



## SPECIAL REPORT

# The cannabinoid CB<sub>1</sub> receptor antagonist, SR141716A, selectively facilitates nociceptive responses of dorsal horn neurones in the rat

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The effect of spinal administration of the selective cannabinoid CB<sub>1</sub> receptor antagonist, SR141716A, and the selective CB<sub>2</sub> receptor antagonist, SR144528, on innocuous versus noxious evoked responses of dorsal horn neurones in the spinal cord of the anaesthetized rat was investigated. SR141716A (0.001–1 ng 50  $\mu\text{l}^{-1}$ ) dose-relatedly facilitated the non-potentiated component of the electrical C-fibre mediated neuronal response ( $120 \pm 6$ ,  $156 \pm 13$ ,  $192 \pm 33$  and  $192 \pm 31\%$  of control respectively;  $n=6$ ). In contrast, SR144528 (0.001–1 ng 50  $\mu\text{l}^{-1}$ ) did not influence the non-potentiated component of the C-fibre evoked neuronal response ( $n=5$ ). The electrical evoked A $\beta$ -fibre mediated neuronal responses were not influenced by SR141716A or SR144528. The results of this study provide evidence that tonic cannabinoid CB<sub>1</sub> receptor activation, but not CB<sub>2</sub> receptor activation, attenuates acute nociceptive transmission, at the level of the spinal cord. These results suggest a selective antinociceptive role of the endogenous cannabinoids at spinal CB<sub>1</sub> receptors.

**Keywords:** Nociception; spinal neurones; cannabinoid receptor antagonism

**Abbreviations:** CB<sub>1</sub> receptor, cannabinoid<sub>1</sub> receptor; CB<sub>2</sub> receptor, cannabinoid<sub>2</sub> receptor; SR141716A, N-(piperidin-1-yl)-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide hydrochloride; SR144528, N-[(1S)-endo-1,3,3-trimethyl bicyclo[2.2.1]heptan-2-yl]-5-(4-chloro-3-methylphenyl)-1-(4-methylbenzyl)-pyrazole-3-carboxamide; s.e.mean; standard error of mean

**Introduction** The localization of spinal cannabinoid CB<sub>1</sub> receptors (Herkenham *et al.*, 1991; Tsou *et al.*, 1997) and an endogenous cannabinoid receptor ligand at this level (see Di Marzo *et al.*, 1998) implicates a functional role of the endogenous spinal cannabinoids at the level of the spinal cord.

SR141716A has been shown to be a potent and selective antagonist of CB<sub>1</sub> receptors in rat brain (Rinaldi-Carmona *et al.*, 1994). However, it has also been suggested that SR141716A acts as an inverse agonist at human CB<sub>1</sub> and CB<sub>2</sub> receptors in transfected Chinese hamster ovary cells (MacLennan *et al.*, 1998).

Spinal administration of SR141716A has been shown to result in thermal hyperalgesia in mice (Richardson *et al.*, 1997, 1998a) and to facilitate formalin evoked pain behaviour in rats (Strangman *et al.*, 1998). These behavioural studies suggest there is tonic control of spinal nociceptive processing by endogenous cannabinoids acting at the CB<sub>1</sub> receptor. In contrast, the nociceptive threshold of CB<sub>1</sub> receptor knockout mice has been shown to be similar to those of wild-type mice (Ledent *et al.*, 1999). Thus the importance of a tonic control of nociceptive thresholds/responses by the endogenous cannabinoids is unclear.

To our knowledge there have been no reports on the effects of blockade of the endogenous cannabinoids on neuronal measures of nociceptive activity. The effect of spinal administration of SR144528, a selective CB<sub>2</sub> receptor antagonist (Rinaldi-Carmona *et al.*, 1998), on nociceptive processing is unknown. The aim of the present study was to ascertain the selectivity of the tonic effects of the endogenous cannabinoids on nociceptive versus non-nociceptive activity. To this end, the effects of spinal administration of SR141716A

and SR144528 on A $\beta$ -fibre versus C-fibre evoked responses of dorsal horn neurones have been studied.

**Methods** The techniques used have been described previously (Chapman *et al.*, 1994). Extracellular recordings of convergent dorsal horn neurones (depth 500–1000  $\mu\text{m}$ ) were made in anaesthetized (1.5% halothane in 66% N<sub>2</sub>O/33% O<sub>2</sub>) Sprague-Dawley rats (200–250 g,  $n=11$ ). Neuronal responses to transcutaneous electrical stimulation ( $3 \times$  C-fibre threshold, trains of 16 stimuli at 0.5 Hz) of the peripheral receptive field were recorded and post-stimulus histograms were constructed. Evoked responses were separated and quantified on the basis of latencies: A $\beta$ -fibre: 0–20 ms post-stimulus; C-fibre: 90–300 ms post-stimulus and post-discharge: 300–800 ms post-stimulus. The non-potentiated C-fibre evoked neuronal response was calculated as the number of action potentials evoked by the first stimulus multiplied by the total number of stimuli (16). The non-potentiated component of the C-fibre evoked response is reflective of the C-fibre input into the dorsal horn prior to the activation of post-synaptic NMDA receptor mediated events and the facilitation of C-fibre evoked responses.

Control responses (less than 10% variance) were established. The effects of spinal administration of SR141716A (0.001–1 ng 50  $\mu\text{l}^{-1}$  [0.042–42 nM]  $n=6$ ) and SR144528 (0.001–1 ng 50  $\mu\text{l}^{-1}$ ,  $n=5$ ) on evoked neuronal responses were measured. SR141716A and SR144528 were dissolved in distilled H<sub>2</sub>O and ethanol [final concentration for highest dose of drugs studied <0.1% ethanol]. Drugs were given cumulatively and effects were measured at 5, 10, 20, 30 and 40 min post-drug administration. Data are presented as percentage of the control response  $\pm$  s.e.mean, statistical analysis was performed with repeated measures (ANOVA) and Dunnett's multiple comparison test.

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**Results** All of the neurones studied were convergent, located in the dorsal horn of the spinal cord. The mean thresholds for electrical stimulation of C-fibres for the two groups of neurones studied were  $1.7 \pm 0.2$  mA (SR141716A,  $n=6$ ) and  $1.5 \pm 0.3$  mA (SR144528,  $n=5$ ).

Spinal administration of SR141716A significantly facilitated the non-potentiated component of the C-fibre evoked neuronal responses in a dose-related manner (Figure 1). Maximal effects of the lower doses of SR141716A (0.001, 0.01 and 0.1 ng  $50 \mu\text{l}^{-1}$ ) were observed at  $28 \pm 5$ ,  $27 \pm 4$  and  $30 \pm 5$  min post-drug administration. The maximal effect of the highest dose of SR141716A (1 ng  $50 \mu\text{l}^{-1}$ ) was observed at  $18 \pm 5$  min post-drug administration. SR141716A produced a non-significant facilitation of the post-discharge response, a measure of the level of the hyperexcitability of the neurone (Figure 1).

The effects of SR141716A (0.001, 0.01, 0.1 and 1 ng  $50 \mu\text{l}^{-1}$ ) on the overall C-fibre evoked neuronal responses (mean control value  $370 \pm 51$  action potentials) were small and non-significant ( $101 \pm 6$ ,  $116 \pm 5$ ,  $120 \pm 11$  and  $126 \pm 11\%$  of control respectively). The same concentrations of SR141716A (0.001–1 ng  $50 \mu\text{l}^{-1}$ ) did not influence the A $\beta$ -fibre evoked responses (mean control value  $90 \pm 8$  action potentials) of the dorsal horn neurones ( $80 \pm 6$ ,  $92 \pm 7$ ,  $101 \pm 13$  and  $100 \pm 15\%$  of control respectively).

Spinal administration of SR144528 (0.001–1 ng  $50 \mu\text{l}^{-1}$ ) did not influence the evoked responses of dorsal horn neurones. The mean maximal effect of the highest concentration of SR144528 studied on the non-potentiated component of the C-fibre evoked neuronal response and the post-discharge was  $93 \pm 18\%$  of control and  $103 \pm 17\%$  of control respectively. Finally, the mean maximal effect of SR144528 (1 ng  $50 \mu\text{l}^{-1}$ ) on the overall C-fibre and A $\beta$ -fibre evoked neuronal response was  $82 \pm 7$  and  $85 \pm 9\%$  of control respectively.

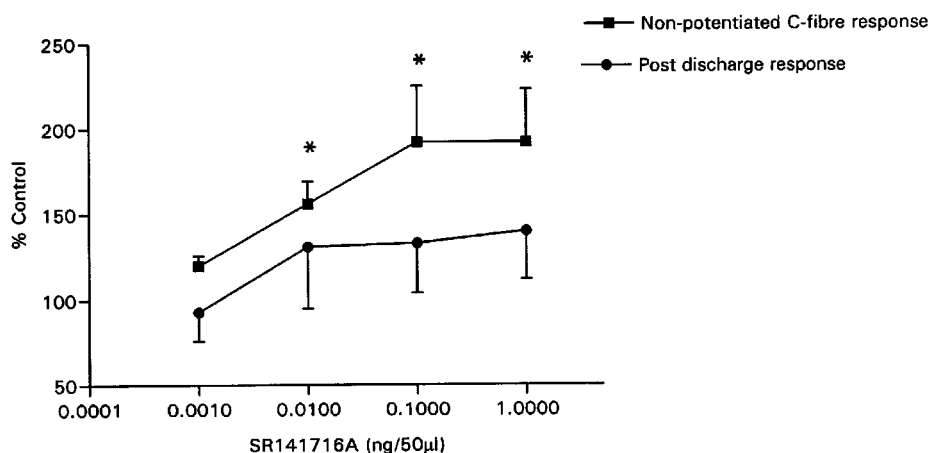
**Discussion** This study has clearly shown for the first time that the cannabinoid CB<sub>1</sub> receptor antagonist, SR141716A, facilitates noxious evoked (C-fibre mediated) responses of spinal neurones. The non-potentiated component of the overall C-fibre evoked neuronal response was most strongly,

and significantly, facilitated by SR141716A. There was a tendency towards a facilitation of the post-discharge response by SR141716A. Innocuous evoked neuronal responses (A $\beta$ -fibre mediated) were not influenced by SR141716A. Spinal administration of the CB<sub>2</sub> receptor antagonist, SR144528, did not influence the C- or A $\beta$ -fibre evoked neuronal responses.

The concentrations of SR141716A used in this study are within the range of concentrations originally shown to be selective for the CB<sub>1</sub> receptor and showing no affinity for an array of other receptors and channels studied (Rinaldi-Carmona *et al.*, 1994). Taken with the finding that the CB<sub>2</sub> receptor antagonist, SR144528, does not influence spinal nociceptive transmission, the results of this study with SR141716A suggest a tonic control of nociceptive evoked activity of spinal neurones by endogenous inhibitory cannabinoids acting at the CB<sub>1</sub> receptor.

The location of the cannabinoid CB<sub>1</sub> receptors is an important determinant of the selective effect of this endogenous inhibitory system on nociceptive transmission. A recent autoradiography study has demonstrated that only 16% of spinal CB<sub>1</sub> receptors are located on C-fibre afferent endings (Hohmann & Herkenham, 1998). Furthermore, a recent *in situ* hybridization study has shown that only 10–15% of dorsal root ganglion (DRG) neurones express mRNA for the CB<sub>1</sub> receptor and a small percentage of the CB<sub>1</sub> receptors in the DRG reside on C-fibres (Hohmann & Herkenham, 1999). Although anatomical data suggests a predominant post-synaptic location of the CB<sub>1</sub> receptors, a functional role of CB<sub>1</sub> receptors located on C-fibre afferent endings has been demonstrated (Richardson *et al.*, 1998b). Thus there are clear differences in the pre- versus postsynaptic location of the CB<sub>1</sub> receptors and opioid receptors which also have anti-nociceptive actions, but are pre-dominantly located presynaptically on C-fibre afferent endings (Besse *et al.*, 1990).

The results of this study are in agreement with previous behavioural studies of the effect of SR141716A and CB<sub>1</sub> receptor antisense oligonucleotides, but are not in agreement with a recent study of nociceptive thresholds in CB<sub>1</sub> knockout mice (see Introduction). Thus there is some disparity between the effect of acutely blocking spinal cannabinoid CB<sub>1</sub> receptors and global CB<sub>1</sub> receptor knockout on nociceptive transmission. Compensatory mechanisms in CB<sub>1</sub> receptor knockout



**Figure 1** The effect of intrathecal administration of SR141716A on the non-potentiated component of the C-fibre evoked response of dorsal horn neurones (mean control value  $320 \pm 81$  action potentials) and the post-discharge response (mean control value  $220 \pm 73$  action potentials) ( $n=6$  for each dose). Note the significant excitatory effect of SR141716A on the non-potentiated response as compared to control. Data shown are means and s.e.means. Statistical analysis: repeated measures (ANOVA) and Dunnett's multiple comparison test, \* $P < 0.01$ .

mice may mask the novel tonic inhibitory control of nociceptive transmission by endogenous cannabinoids acting at the spinal CB<sub>1</sub> receptor reported here.

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