

A second case of human C3b inhibitor (KAF) deficiency

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SUMMARY

The second case of C3b inhibitor deficiency is described in an 11-year-old girl who presented with recurrent attacks of meningitis, in between which she was well. Her serum showed all of the complement component changes noted in the first described case, although showing only a relatively slight defect in its ability to opsonize bacteria for phagocytosis and killing by polymorphonuclear leucocytes. This correlated with the patient's freedom from other infections.

INTRODUCTION

The clinical syndromes associated with genetic defects of individual complement components in man fall into three groups: (i) an increased tendency to bacterial infections as a consequence of a deficiency of C3, either primary (Alper *et al.*, 1972; Ballow, Shira, Harden, Soo and Day, 1975) or secondary (Alper *et al.*, 1970; Alper, Block & Rosen, 1973); (ii) an increased tendency to immune complex diseases; possibly because of poor antigen clearance, and usually associated with defects of the early classical pathway components C1 (De Braco *et al.*, 1974; Pickering *et al.*, 1971), C4 (Hauptmann *et al.*, 1974) and C2 (Ruddy *et al.*, 1970; Agnello, De Braco & Kunkel, 1972) and (iii) angio-neurotic oedema due to C1 esterase deficiency (Donaldson & Evans, 1963; Alper & Rosen, 1971) in which there is also an increased incidence of immune complex disease related to the secondary decrease in serum C4 and C2. Individuals with defects of the later components C5, C6, C7 and C8 have been described, and generally show little evidence of ill-health although they may show a tendency to Neisserian infections (Lim *et al.*, 1976; Peterson, Graham & Brooks, 1976).

In the only previous case of C3b inhibitor deficiency (Alper *et al.*, 1970) the lack of the inhibitor resulted in a continuing activation of the C3 feedback mechanism of the alternative pathway of complement (Alper, Rosen & Lachmann, 1972) so as to lower the C3 and factor B levels, and to exhaust the mechanism for enhancing C3 deposition, which is necessary for bacterial phagocytosis and killing (Fig. 1). The patient, who also had Klinefelter's syndrome and was mentally subnormal, had experienced recurrent upper and lower respiratory infections from an early age.

The present case, an 11-year-old girl, came to light because of recurrent attacks of meningitis from which she made a full and rapid recovery on antibiotic treatment, and in between which she was perfectly healthy. Initial investigations had shown that her serum immunoglobulin levels were normal, but that the levels of C3 and Factor B in her serum and plasma were considerably reduced, with appreciable amounts in the altered or activated form.

CASE HISTORY

The patient was a full-term normal delivery, with normal childhood development. At 4 months of age, she developed bacterial meningitis (*Streptococcus pneumoniae*), with a recurrence shortly after discharge from hospital.

At 1½ years she developed an ear infection which responded to treatment. At 7 years—bacterial meningitis again (*Neisseria meningitidis*). At 11 years—bacterial meningitis again (no organism isolated—associated with a purpuric skin rash). At 11½ years—bacterial meningitis again (*N. meningitidis*). Recovery from the infections has always been rapid and complete.

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TABLE 1. Summary of immunological investigations*

Serum immunoglobulins (g/l):	IgG—14.0 (6.0–16.0):	IgA—2.2 (0.9–5.4):	IgM—1.1 (0.4–1.8)
Serum IgE (IU/ml):	160 (50–500)		
Parotid saliva IgA (mg/l):	36 (30–68)		
Complement:	Tables 2 and 3		
Phagocytic (polymorph) function tests:			
Nitroblue tetrazolium reduction:		spontaneous 6% (up to 10)	stimulated 20% (20–50)
Chemotaxis:		see Table 4	
Bactericidal Index (by the method of Quie <i>et al.</i> , 1967):		0.04 (< 0.2)	
Peripheral Blood lymphocytes:			
Spontaneous sheep cell (E) rosettes:	58% (55–70)		
Complement sheep cell (EAC) rosettes:	25% (20–35)		
Surface immunofluorescence (anti-Fab):	27% (16–30)		
Lymphocyte transformation (Ratio of H3-thymidine uptake in test to control)			
Phytohaemagglutinin	2.0 µg/ml—116 (93–918)		
	0.2 µg/ml— 47 (30–700)		
Antibody activity: isoagglutinins present			
<i>E. coli</i> antibodies† positive 1/64 (> 1/32)			
Viral complement fixation tests		—positive for <i>Mycoplasma pneumoniae</i> , measles, mumps	
		—negative herpes simplex, influenza A, B and C	
Meningococcal complement fixation test—negative			

* Normal ranges in brackets.

† By the method of Webster, Effer & Asherson (1974).

TABLE 2. Complement (normal ranges in brackets)

	Functional	Immunochemical (% NHS standard)
CH ₅₀ (u/ml)*	10; 14 (28–45)	
Clq		93 (75–130)
C1	100% NHS†	
C1 INH		96 (70–140)
C4	* 16,000 u/ml (10–30 × 10 ³) 92% NHS†	110 (55–180)
C2	* 3500 u/ml (1–3 × 10 ³) 100% NHS†	
C3		30; iu35 (60–150)
C5 (% NHS)†	32: 38	
C6 (% NHS)	80†	100 (80–180)
C7 (% NHS)†	13: 25 (75–140)	
C8		95 (90–140)

NHS = Normal human serum. Repeat values given in some cases

* Tube assay.

† Agarose-gel cell red assay.

patients with low C3 levels. C5 was reduced, as was C7, but C6 and C8 were normal and C9 was not tested.

Of the alternative pathway components, Factor B was much reduced, and electrophoresis of fresh plasma or serum showed that it was all in the B_b state (Fig. 3). There was no functional Factor B activity as determined by the ability to combine with purified cobra venom factor to form a C3 splitting enzyme, or to lyse guinea-pig red cells in the presence of human EDTA serum. The properdin level was also

TABLE 3. Alternate pathway (% standard serum)

Factor B	*10 (Functional test*—nil)
Factor P (proderdin)	*25 (70–120)
Factor D	†72
'Total' alternate pathway functional activity† (lysis)—nil	

* Immunochemical quantitation by single radial diffusion.

† Agarose-gel red cell test.

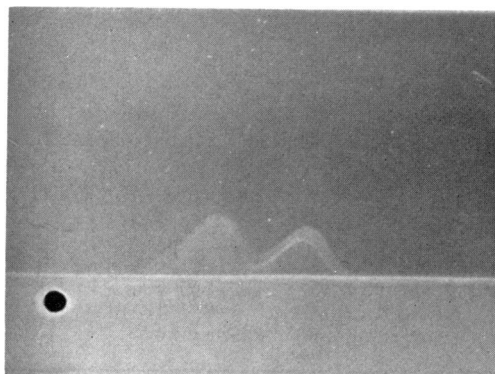


FIG. 2. Two-dimensional electrophoresis in agarose of fresh EDTA plasma from the patient. Anti-C3 anti-serum was present in the agarose for the second dimension. Normal plasma gave a single peak.

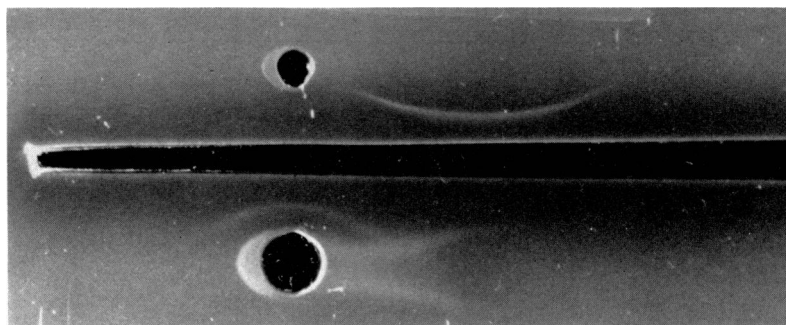


FIG. 3. Immunoelectrophoresis in agarose of fresh normal serum (top) and fresh serum from the patient (bottom). The trough contained anti-factor B, showing all in the Bb position in the patient. Anode to the right.

low although no assessment was made of its functional capacity. Factor D was probably normal. When purified Factor B was incubated with fresh patient's serum for 20 min at 37°C, there was complete conversion to Bb, while similar incubation with normal sera produced negligible conversion (Fig. 4). This had been reported in the original case and shows the presence in her serum of active C3b together with factor D. Tests for total alternative pathway activity, as detected by the lysis of unsensitized guinea-pig erythrocytes in agarose-gel containing MgEGTA, and by B antigen consumption on incubation of serum with zymosan, were negative.

The C3b inhibitor was completely absent on immunochemical testing of several specimens of serum, obtained over a 6 month period (Fig. 5). It was also not detected by more sensitive functional tests.

The serum showed a reduced ability to generate chemotactic factors for the patient's own and normal polymorphs, although the patient's cells responded normally to chemotactic stimuli (Table 4). There

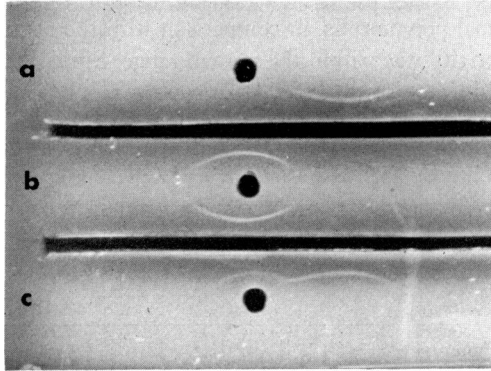


FIG. 4. Electrophoresis in agarose of a preparation of purified factor B, after incubation at 37°C with a fifth of its volume of (a) saline, (b) serum from the patient, (c) normal serum. Anti-factor B was in the troughs. Anode to the right.

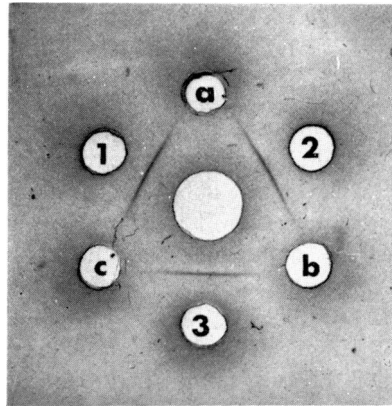


FIG. 5. Double diffusion in agarose-gel of three sera obtained on different occasions from the patient (a, b and c separated by intervals of 2 months and 3½ months respectively), and three sera from normal subjects (1, 2, and 3). The central well contained anti C3b inhibitor (KAF).

TABLE 4. Polymorph chemotaxis

Chemotactic stimulus	Responding cells		
	From patient	From normal subjects	
		A	B
Patients serum	80	85	42
Patients serum + zymosan			23
Normal serum	368	495	297
Normal serum + zymosan			324

* Method modified from Kay (1970) in which 10^6 purified peripheral blood polymorphonuclear leucocytes are suspended in balanced salt solution in a chamber on one side of an $8.0 \mu\text{m}$ Millipore filter with serum or activated serum on the other side, the whole being incubated at 37°C for 90 min, after which the filters are stained and the cells which have migrated through are counted (total cells in eight random high power fields). Mean normal cell count in response to chemotactic factors from normal sera is 243 ± 58 .

was slightly impaired opsonization by the patient's serum of both *Staphylococcus aureus* and *Candida albicans* for ingestion by normal polymorphs, in comparison with the control serum tested at the same time, although the level of activity was within the normal range. Similarly, the killing by normal polymorphs of *S. aureus* ingested in the presence of the patient's serum was slightly less than when normal serum cofactors were present, but the results were within the normal ranges for this test (Table 5).

TABLE 5. Bacterial killing and opsonization (with normal polymorphs)

	Patient's serum	Normal serum	
		(1)	(2)
<i>S. aureus</i> ($\times 10^3$) ingested after 20 min	22	43	35
Proportion (%) killed after 140 min	89	96	98
<i>Candida albicans</i> ingestion (mean organisms/cell)	Normal cells (1)	4.0	5.8
	Normal cells (2)	2.1	3.1 (No serum 0.1)

DISCUSSION

This patient is the second case of C3b inhibitor (KAF) deficiency to be reported. The defect is likely to be genetically transmitted since other members of her immediate family show reduced levels, although in none is the inhibitor completely absent. Full family studies of this and other components of the complement system, as well as other genetic markers are in hand, in an attempt to identify the mode of inheritance and linkage, if any, of this protein; these will be the subjects of further communications.

The serum shows all of the essential complement changes demonstrated in the previous case, and in the *in vitro* model proposed to explain the consequences of the defect (Nicol & Lachmann, 1973). However, while immune haemolysis was reduced, due to low levels of C3 and later factors, the overall functional activities of bacterial opsonization and killing in the presence of polymorphs were not grossly disturbed. The reason for the relative normality of these serum functions, in contrast to their defectiveness in the first reported case, is uncertain. The degree of alternative pathway 'exhaustion' may be less complete, or serum antibody and the classical pathway may be more effective in this patient. Whatever the reason the findings are in accord with her relative freedom from infections apart from meningitis.

There were no group specific anti-meningococcal antibodies demonstrable in her serum, and this may be why the recurrent infections took the particular form that they did. It is uncertain whether the risk from other infections will increase during adolescence and early adulthood, and her progress will be followed with much interest.

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