Environmental Health Perspectives
Vol. 82, pp. 91-95, 1989

Benzene-Mediated Protein Kinase C Activation

By Corinne Da Silva,*Xiaotang Fan,* and Monique Castagna*

Extracellular ligands transfer information into the cell through several pathways that operate in an integrated fashion. Protein kinase C, and enzyme that plays a pivotal role in signal transduction, is the molecular target for tumor promoters from the series of phorbol esters. A number of structurally unrelated tumor promoters also enhance protein kinase C, interacting or not interacting with the phorbol ester binding site. Evidence is provided that benzene potently activate protein kinase C *in vitro*, as well as in intact platelets. The drug does not compete for the phorbol ester binding site and probably affects the hydrophobic environment requires for full enzyme activation. Toluene is equally active. The relevance of the presented findings in the carcinogenic effects of benzene is discussed.

Introduction

There is clinical as well as experimental evidence that cancer is a multistep process that may involve several causal agents. Two stages have been characterized from the experimental model designed by Berenblum (*1*) and Mottram (*2*) in mouse skin: initiation and promotion. More recently, models of cultured cells have also provided evidence that cell transformation by physical and chemical carcinogens requires at least two steps. The initial one is a frequent event that gives rise to potentially transformed cells or "initiated" cells, whereas the second one is a rare event that may occur each time an initiated cell divides (*3*). The requirement for two genetic events apparently corroborates the result of transfection experiments that indicate that at least two cooperating oncogenes are needed to convert primary embryo fibroblasts into tumor cells (*4*).

The two classes of carcinogenic agents can be distinguished on the basis of their molecular targets. Initiators include chemicals that affect DNA and elicit irreversible effects as well as agents that generate genetic damage (i.e., mutation, deletion, insertion, translocation or amplification) such as UV light, ionizing radiation, various retroviruses and DNA-containing viruses. Promoters evoke epigenetic changes and reversibly modify the cellular phenotype, including chemicals as well as physical agents causing chronic tissue injury.

* Institut de Recherches Scientifiques sur le Cancer, 7 Rue Guy Mocquet, 94802 Villejuif, France.
Address reprint requests to C. Da Silva, Institut de Recherches Scientifiques sur le Cancer, 7 Rue Guy Mocquet, 94802 Villejuif, France.

Tumor promoters are not carcinogenic by themselves but increase the probability of an initiator-treated cell to become malignant. Despite their lack of carcinogenicity, tumor promoters play a key role in the etiology of cancer. In fact, genotoxic carcinogens are generally present at harmless doses in the environment and only become nocive when associated with tumor promoters, which dramatically lower a carcinogen's threshold of action (*5*).

Two findings have led to the indentification of the major molecular target of phorbol esters and have shed light on the mechanism by which these potent tumor promoters modify the cellular phenotype. First, Driedger and Blumberg, using (^{3}H) phorbol esters, were able to detect high-affinity binding sites for phorbol esters in various cell lines (*6*). Second, as a result of a collaborative work with Y. Nishizuka, we have contributed to the indentification of protein kinase C, a phospholipid and Ca^{2+}-activated enzyme involved in signal transduction, as the high-affinity binding site for phorbol esters (*7–9*).

In this presentation we wish to provide evidence that protein kinase C is not an exclusive molecular target for phorbol ester tumor promoter and that its enzyme activity may be affected by benzene and various other tumor promoters.

Protein Kinase C Activation by Tumor-Promoting Phorbol Esters

Signal transduction from extracellular ligands into the cell follows different strategies. The two best known signaling pathways are shown schematically in Figure 1.

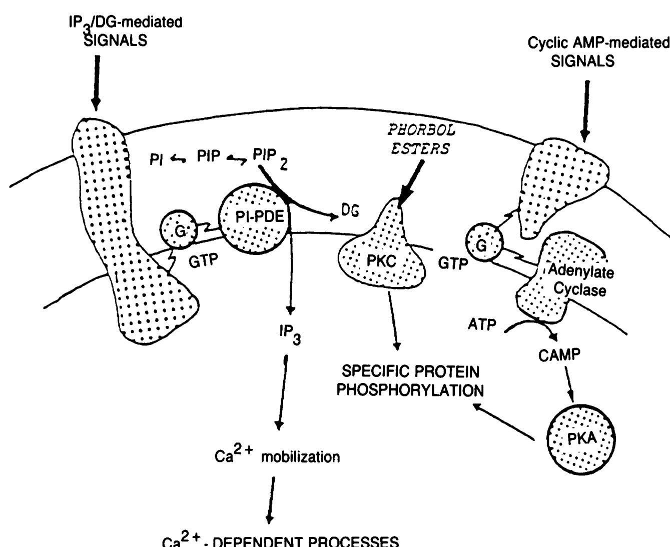

FIGURE 1. Model of cyclic AMP- and IP3/DG-mediated signaling pathways. Abbreviations: PKA, cyclic AMP-dependent protein kinase; PKC, protein kinase C; PI-PDE, phosphatidylinositol phosphodiesterase; PI, phosphatidylinositol; PIP, phosphate dylinositol 4-phosphate; PIP2, phosphatidylinositol 4,5-bisphophate; IP3, inositol 1,4,5-trisphosphate; DG, diacylglycerol.

A number of hormones and neurotransmitters generate cyclic AMP, which serves as a second messenger and triggers protein phosphorylation through activation of a cyclic AMP-dependent protein kinase, while two GTP-binding proteins negatively or positively modulate the response. Alternatively, a large series of ligands that control growth and cellular functions, including neurotransmitters, regulatory peptides, hormones, releasing factors, platelet activators and grow :h factors, proceeds through mediation by two second messengers. Upon ligand-receptor interaction, phosphatidylinositol 4,5-bisphosphate is hydrolyzed and two second messengers are formed: diacylglycerol (DG) and inositol 1,4,5-trisphosphate (IP3). The signaling pathway divides into two branches, the second messengers initiating two sets of cellular events. The former mediates protein kinase C activation, and the latter triggers Ca^{2+} mobilization from internal stores and consequently raises cytosolic Ca^{2+}. A GTP-binding protein functionally similar to those that have been involved in the adenylate cyclase system modulates signal transduction at this step. The two branches act separately but both seem to be required for full cellular response (*8–11*).

The two pathways may not be independent and frequently affect each other. Cyclic AMP-mediated signals negatively control the protein kinase C pathway. Conversely, IP3/DG-mediated signals block adenylate activity. Also, both series of signals potentiate each other in various cell types or cell lines.

Tumor-promoting phorbol esters substitute for the second messenger DG in activating protein kinase C and subsequently bypass the upper part of the pathway. These agents compete with DG for its binding site and display a higher affinity than the natural effector. However, phorbol esters do not fully mimic physiological stimuli. We have recently suggested that these ligands trigger quantitatively and qualitatively different protein kinase C response due to their specific properties which differ in several respects (*9*): a) the half-life of DG is less than 1 min in platelets, whereas phorbol esters like TPA are slowly metabolized in cells; b) phorbol esters do not mobilize Ca^{2+} from internal stores; c) phorbol esters can bypass some regulatory mechanisms that control the IP3/DG pathway, as illustrated in platelets where cyclic nucleotides inhibit thrombin-mediated and not phorbol ester-mediated protein kinase C activation (*12*); and d) phorbol esters can diffuse in all the cellular compartments and as a consequence may evoke a different localization of protein kinase C compared to physiological ligands, thus providing additional substrates to the enzyme. It is likely that these distinctive features might account, at least partly, for their promoting effects.

Benzene and Toluene-Mediated Protein Kinase C Activation

Some reports have supported the possibility that benzene may act as a tumor promoter in its neoplastic effects (*13*). This prompted us to investigate whether this compound affected protein kinase C activity.

The first experiments were designed to test the potency of benzene and its alkylated derivative toluene on protein kinase C activity in intact platelets. In response to physiological ligands, protein kinase C phosphorylates in platelets a set of specific protein substrates. The preferential substrate of the enzyme is a 43 KD protein, which serves as an index of protein kinase C activity in these cells. Figure 2 shows the autoradiogram of SDS-polyacrylamide gel electrophoresis of platelet proteins after the cells were exposed for 2 min to either 12-*O*-tetradecanoylphorbol 13-acetate (TPA), benzene, or

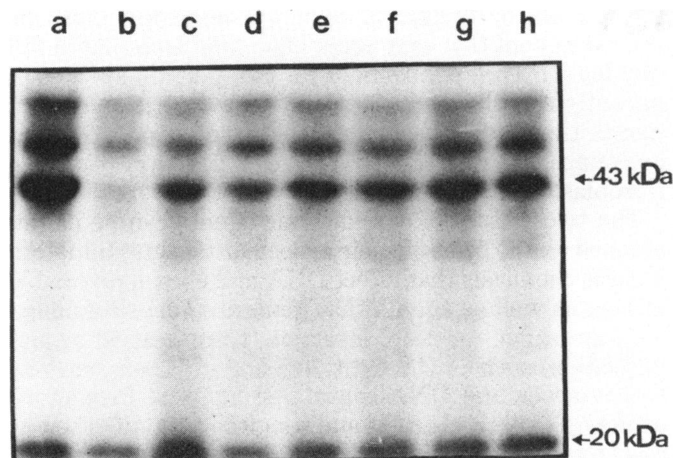

FIGURE 2. Autoradiogram of SDS-polyacrylamide gel electrophoresis of platelet proteins. Cells were stimulated for 2 min with 100 ng/mL TPA (lane a), 5 μL DMSO (lane b), 10% benzene (lanes c,e), 2% benzene (lanes d,f), 10% toluene (lane g), and 2% toluene (lane h). Reprinted from Roghani et al. (*14*).

Table 1. Toluene- and benzene-protein kinase C activity *in vitro*.

Addition, v/v	Enzyme activity, cpm/assay[a]	Increase, %
None	3280 ± 246	100
Toluene		
0.8%	6432 ± 407	196
2.4%	8859 ± 520	270
4.8%	7993 ± 399	243
Benzene		
0.8%	5050 ± 376	154
2.4%	7077 ± 529	215
4.8%	7219 ± 415	220

[a]Assays were carried out in the presence of 0.1 mM $CaCl_2$. Data are means + SE of three experiments in duplicate.

toluene. It can be seen that the two latter compounds activate 43 KD protein phosphorylation and give a phosphorylation pattern similar to that evoked by TPA, suggesting that these phosphorylation events were mediated by the same enzyme.

We have investigated the possibility that benzene interacts with protein kinase C on the phorbol ester/DG binding site. As a result of binding studies conducted on protein kinase C according to a previously described technique using ^{3}H-TPA (*15*), we were unable to show any significant alteration of phorbol ester binding in the presence of either benzene or toluene (data not shown).

To further substantiate these findings, the action of benzene and toluene was studied in the *in vitro* assay described in Roghani et al. (*14*), using a preparation of protein kinase C from mouse brain. Results show that both chemicals directly activated protein kinase C in the presence of phosphatidylserine and Ca^{2+} at concentrations below 1% (Table 1). The dose-response curve of toluene-triggered protein kinase C activation in the presence of 0.5 mM EGTA is shown in Figure 3. Upon addition of TPA at 50 ng/mL, a slight stimulation could also be detected. Benzene and toluene-mediated enzyme activation was not dependent on Ca^{2+}. It should be pointed our that the omission of phospholipid in the incubation mixture prevented enzyme activation, suggesting that the two drugs might interfere with the enzyme-phospholipid interaction.

We have recently shown that mixtures of sodium deoxycholate and phosphatidylserine were much more potent than phosphatidylserine alone in supporting protein kinase C activity and that sodium deoxycholate alone was an enzyme activator (manuscript in preparation). Surprisingly, using such mixtures containing a ratio of phosphatidylserine/deoxycholate of 1/9 (w/w), we could not get any benzene-mediated enzyme activation. Furthermore, benzene inhibited sodium deoxycholate-mediated protein kinase C activation. Results displayed in Figure 4 show the differential effects of benzene on protein kinase C activation depending on the hydrophobic environment. The complexity of the reaction is even more emphasized in experiments where benzene inhibition was studied as a function of deoxycholate concentrations, where at low concentrations of deoxycholate and benzene, an activation of the enzyme can be detected (Fig. 5). The results suggest that benzene and toluene

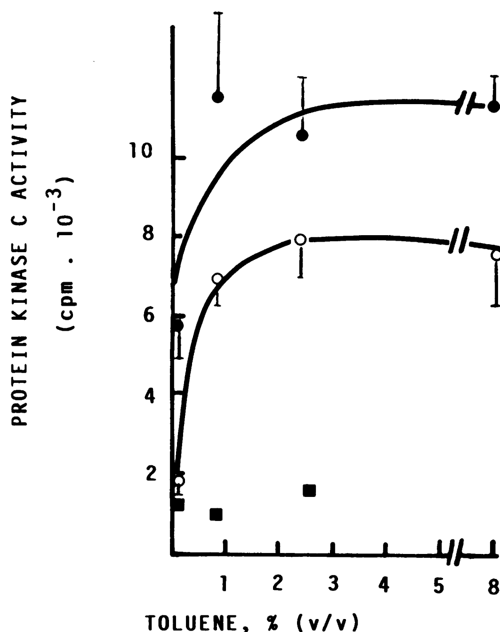

FIGURE 3. Dose-response curve of toluene-mediated protein kinase C activation in the presence of 0.5 mM EGTA (o) or 0.5 mM EGTA + 50 ng/mL TPA (●). In some assays, phospholipid was omitted and 0.5 mM EGTA was added (■). Adapted from Roghani et al. (*14*).

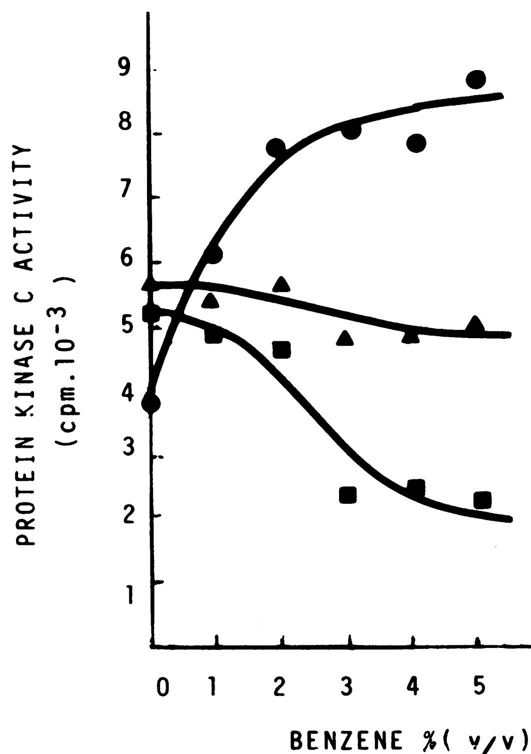

FIGURE 4. Differential effects of benzene on protein kinase C activity in various hydrophobic environments. The enzyme was assayed in 0.5 mM EGTA essentially as described in Roghani et al. (*14*), in the presence of either 80 μg/mL phosphatidylserine (●), 80 μg/mL mixture of phosphatidylserine/sodium deoxycholate in a ratio of 1/9 (w/w) (◄), or 10 μg/mL sodium deoxycholate (■).

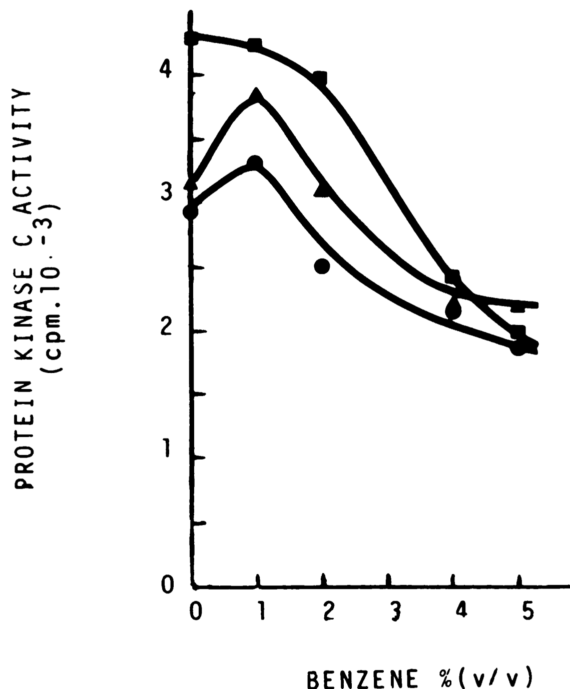

FIGURE 5. Opposing effects of benzene on protein kinase C activity as a function of sodium deoxycholate concentrations. The enzyme was assayed in 0.1 mM $CaCl_2$ as described in Roghani et al. (*14*). Sodium deoxycholate was added at the concentrations of 2 µg/mL (●), 5 µg/mL (◄), or 10 µg/mL (■).

may participate in the hydrophobic environment that is required for maximal enzyme activation. As pointed out recently in several reports, the interaction between enzyme, phospholipid, and substrate is a complex one (*16,17*). The present data suggest that bile salts and benzene may help to elucidate how a hydrophobic domain can activate the enzyme.

Implication of Protein Kinase C in Tumor Promotion

Is tumor promotion associated with protein kinase C activation and more generally with aberrant signal transduction? In order to explore this possibility, several tumor promoters have been tested by us and others (*9*).

Tumor promoters from the series of teleocidin, lyngbyatoxin, and aplysiatoxin isolated from *Lyngbya majuscula* or *Streptomyces mediocidicus* in Sugimura's laboratory (*18*), have been revealed as potent activators of protein kinase C and compete on the phorbol ester binding site. In addition, unsaturated fatty acids, retinoic acid, and lipoxin A are able to activate *in vitro* protein kinase C; this reaction does not require phospholipid. Interestingly, unsaturated fatty acids also enhance enzyme activity in intact platelets (unpublished results). As previously mentioned, solutions of the bile salt sodium deoxycholate potently activated the enzyme without any requirement for phospholipid. Likewise, chloroform and other chlorine-substituted methane derivatives activate protein kinase C *in vitro* and in platelets. These drugs did not compete for the phorbol ester binding site and the *in vitro* reaction requires phospholipid although it was not dependent on Ca^{2+}. Thus, chloroform and its derivatives most likely act in a benzene-like fashion on phospholipid–enzyme interaction.

In summary, several tumor promoters can activate protein kinase C, presumably through interactions at different sites of the molecule. In contrast, well-characterized tumor promoters such as anthralin, tetrachlorodibenzodioxin, phenobarbital, palytoxin, benzoyl peroxyde, and 1,25-dihydroxyvitamin D3 do not directly activate the enzyme. However, the possibility that these tumor promoters act downstream from protein kinase C or through the Ca^{2+} branch of the IP3/DG pathway is currently under investigation.

Discussion and Conclusion

The present results provide evidence that benzene and toluene activate in platelets, as well as in the *in vitro* assay, protein kinase C alone, or in the presence of phorbol ester TPA. The site of action of these organic solvents on the enzyme are presumably different. What is the physiological relevance of these findings? Is benzene-mediated protein kinase C activation relevant in its effect on malignancy?

The effects of benzene on cell proliferation were tested on the fibroblastic cell line Balb/c-3T3. Preliminary results have shown an initial marked increase of ^3H-thymidine incorporation into DNA, up to 0.25%, followed by inhibition of growth at higher concentrations. Phorbol ester TPA at low doses has similar effects on this cell line, suggesting that benzene may similarly affect the IP3/DG pathway.

The concentrations used in our experiments are generally higher than the low daily doses used in experimental carcinogenesis. However, it should be stressed that the real amount of benzene intervening under our conditions is much less, since most of the added benzene is bound to the tube or the dish walls.

Taking phorbol ester-mediated protein kinase C activation as a model, by analogy it is hypothesized that benzene may trigger an enzyme activation qualitatively distinct from that mediated by physiological stimuli. As reviewed by Hunter (*19*), the site of action of oncogene proteins is probably associated with signaling pathways and most likely with the IP3/DG pathway. It is highly probable that the elucidation of how oncogene proteins and tumor promoters cooperate in signal transduction will considerably help to understand how a cell progresses to transformation.

The authors thank Drs. B. Mely and L. Daya Grosjean for helpful suggestions. This work was supported by the Fondation pour la Recherche Medicale.

REFERENCES

1. Berenblum, I. The cocarcinogenic actions of croton resin. Cancer Res. 1: 44–48 (1941).
2. Mottram, J. C. A developing factor in developing blastogenesis. J. Pathol. Bacteriol. 56: 181–187 (1944).

3. Kennedy, A. R., Cairns, J., and Little, J. B. Timing of the steps in transformation of C^3H-10T½ by X-irradiation. Nature 307: 85-86 (1984).
4. Land, H., Parada, L. F., and Weinberg, R. A. Tumorigenic conversion of primary embryo fibroblasts requires at least two cooperating oncogenes. Nature 304: 596-602 (1983).
5. Boutwell, R. K. The function and mechanism of promoters of carcinogenesis. CRC Crit. Rev. Toxicol. 2: 419-422 (1974).
6. Driedger, P., and Blumberg, P. M. Specific binding of phorbol ester tumor promoters. Proc. Natl. Acad. Sci (U.S.) 77: 567-577 (1980).
7. Kikkawa, V., Kaibuchi, K., Castagna, M., Yamanishi, J., Kimihiko, S., Tanaka, Y., Miyake, R., Takai, Y., and Nishizuka, Y. Protein phosphorylation and mechanism of tumor-promoting phorbol esters. In: Advances in Cyclic Nucleotide and Protein Phosphorylation Research, Vol. 17 (P. Greengard, G. A. Robison, R. Paoletti, and S. Nicosia, Eds.), Raven Press, New York, 1984, pp. 437-455.
8. Nishizuka, Y. The role of protein kinase C in cell surface signal transduction and tumor promotion. Nature 308: 693-691 (1984).
9. Castagna, M. Phorbol esters as signal transducers and tumor promoters. Biol. Cell 59: 3-14 (1987).
10. Berridge, M. J. Inositol triphosphate and diacylglycerol as second messengers. Biochem. J. 220: 345-360 (1984).
11. Nishizuka, Y. Studies and perspectives of protein kinase C. Science 233: 305-312 (1986).
12. Yamanishi, J., Takai, Y., Kaibuchi, K., Sano, K., Castagna, M., and Nishizuka, Y. Synergistic functions of phorbol ester and calcium in serotonine release. Biochem. Biophys. Res. Commun. 112: 778-786 (1983).
13. Dean, B. J. Recent findings on the genetic toxicology of benzene, toluene, xylenes and phenols. Mutat. Res. 154: 153-181 (1985).
14. Roghani, M., Da Silva, C., Guvelli, D., and Castagna, M. Benzene and toluene activate protein kinase C. Carcinogenesis 8: 1105-1107 (1987).
15. Aschendel, C. L., Staller, J. M., and Boutwell, R. K. Identification of a calcium and phospholipid-dependent phorbol ester binding activity in the soluble fraction of mouse tissues. Biochem. Biophys. Res. Commun. 121: 340-345 (1983).
16. Bazzi, M. D., and Nelsestuen, G. L., Role of substrate in determining, the phospholipid specificity of protein kinase C activation. Biochemistry 26: 5002-5008 (1987).
17. Nakadate, T., Jeng, A. Y., and Blumberg, P. Effect of phospholipid on substrate phosphorylation by a catalytic fragment of protein kinase C. J. Biol. Chem. 262: 11507-11513 (1987).
18. Sugimura, T. Studies on environmental chemical carcinogenesis in Japan. Science 233: 312-318 (1986).
19. Hunter, T. Oncogenes and growth control. Trends Biochem. Sci. 6: 275-280 (1985).