www.nature.com/bjp

Prostanoid EP_1 - and TP-receptors involved in the contraction of human pulmonary veins

¹Laurence Walch, ²Vincent de Montpreville, ¹Charles Brink & ^{*,1}Xavier Norel

¹CNRS ESA 8078, 133 av. de la résistance, 92350 Le Plessis-Robinson, France and ²Laboratoire Anatomopathologique, Centre Chirurgical Marie Lannelongue, 133 av. de la résistance, 92350 Le Plessis-Robinson, France

1 To characterize the prostanoid receptors (TP, FP, EP_1 and/or EP_3) involved in the vasoconstriction of human pulmonary veins, isolated venous preparations were challenged with different prostanoid-receptor agonists in the absence or presence of selective antagonists.

2 The stable thromboxane A₂ mimetic, U46619, was a potent constrictor agonist on human pulmonary veins ($pEC_{50}=8.60\pm0.11$ and $E_{max}=4.61\pm0.46$ g; n=15). The affinity values for two selective TP-antagonists (BAY u3405 and GR32191B) versus U46619 were BAY u3405: $pA_2=8.94\pm0.23$ (n=3) and GR32191B: apparent $pK_B=8.25\pm0.34$ (n=3), respectively. These results are consistent with the involvement of TP-receptor in the U46619 induced contractions.

3 The two EP₁-/EP₃- agonists (17-phenyl-PGE₂ and sulprostone) induced contraction of human pumonary veins (pEC₅₀=8.56±0.18; $E_{max}=0.56\pm0.24$ g; n=5 and pEC₅₀=7.65±0.13; $E_{max}=1.10\pm0.12$ g; n=14, respectively). The potency ranking for these agonists: 17-phenyl-PGE₂ > sulprostone suggests the involvement of an EP₁-receptor rather than EP₃. In addition, the contractions induced by sulprostone, 17-phenyl-PGE₂ and the IP-/EP₁- agonist (iloprost) were blocked by the DP-/EP₁-/EP₂-receptor antagonist (AH6809) as well as by the EP₁ antagonist (SC19220).

4 $PGF_{2\alpha}$ induced small contractions which were blocked by AH6809 while fluprostenol was ineffective. These results indicate that FP-receptors are not implicated in the contraction of human pulmonary veins.

5 These data suggest that the contractions induced by prostanoids involved TP- and EP_1 -receptors in human pulmonary venous smooth muscle.

British Journal of Pharmacology (2001) 134, 1671-1678

Keywords: AH6809; BAY u3405; contraction; human pulmonary veins; GR32191B; iloprost; prostaglandin; prostanoid receptors; SC19220; sulprostone; U46619

Abbreviations: E_{max}: maximal contraction; K_B values: equilibrium dissociation constant for the antagonist; L-NOARG: N^Gnitro-L-arginine; NC: not calculable; PG: prostaglandin

Introduction

Prostanoids may contract or relax smooth muscle by activating different prostanoid receptors (Coleman et al., 1994). Results derived from isolated tissues showed that prostanoid activation of TP-, FP-, EP1- or EP3-receptors produced smooth muscle contraction by increasing Ca²⁺ or reducing cyclic AMP intracellular levels (Negishi et al., 1995). The preferential receptor for thromboxane A₂ (TPreceptor) has been extensively described in platelet aggregation and in smooth muscle contraction (Shen & Tai, 1998). In most of the human arteries, U46619 the TP selective agonist induced contraction (Maddox et al., 1985; Ohlstein et al., 1988; Uski et al., 1984; Baxter et al., 1995; Templeton et al., 1991), while studies on human veins have been rarely reported. However, the TP-receptor has been described in vasoconstriction of veins in the hand, the placenta and the leg (Arner et al., 1991; Boura et al., 1986; Mais et al., 1985). The involvement of FP-, EP1- or EP3-receptors in the contraction of numerous non-human smooth muscle preparations is frequently reported. The activation of FP-

receptors by prostanoids induced contraction of cat iris sphincter, bovine ciliary muscle, rabbit uterus and ewe myometrium (Woodward et al., 1989; Krauss et al., 1997; Chen et al., 1998; Crankshaw & Gaspar, 1995). In mammals, activation of EP3-receptor induced contraction of smooth muscles present in ileum, colon, myometrium and corpus luteum (Botella et al., 1993; 1995; Crankshaw & Gaspar, 1995; Sharif et al., 1998). The EP1-receptor has been classically described using selective antagonist (SC19220) against PGE₂-induced contractions in preparations derived from either the guinea-pig trachea or gastrointestinal tract (Kennedy et al., 1982; Coleman & Kennedy, 1985). However, few studies characterizing FP-, EP1- or EP3- receptors involved in the control of human vascular tone have been reported. The FP-receptor has been described in different non-vascular human smooth muscle: urinary bladder, and myometrium (Palea et al., 1998; Senior et al., 1992). Furthermore, EP₁-receptors have not been associated with any human smooth muscle contraction and there is limited data on human pulmonary artery and myometrium contractions due to EP₃-receptor activation (Qian et al., 1994; Senior et al., 1993).

^{*}Author for correspondence: E-mail: xnorel@hotmail.com

The aim of the present study was to investigate not only the TP but also the FP-, EP₁- or EP₃-receptors associated with the contractions induced by prostanoids in human pulmonary veins.

Methods

Isolated preparations

Human lung tissues were obtained from patients (21 male and four female) who had undergone surgery for lung carcinoma. The mean age was 58 ± 2 years. Pulmonary venous preparations were removed, dissected free from adjoining connective tissue and lung parenchyma, placed in Tyrode's solution (concentration mM): NaCl 139.2, KCl 2.7, CaCl₂ 1.8, MgCl₂ 0.49, NaHCO₃ 11.9, NaH₂PO₄ 0.4 and glucose 5.5; pH 7.4 and maintained at 4°C. All preparations were used within 1–12 h postsurgery. Vascular preparations with intact endothelium were cut as rings (3–6 mm internal diameter, 3–5 mm in length). The rings were then set up in 10 ml organ baths containing Tyrode's solution, gassed with 95% O₂/5% CO₂ and maintained at 37°C. An optimal load (1.5 g), which ensured maximal physiological responses to the agonists, was applied to each ring.

Changes in force were recorded by isometric force displacement transducers (Narco F-60) and physiographs (Linseis). Subsequently, preparations were allowed to equilibrate for 90 min with bath fluid changes taking place every 10 min.

Experimental protocol

After the equilibration period, the venous preparations were incubated 30 min with indomethacin (1.7 μ M) and 15 min with N^G-nitro-L-arginine (L-NOARG; 0.1 mM). These agents were used to avoid any physiological effect induced by the release of endogenous prostanoids and nitric oxide. The preparations were then contracted with increasing concentrations of prostanoids or selective agonists applied in a cumulative fashion. In some experiments, during the incubation period, different antagonists: BAY u3405 (TP), GR32191B (TP), AH6809 (DP/EP₁/EP₂) or SC19220 (EP₁) were added with indomethacin and L-NOARG. These antagonist were used to determine either their affinity values or to illustrate one response through activation of a single receptor subtype for the agonists which may act on different receptor subtypes. The maximal contraction of each preparation with norepinephrine (10 μ M) was obtained at the end of each experimental protocol.

Data analysis

The changes in force were measured from isometric recordings in grams (g). The contractions produced with the different agonists were expressed either as grams or as per cent of the contraction induced with norepinephrine. The maximal contraction (E_{max} value) produced with an agonist and the half-maximum effective concentration value (EC₅₀ value) were interpolated from the individual concentration-effect curves. The pEC₅₀ values were calculated as the negative log of EC₅₀ values. When the pEC₅₀ values obtained in the absence and presence of antagonist were significantly

different and the tentative assumption was made that the Schild equation held in our experiments, then the apparent pKB value was calculated as the negative log of the equilibrium dissociation constant for the antagonist (KB value). The K_B value was determined using the Schild equation: $K_B = [B]/(DR-1)$, where [B] is the concentration of the antagonist and DR (dose ratio) is the ratio of EC₅₀ values of agonist in the presence and absence of antagonist. In studies on veins with the contractile agonist U46619, different concentrations of BAY u3405 were used to determine the pA₂ value according to the method of Arunlakshana & Schild (1959). For each lung sample, Schild plot analysis was performed, the slope and pA₂ value were determined by least square fitting of a regression line to the points. All results are expressed as means + s.e. mean and were derived from different lung samples (n). Statistical analysis was performed using Student's paired or unpaired *t*-test and Mann-Whitney rank sum test with a confidence level of 95%.

Compounds

U46619 (9,11-dideoxy-11 α ,9 α -methanoepoxy PGF_{2 α}), PGE₂, $PGF_{2\alpha}$, fluprostenol ((±) 16-m-trifluoromethylphenoxy tetranor $PGF_{2\alpha}$) and 17-phenyl-trinor-PGE₂, were purchased from Cayman Chemical Company, Ann Arbor, MI, U.S.A. Iloprost (5-[(E)-(1S,5S,6R,7R)-7-hydroxy-6-[(E)-(3S,4RS)-3-hydroxy-4-methyl-1-octen-6-inyl]bicyclo[3.3.0]-octan-3-ylidene] pentanoic acid) and sulprostone (N-(methylsulphonyl)-9-oxo-11a,15R-dihydroxy-16-phenoxy-17,18,19,20tetranor-prosta-5Z, 13E-dien-1-amide) were a gift from Schering AG, Berlin, Germany. AH6809 (6-isopropoxy-9oxaxanthene-2-carboxylic acid) and GR32191B ([1R- $[1\alpha(z), 2\beta, 3\beta, 5\alpha]]$ -(+)-7-[5[[(1,1'-biphenyl)-4-yl] methoxy]-3-hydroxy-2-(1-piperidinyl) cyclopentyl]-4-heptenoic acid, hydrochloride) were a gift from Glaxo Wellcome, U.K. BAY u3405 (3(R)-3-(4-fluorophenylsulphonamido)-1,2,3,4-tetrahydro-9-carbazole propanoic acid) was a gift from Bayer, Stokes Poges, U.K. SC19220 (8-chlorodibenz [b,f][1,4] oxazepine-10(11H)-carboxy-(2-acetyl)hydrazide) was a gift from Searle Research and Development, Skokie, IL, U.S.A. Norepinephrine, L-NOARG (N^G-nitro-L-arginine) and indomethacin were purchased from Sigma Chemical Co., St. Louis, MO, U.S.A.

Results

The cocktail of inhibitors (indomethacin, L-NOARG) and antagonists with which the venous preparations were incubated induced a small contraction on the resting tone of these preparations (0.07 ± 0.06 g; n = 18). At the end of the protocols, venous preparations were contracted with nor-epinephrine ($10 \ \mu$ M: 3.10 ± 0.44 g; n = 25).

Concentration-dependent contractions of human pulmonary venous preparations produced by U46619 are shown Figure 1a and Table 1. U46619 was a potent constrictor with a maximal effect which was 50% greater than that induced by norepinephrine (10 μ M). BAY u3405 (0.03; 0.1; 0.3 μ M) caused a concentration-related rightward shift of the U46619 concentration-effect curves with a pA₂ of 8.94±0.23 (*n*=3) and a Schild plot slope of 1.07±0.03, which was not significantly different from unity. These values were derived from Schild plots (Figure 1b). In additional lung samples, BAY u3405 and GR32191B (1 μ M) reduced the E_{max} and/or the pEC₅₀ values obtained from U46619 concentration-effect curves (Table 1). In presence of BAY



Figure 1 (a) Contraction of human isolated pulmonary veins induced by U46619. Responses were expressed as per cent of the norepinephrine (10 μ M) contraction. Values are means ± s.e.mean. (b) Schild-plot analysis of the antagonism by BAY u3405 of U46619contractions in veins derived from three human lung samples (each lung sample is presented as a different symbol). Analysis was based on pEC₅₀ values calculated in treated preparations which were significantly different from control values. The linear regression presented was performed using all data points. The calculated average of pA₂ values, slopes and regression coefficients performed with data derived from each lung sample are indicated.

u3405 (1 μ M) the U46619 concentration-effect curves were difficult to restore and no plateau was reached even at the maximal concentration available for U46619 (10 μ M). When possible, the apparent pK_B values for each concentration of BAY u3405 and GR32191B were calculated and these values are presented Table 1.

Concentration-effect curves induced by sulprostone on human pulmonary venous preparations in the absence of antagonist were biphasic, the average curve failed to reach a plateau and was not sigmoïdal (Figure 2). The contraction obtained with the highest available concentration of sulprostone was $54\pm09\%$ (n=5) of norepinephrine contraction (Figure 2). Sigmoïdal curves were obtained with sulprostone, in the presence of either BAY u3405 (1; 10 μ M) or GR32191B (10 μ M) and the E_{max} did not exceed 50% of the norepinephrine contraction (Figure 2 and Table 1).

In presence of BAY u3405 (1 μ M), the contractions induced by sulprostone on venous preparations were inhibited by AH6809 (10; 30 or 100 µM; Figure 3a and Table 1). AH6809 (30, 100 μ M) had a non-competitive behaviour on sulprostone responses, the antagonism was not reversible. Under the same conditions (presence of BAY u3405, 1 µM), SC19220 (100 μ M) significantly inhibited the vasoconstrictions induced by sulprostone (Figure 3a). In presence of this latter treatment, the contraction obtained with the highest available concentration of sulprostone was $32\pm09\%$ (n=3) of norepinephrine contraction, the derived estimations of pEC₅₀ and apparent pK_B were ($<6.97\pm0.41$, $>4.75\pm0.36$), respectively. AH6809 (10 μ M) significantly inhibited the vasoconstrictions induced by 17-phenyl-PGE₂ in the human pulmonary veins treated with BAY u3405, 1 μ M, (Figure 3b and Table 1). In this latter protocol, the contraction obtained

| | Concentration | | Agonist | | Apparent |
|------------|---------------|----|-----------------------------------|------------------|-----------------|
| Treatment | (µM) | n | E_{max} (%) | pEC_{50} | pK_B |
| | | | U4 | 6619 | |
| Tyrode | | 15 | 146 ± 15 | 8.60 ± 0.11 | |
| BAY u3405 | 0.03 | 3 | 149 ± 04 | $7.17 \pm 0.03*$ | 8.93 ± 0.25 |
| BAY u3405 | 0.1 | 3 | 150 ± 06 | $6.35 \pm 0.18*$ | 9.24 ± 0.14 |
| BAY u3405 | 0.3 | 3 | 153 ± 08 | $6.13 \pm 0.06*$ | 8.99 ± 0.24 |
| BAY u3405 | 1 | 4 | $39 \pm 09^*$ | NC | NC |
| GR32191B | 1 | 3 | $133 \pm 06*$ | $6.14 \pm 0.08*$ | 8.25 ± 0.34 |
| | | | Sulprostone | | |
| BAY u3405 | 1 | 14 | 41 ± 04 | 7.65 ± 0.13 | |
| BAY u3405 | 10 | 6 | 37 ± 05 | 7.29 ± 0.05 | |
| GR32191B | 10 | 3 | 40 ± 10 | 7.40 ± 0.01 | |
| AH6809+B | 10 | 5 | 36+03 | 7.14+0.08* | 5.52 ± 0.31 |
| AH6809 + B | 30 | 4 | 10 + 05* | NC | _ |
| AH6809 + B | 100 | 3 | 00 + 00* | NC | |
| | | | - 17-phenyl-PGE2 | | |
| BAY u3405 | 1 | 5 | 17 ± 02 | 8.56 ± 0.18 | |
| | | | $-$ PGF ₂ α $-$ | | |
| BAY u3405 | 1 | 5 | 22 ± 04 | 7.61 ± 0.37 | |
| AH6809 + B | 10 | 5 | $01 \pm 03^*$ | NC | |
| | | | Fluprostenol | | |
| BAY u3405 | 1 | 5 | 08 ± 03 | NC | |
| | | | | | |

Table 1 Effect of TP-, EP- and FP- receptor agonists or antagonists on human isolated pulmonary venous preparations

The maximal contraction (E_{max}) produced with an agonist was expressed as per cent of the contraction induced by norepinephrine (10 μ M). See methods for definitions of pEC₅₀ or apparent pK_B values. Values are means ± s.e.mean and (*n*) indicates the number of lung samples used. NC=not calculable. *Values significantly different (Student *t*-test or Mann-Whitney rank sum test) from appropriate control (Tyrode or BAY u3405 (1 μ M)). + B indicates a co-treatment with BAY u3405 (1 μ M).

with the highest available concentration of 17-phenyl-PGE₂ was $15\pm05\%$ (n=4) of norepinephrine contraction and the derived estimations of pEC₅₀ and the apparent pK_B were ($<7.66\pm0.13$, $>5.88\pm0.20$), respectively. Venous preparations derived from three human lung samples, treated with BAY u3405 (1 μ M), contracted when challenged with iloprost (Figure 4) this response was abolished in the presence of AH6809 (10 μ M). In the presence of BAY u3405 (1 μ M), the



Figure 2 Effect of BAY u3405 (1; 10 μ M) or GR32191B (10 μ M) on the contraction of human isolated pulmonary veins induced by sulprostone. Control and treated preparations were incubated with indomethacin (1.7 μ M) and L-NOARG (0.1 mM). Responses were expressed as per cent of the norepinephrine (10 μ M) contraction. Values are means ± s.e.mean, number of lung samples used are indicated in Table 1 and in the Results section.



Figure 3 Effect of AH6809 (10; 30; 100 μ M) or SC19220 (100 μ M) on contraction of human isolated pulmonary veins induced by sulprostone (a) or 17-phenyl-PGE₂ (b). Control and treated preparations were incubated with indomethacin (1.7 μ M), L-NOARG (0.1 mM) and BAY u3405 (1 μ M). Responses were expressed as per cent of the norepinephrine (10 μ M) contraction. Values are means \pm s.e.mean, number of lung samples used are indicated in Table 1 and in the Results section.

PGE₂ concentration-effect curves were quite variable, PGE₂ either had no effect, or induced contraction or relaxation of human pulmonary venous preparations (Figure 5). These contractions induced by PGE₂ were inhibited by AH6809 (10 μ M, n=3; data not shown).

In human pulmonary veins pre-treated with BAY u3405 (1 μ M), the concentration-effect curves produced with PGF_{2 α} were small while fluprostenol failed to contract these tissues (Table 1). The PGF_{2 α} induced curves were abolished in the presence of BAY u3405 (1 μ M) and AH6809 (10 μ M; Table 1).

Discussion

The present report suggests the involvement of TP- and EP₁receptors in the prostanoid induced contraction of human pulmonary venous preparations.

The contractions observed with U46619 (stable thromboxane A2 mimetic) and their inhibition by the selective TPantagonists (GR32191B, Lumley et al., 1989 and BAY u3405, McKenniff et al., 1991) suggest the presence of a TP-receptor in human pulmonary veins. These results have not previously been reported although the TP-receptor has been frequently described in human pulmonary arteries (Maddox et al., 1985; Sjoberg & Steen, 1989; Lumley et al., 1989; Norel et al., 1991; Ellis & Muller-Schweinitzer, 1991; Qian et al., 1994; Jino et al., 1996). Pharmacological studies as well as investigations using molecular biology have suggested heterogeneity among thromboxane receptors (Lumley et al., 1989; Tymkewycz et al., 1991; Furci et al., 1991; Pierce & Regan, 1998). Actually, the affinity values calculated in human pulmonary veins for GR32191B (8.25 ± 0.34) and BAY u3405 (8.94 ± 0.23) are comparable to those found in human pulmonary arteries: GR32191B (8.18-8.3; Lumley et al., 1989; Qian et al., 1994) and BAY u3405 (9.25; Norel et al., 1991), respectively. These data suggest that the TP-receptor present in both human pulmonary arteries and veins may be the same. In addition, the apparent pKB values obtained with GR32191B (present study) were also in accordance with those derived from pharmacological studies performed on human umbilical



lloprost [-log M]

Figure 4 Representative tracing of the contraction induced by iloprost observed in isolated pulmonary veins derived from three human lung samples. Experiments were performed in presence of indomethacin (1.7 μ M), L-NOARG (0.1 mM) and BAY u3405 (1 μ M).



Figure 5 Variable effects induced by PGE₂ in human isolated pulmonary veins. Experiments were performed in presence of indomethacin (1.7 μ M), L-NOARG (0.1 mM) and BAY u3405 (1 μ M). Responses were expressed as per cent of the norepinephrine (10 μ M) contraction. Each symbol indicates data obtained with one lung sample.

artery (8.04; Templeton *et al.*, 1991), uterine artery (8.5; Baxter *et al.*, 1995) or human platelet (8.2–8.8; Lumley *et al.*, 1989).

The TP-receptor in human pulmonary veins was also activated by the high concentration (>1 μ M) of sulprostone and the TP antagonists eliminated this response. Sulprostone is classically described as a selective agonist for EP₁- and EP₃- receptors (Coleman *et al.*, 1987a, b; 1988; Reeves *et al.*, 1988), however high concentrations (30 μ M) have been reported to induce contractions of the human bronchial preparations and isolated uterine artery where TP-receptors are the only excitatory prostanoid-receptors (Coleman & Sheldrick, 1989; Baxter *et al.*, 1995). The findings (present study) are in agreement with this previous observation, high concentrations of sulprostone may act on the TP-receptor and for this reason all the experiments involving this compound were carried out in the presence of BAY u3405.

In human pulmonary veins, sulprostone and 17-phenyl-PGE₂ (in presence of BAY u3405) induced smaller contractions than the response induced by U46619. The contractions of human pulmonary veins induced by sulprostone or 17phenyl-PGE₂, suggest the involvement of EP₁- or EP₃receptors. The sensitivities (pEC₅₀ value) of the venous preparations to sulprostone and 17-phenyl-PGE₂ were comparable to those determined in standard functional assays for EP1-receptor, namely, the contractions of guineapig fundus (Coleman et al., 1987a) and trachea (Lawrence et al., 1992). In the present study, the potency ranking for these agonist, 17-phenyl-PGE₂>sulprostone (equi-effective molar ratio = 8 and 1, respectively), was similar to the one observed in the previous standard EP1-tissues. In these later tissues or in binding studies with cloned EP1-receptor (rat, Boie et al., 1997; mouse, Watabe et al., 1993; mouse, Kiriyama et al., 1997), 17-phenyl-PGE₂ was 1.5-4 fold more potent than sulprostone. A greater potency, 10 fold, was found for 17phenyl-PGE₂ when compared with sulprostone to increase intracellular Ca²⁺ in rabbit cortical collecting duct via the activation of EP₁-receptor (Guan *et al.*, 1998). In contrast, the rank order of potency for these agonists was reversed in studies where the effects were mediated by the activation of EP₃-receptors. Sulprostone was a more potent contractile agonist, 30-45 fold greater than 17-phenyl-PGE₂ either in human pulmonary artery (Qian *et al.*, 1994) or when inhibiting the twitch contraction of guinea-pig vas deferens (Lawrence *et al.*, 1992). These results (present study) support the presence of an EP₁-receptor associated with contraction in human pulmonary veins rather than an EP₃-receptor. In addition, iloprost, a selective agonist for IP and EP₁-receptors (Schrör *et al.*, 1981; Dong & Jones, 1982; Dong *et al.*, 1986) induced contraction of these preparations even though IP-receptors responsible for relaxation are present (Walch *et al.*, 1999).

The effects of either the DP-/EP1-/EP2- receptor antagonist (AH6809; Coleman et al., 1985; Keery & Lumley, 1988; Woodward et al., 1995) or the EP1-receptor antagonist (SC19220; Sanner, 1969; Kennedy et al., 1982; Coleman et al., 1987a, b) against the concentration-effect curves (present study) produced by both sulprostone and 17-phenyl-PGE₂ also suggest the involvement of the EP1-receptor rather than EP₃. However, the affinity values calculated or estimated for AH6809 in preparations derived from human lung (apparent pK_B of 5.52 and >5.88) are lower than those obtained in similar physiological experiments performed in guinea-pig or dog EP₁-preparations. Lawrence et al. (1992) using sulprostone or 17-phenyl-PGE₂ as the contractile agonist, found the AH6809 affinity values ranking from 6.1-7.35 in either guinea-pig ileum or trachea. In other studies using PGE₂ with the EP₁-preparations (guinea-pig fundus, ileum and dog fundus), the affinity values for AH6809 ranked from 6.6-7.4 (Eglen & Whiting 1988; Coleman et al., 1985). Similarly, the estimated affinity value found for SC19220 in the present study (apparent $pK_B > 4.75$) was lower than the pA_2 of 5.6 calculated in experiments performed with the previous EP1preparations (Coleman et al., 1985; Coleman & Kennedy, 1985). The reason for this discrepancy remains to be established. However, when EP1-receptors were assessed in either physiological or binding studies, the affinity values for AH6809 were always one order of magnitude greater than those for SC19220 (present study; Coleman et al., 1985; Boie et al., 1997; Funk et al., 1993). Since high concentrations of AH6809 (5-100 μM) or SC19220 (100-300 μM) did not block the EP₃ mediated effects in many of the EP₃ biological models (Table 2), the inhibitory effect of these antagonists in human pulmonary vein would suggest that the EP₃-receptor is not involved in contractions.

The variable effects induced by PGE_2 in the human pulmonary venous preparations in presence of the TPantagonist may be explained by two opposing effects of this prostaglandin in these preparations. A contraction *via* the EP₁-receptor and a relaxation *via* another EP-receptor subtype as has been suggested by Walch *et al.* (1999). A similar paradoxical effect was observed in guinea-pig trachea as well as in human bronchial preparations where PGE₂ may act on EP₁- and/or TP- receptors to induce contraction while the activation of the EP₂-receptor provokes the relaxation (Gardiner, 1975; Coleman & Kennedy, 1980; McKenniff *et al.*, 1988).

The results obtained in this report with the EP- agonists or antagonists suggest a role for EP_1 - and not EP_3 - receptor in

| <i>Biological effect mediated</i> via <i>EP</i> ₃ -receptor | <i>Tissue receiving</i> <i>agonist/antagonist</i> | Inhibition | Reference |
|--|--|------------|-------------------------|
| Presynaptique inhibition of | Guinea-pig vas deferens | no | Coleman et al., 1987b |
| norepinephrine release | Rat vena cava | no | Molderings et al., 1992 |
| | Mouse brain cortex | no | Exner & Schlicker, 1995 |
| Inhibition of gastric secretion | Rat cerebral ventricle | no | Yokotani et al., 1996 |
| - | Rat gastric mucosa | no | Reeves et al., 1988 |
| Smooth muscle contraction | Chick ileum | no | Coleman et al., 1987a,b |
| | Pig ileum | no | Botella et al., 1993 |
| | Human pulmonary artery | no | Qian et al., 1994 |

Table 2 EP3-receptor biological models: Absence of antagonism of AH6809 and SC19220

The EP₃-receptor was activated by either PGE₂, enprostil, sulprostone or 17-phenyl-PGE₂.

the contraction of human pulmonary vein. The involvement of EP₁- or EP₃- receptors in the control of vascular tone has been principally investigated in the ocular vascular bed. The EP- agonists decrease the intraocular pressure in various animal models of glaucoma (Woodward et al., 1993; 1994; Bhattacherjee et al., 1999; Waterbury et al., 1990) and contract the pig retinal vessels (Abran et al., 1994). However, the EP₃-receptor is involved in vasoconstriction of guinea-pig aorta (Jones et al., 1998), rat renal afferent arteriole (Tang et al., 2000) and human pulmonary artery (Qian et al., 1994). Arner & Högestatt (1991) showed that iloprost contracted the human hand vein. One could associate this response with activation of an EP_1 -receptor. The data (present study) demonstrated a similar response to that of the hand veins whereas human arterial preparations did not exhibit a contractile response to this agonist (Arner & Högestatt, 1991; Qian et al., 1994). Therefore, the presence of EP1receptors may be found only in human venous preparations.

The low potency of $PGF_{2\alpha}$ or fluprostenol, the selective agonist for FP-receptor, suggests that the FP-receptor is not involved in the contraction of human pulmonary veins. In

References

- ABRAN, D., VARMA, D.R., LI, D.Y. & CHEMTOB, S. (1994). Reduced responses of retinal vessels of the newborn pig to prostaglandins but not to thromboxane. *Can. J. Physiol. Pharmacol.*, **72**, 168–173.
- ARNER, M. & HÖGESTATT, E.D. (1991). Endothelium-dependent relaxation and effects of prostacyclin, endothelin and plateletactivating factor in human hand veins and arteries. *Acta Physiol. Scand.*, **142**, 165–172.
- ARNER, M., HÖGESTATT, E.D. & USKI, T.K. (1991). Characterization of contraction-mediating prostanoid receptors in human hand veins: effects of the thromboxane receptor antagonists BM13,505 and AH23848. Acta Physiol. Scand., 141, 79–86.
- ARUNLAKSHANA, O. & SCHILD, H.O. (1959). Some quantitative uses of drug antagonists. *Br. J. Pharmacol.*, **14**, 48-58.
- BAXTER, G.S., CLAYTON, J.K., COLEMAN, R.A., MARSHALL, K., SANGHA, R. & SENIOR, J. (1995). Characterization of the prostanoid receptors mediating constriction and relaxation of human isolated uterine artery. Br. J. Pharmacol., 116, 1692– 1696.
- BHATTACHERJEE, P., WILLIAMS, B.S. & PATERSON, C.A. (1999). Responses of intraocular pressure and the pupil of feline eyes to prostaglandin EP₁ and FP receptor agonists. *Invest. Ophthalmol. Vis. Sci.*, **40**, 3047–3053.
- BOIE, Y., STOCCO, R., SAWYER, N., SLIPETZ, D.M., UNGRIN, M.D., NEUSCHAFER-RUBE, F., PUSCHEL, G.P., METTERS, K.M. & ABRAMOVITZ, M. (1997). Molecular cloning and characterization of the four rat prostaglandin E2 prostanoid receptor subtypes. *Eur. J. Pharmacol.*, **340**, 227-241.

addition, the small contractions induced by $PGF_{2\alpha}$ were inhibited by AH6809 suggesting that $PGF_{2\alpha}$ activates an EP_1 -rather than FP- receptor.

In summary, the findings in the present study are consistent with the presence of TP- and EP₁- receptors mediating constriction of human pulmonary veins. These findings may be relevant to the pulmonary circulation. Sulprostone is used in obstetrics and gynaecology and one of the clinical side effects observed with this compound is pulmonary oedema (Stock *et al.*, 1995; Levy *et al.*, 1994; Puura *et al.*, 1995). The present study and the work of Qian *et al.* (1994) suggest that this side effect may involve vasospasm of the whole pulmonary vasculature. Such EP-agonist may activate at the same time TP-, EP₃- receptors in the arteries and TP-, EP₁- receptors in the veins.

The authors would like to thank Yvette Le Treut and Ginette Brille for excellent technical assistance.

- BOTELLA, A., DELVAUX, M., FIORAMONTI, J., FREXINOS, J. & BUENO, L. (1995). Receptor subtypes involved in dual effects induced by prostaglandin E2 in circular smooth muscle from dog colon. J. Pharmacol. Exp. Ther., 273, 1008–1014.
- BOTELLA, A., DELVAUX, M., FIORAMONTI, J., FREXINOS, J. & BUENO, L. (1993). Stimulatory (EP₁ and EP₃) and inhibitory (EP₂) prostaglandin E2 receptors in isolated ileal smooth muscle cells. *Eur. J. Pharmacol.*, 237, 131–137.
- BOURA, A.L., GUDE, N.M., KING, R.G., MAK, K.K. & WALTERS, W.A. (1986). Characterization of thromboxane A2 receptors in the human fetal placental vessels and umbilical vein. *Clin. Exp. Pharmacol. Physiol.*, **13**, 83–86.
- CHEN, J., WOODWARD, D.F., YUAN, Y.D., MARSHALL, K. & SENIOR, J. (1998). Prostanoid-induced contraction of the rabbit isolated uterus is mediated by FP receptors. *Prostaglandins Other Lipid Mediat.*, 55, 387–394.
- COLEMAN, R.A. & KENNEDY, I. (1980). Contractile and relaxant actions of prostaglandins on guinea-pig isolated trachea. *Br. J. Pharmacol.*, **68**, 533-539.
- COLEMAN, R.A. & KENNEDY, I. (1985). Characterisation of the prostanoid receptors mediating contraction of guinea-pig isolated trachea. *Prostaglandins*, **29**, 363-375.
- COLEMAN, R.A., DENYER, L.H. & SHELDRICK, R.L.G. (1985). AH6809, a prostanoid EP₁ receptor bloking drug. *Br. J. Pharmacol.*, **85**, 273P.
- COLEMAN, R.A., KENNEDY, I. & SHELDRICK, R.L.G. (1987a). Evidence for the existence of three subtypes of PGE2 sensitive (EP) receptors in smooth muscle. Br. J. Pharmacol., 91, 323P.

- COLEMAN, R.A., KENNEDY, I., SHELDRICK, R.L.G. & TOLOWINS-KA, I.Y. (1987b). Further evidence for the existence of three subtypes of PGE2- sensitive (EP-) receptors in smooth muscle. *Br. J. Pharmacol.*, **91**, 407P.
- COLEMAN, R.A., HUMPHRAY, J.M., SHELDRICK, R.L.G. & WHITE, B.P. (1988). Gastric antisecretory prostanoids: actions at different prostanoid receptors. *Br. J. Pharmacol.*, 95, 724P.
- COLEMAN, R.A. & SHELDRICK, R.L.G. (1989). Prostanoid-induced contraction of human bronchial smooth muscle is mediated by TP-receptors. *Br. J. Pharmacol.*, **96**, 688–692.
- COLEMAN, R.A., SMITH, W.L. & NARUMIYA, S. (1994). International union of pharmacology classification of prostanoid receptors: properties, distribution, and structure of the receptors and their subtypes. *Pharmacol. Rev.*, **46**, 205–229.
- CRANKSHAW, D.J. & GASPAR, V. (1995). Pharmacological characterization in vitro of prostanoid receptors in the myometrium of nonpregnant ewes. J. Reprod. Fertil., **103**, 55-61.
- DONG, Y.J. & JONES, R.L. (1982). Effects of prostaglandins and thromboxane analogues on bullock and dog iris sphincter preparations. *Br. J. Pharmacol.*, **76**, 149–155.
- DONG, Y.J., JONES, R.L. & WILSON, N.H. (1986). Prostaglandin E receptor subtypes in smooth muscle: agonist activities of stable protacyclin analogues. Br. J. Pharmacol., 87, 97-107.
- EGLEN, R.M. & WHITING, R.L. (1988). The action of prostanoid receptor agonists and antagonists on smooth muscle and platelets. *Br. J. Pharmacol.*, **94**, 591–601.
- ELLIS, P. & MULLER-SCHWEINITZER, E. (1991). Maintenance of functional activity of human pulmonary arteries after cryopreservation. *Br. J. Pharmacol.*, **103**, 1377–1380.
- EXNER, H.J. & SCHLICKER, E. (1995). Prostanoid receptors of the EP₃ subtype mediate the inhibitory effect of prostaglandin E2 on noradrenaline release in the mouse brain cortex. *Naunyn Schmiedebergs Arch. Pharmacol.*, **351**, 46–52.
- FUNK, C.D., FURCI, L., FITZGERALD, G.A., GRYGORCZYK, R., ROCHETTE, C., BAYNE, M.A., ABRAMOVITZ, M., ADAM, M. & METTERS, K.M. (1993). Cloning and expression of a cDNA for the human prostaglandin E receptor EP₁ subtype. *J. Biol. Chem.*, 268, 26767–26772.
- FURCI, L., FITZGERALD, D.J. & FITZGERALD, G.A. (1991). Heterogeneity of prostaglandin H2/thromboxane A2 receptors: distinct subtypes mediate vascular smooth muscle contraction and platelet aggregation. J. Pharmacol. Exp. Ther., 258, 74–81.
- GARDINER, P.J. (1975). The effects of some natural prostaglandins on isolated human circular bronchial muscle. *Prostaglandins*, **10**, 607–616.
- GUAN, Y., ZHANG, Y., BREYER, R.M., FOWLER, B., DAVIS, L., HEBERT, R.L. & BREYER, M.D. (1998). Prostaglandin E2 inhibits renal collecting duct Na+absorption by activating the EP₁ receptor. J. Clin. Invest., 102, 194–201.
- JINO, H., KURAHASHI, K., USUI, H., NAKATA, Y., SHIMIZU, Y. & TEMMA, S. (1996). Pharmacological nature of TP receptor mediated contraction in human intrapulmonary artery. *Life Sci.*, 59, 2059–2065.
- JONES, R.L., QIAN, Y.M., CHAN, K.M. & YIM, A.P. (1998). Characterization of a prostanoid EP₃-receptor in guinea-pig aorta: partial agonist action of the non-prostanoid ONO-AP-324. Br. J. Pharmacol., 125, 1288-1296.
- KEERY, R.J. & LUMLEY, P. (1988). AH6809, a prostaglandin DPreceptor blocking drug on human platelets. *Br. J. Pharmacol.*, 94, 745-754.
- KENNEDY, I., COLEMAN, R.A., HUMPHREY, P.P.A., LEVY, G.P. & LUMLEY, P. (1982). Studies on the characterisation of prostanoid receptors: a proposed classification. *Prostaglandins*, **24**, 667–689.
- KIRIYAMA, M., USHIKUBI, F., KOBAYASHI, T., HIRATA, M., SUGIMOTO, Y. & NARUMIYA, S. (1997). Ligand binding specificities of the eight types and subtypes of the mouse prostanoid receptors expressed in Chinese hamster ovary cells. *Br. J. Pharmacol.*, **122**, 217–224.
- KRAUSS, A.H., WIEDERHOLT, M., STURM, A. & WOODWARD, D.F. (1997). Prostaglandin effects on the contractility of bovine trabecular meshwork and ciliary muscle. *Exp. Eye Res.*, 64, 447-453.

- LAWRENCE, R.A., JONES, R.L. & WILSON, N.H. (1992). Characterization of receptors involved in the direct and indirect actions of prostaglandins E and I on the guinea-pig ileum. Br. J. Pharmacol., 105, 271–278.
- LEVY, D.M., HINSHAW, K., KNOX, F.M. & CAMPBELL, D.M. & SUTHERLAND, H.W. (1994). Cardiogenic pulmonary oedema: presentation of pre-eclampsia exacerbated by prostaglandin abortifacients. *Br. J. Obstet. Gynaecol.*, **101**, 263–265.
- LUMLEY, P., WHITE, B.P. & HUMPHREY, P.P.A. (1989). GR32191, a highly potent and specific thromboxane A₂ receptor blocking drug on platelets and vascular and airways smooth muscle *in vitro*. *Br. J. Pharmacol.*, **97**, 783–794.
- MADDOX, Y., CUNARD, C.M., SHAPIRO, R., KAWAGUCHI, A., GOLDMAN, M., LOWER, R.R. & RAMWELL, P.W. (1985). A comparison of the contractile responses of rodent and human pulmonary vascular segments to eicosanoids. In *Prostaglandins*, *Leukotrienes and Lipoxins*. pp. 267–272. Elsevier, Martin Bailey-J.
- MAIS, D.E., SAUSSY, JR. D.L., CHAIKHOUNI, A., KOCHEL, P.J., KNAPP, D.R., HAMANAKA, N. & HALUSHKA, P.V. (1985). Pharmacologic characterization of human and canine thromboxane A2/prostaglandin H2 receptors in platelets and blood vessels: evidence for different receptors. J. Pharmacol. Exp. Ther., 233, 418–424.
- MCKENNIFF, M., RODGER, I.W., NORMAN, P. & GARDINER, P.J. (1988). Characterisation of receptors mediating the contractile effects of prostanoids in guinea-pig and human airways. *Eur. J. Pharmacol.*, **153**, 149–159.
- MCKENNIFF, M.G., NORMAN, P., CUTHBERT, N.J. & GARDINER, P.J. (1991). BAY u3405, a potent and selective thromboxane A2 receptor antagonist on airway smooth muscle in vitro. *Br. J. Pharmacol.*, **104**, 585–590.
- MOLDERINGS, G., MALINOWSKA, B. & SCHLICKER, E. (1992). Inhibition of noradrenaline release in the rat vena cava via prostanoid receptors of the EP₃-subtype. *Br. J. Pharmacol.*, **107**, 352–355.
- NEGISHI, M., SUGIMOTO, Y. & ICHIKAWA, A. (1995). Molecular mechanisms of diverse actions of prostanoid receptors. *Biochim. Biophys. Acta*, **1259**, 109–120.
- NOREL, X., LABAT, C., GARDINER, P.J. & BRINK, C. (1991). Inhibitory effects of BAY u3405 on prostanoid-induced contractions in human isolated bronchial and pulmonary arterial muscle preparations. *Br. J. Pharmacol.*, **104**, 591–595.
- OHLSTEIN, E.H., KOPIA, G.A., ZEID, R.L., VALOCIK, R.W., HOR-OHONICH, S., HIEBLE, J.P. & WASSERMAN, M.A. (1988). Effects of the thromboxane receptor antagonist SK&F 88046 in the canine, monkey and human coronary vasculature. *Prostaglandins*, **36**, 69-84.
- PALEA, S., TOSON, G., PIETRA, C., TRIST, D.G., ARTIBANI, W., ROMANO, O. & CORSI, M. (1998). Pharmacological characterization of thromboxane and prostanoid receptors in human isolated urinary bladder. *Br. J. Pharmacol.*, **124**, 865–872.
- PIERCE, K.L. & REGAN, J.W. (1998). Prostanoid receptor heterogeneity through alternative mRNA splicing. *Life Sci.*, 62, 1479– 1483.
- PUURA, A., SCHAVIKIN, L., YLI-KESTI, O., YLA-OUTINEN, A., VIRTANEN, V. & KAUKINEN, S. (1995). [Critical pulmonary edema following cesarean section]. *Duodecim.*, **111**, 249–252.
- QIAN, Y.M., JONES, R.L., CHAN, K.M., STOCK, A.I. & HO, J.K.S. (1994). Potent contractile actions of prostanoid EP₃-receptor agonists on human isolated pulmonary artery. *Br. J. Pharmacol.*, **113**, 369–374.
- REEVES, J.J., BUNCE, K.T., SHELDRICK, R.L.G. & STABLES, R. (1988). Evidence for the PGE receptor subtype mediating inhibition of acid secretion in the rat. Br. J. Pharmacol., 95, 805P.
- SANNER, J.H. (1969). Antagonism of prostaglandin E2 by 1-acetyl-2-(8-chloro-10,11-dihydrodibenz (b,f) (1,4) oxazepine-10-carbonyl) hydrazine (SC-19220). Arch. Int. Pharmacodyn. Ther., 180, 46-56.
- SCHRÖR, K., DARIUS, H., MATZKY, R. & OHLENDORF, R. (1981). The antiplatelet and cardiovascular actions of a new carbacyclin derivative (ZK36374) equi-potent to PGI₂ in vitro. Naunyn-Schmiedebergs Arch. Pharmacol., **316**, 252–255.

- SENIOR, J., MARSHALL, K., SANGHA, R. & CLAYTON, J.K. (1993). In vitro characterization of prostanoid receptors on human myometrium at term pregnancy. *Br. J. Pharmacol.*, **108**, 501– 506.
- SENIOR, J., SANGHA, R., BAXTER, G.S., MARSHALL, K. & CLAYTON, J.K. (1992). In vitro characterization of prostanoid FP-, DP-, IP- and TP-receptors on the non-pregnant human myometrium. *Br. J. Pharmacol.*, **107**, 215–221.
- SHARIF, N.A., XU, S.X., WILLIAMS, G.W., CRIDER, J.Y., GRIFFIN, B.W. & DAVIS, T.L. (1998). Pharmacology of [3H]prostaglandin E1/[3H]prostaglandin E2 and [3H]prostaglandin F₂alpha binding to EP₃ and FP prostaglandin receptor binding sites in bovine corpus luteum: characterization and correlation with functional data. J. Pharmacol. Exp. Ther., 286, 1094-1102.
- SHEN, R.F. & TAI, H.H. (1998). Thromboxanes: synthase and receptors. J. Biomed. Sci., 5, 153-172.
- SJOBERG, T. & STEEN, S. (1989). The strong contractile effect of the thromboxane receptor agonist U-46619 in isolated human pulmonary arteries and its competitive antagonism by BM-13.505. Acta. Physiol. Scand., 136, 161–165.
- STOCK, A., JONES, R., CHUNG, T. & FUNG, H.Y. (1995). Pulmonary edema in association with an intravenous infusion of sulprostone. Acta Obstet. Gynecol. Scand., 74, 156–158.
- TANG, L., LOUTZENHISER, K. & LOUTZENHISER, R. (2000). Biphasic actions of prostaglandin E(2) on the renal afferent arteriole : role of EP(3) and EP(4) receptors. *Circ. Res.*, **86**, 663– 670.
- TEMPLETON, A.G., MCGRATH, J.C. & WHITTLE, M.J. (1991). The role of endogenous thromboxane in contractions to U46619, oxygen, 5-HT and 5-CT in the human isolated umbilical artery. *Br. J. Pharmacol.*, **103**, 1079–1084.
- TYMKEWYCZ, P.M., JONES, R.L., WILSON, N.H. & MARR, C.G. (1991). Heterogeneity of thromboxane A2 (TP-) receptors: evidence from antagonist but not agonist potency measurements. *Br. J. Pharmacol.*, **102**, 607–614.
- USKI, T.K., ANDERSSON, K.E., BRANDT, L. & LJUNGGREN, B. (1984). Characterization of the prostanoid receptors and of the contractile effects of prostaglandin F_2 alpha in human pial arteries. *Acta Physiol. Scand.*, **121**, 369–378.

- WALCH, L., LABAT, C., GASCARD, J.P., DE MONTPREVILLE, V., BRINK, C. & NOREL, X. (1999). Prostanoid receptors involved in the relaxation of human pulmonary vessels. *Br. J. Pharmacol.*, 126, 859–866.
- WATABE, A., SUGIMOTO, Y., HONDA, A., IRIE, A., NAMBA, T., NEGISHI, M., ITO, S., NARUMIYA, S. & ICHIKAWA, A. (1993). Cloning and expression of cDNA for a mouse EP₁ subtype of prostaglandin E receptor. J. Biol. Chem., 268, 20175-20178.
- WATERBURY, L.D., EGLEN, R.M., FAUROT, G.F. & COOPER, G.F. (1990). EP₃, but not EP₂, FP, or TP prostanoid-receptor stimulation may reduce intraocular pressure. *Invest Ophthalmol. Vis. Sci.*, **31**, 2560–2567.
- WOODWARD, D.F., BURKE, J.A., WILLIAMS, L.S., PALMER, B.P., WHEELER, L.A., WOLDEMUSSIE, E., RUIZ, G. & CHEN, J. (1989).
 Prostaglandin F₂ alpha effects on intraocular pressure negatively correlate with FP-receptor stimulation. *Invest. Ophthalmol. Vis. Sci.*, **30**, 1838–1842.
- WOODWARD, D.F., CHAN, M.F., BURKE, J.A., CHENG-BENNETT,
 A., CHEN, G., FAIRBAIRN, C.E., GAC, T., GARST, M.E.,
 GLUCHOWSKI, C., KAPLAN, L.J., LAWRENCE, R.A., ROOF, M.,
 SACHS, G., SHAN, T., WHEELER, L.A. & WILLIAMS L.S. (1994).
 Studies on the ocular hypotensive effects of prostaglandin F₂
 alpha ester prodrugs and receptor selective prostaglandin
 analogs. J. Ocul. Pharmacol., 10, 177–193.
- WOODWARD, D.F., LAWRENCE, R.A., FAIRBAIRN, C.E., SHAN, T. & WILLIAMS, L.S. (1993). Intraocular pressure effects of selective prostanoid receptor agonists involve different receptor subtypes according to radioligand binding studies. J. Lipid Mediat., 6, 545-553.
- WOODWARD, D.F., PEPPERL, D.J., BURKEY, T.H. & REGAN, J.W. (1995). 6-isopropoxy-9-oxoxanthene-2-carboxylic acid (AH6809) a human EP_2 receptor antagonist. *Biochem. Pharmacol.*, **50**, 1731–1733.
- YOKOTANI, K., OKUMA, Y. & OSUMI, Y. (1996). Inhibition of vagally mediated gastric acid secretion by activation of central prostanoid EP₃ receptors in urethane-anaesthetized rats. *Br. J. Pharmacol.*, **117**, 653–656.

(Received December 15, 2000 Revised September 18, 2001 Accepted October 2, 2001)