Med.* **224**:141–146.
76. Haverkos, H. W. 1988. Epidemiologic studies—Kaposi's sarcoma vs. opportunistic infections among homosexual men with AIDS. *NIDA Res. Monogr.* **83**:96–105.
77. Haverkos, H. W., and J. W. Curran. 1982. The current outbreak of Kaposi's sarcoma and opportunistic infections. *CA Cancer J. Clin.* **32**:330–339.
78. Hernandez, M. C., L. R. Flores, and B. M. Bayer. 1993. Immunosuppression by morphine is mediated by central pathways. *J. Pharmacol. Exp. Ther.* **267**:1336–1341.
79. Herring, A. C., and N. E. Kaminski. 1999. Cannabinol-mediated inhibition of nuclear factor-kappaB, cAMP response element-binding protein, and interleukin-2 secretion by activated thymocytes. *J. Pharmacol. Exp. Ther.* **291**:1156–1163.
80. Hiemke, C., M. Stolp, S. Reuss, A. Wevers, S. Reinhardt, A. Maelicke, S. Schlegel, and H. Schroder. 1996. Expression of alpha subunit genes of nicotinic acetylcholine receptors in human lymphocytes. *Neurosci. Lett.* **214**:171–174.
81. Hilburger, M. E., M. W. Adler, A. L. Truant, J. J. Meissler, Jr., V. Satishchandran, T. J. Rogers, and T. K. Eisenstein. 1997. Morphine induces sepsis in mice. *J. Infect. Dis.* **176**:183–188.
82. Hoffman, K. E., K. A. Maslonek, L. A. Dykstra, and D. T. Lysle. 1995. Effects of central administration of morphine on immune status in Lewis and Wistar rats. *Adv. Exp. Med. Biol.* **373**:155–159.
83. Houghtling, R. A., and B. M. Bayer. 2002. Rapid elevation of plasma interleukin-6 by morphine is dependent on autonomic stimulation of adrenal gland. *J. Pharmacol. Exp. Ther.* **300**:213–219.
84. Houghtling, R. A., R. D. Mellon, R. J. Tan, and B. M. Bayer. 2000. Acute effects of morphine on blood lymphocyte proliferation and plasma IL-6 levels. *Ann. N. Y. Acad. Sci.* **917**:771–777.
85. Howlett, A. C. 1995. Pharmacology of cannabinoid receptors. *Annu. Rev. Pharmacol. Toxicol.* **35**:607–634.
86. Howlett, A. C., F. Barth, T. I. Bonner, G. Cabral, P. Casellas, W. A. Devane, C. C. Felder, M. Herkenham, K. Mackie, B. R. Martin, R. Mechoulam, and R. G. Pertwee. 2002. International Union of Pharmacology. XXVII. Classification of cannabinoid receptors. *Pharmacol. Rev.* **54**:161–202.
87. Huber, G. L., V. E. Pochay, W. Pereira, J. W. Shea, W. C. Hinds, M. W. First, and G. C. Sornberger. 1980. Marijuana, tetrahydrocannabinol, and pulmonary antibacterial defenses. *Chest* **77**:403–410.
88. Hudolin, V. 1975. Tuberculosis and alcoholism. *Ann. N. Y. Acad. Sci.* **252**:353–364.
89. Hussey, H. H., and S. Katz. 1950. Infections resulting from narcotic addiction. *Am. J. Med.* **9**:186.
90. Hussey, H. H., T. F. Keliber, B. B. Schaefer, and B. J. Walsh. 1944. Septicemia and bacterial endocarditis resulting from heroin addiction. *JAMA* **126**:535.
91. Jaffe, J. H., and W. R. Martin. 1990. Opioid analgesics and antagonists. Pergamon Press, Elmsford, N.Y.
92. Jerrells, T. R. 2002. Association of alcohol consumption and exaggerated immunopathologic effects in the liver induced by infectious organism. *Front. Biosci.* **7**:d1487–d1493.
93. Jerrells, T. R. 2002. Role of activated CD8(+) T cells in the initiation and continuation of hepatic damage. *Alcohol* **27**:47–52.
94. Jerrells, T. R., A. J. Saad, and R. Domiati-Saad. 1992. Effects of ethanol on parameters of cellular immunity and host defense mechanisms to infectious agents. *Alcohol* **9**:459–463.
95. Jerrells, T. R., and D. Sibley. 1995. Effects of ethanol on cellular immunity to facultative intracellular bacteria. *Alcohol Clin. Exp. Res.* **19**:11–16.
96. Jerrells, T. R., D. A. Sibley, I. I. Slukvin, and K. A. Mitchell. 1998. Effects of ethanol consumption on mucosal and systemic T-cell-dependent immune responses to pathogenic microorganisms. *Alcohol Clin. Exp. Res.* **22**:212S–215S.
97. Jerrells, T. R., I. Slukvin, D. Sibley, and J. Fuseler. 1994. Increased susceptibility of experimental animals to infectious organisms as a consequence of ethanol consumption. *Alcohol Suppl.* **2**:425–430.
98. Jessop, J. J., and M. S. Taplits. 1991. Effect of high doses of morphine on Con-A induced lymphokine production *in vitro*. *Immunopharmacology* **22**:175–184.
99. Joy, J. E., S. J. Watson, and J. A. Benson. 1999. Marijuana and medicine: assessing the science base. National Academy Press, Washington, D.C.
100. Juel-Jensen, B. E. 1972. Cannabis and recurrent herpes simplex. *Br. Med. J.* **4**:296.
101. Kalra, R., S. P. Singh, S. M. Savage, G. L. Finch, and M. L. Sopori. 2000. Effects of cigarette smoke on immune response: chronic exposure to cigarette smoke impairs antigen-mediated signaling in T cells and depletes IP3-sensitive Ca(2+) stores. *J. Pharmacol. Exp. Ther.* **293**:166–171.
102. Kaminski, N., W. S. Koh, K. H. Yang, M. Lee, and F. K. Kessler. 1994. Suppression of the humoral immune response by cannabinoids is partially mediated through inhibition of adenylate cyclase by a pertussis toxin-sensitive G-protein coupled mechanism. *Biochem. Pharmacol.* **48**:1899–1908.
103. Karch, S. R. (ed.). 1993. CRC Press, Inc., Boca Raton, Fla.
104. Kawakami, Y., T. W. Klein, C. A. Newton, C. A. McCarthy, J. Djeu, G. Denner, S. Specter, and H. Friedman. 1988. Suppression by cannabinoids of a cloned cell line with natural killer cell activity. *Proc. Soc. Exp. Biol. Med.* **187**:355–359.
105. Kee, T. H. 1908. The habitual use of opium as a factor in the production of diseases. *Philipp. J. Sci.* **6**:63.
106. Klein, T., H. Friedman, and S. Specter. 1998. Marijuana, immunity and infection. *J. Neuroimmunol.* **83**:102–115.
107. Klein, T. W., and H. Friedman. 1990. Modulation of murine immune cell function by marijuana components, p. 87–111. *In* R. Watson (ed.), *Drugs of abuse and immune function*. CRC Press, Inc., Boca Raton, Fla.
108. Klein, T. W., B. Lane, C. A. Newton, and H. Friedman. 2000. The cannabinoid system and cytokine network. *Proc. Soc. Exp. Biol. Med.* **225**:1–8.
109. Klein, T. W., K. Matsui, C. A. Newton, J. Young, R. E. Widen, and H. Friedman. 1993. Cocaine suppresses proliferation of phytohemagglutinin-activated human peripheral blood T-cells. *Int. J. Immunopharmacol.* **15**:77–86.
110. Klein, T. W., C. Newton, and H. Friedman. 1987. Inhibition of natural killer cell function by marijuana components. *J. Toxicol. Environ. Health* **20**:321–332.
111. Klein, T. W., C. Newton, R. Widen, and H. Friedman. 1993. Δ^9 -Tetrahydrocannabinol injection induces cytokine-mediated mortality of mice infected with *Legionella pneumophila*. *J. Pharmacol. Exp. Ther.* **267**:635–640.
112. Klein, T. W., C. A. Newton, N. Nakachi, and H. Friedman. 2000. Δ^9 -tetrahydrocannabinol treatment suppresses immunity and early IFN- γ , IL-12, and IL-12 receptor β 2 responses to *Legionella pneumophila* infection. *J. Immunol.* **164**:6461–6466.
113. Klein, T. W., C. A. Newton, R. Widen, and H. Friedman. 1985. The effect of delta-9-tetrahydrocannabinol and 11-hydroxy-delta-9-tetrahydrocannabinol on T lymphocyte and B lymphocyte mitogen responses. *J. Immunopharmacol.* **7**:451–466.
114. Kolls, J. K., J. Xie, D. Lei, S. Greenberg, W. R. Summer, and S. Nelson. 1995. Differential effects of *in vivo* ethanol on LPS-induced TNF and nitric oxide production in the lung. *Am. J. Physiol.* **268**:L991–L998.
115. Kraft, A., and N. M. Leitch. 1921. The action of drugs in infection, the influence of morphine in experimental septicemia. *J. Pharmacol. Exp. Ther.* **17**:377.
116. Kruegar, H., N. B. Eddy, and M. Sumwalt. 1941. The pharmacology of opium alkaloids. *Public Health Rep.* **165**(Suppl.):415.
117. Lange, W. R., J. C. Ball, M. B. Pfeiffer, F. R. Snyder, and E. J. Cone. 1989. The Lexington addicts, 1971–1972: demographic characteristics, drug use patterns, and selected infectious disease experience. *Int. J. Addict.* **24**:609–626.
118. Lebargy, F., K. Benhammou, D. Morin, R. Zini, S. Urien, F. Bree, J. Bignon, A. Branellec, and G. Lagrue. 1996. Tobacco smoking induces expression of very-high-affinity nicotine binding sites on blood polymorphonuclear cells. *Am. J. Respir. Crit. Care Med.* **153**:1056–1063.
119. Lee, S. F., C. Newton, R. Widen, H. Friedman, and T. W. Klein. 2001. Differential expression of cannabinoid CB2 receptor mRNA in mouse im-

- mune cell subpopulations and following B cell stimulation. *Eur. J. Pharmacol.* **423**:235–241.
120. **Lefkowitz, S. S., and C. Y. Chiang.** 1975. Effects of certain abused drugs on hemolysin forming cells. *Life Sci.* **17**:1763–1767.
 121. **Liu, Y., D. J. Blackburn, L. F. Chuang, K. F. Killam, Jr., and R. Y. Chuang.** 1992. Effects of *in vivo* and *in vitro* administration of morphine sulfate upon rhesus macaque polymorphonuclear cell phagocytosis and chemotaxis. *J. Pharmacol. Exp. Ther.* **263**:533–539.
 122. **Liu, Y., B. B. Whitlock, J. A. Pultz, and S. A. Wolfe, Jr.** 1995. Sigma-1 receptors modulate functional activity of rat splenocytes. *J. Neuroimmunol.* **59**:143–154.
 123. **Louria, D. B., T. Hensle, and J. Rose.** 1967. The major medical complications of heroin addiction. *Ann. Intern. Med.* **67**:1–22.
 124. **Lyman, W. D.** 1993. Perinatal AIDS: drugs of abuse and transplacental infection. *Adv. Exp. Med. Biol.* **335**:211–217.
 125. **Lysle, D. T., M. E. Coussons, V. J. Watts, E. H. Bennett, and L. A. Dykstra.** 1993. Morphine-induced alterations of immune status: dose dependency, compartment specificity and antagonism by naltrexone. *J. Pharmacol. Exp. Ther.* **265**:1071–1078.
 126. **Lysle, D. T., K. E. Hoffman, and L. A. Dykstra.** 1996. Evidence for the involvement of the caudal region of the periaqueductal gray in a subset of morphine-induced alterations of immune status. *J. Pharmacol. Exp. Ther.* **277**:1533–1540.
 127. **MacFarlane, A. S., X. Peng, J. J. Meissler, Jr., T. J. Rogers, E. B. Geller, M. W. Adler, and T. K. Eisenstein.** 2000. Morphine increases susceptibility to oral *Salmonella typhimurium* infection. *J. Infect. Dis.* **181**:1350–1358.
 128. **MacGregor, R. R., and D. B. Louria.** 1997. Alcohol and infection. *Curr. Clin. Top. Infect. Dis.* **17**:291–315.
 129. **Mandrekar, P., D. Catalano, and G. Szabo.** 1999. Inhibition of lipopolysaccharide-mediated NFκappaB activation by ethanol in human monocytes. *Int. Immunol.* **11**:1781–1790.
 130. **Mansell, P. W.** 1984. Acquired immune deficiency syndrome, leading to opportunistic infections, Kaposi's sarcoma, and other malignancies. *Crit. Rev. Clin. Lab. Sci.* **20**:191–204.
 131. **Mao, J. T., M. Huang, J. Wang, S. Sharma, D. P. Tashkin, and S. M. Dubinett.** 1996. Cocaine down-regulates IL-2-induced peripheral blood lymphocyte IL-8 and IFN-γ production. *Cell. Immunol.* **172**:217–223.
 132. **Mao, J. T., L. X. Zhu, S. Sharma, K. Chen, M. Huang, S. J. Santiago, J. Gulsurd, D. P. Tashkin, and S. M. Dubinett.** 1997. Cocaine inhibits human endothelial cell IL-8 production: the role of transforming growth factor-beta. *Cell. Immunol.* **181**:38–43.
 133. **Massi, P., D. Fuzio, D. Viganò, P. Sacerdote, and D. Parolaro.** 2000. Relative involvement of cannabinoid CB(1) and CB(2) receptors in the Delta (9)-tetrahydrocannabinol-induced inhibition of natural killer activity. *Eur. J. Pharmacol.* **387**:343–347.
 134. **Matsuda, L. A., S. J. Lolait, M. J. Brownstein, A. C. Young, and T. I. Bonner.** 1990. Structure of cannabinoid receptor and functional expression of the cloned cDNA. *Nature* **346**:561–564.
 135. **Matsumoto, R. R., K. A. McCracken, B. Pouw, Y. Zhang, and W. D. Bowen.** 2002. Involvement of sigma receptors in the behavioral effects of cocaine: evidence from novel ligands and antisense oligodeoxynucleotides. *Neuropharmacology* **42**:1043–1055.
 136. **Matsunaga, K., T. W. Klein, H. Friedman, and Y. Yamamoto.** 2001. Involvement of nicotinic acetylcholine receptors in suppression of antimicrobial activity and cytokine responses of alveolar macrophages to *Legionella pneumophila* infection by nicotine. *J. Immunol.* **167**:6518–6524.
 137. **Maurice, T., R. Martin-Fardon, P. Romieu, and R. R. Matsumoto.** 2002. Sigma(1) (sigma(1)) receptor antagonists represent a new strategy against cocaine addiction and toxicity. *Neurosci. Biobehav. Rev.* **26**:499–527.
 138. **McAllister, S. D., and M. Glass.** 2002. CB(1) and CB(2) receptor-mediated signalling: a focus on endocannabinoids. Prostaglandins Leukotrienes Essent. Fatty Acids **66**:161–171.
 139. **McCarthy, L., M. Wetzel, J. K. Sliker, T. K. Eisenstein, and T. J. Rogers.** 2001. Opioids, opioid receptors, and the immune response. *Drug Alcohol Depend.* **62**:111–123.
 140. **McKallip, R. J., C. Lombard, B. R. Martin, M. Nagarkatti, and P. S. Nagarkatti.** 2002. Delta(9)-tetrahydrocannabinol-induced apoptosis in the thymus and spleen as a mechanism of immunosuppression *in vitro* and *in vivo*. *J. Pharmacol. Exp. Ther.* **302**:451–465.
 141. **Meadows, G. G., S. E. Blank, and D. D. Duncan.** 1989. Influence of ethanol consumption on natural killer cell activity in mice. *Alcohol Clin. Exp. Res.* **13**:476–479.
 142. **Mechoulam, R., S. Ben-Shabat, L. Hanus, M. Ligumsky, N. E. Kaminski, A. R. Schatz, A. Gopher, S. Almog, B. R. Martin, D. R. Compton, R. G. Pertwee, G. Griffin, M. Bayewitch, J. Barg, and Z. Vogel.** 1995. Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. *Biochem. Pharm.* **50**:83–90.
 143. **Mechoulam, R., Z. Ben-Zvi, B. Yagnitinsky, and A. Shani.** 1969. A new tetrahydrocannabinolic acid. *Tetrahedron. Lett.* **28**:2339–2341.
 144. **Mellon, R. D., and B. M. Bayer.** 1998. Evidence for central opioid receptors in the immunomodulatory effects of morphine: review of potential mechanism(s) of action. *J. Neuroimmunol.* **83**:19–28.
 145. **Mendenhall, C. L., C. J. Grossman, G. A. Roselle, S. Ghosh, P. S. Gartside, S. D. Rouster, P. V. Chalasani, G. Schmitt, K. Martin, and K. Lamping.** 1990. Host response to mycobacterial infection in the alcoholic rat. *Gastroenterology* **99**:1723–1726.
 146. **Meyerhoff, D. J.** 2001. Effects of alcohol and HIV infection on the central nervous system. *Alcohol Res. Health* **25**:288–298.
 147. **Mishkin, E. M., and G. A. Cabral.** 1985. Delta-9-tetrahydrocannabinol decreases host resistance to herpes simplex virus type 2 vaginal infection in the B6C3F1 mouse. *J. Gen. Virol.* **66**:2539–2549.
 148. **Molitor, T. W., A. Morilla, J. M. Risdahl, M. P. Murtaugh, C. C. Chao, and P. K. Peterson.** 1992. Chronic morphine administration impairs cell-mediated immune responses in swine. *J. Pharmacol. Exp. Ther.* **260**:581–586.
 149. **Morahan, P. S., P. C. Klykken, S. H. Smith, L. S. Harris, and A. E. Munson.** 1979. Effects of cannabinoids on host resistance to *Listeria monocytogenes* and herpes simplex virus. *Infect. Immun.* **23**:670–674.
 150. **Moskowitz, L. B., P. Kory, J. C. Chan, H. W. Haverkos, F. K. Conley, and G. T. Hensley.** 1983. Unusual causes of death in Haitians residing in Miami. High prevalence of opportunistic infections. *JAMA* **250**:1187–1191.
 151. **Munro, S., K. L. Thomas, and M. Abu-Shaar.** 1993. Molecular characterization of a peripheral receptor for cannabinoids. *Nature* **365**:61–65.
 152. **Mutchnick, M. G., and H. H. Lee.** 1988. Impaired lymphocyte proliferative response to mitogen in alcoholic patients. Absence of a relation to liver disease activity. *Alcohol Clin. Exp. Res.* **12**:155–158.
 153. **Nahas, G. G., A. Morishima, and B. Desoize.** 1977. Effects of cannabinoids on macromolecular synthesis and replication of cultured lymphocytes. *Fed. Proc.* **36**:1748–1752.
 154. **Nahas, G. G., N. Suci-Foca, J.-P. Armand, and A. Morishima.** 1974. Inhibition of cellular mediated immunity in marijuana smokers. *Science* **183**:419–420.
 155. **Nahas, G. G., D. Zagury, and I. W. Schwartz.** 1973. Evidence for the possible immunogenicity of Δ⁹-tetrahydrocannabinol (THC) in rodents. *Nature* **243**:407–408.
 156. **Nair, M. P., Z. A. Kronfol, and S. A. Schwartz.** 1990. Effects of alcohol and nicotine on cytotoxic functions of human lymphocytes. *Clin. Immunol. Immunopathol.* **54**:395–409.
 157. **Nalpas, B., S. Pol, V. Thepot, H. Zylberberg, P. Berthelot, and C. Brechot.** 1998. ESBRA 1997 Award lecture: relationship between excessive alcohol drinking and viral infections. *Alcohol* **33**:202–206.
 158. **Nelson, S., G. Bagby, and W. R. Summer.** 1989. Alcohol suppresses lipopolysaccharide-induced tumor necrosis factor activity in serum and lung. *Life Sci.* **44**:673–676.
 159. **Nelson, S., and J. K. Kolls.** 2002. Alcohol, host defense and society. *Nat. Rev. Immunol.* **2**:205–209.
 160. **Nelson, S., C. Mason, G. Bagby, and W. Summer.** 1995. Alcohol, tumor necrosis factor, and tuberculosis. *Alcohol Clin. Exp. Res.* **19**:17–24.
 161. **Newell, G. R., P. W. Mansell, M. R. Spitz, J. M. Reuben, and E. M. Hersh.** 1985. Volatile nitrites. Use and adverse effects related to the current epidemic of the acquired immune deficiency syndrome. *Am. J. Med.* **78**:811–816.
 162. **Newton, C., T. Klein, and H. Friedman.** 1998. The role of macrophages in THC-induced alteration of the cytokine network. *Adv. Exp. Med. Biol.* **437**:207–214.
 163. **Newton, C. A., T. W. Klein, and H. Friedman.** 1994. Secondary immunity to *Legionella pneumophila* and Th1 activity are suppressed by delta-9-tetrahydrocannabinol injection. *Infect. Immun.* **62**:4015–4020.
 164. **Nieman, R. B., J. Fleming, R. J. Coker, J. R. Harris, and D. M. Mitchell.** 1993. The effect of cigarette smoking on the development of AIDS in HIV-1-seropositive individuals. *Aids* **7**:705–710.
 165. **Osler, W.** 1880. Oedema of the left lung in morphia poisoning. *Montreal Gen. Hosp. Rep.* **1**:291.
 166. **Ou, D. W., M. L. Shen, and Y. D. Luo.** 1989. Effects of cocaine on the immune system of Balb/C mice. *Clin. Immunol. Immunopathol.* **52**:305–312.
 167. **Ouyang, Y., N. Virasch, P. Hao, M. T. Aubrey, N. Mukerjee, B. E. Bierer, and B. M. Freed.** 2000. Suppression of human IL-1β, IL-2, IFN-γ, and TNF-α production by cigarette smoke extracts. *J. Allergy Clin. Immunol.* **106**:280–287.
 168. **Pacifici, R., S. Di Carlo, A. Bacosi, and P. Zuccaro.** 1993. Macrophage functions in drugs of abuse-treated mice. *Int. J. Immunopharmacol.* **15**:711–716.
 169. **Panaslak, W., S. Gumulka, M. Kobus, and M. Luczak.** 1990. The influence of morphine on development of HSV-1 and M-MSV virus infection in mice. *Acta Microbiol. Pol.* **39**:215–218.
 170. **Paradise, L. J., and H. Friedman.** 1993. Syphilis and drugs of abuse. *Adv. Exp. Med. Biol.* **335**:81–87.
 171. **Pellegrino, T. C., K. L. Dunn, and B. M. Bayer.** 2001. Mechanisms of cocaine-induced decreases in immune cell function. *Int. Immunopharmacol.* **1**:665–675.
 172. **Peng, X., J. J. Cebra, M. W. Adler, J. J. Meissler, Jr., A. Cowan, P. Feng, and T. K. Eisenstein.** 2001. Morphine inhibits mucosal antibody responses and TGF-beta mRNA in gut-associated lymphoid tissue following oral cholera toxin in mice. *J. Immunol.* **167**:3677–3681.

173. Peng, X., D. M. Mosser, M. W. Adler, T. J. Rogers, J. J. Meissler, Jr., and T. K. Eisenstein. 2000. Morphine enhances interleukin-12 and the production of other pro-inflammatory cytokines in mouse peritoneal macrophages. *J. Leukoc. Biol.* **68**:723–728.
174. Pertwee, R. G. 1997. Pharmacology of cannabinoid CB1 and CB2 receptors. *Pharmacol. Ther.* **74**:129–180.
175. Peterson, J. D., K. Vasquez, and C. Waltenbaugh. 1998. Interleukin-12 therapy restores cell-mediated immunity in ethanol-consuming mice. *Alcohol Clin. Exp. Res.* **22**:245–251.
176. Peterson, P. K., G. Gekker, C. Brummitt, P. Pentel, M. Bullock, M. Simpson, J. Hitt, and B. Sharp. 1989. Suppression of human peripheral blood mononuclear cell function by methadone and morphine. *J. Infect. Dis.* **159**:480–487.
177. Peterson, P. K., G. Gekker, C. C. Chao, R. Schut, T. W. Molitor, and H. H. Balfour, Jr. 1991. Cocaine potentiates HIV-1 replication in human peripheral blood mononuclear cell cocultures. Involvement of transforming growth factor-beta. *J. Immunol.* **146**:81–84.
178. Peterson, P. K., G. Gekker, S. Hu, W. S. Sheng, T. W. Molitor, and C. C. Chao. 1995. Morphine stimulates phagocytosis of *Mycobacterium tuberculosis* by human microglial cells: involvement of a G protein-coupled opiate receptor. *Adv. Neuroimmunol.* **5**:299–309.
179. Peterson, P. K., B. Sharp, G. Gekker, C. Brummitt, and W. F. Keane. 1987. Opioid-mediated suppression of interferon-gamma production by cultured peripheral blood mononuclear cells. *J. Clin. Investig.* **80**:824–831.
180. Pontieri, F. E., G. Tanda, F. Orzi, and G. Di Chiara. 1996. Effects of nicotine on the nucleus accumbens and similarity to those of addictive drugs. *Nature* **382**:255–257.
181. Pross, S. H., T. W. Klein, C. A. Newton, J. Smith, R. Widen, and H. Friedman. 1990. Differential suppression of T-cell subpopulations by THC (delta-9-tetrahydrocannabinol). *Int. J. Immunopharmacol.* **12**:539–544.
182. Pross, S. H., Y. Nakano, R. Widen, S. McHugh, C. A. Newton, T. W. Klein, and H. Friedman. 1992. Differing effects of delta-9-tetrahydrocannabinol (THC) on murine spleen cell populations dependent upon stimulators. *Int. J. Immunopharmacol.* **14**:1019–1027.
183. Pruett, S. B., Y. C. Han, and B. A. Fuchs. 1992. Morphine suppresses primary humoral immune responses by a predominantly indirect mechanism. *J. Pharmacol. Exp. Ther.* **262**:923–928.
184. Rahim, R. T., J. J. Meissler, Jr., A. Cowan, T. J. Rogers, E. B. Geller, J. Gaughan, M. W. Adler, and T. K. Eisenstein. 2001. Administration of mu-, kappa- or delta2-receptor agonists via osmotic mini pumps suppresses murine splenic antibody responses. *Int. Immunopharmacol.* **1**:2001–2009.
185. Reichman, L. B., C. P. Felton, and J. R. Edsall. 1979. Drug dependence, a possible new risk factor for tuberculosis disease. *Arch. Intern. Med.* **139**:337–339.
186. Reisine, T., and G. I. Bell. 1993. Molecular biology of opioid receptors. *Trends Neurosci.* **16**:506–510.
187. Reynolds, L., and B. C. Cantab. 1910. The influence of narcotics on phagocytosis. *Lancet* **178**:569.
188. Risdahl, J. M., K. V. Khanna, P. K. Peterson, and T. W. Molitor. 1998. Opiates and infection. *J. Neuroimmunol.* **83**:4–18.
189. Risdahl, J. M., P. K. Peterson, C. C. Chao, C. Pijoan, and T. W. Molitor. 1993. Effects of morphine dependence on the pathogenesis of swine herpesvirus infection. *J. Infect. Dis.* **167**:1281–1287.
190. Risdahl, J. M., P. K. Peterson, and T. W. Molitor. 1996. Opiates, infection and immunity, p. 1–42. *In* H. Friedman, T. W. Klein, and S. Specter (ed.), *Drugs of abuse, immunity, and infections*. CRC Press, Inc., Boca Raton, Fla.
191. Rojavin, M., I. Szabo, J. L. Bussiere, T. J. Rogers, M. W. Adler, and T. K. Eisenstein. 1993. Morphine treatment *in vitro* or *in vivo* decreases phagocytic functions of murine macrophages. *Life Sci.* **53**:997–1006.
192. Roth, M. D., D. P. Tashkin, R. Choi, B. D. Jamieson, J. A. Zack, and G. C. Baldwin. 2002. Cocaine enhances human immunodeficiency virus replication in a model of severe combined immunodeficient mice implanted with human peripheral blood leukocytes. *J. Infect. Dis.* **185**:701–705.
193. Roy, S., S. Balasubramanian, S. Sumandep, R. Charboneau, J. Wang, D. Melnyk, G. J. Beilman, R. Vatassery, and R. A. Barke. 2001. Morphine directs T cells toward T(H2) differentiation. *Surgery* **130**:304–309.
194. Roy, S., R. A. Barke, and H. H. Loh. 1998. Mu-opioid receptor-knockout mice: role of mu-opioid receptor in morphine mediated immune functions. *Brain Res. Mol. Brain Res.* **61**:190–194.
195. Roy, S., R. G. Charboneau, and R. A. Barke. 1999. Morphine synergizes with lipopolysaccharide in a chronic endotoxemia model. *J. Neuroimmunol.* **95**:107–114.
196. Ruiz, M., S. Ewig, M. A. Marcos, J. A. Martinez, F. Arancibia, J. Mensa, and A. Torres. 1999. Etiology of community-acquired pneumonia: impact of age, comorbidity, and severity. *Am. J. Respir. Crit. Care Med.* **160**:397–405.
197. Saad, A. J., R. Domiati-Saad, and T. R. Jerrells. 1993. Ethanol ingestion increases susceptibility of mice to *Listeria monocytogenes*. *Alcohol Clin. Exp. Res.* **17**:75–85.
198. Schatz, A. R., W. S. Koh, and N. E. Kaminski. 1993. Δ^9 -Tetrahydrocannabinol selectively inhibits T-cell dependent humoral immune responses through direct inhibition of accessory T-cell function. *Immunopharmacology* **26**:129–137.
199. Scott, J. M. 1969. The white poppy. A history of opium. Cox and Wyman Ltd., London, England.
200. Sei, Y., T. McIntyre, E. Fride, K. Yoshimoto, P. Skolnick, and P. K. Arora. 1991. Inhibition of calcium mobilization is an early event in opiate-induced immunosuppression. *FASEB J.* **5**:2194–2199.
201. Sei, Y., K. Yoshimoto, T. McIntyre, P. Skolnick, and P. K. Arora. 1991. Morphine-induced thymic hypoplasia is glucocorticoid-dependent. *J. Immunol.* **146**:194–198.
202. Sepulveda, R. T., S. Jiang, D. G. Besselsen, and R. R. Watson. 2002. Alcohol consumption during murine acquired immunodeficiency syndrome accentuates heart pathology due to Cocksackievirus. *Alcohol* **37**:157–163.
203. Shahabi, N. A., and B. M. Sharp. 1995. Antiproliferative effects of delta opioids on highly purified CD4+ and CD8+ murine T cells. *J. Pharmacol. Exp. Ther.* **273**:1105–1113.
204. Shahbazian, L. M., H. R. Darban, J. R. Darban, A. M. Stazzone, and R. R. Watson. 1992. Influence of the level of dietary ethanol in mice with murine AIDS on resistance to *Streptococcus pneumoniae*. *Alcohol* **27**:345–352.
205. Sharkey, J., K. A. Glen, S. Wolfe, and M. J. Kuhar. 1988. Cocaine binding at sigma receptors. *Eur. J. Pharmacol.* **149**:171–174.
206. Shavit, Y., A. Depaulis, F. C. Martin, G. W. Terman, R. N. Pechnick, C. J. Zane, R. P. Gale, and J. C. Liebeskind. 1986. Involvement of brain opiate receptors in the immune-suppressive effect of morphine. *Proc. Natl. Acad. Sci. USA* **83**:7114–7117.
207. Shavit, Y., F. C. Martin, R. Yirmiya, S. Ben-Eliyahu, G. W. Terman, H. Weiner, R. P. Gale, and J. C. Liebeskind. 1987. Effects of a single administration of morphine or foot shock stress on natural killer cell cytotoxicity. *Brain Behav. Immun.* **1**:318–328.
208. Sibley, D., and T. R. Jerrells. 2000. Alcohol consumption by C57BL/6 mice is associated with depletion of lymphoid cells from the gut-associated lymphoid tissues and altered resistance to oral infections with *Salmonella typhimurium*. *J. Infect. Dis.* **182**:482–489.
209. Sibley, D. A., J. Fuseler, I. Slukvin, and T. R. Jerrells. 1995. Ethanol-induced depletion of lymphocytes from the mesenteric lymph nodes of C57BL/6 mice is associated with RNA but not DNA degradation. *Alcohol Clin. Exp. Res.* **19**:324–331.
210. Sibley, D. A., N. Osa, C. Kusynski, L. Wilkie, and T. R. Jerrells. 2001. Alcohol consumption is associated with alterations in macrophage responses to interferon- γ and infection by *Salmonella typhimurium*. *FEMS Immunol. Med. Microbiol.* **32**:73–83.
211. Sidney, S., J. E. Beck, I. S. Tekawa, C. P. Quesenberry, and G. D. Friedmen. 1997. Marijuana use and mortality. *Am. J. Public Health* **87**:585–590.
212. Siegel, L. 1986. AIDS: relationship to alcohol and other drugs. *J. Subst. Abuse Treat.* **3**:271–274.
213. Slavinsky, J., III, T. Myers, R. K. Swoboda, J. E. Leigh, S. Hager, and P. L. Fidel, Jr. 2002. Th1/Th2 cytokine profiles in saliva of HIV-positive smokers with oropharyngeal candidiasis. *Oral Microbiol. Immunol.* **17**:38–43.
214. Smith, M. S., Y. Yamamoto, C. A. Newton, H. Friedman, and T. W. Klein. 1997. Psychoactive cannabinoids increase mortality and alter acute phase cytokine responses in mice sublethally infected with *Legionella pneumophila*. *Proc. Soc. Exp. Biol. Med.* **214**:69–75.
215. Smith, S. R., C. Terminelli, and G. Denhardt. 2000. Effects of cannabinoid receptor agonist and antagonist ligands on production of inflammatory cytokines and anti-inflammatory interleukin-10 in endotoxemic mice. *J. Pharmacol. Exp. Ther.* **293**:136–150.
216. Sopori, M. 2002. Effects of cigarette smoke on the immune system. *Nat. Rev. Immunol.* **2**:372–377.
217. Sopori, M. L., and W. Kozak. 1998. Immunomodulatory effects of cigarette smoke. *J. Neuroimmunol.* **83**:148–156.
218. Sopori, M. L., W. Kozak, S. M. Savage, Y. Geng, D. Soszynski, M. J. Kluger, E. K. Perryman, and G. E. Snow. 1998. Effect of nicotine on the immune system: possible regulation of immune responses by central and peripheral mechanisms. *Psychoneuroendocrinology* **23**:189–204.
219. Specter, S., T. W. Klein, C. Newton, M. Mondragon, R. Widen, and H. Friedman. 1986. Marijuana effects on immunity: suppression of human natural killer cell activity by delta-9-tetrahydrocannabinol. *Int. J. Immunopharmacol.* **8**:741–745.
220. Specter, S., and G. Lancz. 1991. Effects of marijuana on human natural killer cell activity. *Adv. Exp. Med. Biol.* **288**:47–56.
221. Specter, S., G. Lancz, and J. Hazelden. 1990. Marijuana and immunity: tetrahydrocannabinol mediated inhibition of lymphocyte blastogenesis. *Int. J. Immunopharmacol.* **12**:261–267.
222. Specter, S., G. Lancz, G. Westrich, and H. Friedman. 1991. Delta-9-tetrahydrocannabinol augments murine retroviral induced immunosuppression and infection. *Int. J. Immunopharmacol.* **13**:411–417.
223. Srivastava, M. D., B. I. S. Srivastava, and B. Brouhard. 1998. Δ^9 tetrahydrocannabinol and cannabidiol alter cytokine production by human immune cells. *Immunopharmacology* **40**:179–185.
224. Starec, M., B. Rouveix, M. Sinet, F. Chau, B. Desforges, J. J. Pocardalo, and P. Lechat. 1991. Immune status and survival of opiate- and cocaine-treated mice infected with Friend virus. *J. Pharmacol. Exp. Ther.* **259**:745–750.

225. **Sternbach, G. L.** 1990. Infections in alcoholic patients. *Emerg. Med. Clin. North Am.* **8**:793–803.
226. **Stoltz, D. A., S. Nelson, J. K. Kolls, P. Zhang, R. P. Bohm, Jr., M. Murphy-Corb, and G. J. Bagby.** 2000. *In vitro* ethanol suppresses alveolar macrophage TNF- α during simian immunodeficiency virus infection. *Am. J. Respir. Crit. Care Med.* **161**:135–140.
227. **Stoneburner, R. L., D. C. Des Jarlais, D. Benezra, L. Gorelkin, J. L. Sotharan, S. R. Friedman, S. Schultz, M. Marmor, D. Mildvan, and R. Maslansky.** 1998. A larger spectrum of severe HIV-1-related disease in intravenous drug users in New York City. *Science* **242**:916–919.
228. **Su, T. P.** 1991. Sigma receptors. Putative links between nervous, endocrine and immune systems. *Eur. J. Biochem.* **200**:633–642.
229. **Swanson, M. A., W. T. Lee, and V. M. Sanders.** 2001. IFN- γ production by Th1 cells generated from naive CD4+ T cells exposed to norepinephrine. *J. Immunol.* **166**:232–240.
230. **Szabo, G.** 1999. Consequences of alcohol consumption on host defence. *Alcohol* **34**:830–841.
231. **Szabo, G., L. Girouard, P. Mandrekar, and D. Catalano.** 1998. Regulation of monocyte IL-12 production: augmentation by lymphocyte contact and acute ethanol treatment, inhibition by elevated intracellular cAMP. *Int. J. Immunopharmacol.* **20**:491–503.
232. **Szabo, G., P. Mandrekar, A. Dolganiuc, D. Catalano, and K. Kodys.** 2001. Reduced alloreactive T-cell activation after alcohol intake is due to impaired monocyte accessory cell function and correlates with elevated IL-10, IL-13, and decreased IFN γ levels. *Alcohol Clin. Exp. Res.* **25**:1766–1772.
233. **Szabo, G., B. Verma, and D. Catalano.** 1993. Selective inhibition of antigen-specific T lymphocyte proliferation by acute ethanol exposure: the role of impaired monocyte antigen presentation capacity and mediator production. *J. Leukoc. Biol.* **54**:534–544.
234. **Szabo, G., B. K. Verma, M. Fogarasi, and D. E. Catalano.** 1992. Induction of transforming growth factor-beta and prostaglandin E2 production by ethanol in human monocytes. *J. Leukoc. Biol.* **52**:602–610.
235. **Szabo, I., M. Rojavin, J. L. Bussiere, T. K. Eisenstein, M. W. Adler, and T. J. Rogers.** 1993. Suppression of peritoneal macrophage phagocytosis of *Candida albicans* by opioids. *J. Pharmacol. Exp. Ther.* **267**:703–706.
236. **Tanda, G., F. E. Pontieri, and G. Di Chiara.** 1997. Cannabinoid and heroin activation of mesolimbic dopamine transmission by a common mu1 opioid receptor mechanism. *Science* **276**:2048–2050.
237. **Taub, D. D., T. K. Eisenstein, E. B. Geller, M. W. Adler, and T. J. Rogers.** 1991. Immunomodulatory activity of mu- and kappa-selective opioid agonists. *Proc. Natl. Acad. Sci. USA* **88**:360–364.
238. **Thomas, P. T., R. V. House, and H. N. Bhargava.** 1995. Direct cellular immunomodulation produced by diacetylmorphine (heroin) or methadone. *Gen. Pharmacol.* **26**:123–130.
239. **Tubaro, E., U. Avico, C. Santiangeli, P. Zuccaro, G. Cavallo, R. Pacifici, C. Croce, and G. Borelli.** 1985. Morphine and methadone impact on human phagocytic physiology. *Int. J. Immunopharmacol.* **7**:865–874.
240. **Tubaro, E., G. Borelli, C. Croce, G. Cavallo, and C. Santiangeli.** 1983. Effect of morphine on resistance to infection. *J. Infect. Dis.* **148**:656–666.
241. **Valjent, E., C. Pages, M. Rogard, M. J. Besson, R. Maldonado, and J. Caboche.** 2001. Delta 9-tetrahydrocannabinol-induced MAPK/ERK and Elk-1 activation *in vivo* depends on dopaminergic transmission. *Eur. J. Neurosci.* **14**:342–352.
242. **Veyries, M. L., M. Sinet, B. Desforges, and B. Rouveix.** 1995. Effects of morphine on the pathogenesis of murine Friend retrovirus infection. *J. Pharmacol. Exp. Ther.* **272**:498–504.
243. **Vlahov, D., J. C. Anthony, D. Celentano, L. Solomon, and N. Chowdhury.** 1991. Trends of HIV-1 risk reduction among initiates into intravenous drug use 1982–1987. *Am. J. Drug Alcohol Abuse* **17**:39–48.
244. **Wagner, F., R. Fink, R. Hart, C. Lersch, H. Dancygier, and M. Classen.** 1992. Ethanol inhibits interferon-gamma secretion by human peripheral lymphocytes. *J. Stud. Alcohol* **53**:277–280.
245. **Waltenbaugh, C., and J. D. Peterson.** 1997. Ethanol impairs the induction of delayed hypersensitivity in C57BL/6 mice. *Alcohol* **14**:149–153.
246. **Waltenbaugh, C., K. Vasquez, and J. D. Peterson.** 1998. Alcohol consumption alters antigen-specific Th1 responses: mechanisms of deficit and repair. *Alcohol Clin. Exp. Res.* **22**:220S–223S.
247. **Wang, J. Y., B. Liang, and R. R. Watson.** 1997. Alcohol consumption alters cytokine release during murine AIDS. *Alcohol* **14**:155–159.
248. **Wang, Y., D. S. Huang, P. T. Giger, and R. R. Watson.** 1993. Ethanol-induced modulation of cytokine production by splenocytes during murine retrovirus infection causing murine AIDS. *Alcohol Clin. Exp. Res.* **17**:1035–1039.
249. **Wang, Y., D. S. Huang, P. T. Giger, and R. R. Watson.** 1994. Influence of chronic dietary ethanol on cytokine production by murine splenocytes and thymocytes. *Alcohol Clin. Exp. Res.* **18**:64–70.
250. **Wang, Y., D. S. Huang, and R. R. Watson.** 1994. *In vivo* and *in vitro* cocaine modulation on production of cytokines in C57BL/6 mice. *Life Sci.* **54**:401–411.
251. **Wang, Y., and R. R. Watson.** 1994. Chronic ethanol consumption before retrovirus infection is a cofactor in the development of immune dysfunction during murine AIDS. *Alcohol Clin. Exp. Res.* **18**:976–981.
252. **Wang, Y., and R. R. Watson.** 1994. Chronic ethanol consumption prior to retrovirus infection alters cytokine production by thymocytes during murine AIDS. *Alcohol* **11**:361–365.
253. **Watson, E. S., J. C. Murphy, H. N. ElSohly, M. A. Elsohly, and C. E. Turner.** 1983. Effects of the administration of coca alkaloids on the primary immune responses of mice: interaction with delta 9-tetrahydrocannabinol and ethanol. *Toxicol. Appl. Pharmacol.* **71**:1–13.
254. **Watzl, B., G. Chen, P. Scuderi, S. Pirozhkov, and R. R. Watson.** 1992. Cocaine-induced suppression of interferon- γ secretion in leukocytes from young and old C57BL/6 mice. *Int. J. Immunopharmacol.* **14**:1125–1231.
255. **Watzl, B., P. Scuderi, and R. R. Watson.** 1991. Influence of marijuana components (THC and CBD) on human mononuclear cell cytokine secretion *in vitro*. *Adv. Exp. Med. Biol.* **288**:63–70.
256. **Weber, R., B. Ledergerber, M. Opravil, W. Siegenthaler, and R. Luthy.** 1990. Progression of HIV infection in misusers of injected drugs who stop injecting or follow a programme of maintenance treatment with methadone. *Br. Med. J.* **301**:1362–1365.
257. **Weber, R. J., L. C. Band, B. deCosta, A. Pert, and K. C. Rice.** 1991. Neural control of immune function: opioids, opioid receptors and immunosuppression. *NIDA Res. Monogr.* **105**:96–102.
258. **Weber, R. J., and A. Pert.** 1989. The periaqueductal gray matter mediates opiate-induced immunosuppression. *Science* **245**:188–190.
259. **Wetzel, M. A., A. D. Steele, T. K. Eisenstein, M. W. Adler, E. E. Henderson, and T. J. Rogers.** 2000. Mu-opioid induction of monocyte chemoattractant protein-1, RANTES, and IFN- γ -inducible protein-10 expression in human peripheral blood mononuclear cells. *J. Immunol.* **165**:6519–6524.
260. **Whipham, T.** 1875. Fatal pleuropneumonia in the case of a man aged 56, addicted to the abuse of morphia, alcohol and bromide of potassium. *Trans. Clin. Soc. Lond.* **8**:108.
261. **Wilson, R. I., and R. A. Nicoll.** 2002. Endocannabinoid signaling in the brain. *Science* **296**:678–682.
262. **Xu, W., T. Flick, J. Mitchel, C. Knowles, and K. Ault.** 1999. Cocaine effects on immunocompetent cells: an observation of *in vitro* cocaine exposure. *Int. J. Immunopharmacol.* **21**:463–472.
263. **Yamamoto, Y., T. W. Klein, and H. Friedman.** 1993. Differential effects of ethanol on permissive versus nonpermissive macrophages infected with *Legionella pneumophila*. *Proc. Soc. Exp. Biol. Med.* **203**:323–327.
264. **Yates, S. L., M. Bencherif, E. N. Fluhler, and P. M. Lippiello.** 1995. Up-regulation of nicotinic acetylcholine receptors following chronic exposure of rats to mainstream cigarette smoke or alpha 4 beta 2 receptors to nicotine. *Biochem. Pharmacol.* **50**:2001–2008.
265. **Yeager, M. P., T. A. Colacchio, C. T. Yu, L. Hildebrandt, A. L. Howell, J. Weiss, and P. M. Guyre.** 1995. Morphine inhibits spontaneous and cytokine-enhanced natural killer cell cytotoxicity in volunteers. *Anesthesiology* **83**:500–508.
266. **Zhang, P., G. J. Bagby, D. M. Boe, Q. Zhong, P. Schwarzenberger, J. K. Kolls, W. R. Summer, and S. Nelson.** 2002. Acute alcohol intoxication suppresses the CXC chemokine response during endotoxemia. *Alcohol Clin. Exp. Res.* **26**:65–73.
267. **Zhang, Z., J. Cork, P. Ye, D. Lei, P. O. Schwarzenberger, W. R. Summer, J. E. Shellito, S. Nelson, and J. K. Kolls.** 2000. Inhibition of TNF- α processing and TACE-mediated ectodomain shedding by ethanol. *J. Leukoc. Biol.* **67**:856–862.
268. **Zhu, L. X., S. Sharma, M. Stolina, B. Gardner, M. D. Roth, D. P. Tashkin, and S. M. Dubinett.** 2000. Δ -9-Tetrahydrocannabinol inhibits antitumor immunity by a CB2 receptor-mediated, cytokine-dependent pathway. *J. Immunol.* **165**:373–380.
269. **Zhu, W., H. Friedman, and T. W. Klein.** 1998. Δ -9-Tetrahydrocannabinol induces apoptosis in macrophages and lymphocytes: involvement of Bcl-2 and caspase-1. *J. Pharmacol. Exp. Ther.* **286**:1103–1109.
270. **Zhu, W., T. Igarashi, H. Friedman, and T. W. Klein.** 1995. Δ -9-Tetrahydrocannabinol (THC) causes the variable expression of IL2 receptor subunits. *J. Pharmacol. Exp. Ther.* **274**:1001–1007.
271. **Zhu, W., T. Igarashi, Z.-T. Qi, C. Newton, R. E. Widen, H. Friedman, and T. W. Klein.** 1993. Delta-9-tetrahydrocannabinol (THC) decreases the number of high and intermediate affinity IL-2 receptors of the IL-2 dependent cell line NKB61A2. *Int. J. Immunopharmacol.* **15**:401–408.