



Mobilization of rat stomach ECL-cell histamine in response to short- or long-term treatment with omeprazole and/or YF 476 studied by gastric submucosal microdialysis in conscious rats

¹T. Konagaya, ¹M. Bernsand, ¹P. Norlén & *¹R. Håkanson

¹Department of Pharmacology, Institute of Physiological Sciences, University of Lund, Sölvegatan 10, S-223 62 Lund, Sweden

1 Mobilization of histamine from the ECL cells was monitored by gastric submucosal microdialysis in conscious rats. The ECL cells are known to operate under gastrin control and the purpose of the present study was to examine their *in situ* response to short-term (12 h) as well as long-term (28 days) hypergastrinaemia, induced by treatment with the proton pump inhibitor omeprazole.

2 Hypergastrinaemia promptly raised the histamine concentration in the microdialysate. The effect was prevented by CCK₂ receptor blockade (YF476). On day 7 of omeprazole treatment the microdialysate histamine concentration reached a peak, five times higher than before treatment. Subsequently (14 and 28 days), less histamine was mobilized.

3 Gastrin infusion (4 h) raised the microdialysate histamine concentration in a dose-dependent manner in fasted rats and freely fed rats and in rats treated with omeprazole for a week. However, while fasted and fed rats responded to low doses of gastrin, the omeprazole-treated rats required large doses of gastrin to respond.

4 When the amount of histamine mobilized was related to the serum gastrin concentration the following EC₅₀ values could be calculated: fasted rats 2.3×10^{-10} M, freely fed rats 2.5×10^{-10} M, omeprazole-treated rats 8.7×10^{-10} M. The maximal histamine responses in the three groups were $18.4 \text{ pmol } 4 \text{ h}^{-1} \pm 0.8$, $21.9 \text{ pmol } 4 \text{ h}^{-1} \pm 1.2$ and $68.0 \text{ pmol } 4 \text{ h}^{-1} \pm 3.5$, respectively.

5 The results suggest that ECL cells, exposed to a high gastrin concentration for a week, respond with a shift in the receptor-ligand binding affinity from high to low. Apparently, CCK₂ receptors of the ECL cells are subject to dynamic changes with respect to ligand-binding affinity.

British Journal of Pharmacology (2001) **133**, 37–42

Keywords: ECL cells; gastrin; CCK₂ receptors; CCK₂ receptor antagonist; histamine; histamine mobilization; omeprazole

Abbreviations: CCK, cholecystokinin; CCK₂ receptor, cholecystokinin 2-type receptor; CI, confidence interval; ECL cell, enterochromaffin-like cell; OPZ, omeprazole; YF, YF476, a CCK₂ receptor blocker

Introduction

The ECL cells constitute the predominant endocrine/paracrine cell type in the oxyntic mucosa of the rat stomach (Håkanson *et al.*, 1994; Chen *et al.*, 1999). They produce and secrete histamine in response to gastrin. In fact, it is believed that gastrin stimulates acid secretion by mobilizing ECL-cell histamine that in turn stimulates histamine H₂ receptors on adjacent parietal cells (Waldum *et al.*, 1991; Andersson *et al.*, 1996).

We have recently implemented a technique for gastric submucosal microdialysis which enables us to monitor the secretion of ECL-cell histamine in intact, conscious rats (Kitano *et al.*, 2000a). The aim of the present study was to use this technique to examine the mobilization of ECL-cell histamine in conscious rats with either hypo- or hypergastrinemia. Hypogastrinemia was induced by food deprivation. Hypergastrinemia was induced by continuous intravenous infusion of gastrin or by treatment with the proton pump inhibitor omeprazole, which causes hypergastrinemia by eliminating the acid feedback inhibition of gastrin release (Ryberg *et al.*, 1989). The receptor responsible for the effect

of gastrin on the ECL cells is of the cholecystokinin 2-type (CCK₂) (Sandvik & Waldum, 1991; Prinz *et al.*, 1994; Ding *et al.*, 1997a; Chen *et al.*, 2000). The effect of CCK₂-receptor blockade was investigated by the use of YF476, a potent and selective CCK₂ receptor antagonist, which is known to prevent gastrin from mobilizing ECL-cell histamine (Lindström *et al.*, 1999).

Methods

Animals

Male-Sprague-Dawley rats, weighing 250–300 g, were used in this study. The rats were housed in plastic cages under conditions of controlled temperature, humidity and 12:12 h light–dark cycle. They received standard laboratory chow and tap water *ad libitum*. Before the experiments, the rats had been thoroughly familiarized with Bollman-type restraining cages for at least 2 weeks. All rats were equipped with a microdialysis probe (MAB 3.8.10, membrane length 10 min, outer diameter 0.57 mm, cut-off 35 kDa, AgnTho's AB, Lidingö, Sweden), inserted in the submucosa of the acid-

*Author for correspondence.

producing part of the stomach as previously described by Kitano *et al.* (2000a). At the same time the rats were fitted with an indwelling catheter in the right jugular vein. After implantation of the microdialysis probe, the rats were left to recover for 3 days in individual wire-mesh-bottom cages with free access to food and water. When rats were to be fasted (48 h) they were denied food but had free access to water. During sampling from the microdialysis probes (see below), they were kept in Bollman-type restraining cages. Blood samples for the determination of serum gastrin were drawn repeatedly from the tail. After completion of the study, the animals were killed by an overdose of chloral hydrate intraperitoneally. The experimental protocol was approved by the local Animal Welfare Committee of Lund/Malmö.

Drugs

Human Leu¹⁵-gastrin-17 was purchased from Research Plus, Bayonne, NJ, U.S.A. It was dissolved in 0.9% saline, containing 0.1% bovine serum albumin (ICN, Aurora, OH, U.S.A.). The proton pump inhibitor omeprazole was obtained from AstraZeneca, Mölndal, Sweden and dissolved in 0.25% Methocel (Dow Corning, Midland, MI, U.S.A.). Omeprazole or vehicle was administered once daily by oral gavage (400 mmol kg⁻¹ day⁻¹ (Larsson *et al.*, 1986) between

0800 and 1000 h. The CCK₂ receptor antagonist YF476, kindly provided by Dr A Harris (Ferring, Vanlose, Denmark), was dissolved in polyethylene glycol 300 (Acros Organics, Geel, Belgium). YF476 (or vehicle) was given by a single subcutaneous injection of 300 mmol kg⁻¹, a dose known to cause sustained CCK₂ receptor blockade for at least a month (Kitano *et al.*, 2000b).

Microdialysis

Microdialysis was performed 3 days after implantation of the probe. All rats were conscious throughout the experiment since anaesthesia is known to suppress ECL-cell histamine mobilization (Nörlén *et al.*, 2000). The inlet tube of the microdialysis probe was connected to a microinfusion pump and the outlet tube was allowed to drain in 300 µl polyethylene vials. Perfusion of the microdialysis probe with 0.9% saline (72 µl h⁻¹) started at 0800 h. After 2 h equilibration, collection of microdialysate commenced.

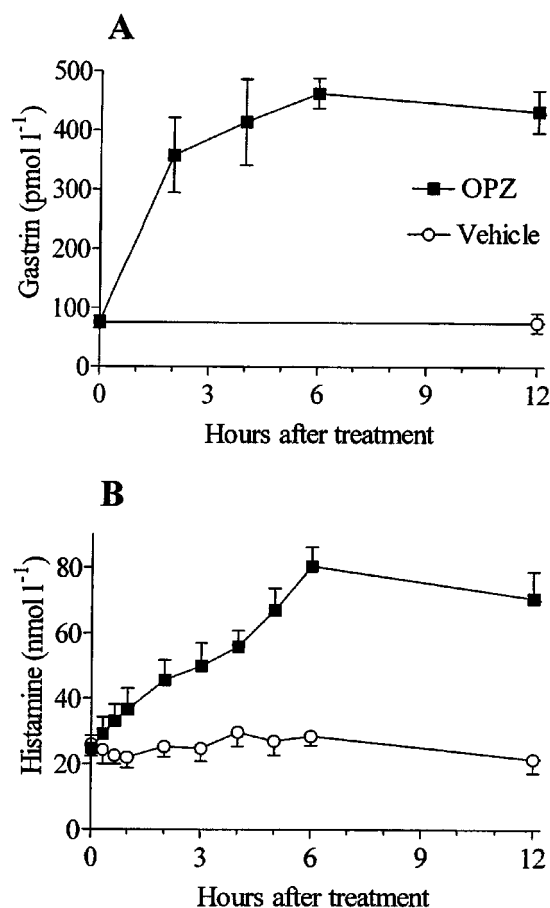


Figure 1 Serum gastrin (A) and microdialysate histamine (B) concentrations were monitored for 12 h after the administration of a single oral dose of omeprazole (OPZ, 400 µmol kg⁻¹) or vehicle (at zero time). Means ± s.e.mean (8).

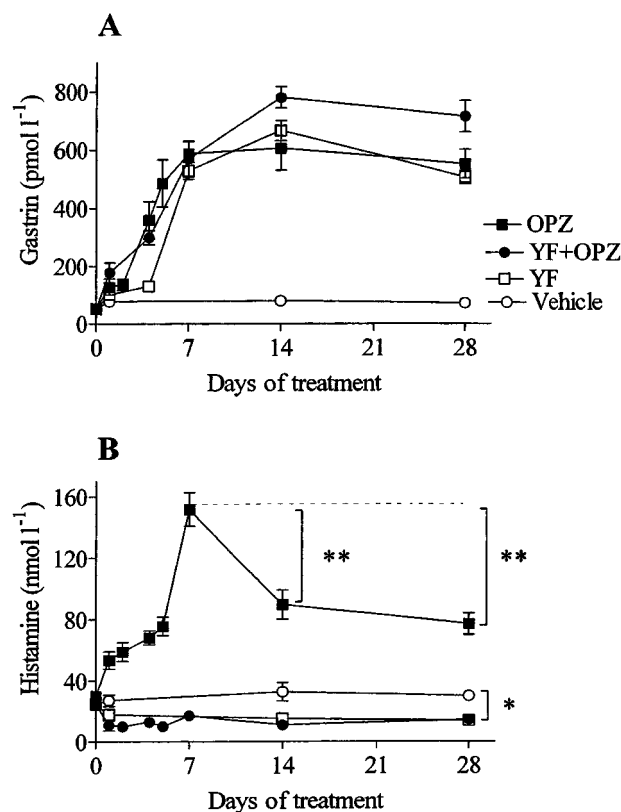


Figure 2 Serum gastrin (A) and microdialysate histamine (B) concentrations in response to treatment with omeprazole (OPZ), YF476 (YF), OPZ+YF or vehicle (control group), starting at zero time. OPZ was given daily by the oral route, YF was deposited subcutaneously once. OPZ, YF and OPZ+YF induced a slow, progressive increase in the serum gastrin concentration for about a week. Subsequently, the elevated serum gastrin concentration remained at a plateau for the duration of the experiment. The microdialysate histamine concentration reached a peak after 7 days of OPZ treatment and declined thereafter. The concentration at day 7 was compared to the concentrations at days 14 and 28, ** $P < 0.01$, (Dunnett's test). The microdialysate histamine concentration of the control rats was reduced by treatment with YF476 (* $P < 0.05$, and the effect of OPZ was prevented by YF. Means ± s.e.mean (8–20).

Determination of gastrin and histamine

Gastrin was determined in serum by radioimmunoassay (Stadil & Rehfeld, 1973). Histamine was determined in the microdialysate samples by radioimmunoassay (Morel & Delaage, 1988). The radioimmunoassay kit was from Immunotech, Marseille, France.

Experimental design

Effect of acute omeprazole Sixteen freely fed rats were used. After equilibration for 2 h, two microdialysis samples were collected with hourly intervals. After the first two microdialysate samples (basal) had been collected, omeprazole or vehicle was administered. Microdialysate samples were then collected every 20 min in the first hour and then hourly for 6 h. After collecting the 6 h samples the rats were returned to their cages until they were again placed in the Bollman cages 1 h before collecting the 12 h samples. Blood samples for determining the serum gastrin concentration were drawn from the tail 0, 2, 4, 6 and 12 h after giving omeprazole or vehicle.

Effect of long-term omeprazole and/or YF476 One hundred and forty-eight fed rats were allocated to four groups; omeprazole treatment, YF476 treatment, treatment with omeprazole + YF476 or vehicle (controls). Rats received

omeprazole (400 mmol kg^{-1}) or vehicle orally every day (between 0900 and 1100 h) except on the day of sampling. Treatment started on day 0 and was discontinued after 1, 2, 4, 5, 7, 14 or 28 days. YF476 (300 mmol kg^{-1}) was given as a single subcutaneous injection on day 0. Histamine in the microdialysate samples and gastrin in the serum were measured (6–8 rats in each group). Serum and microdialysate samples were collected between 0900 and 1100 h. Each rat was used for sampling once only.

Gastrin dose-response curves in hypo-, normo- and hypergastrinemic rats One hundred and sixteen rats were allocated to three groups: fasted rats, freely fed rats and rats treated with omeprazole for 1 week. Omeprazole was not given on the day of the experiment. The rats were placed in Bollman cages at 0800–0900 h with free access to food and water during the experiment (except the fasted rats which were denied food). After collecting basal microdialysate samples for 2 h, continuous intravenous infusion of increasing doses of human Leu¹⁵-gastrin-17 ($0.15, 0.5, 1.5, 5, 15, 50, 150$ or $500 \text{ nmol kg}^{-1} \text{ h}^{-1}$, 1 ml h^{-1}) was initiated. Microdialysate samples were collected every 20 min during the first hour and then hourly for the next 3 h. The basal serum gastrin concentration was determined in samples collected from the tail vein during the equilibration period preceding the first sampling of microdialysate, while the serum gastrin concentration at the conclusion of the 4-h gastrin infusion

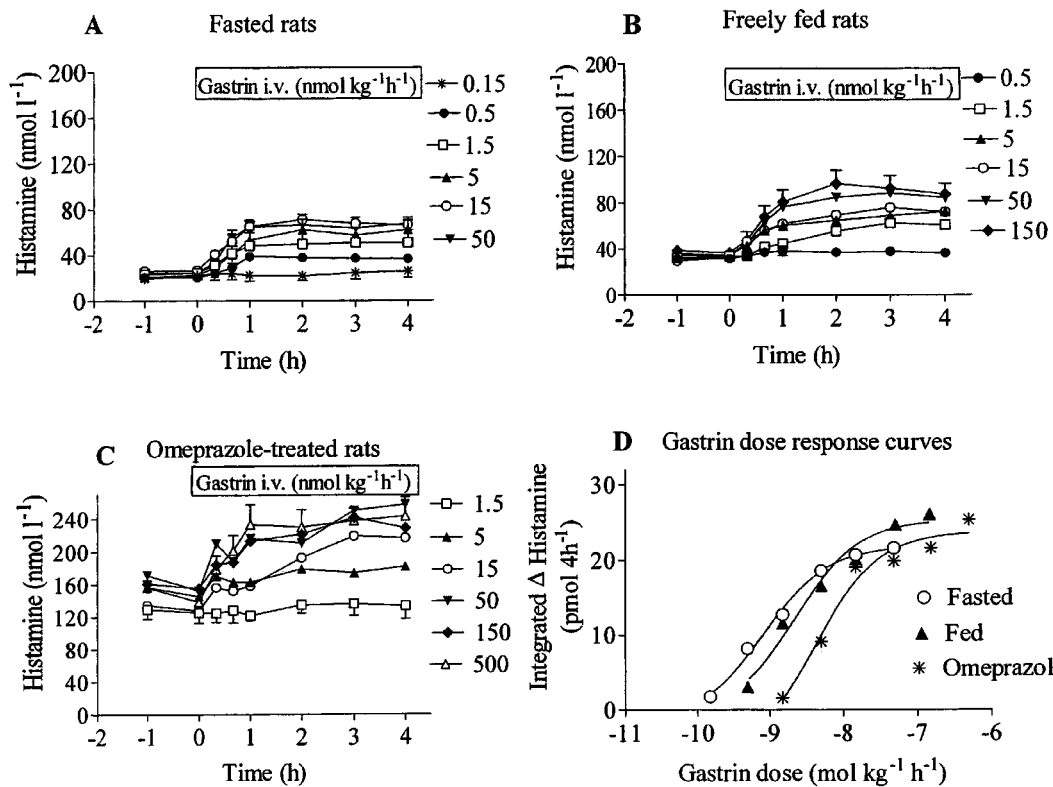


Figure 3 Gastrin-evoked mobilization of histamine in fasted rats (A), freely fed rats (B) and rats treated with omeprazole daily for 1 week (C). Omeprazole was given by the oral route. Gastrin-17 was given by continuous intravenous infusion in doses as indicated. The pre-gastrin level of histamine in the microdialysate of freely fed rats ($34.3 \text{ nmol l}^{-1} \pm 1.5$) was 1.5 times higher than in fasted animals ($22.7 \text{ nmol l}^{-1} \pm 0.1$); in omeprazole-treated rats the pre-gastrin histamine level ($145 \text{ nmol l}^{-1} \pm 6.2$) was seven times higher than in the fasted rats. Means \pm s.e. mean (36–48). Gastrin dose-response curves (D) were constructed (using the GraphPad PRISM program) for the three groups of rats based on 4-h integrated increments in histamine.

was determined after the last microdialysate sample had been collected. The amounts of histamine mobilized are expressed as integrated increment of histamine in the microdialysate over a 4 h time period of gastrin infusion or as integrated total histamine output in the microdialysate during the same time period.

Statistical analysis

All values are expressed as mean \pm s.e.mean. Analysis by ANOVA was followed by Dunnett's *t*-test for unpaired data. *P* values of <0.05 were considered significant. Dose-response curves, concentration-response curves, EC_{50} values (i.e. the concentration producing 50% of the maximal effect), and the 95% confidence interval (CI) were constructed/calculated using a GraphPad PRISM program (version 3.00, GraphPad Software, San Diego, CA, U.S.A.).

Results

Effects of a single dose of omeprazole

Changes in the microdialysate histamine concentration and the serum gastrin concentration were monitored for 12 h after the administration of a single dose of omeprazole. The histamine concentration increased until a plateau was reached after 6 h (3 fold increase). The changes in serum gastrin preceded those in microdialysate histamine with a few hours (Figure 1).

Effects of long-term treatment with omeprazole and/or YF476

Daily omeprazole treatment gradually raised the serum gastrin concentration until a plateau was reached after about 7 days (15 fold elevation). Also the histamine concentration in the gastric submucosal microdialysate increased until day 7 when it reached a peak ($151 \text{ nmol l}^{-1} \pm 10$), about five times higher than before treatment (Figure 2). On day 14 and 28, there was a decline in the amount of histamine mobilized; at this stage the histamine concentration was about three times higher than before treatment ($89 \text{ nmol l}^{-1} \pm 9$). Treatment with YF476 raised the serum gastrin concentration to the same level as after omeprazole and lowered the amount of histamine mobilized compared to vehicle-treated controls already a few days after the start of the experiment. On day 28, the YF476-treated rats had a microdialysate histamine concentration of $15 \text{ nmol l}^{-1} \pm 2$; this value should be compared with that in vehicle-treated controls ($32.6 \text{ nmol l}^{-1} \pm 6.1$) ($P < 0.05$). YF476 also prevented the omeprazole-induced increase in microdialysate histamine (Figure 2).

Gastrin dose-response curves in hypo-, normo- and hypergastrinemic rats

Fasted rats had lower serum gastrin concentration ($11.0 \text{ pmol l}^{-1} \pm 0.7$, hypogastrinemia) than freely fed rats ($76.6 \text{ pmol l}^{-1} \pm 6.7$, normogastrinemia). Rats treated with omeprazole for 1 week had greatly elevated serum gastrin concentration ($589 \text{ pmol l}^{-1} \pm 40$, hypergastrinemia). Gastrin

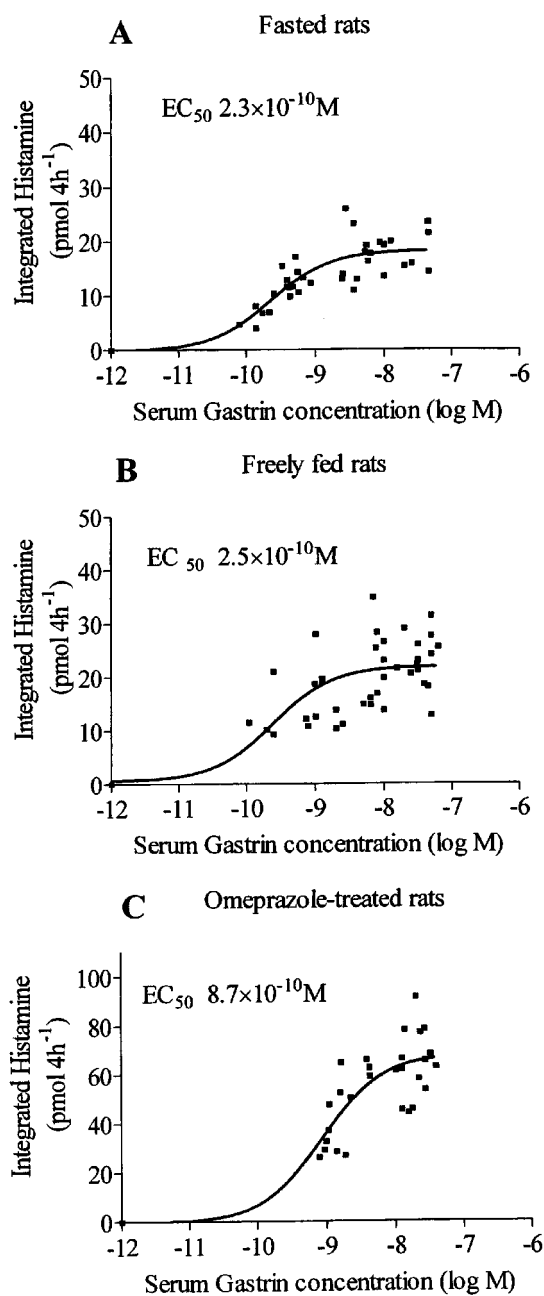


Figure 4 Gastrin concentration-response curves in hypo- (fasted, A), normo- (freely fed, B) and hypergastrinemic (1-week omeprazole treatment, C) rats. The curves were constructed from the serum gastrin and microdialysate histamine concentrations of the experiments in Figure 3 and from the assumption that no histamine would be released from the ECL cells at a gastrin concentration of 10^{-12} M. The EC_{50} values were calculated by identifying the gastrin concentration at the point of intersection of the concentration-response curve with the half-maximal level of histamine release. The EC_{50} values were 2.3×10^{-10} M in the fasted rats (95% confidence interval, CI: 1.3×10^{-10} - 3.0×10^{-10}), 2.5×10^{-10} M in the freely fed rats (CI: 9.8×10^{-11} - 6.7×10^{-10}) and 8.7×10^{-10} M in the omeprazole-treated rats (CI: 4.6×10^{-10} - 1.6×10^{-9}). The maximal histamine response was $18.4 \text{ pmol } 4 \text{ h}^{-1} \pm 0.8$, $21.9 \text{ pmol } 4 \text{ h}^{-1} \pm 1.2$ and $68.0 \text{ pmol } 4 \text{ h}^{-1} \pm 3.5$ in hypo-, normo- and hypergastrinemic rats, respectively.

infusion promptly raised the microdialysate histamine concentration in a dose-dependent manner in fasted, freely

fed and omeprazole-treated rats (Figure 3). Fasted and fed rats responded to quite low doses of gastrin, while omeprazole-treated rats required large doses of gastrin to respond. The data were used to plot the amount of histamine mobilized *versus* the serum gastrin concentration, and the EC₅₀ values were calculated (Figure 4). The EC₅₀ value in the omeprazole-treated rats was significantly higher ($P < 0.05$) than in the fasted rats, and the maximal histamine response was much higher in the omeprazole-treated rats than in the two other groups ($P < 0.05$).

Discussion

Using the technique of gastric submucosal microdialysis in conscious rats we were able to show that omeprazole mobilized gastric histamine. A single dose of omeprazole promptly raised the serum gastrin concentration and the microdialysate histamine concentration for up to 12 h. Thus, the omeprazole-evoked decrease in gastric acid secretion stimulated gastrin secretion and consequently also ECL-cell histamine mobilization. Long-term omeprazole treatment induced a progressive rise in the serum gastrin concentration until a plateau was reached 7 days after start of treatment. In this experiment serum samples were collected shortly before giving the subsequent dose, and the slow progressive rather than immediate rise in serum gastrin probably reflects the fact that it takes time to build up long-lasting acid inhibition by omeprazole given once a day. On the whole, the microdialysate histamine concentration changed according to the serum gastrin concentration with an initial slow rise and a peak after 7 days. That omeprazole-evoked mobilization of ECL-cell histamine depends on gastrin was supported by the finding that the CCK₂ receptor antagonist YF476 prevented the histamine response to omeprazole. Gastrin interacts with two distinct receptor types, referred to as CCK₁ and CCK₂, which have been cloned and characterized (Kopin *et al.*, 1992; Wank *et al.*, 1994). ECL cells, which represent an important gastrin target, seem to have only CCK₂ receptors (Chiba *et al.*, 1991; Asahara *et al.*, 1994; Ding & Håkanson, 1996; Ding *et al.*, 1997b,c). YF476 is a potent and quite selective CCK₂ receptor antagonist, which inhibits gastrin-induced gastric acid secretion (Kitano *et al.*, 2000b) and also prevents gastrin-induced histamine mobilization in isolated ECL cells (Lindström *et al.*, 1999) as well as in ECL cells *in situ* (Chen *et al.*, 2000; Kitano *et al.*, 2000a,b). The suppression of ECL-cell histamine mobilization in YF476-treated rats compared to vehicle-treated controls means that

the drug blocks not only the effect of omeprazole-induced hypergastrinemia but also that of physiologically secreted gastrin in freely fed rats.

The microdialysate histamine concentration is likely to reflect the activity (and histamine content) of the ECL cells. Although the mobilization of histamine was stimulated as a result of omeprazole treatment, with a peak after 7 days, the histamine response was lower after 14 and 28 days of omeprazole treatment than after 7 days. We have previously examined the time course of omeprazole-induced effects on the ECL cells in intact rats and shown the ECL-cell HDC activity to be maximally stimulated after 7 days of omeprazole treatment followed by a progressive down-regulation (Kimura *et al.*, 1997). Based on these results we suggested that sustained hypergastrinemia leads to a reduced ability of the ECL cells to respond to gastrin, perhaps through a reduced affinity of the CCK₂ receptor for gastrin. In the present study, we made an attempt to explore the mechanism behind the reduced mobilization of histamine in response to omeprazole. Gastrin was infused intravenously in increasing doses to rats with varying serum gastrin concentrations: hypo- (fasted), normo- (fed), and hypergastrinemic (omeprazole-treated) rats. Gastrin dose-response and concentration-response curves were constructed, showing that the EC₅₀ value of gastrin-induced histamine mobilization depended on the serum gastrin concentration prior to the administration of exogenous gastrin. In hypergastrinemic rats the EC₅₀ value was 3.5 times higher than in hypogastrinemic (fasted) rats ($P < 0.05$). Interestingly, the maximal histamine response was more than three times greater in the hypergastrinemic rats than in the hypo- and normogastrinemic rats. The increased histamine response (after 1 week of omeprazole treatment) does not reflect an increase in the number of ECL cells (Larsson *et al.*, 1986; Kitano *et al.*, 2000b) but reflects either an increased amount of releasable ECL-cell histamine or an increased receptor number per individual ECL cell. From the kinetic data we propose also that the CCK₂ receptors of the ECL cells change from a high to a low affinity state when exposed to high concentrations of gastrin (see also Chen *et al.*, 1998). Our observations indicate that the CCK₂ receptors of the ECL cells are subject to dynamic changes with respect to ligand-binding affinity and that the serum gastrin concentration is a controlling factor.

This study was supported by grants from the Swedish MRC (04X-1007) and the A. Pahlsson Foundation.

References

- ANDERSSON, K., CABERO, J.L., MATTSSON, H. & HÅKANSON, R. (1996). Gastric acid secretion after depletion of enterochromaffin-like cell histamine. A study with α -fluoromethylhistidine in rats. *Scand. J. Gastroenterol.*, **31**, 24–30.
- ASAHARA, M., KINOSHITA, Y., WAKATA, H., MATSUSHIMA, Y., NARIBAYASHI, Y., NAKAMURA, A., MATSUI, H., CHIHARA, K., YAMAMOTO, J., ICHIKAWA, A. & CHIBA, T. (1994). Gastrin receptor genes are expressed in gastric parietal cells and enterochromaffin-like cells of *Mastomys natalensis*. *Dig. Dis. Sci.*, **39**, 2149–2156.
- CHEN, D., ZHAO, C.-M., LINDSTRÖM, E. & HÅKANSON, R. (1999). Rat stomach ECL cells: Up-date of biology and physiology. *Gen. Pharmacol.*, **32**, 413–422.
- CHEN, D., ZHAO, C.-M., NORLÉN, P., BJÖRKQVIST, M., DING, X.-Q., KITANO, M. & HÅKANSON, R. (2000). Effect of cholecystokinin-2 receptor blockade on rat stomach ECL cells. A histochemical, electron-microscope and chemical study. *Cell. Tissue Res.*, **299**, 81–95.

- CHEN, D., ZHAO, C.-M., YAMADA, H., NORLÉN, P. & HÅKANSON, R. (1998). Novel aspects of gastrin-induced activation of histidine decarboxylase in rat stomach ECL cells. *Regul. Pept.*, **77**, 169–175.
- CHIBA, T., KINOSHITA, Y., MORISHITA, T., NAKATA, H., NAKAMURA, A. & HOSODA, S. (1991). Receptors for gastrin on gastric carcinoid tumor membrane of *Mastomys natalensis*. *Biochem. Biophys. Res. Commun.*, **177**, 739–744.
- DING, X.-Q. & HÅKANSON, R. (1996). Evaluation of the specificity and potency of a series of cholecystokinin-B/gastrin receptor antagonists *in vivo*. *Pharmacol. Toxicol.*, **79**, 124–130.
- DING, X.-Q., LINDSTRÖM, E. & HÅKANSON, R. (1997a). Time course of deactivation of rat stomach ECL cells following cholecystokinin B/gastrin receptor blockade. *Brit. J. Pharmacol.*, **122**, 1–6.
- DING, X.-Q., LINDSTRÖM, E. & HÅKANSON, R. (1997b). Cholecystokinin-B/gastrin receptor blockade suppresses the activity of rat stomach ECL cells. *Pharmacol. Toxicol.*, **81**, 19–25.
- DING, X.-Q., LINDSTRÖM, E. & HÅKANSON, R. (1997c). Evaluation of three novel cholecystokinin-B/gastrin receptor antagonists: A study of their effects on rat stomach enterochromaffin-like cell activity. *Pharmacol. Toxicol.*, **81**, 232–237.
- HÅKANSON, R., CHEN, D. & SUNDLER, F. (1994). The ECL cells. In: *Physiology of the Gastrointestinal Tract* (ed. LR Johnson) vol 2, 3rd ed. Raven Press, pp 1171–1184.
- KIMURA, K., CHEN, D., LINDSTRÖM, E., YAMADA, H., ZHAO, C.-M. & HÅKANSON, R. (1997). Functional impairment of the individual rat stomach ECL cells in response to sustained hypergastrinemia. *Regul. Pept.*, **72**, 69–77.
- KITANO, M., NORLÉN, P., DING, X.-Q., NAKAMURA, S. & HÅKANSON, R. (2000b). Long-lasting cholecystokinin-2 receptor blockade after a single subcutaneous injection of YF476 or YM022. *Brit. J. Pharmacol.*, **130**, 699–705.
- KITANO, M., NORLÉN, P. & HÅKANSON, R. (2000a). Gastric submucosal microdialysis: a method to study gastrin- and food-evoked mobilization of ECL-cell histamine in conscious rats. *Regul. Pept.*, **86**, 113–124.
- KOPIN, A.S., LEE, Y.-M., MCBRIDE, E.W., MILLER, L.J., LU, M., LIN, H.Y., KOLAKOWSKI, F. & BEINBORN, M. (1992). Expression cloning of the canine parietal cell gastrin receptor. *Proc. Natl. Acad. Sci. U.S.A.*, **89**, 3605–3609.
- LARSSON, H., CARLSSON, E., MATTSSON, H., LUNDELL, L., SUNDLER, F., SUNDELL, G., WALLMARK, B., WATANABE, T. & HÅKANSON, R. (1986). Plasma gastrin and gastric enterochromaffin-like cell activation and proliferation. Studies with omeprazole and ranitidine in intact and antrectomized rats. *Gastroenterology*, **20**, 391–399.
- LINDSTRÖM, E., BJÖRKQVIST, M., DING, X.-Q. & HÅKANSON, R. (1999). Pharmacological analysis of CCK₂ receptor antagonists using isolated ECL cells. *Brit. J. Pharmacol.*, **127**, 530–536.
- MOREL, A.M. & DELAAGE, M.A. (1988). Immunoanalysis of histamine through a novel chemical derivatization. *J. Allergy Clin. Immunol.*, **82**, 646–654.
- NORLÉN, P., KITANO, M., LINDSTRÖM, E. & HÅKANSON, R. (2000). Anaesthetic agents inhibit gastrin-stimulated but not basal histamine release from rat stomach ECL cells. *Brit. J. Pharmacol.*, **130**, 725–730.
- PRINZ, C., SCOTT, D.-R., HURWITZ, D., HELANDER, H.F. & SACHS, G. (1994). Gastrin effects on isolated rat enterochromaffin-like cells in primary culture. *Am. J. Physiol.*, **267**, G663–G672.
- RYBERG, B., MATTSSON, H., LARSSON, H. & CARLSSON, E. (1989). Correlation between inhibition of gastric acid secretion, plasma gastrin and oxyntic mucosal histidine decarboxylase activity in the rat. *Scand. J. Gastroenterol.*, **24**, 287–292.
- SANDVIK, A.K. & WALDUM, H.L. (1991). CCK-B (gastrin) receptor regulates gastric histamine release and acid secretion. *Am. J. Physiol.*, **260**, G925–G928.
- STADIL, F. & REHFELD, J.F. (1973). Determination of gastrin in serum. An evaluation of the reliability of a radioimmunoassay. *Scand. J. Gastroenterol.*, **8**, 101–112.
- WALDUM, H.L., SANDVIK, A.K., BRENNAN, E. & PETERSEN, H. (1991). The gastrin-histamine sequence in the regulation of gastric acid secretion. *Gut*, **32**, 698–700.
- WANK, S.A., PISEGNA, J.R. & DE WEERTH, A. (1994). Cholecystokinin receptor family. Molecular cloning, structure and functional expression in rat, guinea pig and human. *Ann. N.Y. Acad. Sci.*, **713**, 49–66.

(Received December 11, 2000

Revised February 7, 2001

Accepted February 13, 2001)