

An empirical test of partner choice mechanisms in a wild legume–rhizobium interaction

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Mutualisms can be viewed as biological markets in which partners of different species exchange goods and services to their mutual benefit. Trade between partners with conflicting interests requires mechanisms to prevent exploitation. Partner choice theory proposes that individuals might foil exploiters by preferentially directing benefits to cooperative partners. Here, we test this theory in a wild legume–rhizobium symbiosis.

Rhizobial bacteria inhabit legume root nodules and convert atmospheric dinitrogen (N₂) to a plant available form in exchange for photosynthates. Biological market theory suits this interaction because individual plants exchange resources with multiple rhizobia. Several authors have argued that microbial cooperation could be maintained if plants preferentially allocated resources to nodules harbouring cooperative rhizobial strains. It is well known that crop legumes nodulate non-fixing rhizobia, but allocate few resources to those nodules. However, this hypothesis has not been tested in wild legumes which encounter partners exhibiting natural, continuous variation in symbiotic benefit.

Our greenhouse experiment with a wild legume, *Lupinus arboreus*, showed that although plants frequently hosted less cooperative strains, the nodules occupied by these strains were smaller. Our survey of wild-grown plants showed that larger nodules house more *Bradyrhizobia*, indicating that plants may prevent the spread of exploitation by favouring better cooperators.

Keywords: exploitation; cooperation; symbiosis; mutualism; nitrogen fixation; sanctions

1. INTRODUCTION

Mutualisms can be modelled as biological markets in which members of each species exchange resources or services (Noë & Hammerstein 1994). However, markets are vulnerable to exploitation and require mechanisms to promote fair commodity exchange (Bronstein 2001). Both ‘partner choice’ and ‘partner fidelity’ can constrain exploitation (Bull & Rice 1991; Noë *et al.* 1991; Simms & Taylor 2002; Sachs *et al.* 2004). Partner fidelity occurs when individuals receive returned benefits from their investments in others. Such fitness feedbacks can arise from vertical transmission of symbionts (Fine 1975; Axelrod & Hamilton 1981), certain spatial structures (Wilkinson 1997; Doebeli & Knowlton 1998), or other mechanisms that assure long-term reciprocal interactions. However, positive fitness feedback can be weakened by horizontal transmission and/or competition among potential mutualists (Soberon 1985; Frank 1994, 1996*a,b*; West *et al.* 2002*a,b*; Bronstein *et al.* 2003; Wilson *et al.* 2003). In such cases, exploitation may be constrained by partner choice, in which individuals preferentially extend benefits to cooperative members of the partner species (Bull &

Rice 1991; Simms & Taylor 2002; Sachs *et al.* 2004). There are few empirical tests of the partner choice hypothesis (but see Bshary & Grutter 2002; Grutter & Bshary 2003; Kiers *et al.* 2003; Mueller *et al.* 2004).

Root-nodule inhabiting bacteria (hereafter termed rhizobia) are particularly attractive model systems for examining the maintenance of mutualism (Denison 2000; Simms & Taylor 2002). Rhizobia fix atmospheric nitrogen in exchange for photosynthates, but the interaction does not always appear cooperative. Plant and bacteria reproduce and disperse independently (Simms & Taylor 2002) and individual plants usually interact with multiple bacterial genotypes that vary from beneficial to completely ineffective mutualists (Moawad & Beck 1991; Quigley *et al.* 1997; Moawad *et al.* 1998; Denison 2000; Thrall *et al.* 2000). Several authors have suggested that rhizobial cooperation could be promoted by plant traits that preferentially allocate resources to nodules harbouring cooperative strains (Denison 2000; Simms & Taylor 2002; West *et al.* 2002*b*; Sprent 2003). Further, crop legumes are known to sanction (Denison 2000) nodules occupied by ineffective rhizobia (Chen & Thornton 1940; Wadisirisuk & Weaver 1985; Kiers *et al.* 2003). However, the partner choice hypothesis has not been tested in wild legumes interacting with their naturally occurring rhizobia.

Here, we use a greenhouse experiment to test whether yellow bush lupin, *Lupinus arboreus* Sims, a short-lived perennial shrub in the Leguminosae, can allocate resources to more effective symbionts when nodulated

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by mixed populations of bacterial strains with which they naturally occur.

2. MATERIAL AND METHODS

Yellow bush lupin is native to the central California coast and is common throughout Bodega Marine Reserve (BMR; 38°19'01" N, 123°04'18" W). At BMR, *L. arboreus* can occur alone or with up to four other lupin species: *Lupinus nanus*, *Lupinus bicolor*, *Lupinus variicolor* and *Lupinus chamissonis* (Barbour *et al.* 1973). Like other lupins (Bottomley *et al.* 1994; Ludwig *et al.* 1995; Barrera *et al.* 1997; but see Stepkowski *et al.* 2003), lupins at BMR associate predominantly with *Bradyrhizobium* strains that are closely related to the primary soyabean symbiont, *Bradyrhizobium japonicum*, and are generally designated as *Bradyrhizobium* sp. (*Lupinus*) (Taylor & Simms, unpublished data).

Rhizobia fix nitrogen only after differentiating into bacteroids within the plant (Simms & Bever 1998). Although there is debate about the ability of bacteroids to survive nodule senescence, de-differentiate and reproduce, *Bradyrhizobium* bacteroids or vegetative cells of the strain generating bacteroids apparently survive senescence of lupin nodules (Sprent *et al.* 1987). Mycorrhizae are unlikely to be important for the nutrient or water relations of lupins (Trinick 1977; Avio *et al.* 1990; Oba *et al.* 2001).

Seeds and bacteria were collected from six intensively studied sites at BMR (Maron & Simms 2001), three in dunes and three in grasslands, located *ca* 500 m apart. Seeds were collected from two maternal plants sampled from each of three sites: South Grassland, Mid Grassland and Mid Dunes. Ten bacterial isolates were obtained from nodules excised from two naturally occurring lupins at each of five sites: South Grassland, Mid Grassland, South Dunes, Mid Dunes and Mussel Point.

We genotyped bacterial isolates using PCR-RFLPs of the 16S–23S rRNA internal transcribed spacer (ITS) region (Taylor & Simms, unpublished data) and characterized their symbiotic benefit to two lupin species, *L. arboreus* and *L. variicolor* (J. Povich and E. L. Simms, unpublished data). Three strains were selected that differed in RFLP type and represented the observed range of symbiotic benefit to *L. arboreus* (table 1).

To test whether plants preferentially allocate resources to more beneficial bacteria, we established three mixed inoculation treatments (mediocre and poor), (good and poor), (good and mediocre) and an uninoculated control treatment. Each treatment was applied to one randomly assigned seedling from each of six maternal plants in each of five blocks. Within a block, the six seedlings in an inoculation treatment were each grown in a separate pot, with pots racked together to form a subplot within a fully randomized split plot design. We maintained 1 m distances among racks to prevent cross-contamination. Four racks comprised a block. Plants were assigned to blocks by size, which primarily reflected germination date. Blocks varied by plant age, location on the greenhouse bench, inoculation volume and harvest date.

On 25 July 2003, seeds for blocks 1–3 were scarified and surface-sterilized by soaking in concentrated sulphuric acid for 15 min and rinsing in sterile deionized water. The seeds were germinated in autoclaved trays of moist vermiculite under metal halide lamps set to a 23 h photoperiod at room temperature. After 12 days, seedlings were transplanted to sterile Deepots (Steuwe & Sons) filled with autoclaved calcine-

Table 1. Characteristics of *Bradyrhizobium* strains.

strain	wild host ^a	location	shoot mass (g) ^b	ranked symbiotic benefit
LA11-1	<i>L. arboreus</i>	South Grass-land	1.93	poor
LA17-1	<i>L. arboreus</i>	North Dunes	2.14	mediocre
LB16-1	<i>L. bicolor</i>	Mussel Point	2.36	good

^a Host from which strain was isolated.

^b Average aboveground dry mass after 20 weeks of 48 *L. arboreus* plants inoculated with the strain. Strains differed significantly ($F_{2,126} = 12.46$, $p \leq 0.0001$); experiment-wide 95% confidence limit = 0.12 g.

clay (Turface, Profile Products). On 6 August 2003, seeds for blocks 4 and 5 were surface-sterilized for 20 min in bleach, rinsed in sterile water and nick scarified with a razorblade. After imbibing sterile water for 3 days, germinated seeds were planted directly into sterile Deepots of autoclaved Turface. All seedlings were watered four times per day with UV-sterilized carbon-filtered tap water until 20 August 2003, after which they were watered as necessary (usually daily) and fertilized biweekly with nitrogen-free modified Jensen's solution.

Each strain was initiated from a single-plated colony grown in liquid culture (modified arabinose-gluconate (MAG) media, modified by P. van Berkum from Cole & Elkan 1973) and cryopreserved in 60% glycerol. Inoculant was prepared by regrowing on solid media and washing cells with sterile 0.85 M KCl. Cell densities were determined by absorbance and adjusted to 1×10^8 cells ml⁻¹. Cell numbers were calibrated by cytometer and confirmed with plate counts. Mixed inoculants were prepared by combining equal quantities of the two appropriate single-strain inoculants. We pipetted 10 ml of mixed inoculant onto each seedling in blocks 1–3 on 12 August 2003. On 1 September 2003, we inoculated seedlings in blocks 4 and 5 with 7 ml. Control plants within a block received the appropriate volume of sterile 0.85 M KCl.

Block 1 was harvested on 21 September 2003; remaining plants were harvested on 6–7 January 2004. During both harvests, we selected a range of nodule sizes from each plant. To ensure that small nodules were not younger than large nodules, we chose small nodules proximal along the roots from which large nodules were obtained. We measured the diameters of six nodules, then each was excised, surface-sterilized with household bleach and stored at -20°C in $5 \times$ its volume of 20% Chelex 100 resin (Sigma Chemical) in $2 \times$ PCR buffer (40 mM Tris-HCl, pH 8.0; 100 mM KCl). Immediately after thawing, nodules were heated and vortexed at 95°C , centrifuged for 5 min at 16 000g. The resulting crude extract was stored at -20°C .

From each sample, we amplified 1440 bp of the intergenic spacer between the large and small ribosomal subunits (hereafter referred to as ITS) using primers ITS450 and ITS1440 (van Berkum & Fuhrmann 2000). The 50 μl PCR reaction contained 2.0 mM MgCl₂, 0.25 mM dNTPs, 0.5 μM primers, 1.25 units *Taq* polymerase (Invitrogen Life Technologies), $1 \times$ Invitrogen PCR buffer and 10 μl of diluted nodule extract (usually a 25-fold dilution). Reactions were initiated with a 2 min 95°C denaturation and ended with a 10 min extension step at 72°C . Cycling involved 94°C denaturation for 30 s, 70°C annealing for 40 s, reduced by 0.5°C per cycle to 60°C in the touchdown phase, with an additional 30 cycles at 60°C , and 72°C extension for 1.5 min. Aliquots of the

PCR product were digested with endonucleases *Mwo*I and *Hga*I. Fragments were separated on 10 cm gels of 1% regular agarose and 2% high resolution agarose (Sigma Chemical) in tris-acetate-EDTA electrophoresis (TAE) buffer at 170 V for 2–3 h, stained with ethidium bromide and visualized on a UV transilluminator.

Owing to germination problems, each treatment in block 5 lacked one plant. Nineteen plants died during a late August heatwave; five were replaced with substitutes of the same age that had been appropriately inoculated contemporaneously. By harvest, 17 additional plants had died, including nine control plants, reducing the number of harvested plants to 85. Of these, one inoculated plant failed to nodulate and PCR reactions failed on all nodules of three other inoculated plants. Ultimately, we obtained informative nodule occupancy data from 65 plants.

Nodule size and strain identity were analysed with a repeated measures nested analysis of variance (ANOVA) with a compound symmetry covariance structure using the SAS 9.1 MIXED procedure (SAS Institute 2004). The model included nodules as repeated measures within plants, plants as random subjects nested within fixed inoculation treatments and treatments nested within random blocks. Plant family structure was too unbalanced to analyse. The block effect and block by plant (treatment) interaction were non-significant and raised the Akaike Information Criterion score, so these effects were dropped. For each inoculation treatment, we subsequently performed a one-way ANOVA of plant effect on nodule size, used the residuals to classify nodules into two or three size classes and used chi-squared tests to determine whether standardized nodule sizes were distributed independently of strain identity.

To establish the relationship between nodule diameter and *Bradyrhizobium* population size within a nodule, we surveyed six nodules each from nine *L. arboreus* seedlings at four sites across BMR. Nodules were collected, measured and surface-sterilized as described above. Each nodule was macerated in sterile MAG media and the slurry was serially diluted and plated on MAG-agar plates for colony counts. Plate counts were averaged across at least two replicates per nodule to estimate the *Bradyrhizobium* population size. Colony number was natural log-transformed to normalize residuals and regressed on nodule diameter using JMP 5.1.2 (SAS Institute).

3. RESULTS

We obtained readable bands from 301 nodules. Banding patterns from two nodules suggested occupancy by more than one isolate. We found no evidence of cross-contamination among treatments. At harvest, no control plants were nodulated and all identified strains matched at least one of the strains with which their host plant had been inoculated.

Nodule sizes within plants were significantly influenced by occupant identity ($F_{2,25} = 16.42$, $p < 0.0001$). Within the average plant, nodules that were occupied by the more beneficial strain were significantly larger (figure 1*a–c*). However, because strain identity interacted significantly with treatment ($F_{1,25} = 10.14$, $p < 0.004$), we also examined the strain effect in each treatment separately.

Of 24 plants inoculated with both good and poor strains, we detected 11 plants nodulated by both strains. Occupant identity explained a significant component of variation in nodule size within plants ($F_{1,10} = 5.38$,

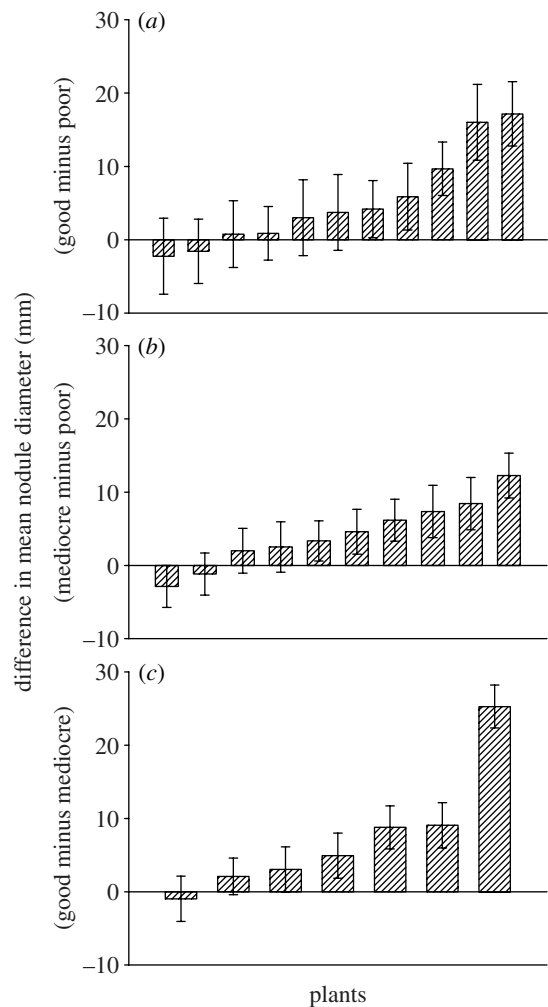


Figure 1. Within plant average difference in sizes of nodules occupied by best versus worst strain. Plants inoculated by (a) strains LB16-1 (good) and LA11-1 (poor), (b) strains LA17-1 (mediocre) and LA11-1 (poor), (c) strains LB16-1 (good) and LA17-1 (mediocre). Bars indicate 1 s.e.

$p = 0.04$), with smaller average size among nodules occupied by the poor strain (figure 1*a*). Further, contingency table analysis indicated that strain distribution between nodule size categories differed from random, with the poor strain occurring more frequently in smaller nodules (table 2*a*, $\chi^2_1 = 5.00$, $p < 0.03$). Of the 14 plants in which we detected only one strain, all were occupied by the poor strain.

Of 22 plants inoculated with both mediocre and poor strains, we found 10 nodulated by both strains. Occupant identity explained a significant component of variation in nodule size within plants ($F_{1,9} = 7.91$, $p = 0.02$), with smaller average size among nodules occupied by the poor strain (figure 1*b*). Strain distribution among nodule size categories differed from random (table 2*b*, $\chi^2_2 = 12.36$, $p < 0.002$), with the poor strain occurring more frequently in smaller nodules. Of the 12 plants in which we detected only one strain, three contained the poor and nine contained the mediocre strain.

Finally, of 18 plants inoculated with both good and mediocre strains, seven plants were nodulated by both strains and occupant identity explained a significant component of variation in nodule size within plants ($F_{1,6} = 10.48$, $p = 0.02$), with smaller average size among nodules occupied by the mediocre strain (figure 1*c*). As in

Table 2. Contingency tables of strain identity versus nodule size class after accounting for differences among plants.

strain	rank	large	small	medium
<i>(a) strains LB16-1 versus LA11-1</i>				
LB16-1	good	10	8	
LA11-1	poor	8	25	
$\chi^2_1 = 5.00, p < 0.03$				
<i>(b) strains LA17-1 versus LA11-1</i>				
LA17-1	med ^a	11	6	12
LA11-1	poor	0	8	18
$\chi^2_2 = 12.36, p < 0.002$				
<i>(c) strains LB16-1 versus LA17-1</i>				
LB16-1	good	6	2	
LA17-1	med ^a	6	17	
$\chi^2_1 = 5.99, p < 0.02$				

^a med, mediocre.

the previous two treatments, strain distribution between nodule size categories differed from random, with the mediocre strain occurring more frequently in smaller nodules (table 2c, $\chi^2_1 = 5.99, p < 0.02$). Of the 11 plants in which we detected only one strain, all were occupied by the mediocre strain.

Among 33 nodules sampled randomly from nine wild-grown *L. arboreus* seedlings collected at BMR, total population size of *Bradyrhizobium* cells in a nodule regressed positively on nodule size ($r^2 = 0.2, p < 0.01$).

4. DISCUSSION

In lupins experimentally infected by divergent pairs of naturally occurring *Bradyrhizobium* strains, nodule size varied with the symbiotic effectiveness of bacterial occupants. On average, nodules inhabited by the less beneficial strain were smaller. In a separate survey of wild-grown *L. arboreus*, larger nodules contained more *Bradyrhizobium* cells, as previously found for soybean nodules (Kiers *et al.* 2003). These results support the hypothesis that legumes can favour more cooperative rhizobia by manipulating bacterial fitness in the nodule (Denison 2000; Simms & Taylor 2002; West *et al.* 2002b; Sprent 2003).

Although our experiment does not specifically distinguish between plant or bacterial control of allocation to nodules, we argue that the original hypothesis is more parsimonious. Our data complement the findings of Kiers *et al.* (2003), who found that the fresh weights of soybean nodules deprived of atmospheric dinitrogen were smaller than those of unmanipulated nodules occupied by the same strain on the same plant. They suggested that plants might control nodule size in part by altering oxygen supply. Other mechanisms might also contribute to the effect (Simms & Taylor 2002).

Among the 37 plants in which we found only one strain, 25 had been inoculated with a mix including the good strain, yet we detected this strain in none of these plants. This result corroborates other studies, which have shown that the ability of rhizobia to compete for nodulation may be uncorrelated with symbiotic benefit (Triplett & Sadowsky 1992; Vasquez-Arroyo *et al.* 1998; Bloem & Law 2001; Hafeez *et al.* 2001).

For nodule-specific plant responses to constrain cheating effectively, individual nodules must be occupied by single bacterial genotypes (Denison 2000; West *et al.* 2002b; Kiers *et al.* 2003; Denison & Kiers 2004). Among the 301 nodules from which we obtained readable RFLP bands were two that showed a banding pattern suggestive of occupancy by more than one isolate. These data suggest that multiply infected nodules are rare.

We believe that our study provides the first evidence that nodule size scales with naturally occurring variation in symbiotic effectiveness of the bacterial occupant. Past workers on agricultural legumes have noted that ineffective strains often result in small nodules (Simms & Taylor 2002), but, with a few notable exceptions (Chen & Thornton 1940; Singleton & Stockinger 1983; Kiers *et al.* 2003), the phenomenon has rarely been quantified.

In summary, our results suggest that post-infection partner choice could be an important mechanism constraining exploitation by rhizobia in this legume–rhizobium mutualism. We look forward to additional tests of the assumptions of this theory.

We are most grateful to C. Coleman, M. Horn and T.F. Colton for assistance with harvests and data entry, and to C.T. Hsu and A. Jain, who measured nodules and censused bacterial densities. Major funding was provided by DEB-0108708 and DEB-9996236 to ELS from the USA NSF. Funds were also provided by the Miller Institute for Basic Research, the UC Berkeley Committee on Research and the UC Berkeley Faculty Research Fund for the Biological Sciences. Mona Urbina was supported by GM48983 from the US NIH to D.A. Gailey of CSU East Bay. We are grateful to two anonymous reviewers for helping to improve the manuscript. We thank Bodega Marine Reserve for access to field sites, and Reserve Manager P. Conners for his insights and assistance.

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