

## REVIEW

## NO as a signalling molecule in the nervous system

\*<sup>1</sup>Juan V. Esplugues<sup>1</sup>Departamento de Farmacología, Facultad de Medicina, Universidad de Valencia, Spain*British Journal of Pharmacology* (2002) 135, 1079–1095**Keywords:** NO; CNS; NANC; NOS synthases; stroke; neurotransmission; signalling**Abbreviations:** CAM, calmodulin; CAPON, carboxy-terminal PDZ ligand of nNOS; cyclic AMP, cyclic adenosin monophosphate; cyclic GMP, cyclic guanosin monophosphate; eNOS, endothelial NO synthase; iNOS, inducible NO synthase; LPS, lipopolysaccharides; LTD, long-term depression; LTP, long-term potentiation; NADPH,  $\alpha$ -nicotinamide adenine dinucleotide phosphate; NANC, non-adrenergic non-cholinergic; NO, nitric oxide; nNOS, neuronal NO synthase; ONOO<sup>-</sup>, peroxynitrite; PIN, protein inhibitor of nNOS; PSD, postsynaptic density protein; sGC, soluble guanylyl cyclase; SOD, superoxide dismutase; tSXV, Ser/Thr-X-Val motif; VIP, vasoactive intestinal peptide

The discovery that nitric oxide (NO) functions as a signalling molecule in the nervous system has radically changed the concept of neural communication. Indeed, the adoption of the term nitergic for nerves whose transmitter function depends on the release of NO or for transmission mechanisms brought about by NO (Moncada *et al.*, 1997) emphasizes the specific characteristics of this mediator. The physical properties of NO prevent its storage in lipid-lined vesicles and metabolism by hydrolytic degradatory enzymes. Therefore, unlike established neurotransmitters, NO is synthesized on demand and is neither stored in synaptic vesicles nor released by exocytosis, but simply diffuses from nerve terminals. The distance of this NO diffusion (40–300  $\mu\text{m}$  in diameter) implies that structures in the vicinity of the producing cell, both neuronal and non-neuronal, are influenced following its release. This suggests that, as well as acting as a neurotransmitter, NO has a neuromodulatory role (Garthwaite & Boulton, 1995). In addition, it diffuses into rather than binds with protein receptors on adjacent cells, and most of its known actions are the consequence of interplay with intracellular targets that would usually be regarded as secondary messengers. The activity of conventional neurotransmitters is terminated either by re-uptake mechanisms or enzymatic degradation while inactivation of NO follows reaction with a substrate. There are multiple points at which biological control can be exerted over the production and activity of conventional neurotransmitters. However, control of the synthesis of NO is the key to regulating its activity.

Endothelial NOS (eNOS) and inducible NOS (iNOS) are present in the nervous system and will be duly addressed here. However, neuronal NOS (nNOS) is the principal isoform present in said system and will be the main focus of this review. All nNOS positive neurones exhibit  $\alpha$ -nicotinamide adenine dinucleotide phosphate (NADPH)-diaphorase activity, which has become the histochemical marker of nitergic neurones. However, early results

demonstrating this may have been limited by inappropriate fixation procedures and should be viewed with caution (Wolf, 1997). The original cloning of full-length nNOS produced what is now designated as nNOS $\alpha$ , and which accounts for the majority of nNOS activity in nervous tissue (Bredt *et al.*, 1991). In addition, four splice variants have recently been identified (nNOS $\beta$ , nNOS $\gamma$ , nNOS $\mu$  and nNOS-2) and these appear to exhibit distinct cellular and tissue locations (Gibson, 2001; Nakane *et al.*, 1993; Silvagno *et al.*, 1996; Alderton *et al.*, 2001). In particular, there is growing evidence that nNOS biosynthesis in excitable tissues is not restricted to neurones while substantial amounts of this enzyme have been identified in skeletal muscle, where it is involved in the regulation of metabolism and muscle contractility (Stamler & Meissner, 2001).

The magnitude of literature dealing with the role of NO in the nervous system is so great that it would be impossible to include in this review the entirety of the research carried out. For logistical reasons, only groundbreaking references have been quoted, but when necessary, recent reviews dealing with specific areas within the field have been included and are intended to act as a guideline for further reading.

## Regulation of nNOS

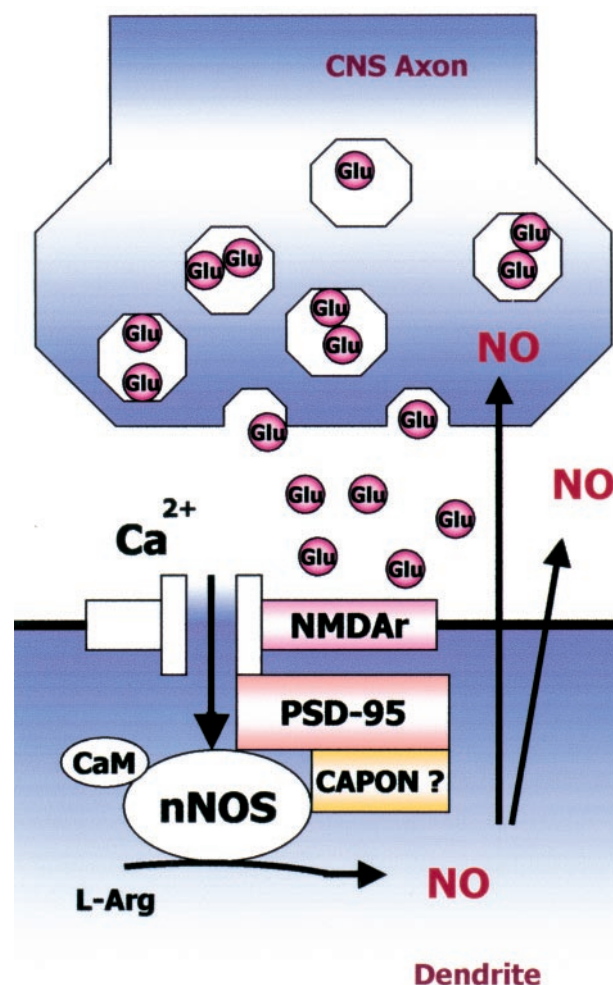
The most important regulator of nNOS activity seems to be free cytosolic Ca<sup>2+</sup>, which stimulates nNOS through interaction with calmodulin. Arrival of action potentials activates voltage-dependent Ca<sup>2+</sup> channels situated in the neurolemma, and stimulates the release of Ca<sup>2+</sup> from intracellular stores. This elevates cytosolic Ca<sup>2+</sup> concentrations above the 400 nM required for calmodulin to bind to nNOS, thereby activating the enzyme. When the concentration of Ca<sup>2+</sup> falls, it dissociates from the calmodulin, which in turn dissociates from the nNOS, thus acting as a switch that turns the enzyme on and off (Knowles *et al.*, 1989; Sheng *et al.*, 1992). Phosphorylation, although less well analysed, constitutes an additional mechanism for regulating nNOS activity. The catalytic activity of the enzyme is decreased following phosphorylation by cyclic adenosin monophosphate (cyclic

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AMP)-dependent protein kinase (Bredt *et al.*, 1992; Brüne & Lapetina, 1991), protein kinase C (Bredt *et al.*, 1992; Nakane *et al.*, 1991) or  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase II (Bredt *et al.*, 1992; Hayashi *et al.*, 1999; Komeima *et al.*, 2000; Nakane *et al.*, 1991; Schmidt *et al.*, 1992b).

This process occurs in the majority of peripheral and in some central nitricergic neurones (Figures 1 and 2). However, in the CNS, NO synthesis seems predominantly regulated by the influx of  $\text{Ca}^{2+}$  through receptor-dependent channels, in particular following postsynaptic stimulation of NMDA receptors by the excitatory neurotransmitter, glutamate (Bredt & Snyder, 1989; Garthwaite *et al.*, 1989). The amino acid terminal of nNOS possesses a PDZ domain which is not present in the  $\beta$  and  $\gamma$  splice variants (Alderton *et al.*, 2001; Brenman *et al.*, 1996). The aforementioned domains are modular structures of approximately 100 amino acids, which occur in a number of proteins, anchoring them to cytoskeletal elements such as synaptic densities and related membrane-associated guanylate kinases (Tomita *et al.*, 2001). In the case of nNOS, its PDZ domain interacts with the postsynaptic density protein PSD-95, whereas the N-Methyl-D-Aspartate (NMDA) receptor contains a Ser/Thr-X-Val motif (tSXV) that also binds with PSD95. By facilitating the proximity of NMDA receptors to the enzyme, the scaffolding protein, PSD95 directly exposes nNOS to the flux of  $\text{Ca}^{2+}$  entering the ion channel of activated NMDA receptors (Kornau *et al.*, 1995; Tomita *et al.*, 2001). Transient  $\text{Ca}^{2+}$  fluxes following the activation of other receptors would be too diluted to have a similar effect by the time they reach the vicinity of nNOS. It is possible that NO bioactivity feeds back to control the activity of the channel as S-nitrosylation of critical cysteines seems to down-regulate the NMDA receptor (Choi *et al.*, 2000). There are many other potential regulators of the NMDA receptor/nNOS coupling and downstream signalling pathways. The protein carboxy-terminal PDZ ligand of nNOS (CAPON) is thought to be selectively associated with nNOS and to exhibit a similar regional distribution. CAPON competes with nNOS for PDZ domains, binding to the enzyme and forcing it to disassociate itself from the plasma membrane (Jaffrey *et al.*, 1998). Therefore, CAPON determines the amount of nNOS tethered to the plasma membrane and, in this way, regulates NO formation in neurones of the CNS. Furthermore, CAPON anchors nNOS to other macromolecules, such as the small G-protein Dexas-1 (Fang *et al.*, 2000), although the importance of this relationship remains to be determined. At this point, it must be said that recent proteomic analysis has not identified CAPON in the vicinity of the NMDA receptor, which raises some doubts about the above hypotheses (Husi *et al.*, 2000).

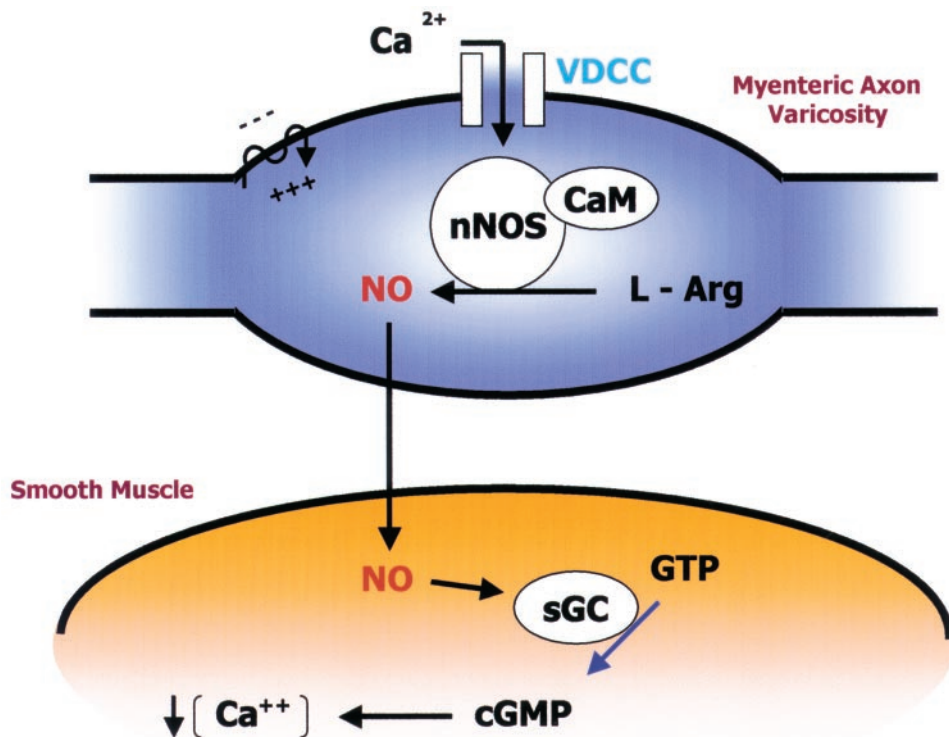
Various other receptors and domains contain the tSXV motif and are also potentially associated with central nNOS and regulated by this multifunctional protein-protein interaction (Tomita *et al.*, 2001). nNOS may also be inhibited through an interaction with protein inhibitor of nNOS (PIN), a highly conserved small protein that was originally thought to destabilize nNOS dimers and thus act as an endogenous inhibitor of nNOS (Jaffrey & Snyder, 1996). However, recent reports suggest that PIN is an axonal transport protein for nNOS, rather than its regulator (Hemmens *et al.*, 1998; Rodriguez-Crespo *et al.*, 1998). nNOS may also be inhibited through an interaction with caveolin-1 and caveolin-3 that, in a way similar to the effect of caveolin-1 on eNOS, could



**Figure 1** Activation of nNOS in the CNS. Release of glutamate activates NMDA receptors (NMDAr), and the consequent flux of  $\text{Ca}^{2+}$  entering the ion channel activates nNOS, which is linked to the receptor via the postsynaptic density protein PSD-95. It is possible that NO bioactivity feeds back to control the presynaptic neuron and the activity of the channel. The protein CAPON is thought to be selectively associated with nNOS and regulates NO formation in neurones.

displace calmodulin from nNOS (García-Cerdeña *et al.*, 1997; Venema *et al.*, 1997). Furthermore, these members of the caveolin family interact with other signalling molecules such as *c-src*, Ha-ras and  $\text{G}_{\text{s}\alpha}$ , which suggests a potential role for nNOS in some signalling complexes (Couet *et al.*, 1997). Finally, a role for heat shock protein NOS-hsp90/hetero-complexes in the modulation of the haem's interaction with nNOS has recently been suggested (Bender *et al.*, 1999).

In skeletal muscle, nNOS activity is related to muscle ACh receptors and membrane depolarization (Figure 3). Again, nNOS is targeted to membrane structures due to the association of its PDZ domain with  $\alpha_1$ -syntrophin, a dystrophin-associated protein that shares homology with postsynaptic density proteins PDS95 y PDS93. Interactions with nNOS, highly expressed in skeletal muscle, and with caveolin-3 are also possible (Brenman *et al.*, 1995; Chao *et al.*, 1996; Stamler & Meissner 2001; Venema *et al.*, 1997).



**Figure 2** nNOS in myenteric neurones is regulated by the flux of  $\text{Ca}^{2+}$  through voltage-dependent calcium channels (VDCC). NO relaxes the adjacent smooth muscle following activation of sGC.

Although considered to be constitutive, levels of nNOS activity and expression appear to be subject to dynamic up- or down-regulation induced by a large variety of stimuli, including nerve (Steel *et al.*, 1994; Verge *et al.*, 1992) and brain injury (Kitchener *et al.*, 1993; Regidor *et al.*, 1993), aging (Carrier *et al.*, 1997; Mollace *et al.*, 1995), pharmacological treatment (Bagetta *et al.*, 1993), lactation (Ceccatelli & Eriksson, 1993), hypoxia (Guo *et al.*, 1997), stress (Cazal *et al.*, 1993), gonadectomy (Ceccatelli *et al.*, 1993), light exposure (Schaad *et al.*, 1994) and exercise (Tidball *et al.*, 1998).

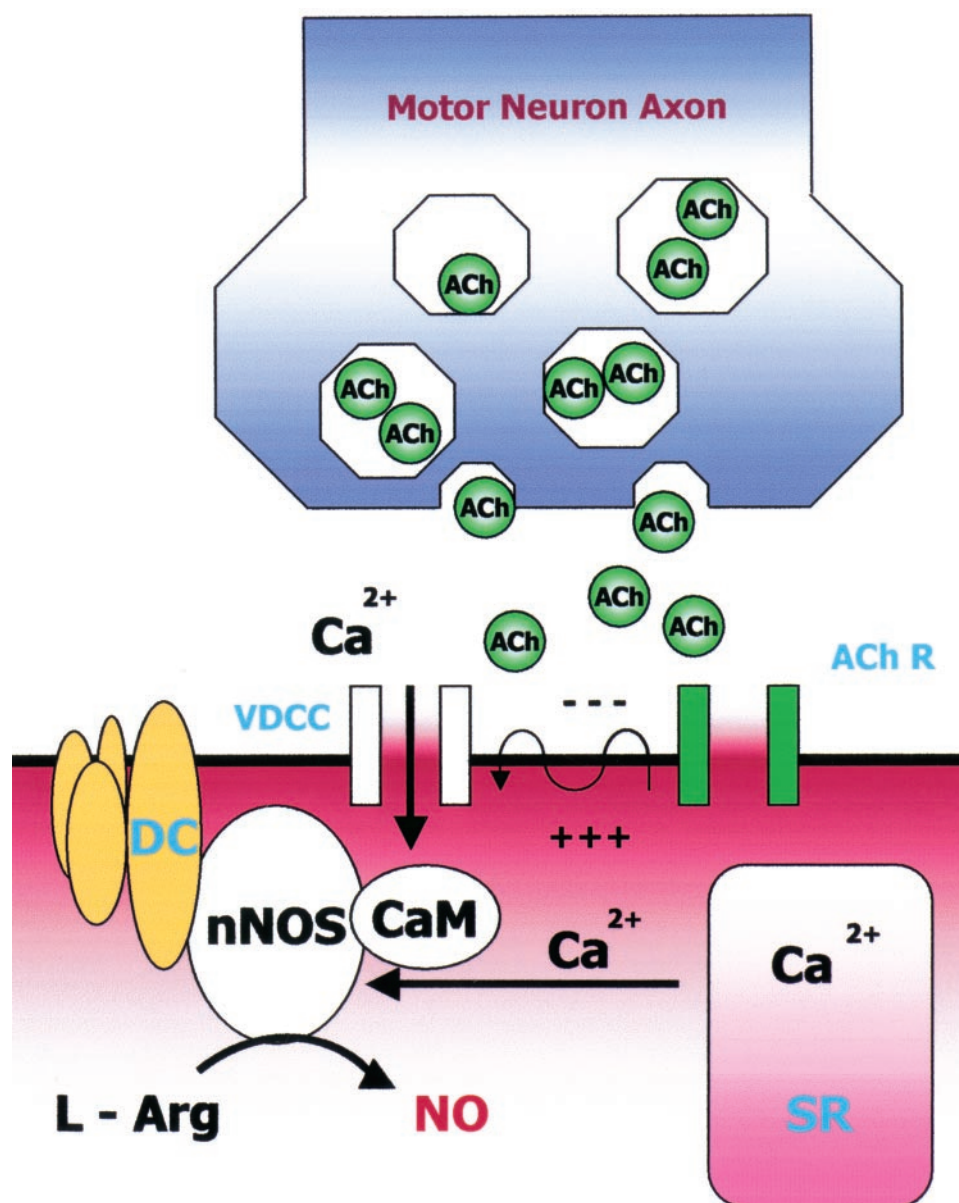
The existence of presynaptic automodulation in nitrenergic neurones has also been proposed. Although not yet fully characterized, this action may result from the combination of NO with the haem group of NOS, which inhibits the enzyme (Klatt *et al.*, 1992; Rogers & Ignarro, 1992). Finally, there is evidence that cells maintain low levels of cyclic guanosin monophosphate (cyclic GMP) while producing NO. This occurs because increases in  $\text{Ca}^{2+}$  levels, similar to those needed to stimulate nNOS, also activate a  $\text{Ca}^{2+}$ /calmodulin-dependent cyclic GMP phosphodiesterase that facilitates the degradation of cyclic GMP (Mayer *et al.*, 1992).

## Signalling

The actions of NO are a consequence of its influence on a variety of protein functions which it exerts through its reaction with cysteine thiol, S-nitrosylation, and transition metal centres (Drapier & Bouton, 1996; Jaffrey *et al.*, 2001; Lane *et al.*, 2001). The enzyme soluble guanylyl cyclase (sGC)

has long been considered to be the major physiological target for neuronal NO, and there is ample evidence that increases in cyclic GMP levels mediate a large number of the physiological actions of NO. Thus, immunohistochemical techniques have found that the distribution of sGC and cyclic GMP is complementary to that of nNOS (Schmidt *et al.*, 1992a; Southam & Garthwaite, 1993; Young *et al.*, 1993). Functionally, either nitrenergic nerve stimulation or administration of NO-donors increases intracellular cyclic GMP concentrations (Bredt & Snyder, 1989; Torphy *et al.*, 1986). In both cases, these responses are mimicked by analogues of cyclic GMP (Gibson & Mirzazadeh, 1989), whereas inhibition of the destruction of this intracellular mediator potentiates the results of nitrenergic stimulation (Barbier & Lefebvre, 1995; Bayguinov & Sanders, 1993). The mechanisms linking the rise in cyclic GMP content to the various effects of NO in the CNS and peripheral smooth muscle are not fully understood, although in both cases the final step seems to be a reduction of  $[\text{Ca}^{2+}]_i$ . Alternative targets of cyclic GMP may involve direct channel gating with the opening of inward  $\text{Ca}^{2+}$  and  $\text{Na}^{2+}$  channels, activation of cyclic GMP-dependent kinases, actions related to cyclic adenosin diphosphate (ADP) ribose, and interactions with cyclic AMP resulting from regulation of cyclic GMP-dependent phosphodiesterases (Hunter, 2000; Jaffrey *et al.*, 2001; Lincoln *et al.*, 2001).

NO modulates oxygen consumption in the mitochondria. In particular, nanomolar concentrations of NO inhibit cytochrome oxidase, the terminal haem-containing enzyme in the mitochondrial respiratory chain. Recent evidence demonstrates that this effect is reversible and competitive



**Figure 3** Activation of nNOS in the skeletal muscle follows the influx of  $\text{Ca}^{2+}$  through voltage-dependent calcium channels (VDCC) induced by activation of ACh receptors (AChR) and membrane depolarization. The release of  $\text{Ca}^{2+}$  from the sarcoplasmic reticulum (SR) is also implicated. nNOS targets the membrane due to its association with  $\alpha_1$ -syntrophin, a component of the dystrophin complex (DC).

with oxygen, and suggests that NO is a crucial regulator in the generation of energy and the mediation of cell death by mitochondria (Beltrán *et al.*, 2000). The consequences of such an activity are still to be evaluated but, obvious physiological implications aside, they could clarify the mechanisms by which NO is involved in cell or tissue damage. Furthermore, and in conjunction with differences in glycolytic capacity, this activity may explain why neurones and glia show variations in sensitivity to NO-induced damage (Brown, 2000; Almeida *et al.*, 2001).

NO has been linked to the release (Meffert *et al.*, 1996) of other neurotransmitters and the effects which they produce, in particular acetylcholine (Gustafsson *et al.*, 1990; Li & Rand, 1989b), noradrenaline (Boeckxstaens *et al.*, 1993; Li & Rand, 1989a) dopamine (Hanbauer *et al.*, 1992), glutamate

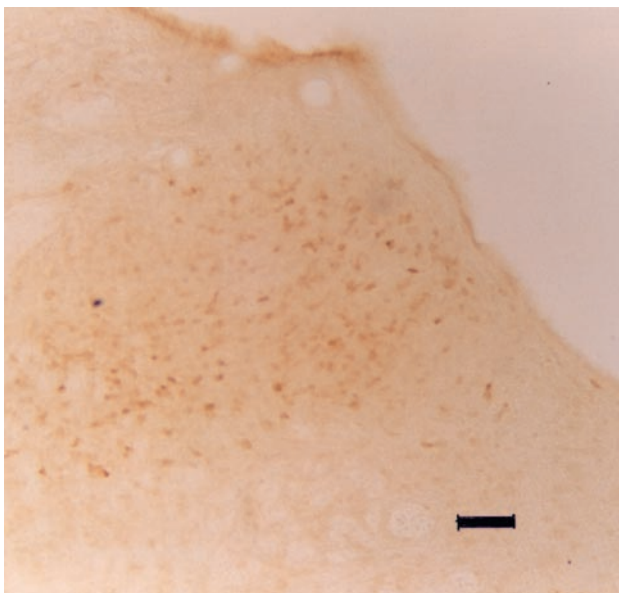
(Montague *et al.*, 1994; Sorkin, 1993),  $\gamma$ -aminobutyric acid (GABA) (Beltran *et al.*, 1990; Kuriyama & Ohkuma, 1995), serotonin (Bogers *et al.*, 1991; Reiser, 1990b), adenosin triphosphate (ATP) (Boeckxstaens *et al.*, 1991a), bombesin (Beltran *et al.* 1999), carbon monoxide (Xue *et al.*, 2000), opioids (Barnette *et al.*, 1990) and endothelin (Reiser, 1990a). The mechanisms responsible for these interactions are still not fully understood, but direct S-nitrosylation of receptors, activation of cyclic GMP-dependent protein phosphorylation cascades, regulation of neuronal energy dynamics and a modulating effect on transporters are potentially involved (Choi *et al.*, 2000; Kiss & Vizi, 2001; Pieper *et al.*, 2000). In addition, a presynaptic modulation of NO release through the activation of  $\alpha_2$ -adrenoceptors, nicotinic receptors, purinergic receptors etc. has also been

proposed (Boeckxstaens *et al.*, 1993). Finally, it has been suggested that NO modulates gene transcription and translation in neurones and glia (Hess *et al.*, 1993; Peunova & Enikolopov, 1993; 1995). However, these effects would seem to be indirect since there is little evidence of the existence of DNA elements within the promotor regions of eukaryotic cells that respond directly to NO (Morris, 1995).

## Nitric oxide in the central nervous system

### Introduction

NO was first characterized in the CNS as the intercellular messenger mediating the increase in cyclic GMP levels that follows activation of glutamate receptors (Garthwaite *et al.*, 1988). The majority of the information available deals with nNOS, of which the brain contains the highest activity found in any tissue, and which, although present in some cerebral vessels and in glial cells, is predominantly found in neurones (Bredt *et al.*, 1990; Salter *et al.*, 1991). nNOS-containing neurones are present in many areas of the CNS (Figure 4), with the highest densities occurring in the accessory olfactory bulb and granule cells of the cerebellum. Although nNOS neurones represent only roughly 1% of cell bodies in the cerebral cortex, virtually every neurone in the cortex is exposed to nNOS nerve terminals. From a morphological point of view nNOS neurones display a great heterogeneity in their localization within the CNS, constituting a small population of varying interneurones. Furthermore, the number and chemical characteristics of nNOS neurones vary considerably depending on the area of the brain while the enzyme itself does not co-localize with any single neurotransmitter (Braissant *et al.*, 1999; Iwase *et al.*, 1998; Vincent, 1995; Wolf, 1997). nNOS can be located either pre- or post-synaptically and is particularly implicated in neural signal-



**Figure 4** Representative microphotograph of basal nNOS immunoreactivity (monoclonal antibody) in the dorsal vagal complex (DVC) of the brainstem. Scale bar = 100  $\mu$ m.

ling, neurotoxicity, synaptic plasticity and modulation of behavioural pathways such as learning or expression of pain.

eNOS is mainly involved in the regulation of vascular function and, although also present in some populations of neurones (Dinerman *et al.*, 1994) and glia (Wiencken & Casagrande, 1999), is predominantly located in the endothelial cells of cerebral vessels. Finally, induction of iNOS in glial cells is implicated in the unspecific immune response of the brain and is usually associated with pathological conditions (Murphy, 2000).

### Effects of centrally released NO

**Modulation of synaptic plasticity** NO has been proposed as the retrograde messenger which co-ordinates the enhancement of both pre- and post-synaptic mechanisms involved in two forms of synaptic plasticity; namely long-term potentiation (LTP) and long-term depression (LTD). LTP is a property of many central excitatory synapses characterized by a prolonged enhancement of synaptic transmission, or an activity-dependent increase in synaptic strength, lasting from hours to weeks or even longer. The process by which LTP is induced is not completely clear, but it involves glutamate acting on amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA) or NMDA-receptors. This activates a series of events in which  $Ca^{2+}$ /calmodulin-dependent protein kinase II, NOS and protein tyrosine kinases are implicated. LTP is thought to be a synaptic correlate of learning and memory, and is most pronounced in higher brain centres involved in cognitive functions, particularly in the cerebral cortex and hippocampus. The basic evidence for said involvement in LTP stems from *in vitro* studies in which inhibition of NOS prevented the development of LTP (Bohme *et al.*, 1991; O'Dell *et al.*, 1991; Schuman & Madison, 1991). Gene targeting suggests that both nNOS and neuronally located eNOS are implicated in LTP. Thus, while LTP is only slightly reduced in nNOS or eNOS null mice (O'Dell *et al.*, 1994), animals deficient in both NOS isozymes exhibit a substantially decreased LTP (Son *et al.*, 1996). Guanylate cyclase seems to be the main effector of NO in the induction of LTP (Bohme *et al.*, 1991; Haley *et al.*, 1992), however, ADP-ribosylation (Brüne & Lapetina, 1989) and activation of calmodulin-dependent kinases (Soderling, 2000; Tomita *et al.*, 2001) have also been implicated.

LTD is characterized by a long lasting depression of parallel fibre synapses, which follows repeated excitation of the climbing fibres of Purkinje cells. The reduction in synaptic strength appears to result from a diminished sensitization of postsynaptic AMPA receptors which is mediated by activation of protein kinases C and G and of the NO-cyclic GMP signalling pathway (Daniel *et al.*, 1993; Shibuki & Okada, 1991). LTD can be observed in higher regions of the brain, although it has been particularly well studied in the cerebellum where it has been proposed as a model for the learning of motor movements. Finally, the role of NO in both forms of synaptic plasticity involves interaction with PSD-95 and related membrane-associated guanylate kinases, underlined by the fact that both LTP and LTD are significantly modified in mice with targeted disruption of PSD-95 (Migaud *et al.*, 1998).

### *Involvement in central and peripheral functions*

NO has complex influences on brain development, memory formation and behaviour through regulation of synaptic plasticity. Inhibition of NO synthesis produces amnesia (Holscher & Rose, 1992), disrupts spatial learning and olfactory memory (Bohme *et al.*, 1993; Kendrick *et al.*, 1997), blunts behavioural performance during task acquisition and decreases locomotor activity in habituation tasks (Yamada *et al.*, 1995). NO has also been implicated in neuronal targeting and brain development (Contestabile, 2000; Okere & Kaba, 2000; Wu *et al.*, 1994a), visual processing (Cudeiro & Rivadulla, 1999), discriminative learning (Groll-Knapp *et al.*, 1988), food and drinking behaviour (Calapai *et al.*, 1992; Morley & Flood, 1992), thermoregulation (De Luca *et al.*, 1995), opiate tolerance and withdrawal (Mao, 1999; Zhu & Barr, 2001), circadian rhythm (Watanabe *et al.*, 1995), sleep (Kapas *et al.*, 1994) and respiratory pattern generation (Ling *et al.*, 1992). Likewise, behavioural responses mediated by oxytocinergic and serotonergic pathways are thought to involve NO generation or stimulation of central nitric neurons (Melis *et al.*, 1994; 1995). The participation of NO in behavioural mechanisms seems likely after being confirmed in knockout nNOS mice. These animals show no overt behavioural disorders when housed individually, but if kept together males exhibit exceptionally aggressive and hypersexual behaviour. In contrast, nNOS deficiency reduces aggression in female mice (Mani *et al.*, 1994; Nelson *et al.*, 1995).

There is an ever-growing list of peripheral functions in which a role for central NO has been proposed, although the exact physiological relevance of these observations warrants further investigation. Without intending to provide an exhaustive list, there is evidence implicating central NO in the regulation of blood pressure (Togashi *et al.*, 1992), heart rate (Sakuma *et al.*, 1992), stimulated renal sympathetic nerve activity (Sakuma *et al.*, 1992), gastric acid secretion and motility (Esplugues *et al.*, 1996; Garcia-Zaragoza *et al.*, 2000; Quintana *et al.*, 2001) and motor disruption associated with alcohol abuse (Sandor *et al.*, 1995). Likewise, NO in the CNS appears to be involved in reflexes leading to a diminished sympathetic output to the periphery and the modulation of various neuroendocrine responses, including the production of oxytocin, luteinizing hormone-releasing hormone, osmoregulator peptide corticotropin-releasing hormone and adrenocorticotrophic hormone (Aguilla, 1994; Ceccatelli *et al.*, 1993; Costa *et al.*, 1993; Rivier & Shen, 1994).

### *Perception of pain*

NO has been implicated at various levels of the nociceptive neural pathways, both peripherally (primary afferent neurons and dorsal root ganglia) and centrally (brainstem and several sensory structures of the thalamus) (Mao, 1999). Functionally, most nociceptive reflexes involve the interaction of NO and NMDA receptors, and it is established that synthesis of NO enhances spinal facilitation of the afferent input conveyed to the cortex and subsequently manifested in behavioural responses (Mayer *et al.*, 1999). However, the role of NO changes according to the pain stimuli. Inhibition of NO has antinociceptive effects when pain stems from chemically stimulated peripheral nerve terminals and in

models of thermal hyperalgesia or visceral pain (Kitto *et al.*, 1992; Malmberg & Yaksh, 1993; Moore *et al.*, 1991; 1993), while intrathecal administration of L-arginine induces allodynia by converting a non-noxious to a noxious mechanical stimulus (Minami *et al.*, 1995). In contrast, blockade of NO synthesis exacerbates pain in models of mechanical hyperalgesia (Zhuo *et al.*, 1993). The use of nNOS knockout mice has not clarified such contradictory results; for example, these animals display a normal sensitization to some types of damage unmodified by NOS inhibitors (Crosby *et al.*, 1995). The implication of NO in the antinociceptive effects of drugs is also controversial. For instance, inhibitors of the NO/guanylate cyclase system potentiate the antinociceptive actions of morphine while attenuating the antinociceptive effects of  $\beta$ -endorphin (Xu & Tseng, 1995; Zhu & Barr, 2001). Furthermore, there is evidence which suggests that the splice variants of nNOS, nNOS-2 modulates morphine analgesia but not morphine tolerance (Kolesnikov *et al.*, 1997).

### *Neuronal damage and protection*

The neuronal damage that accompanies cerebral ischaemia involves an excessive release of glutamate and a subsequent activation of NMDA receptors that, if maintained for a sufficient period of time, induces a massive influx of  $Ca^{2+}$  into the postsynaptic neurone which, in turn, triggers the activation of nNOS and overproduction of NO. In contrast, NO produced by activation of eNOS (Marks *et al.*, 1996; Stagliano *et al.*, 1997), and even NMDA receptors (Fergus & Lee, 1997; Wilderman & Armstead, 1997), plays a protective role in brain ischaemia by maintaining regional cerebral blood flow. The first indications that NO could mediate neurotoxic effects came with the discovery that inhibition of NOS attenuates glutamate toxicity in primary neuronal cultures from the rat cerebral cortex (Dawson *et al.*, 1991) and induces neuroprotection in animal models of stroke (Nowicki *et al.*, 1991). These studies were soon followed by others which showed that inhibition of NO synthesis attenuated NMDA neurotoxicity both *in vivo* (Nagafuji *et al.*, 1992; Tominaga *et al.*, 1993) and *in vitro* (Kollegger *et al.*, 1993), and which demonstrated enhanced concentrations of NO in various stroke models (Kader *et al.*, 1993; Malinski *et al.*, 1993). These concepts were initially controversial due to the appearance of contradictory *in vitro* and *in vivo* reports in which NO-mediated neurotoxicity was not observed (Bolaños & Almeida, 1999; Samdani *et al.*, 1997). However, such discrepancies stem from variations in experimental conditions and doses of NOS inhibitors. Thus, inhibition of nNOS with concentrations of NOS inhibitors that do not suppress eNOS activity reduces infarct volume, whereas the use of selective nNOS inhibitors is consistently neuroprotective in models of focal ischaemia (Yoshida *et al.*, 1994; Zhang *et al.*, 1996). Exacerbation of injury seems to occur through inhibition of eNOS with high doses of non-selective NOS inhibitors, which results in deleterious alterations of cerebral blood flow and a subsequent increase in infarction volume.

A better understanding of the role of nNOS and eNOS in neuronal damage has been obtained using transgenic animals. nNOS knockout mice develop substantially less brain damage following ischaemia than those of the wild-type strain (Huang *et al.*, 1994), while neuronal cultures from such animals are

more resistant to damage by glutamate and hypoxia hypoglycaemia (Dawson *et al.*, 1996). The reduction of infarct volume in nNOS null transgenic mice does not occur when non-specific NOS inhibitors are administered at concentrations that inhibit NO-dependent relaxation of pial vessels. On the other hand, eNOS knockout mice exhibit more extensive damage following ischaemia, an effect associated with a sharp reduction of blood flow in the affected area, and administration of NOS inhibitors to these animals reduces injury (Huang *et al.*, 1996).

The interactions and signalling mechanisms involved in these NO-related effects are complex. Vascular protection is linked to cyclic GMP-mediated mechanisms (Utepbergenov *et al.*, 1998). In addition, S-nitrosylation of glutathione by NO has been implicated in the antioxidative neuronal defence system, while NO is thought to scavenge reactive oxygen species and partially offset ischaemia induced oxidative damage (Wink *et al.*, 1993). Likewise NO could be directly neuroprotective by interacting with a specific site of the NMDA-receptor channel, resulting in a decreased binding of glutamate or a diminished flow of  $\text{Ca}^{2+}$  through the channel after activation (Choi *et al.*, 2000; Hoyt *et al.*, 1992).

Generation of peroxynitrite seems to be the leading cytotoxic mediator in glutamate-induced damage (Fukuyama *et al.*, 1998; Lafon-Cazal *et al.*, 1993; Tanaka *et al.*, 1997). The production of peroxynitrite has been detected in postischaemic brains (Fukuyama *et al.*, 1998; Tanaka *et al.*, 1997). Mice with overexpression of superoxide dismutase (SOD) and nNOS deletion exhibit decreased neurotoxicity following vascular stroke (Yang *et al.*, 1994), accompanied by a strongly suppressed peroxynitrite ( $\text{ONOO}^-$ ) production (Keller *et al.*, 1998). Damage caused to DNA by NO and peroxynitrite appears to be an important neurotoxic mechanism. This is due to the subsequent activation of the nuclear repair enzyme polyADP-ribose synthase which is capable of triggering massive energy depletion resulting in cellular death if the DNA damage is severe (Pieper *et al.*, 1999; Wallis *et al.*, 1993; Zhang *et al.*, 1994). Inhibition of nNOS reduces polyADP-ribosylation while inhibition of polyADP-ribose synthetase decreases infarction volume. Animals with a genomic deletion of polyADP-ribose synthetase exhibit greater resistance to cerebral ischaemia than nNOS knockout mice or those treated with NOS inhibitors or NMDA antagonists (Eliasson *et al.*, 1997). In addition, inhibition of mitochondrial respiratory chain enzymes exacerbates the depletion of neuronal energy stores (Beltrán *et al.*, 2000; Bolaños *et al.*, 1994; 1995; Clementi *et al.*, 1998; Moncada & Erusalimsky, 2002). High local concentrations of NO may also reduce cellular viability by nitrosylating several enzymes, including phosphokinases C (Hammer *et al.*, 1993) and glyceraldehyde-3-phosphate dehydrogenase (Zhang & Snyder, 1992), or by interacting with the iron present in haem or non-haem complexes associated with enzymes such as cytochrome P450 or aconitases (Drapier & Bouton, 1996).

Inflammatory conditions and transient ischaemic periods induce the expression of iNOS in various populations of cerebral cells. However, iNOS expression occurs later than that of nNOS and eNOS, and the cellular site of this expression is dependent on the nature of injury (Iadecola *et al.*, 1995a). iNOS immunoreactivity is present in the neutrophils which infiltrate the brain after permanent ischaemia, is predominant in vascular cells in transient

ischaemia (Iadecola *et al.*, 1995b), and is abundant in reactive astrocytes in global ischaemia (Endoh *et al.*, 1994). NO produced by iNOS seems to exert a detrimental effect in the ischaemic brain, contributing to the progression of tissue damage and exacerbating glutamate neurotoxicity (Chao *et al.*, 1992). Thus, NO produced *in vitro* after hypoxia induces apoptotic death in neurones (Boje & Akora, 1992), while studies with iNOS knock out mice have confirmed the induction of iNOS in the delayed neuronal damage following ischaemia (Iadecola, 1997).

### *Role in central disorders*

There is growing evidence to support a role for NO in the aetiology of neurologic conditions, including autoimmune and chronic neurodegenerative diseases. Concentrations of NO present in inflamed tissue cause reversible conduction block in normal, demyelinated and early remyelinated axons. Thus, diseases such as multiple sclerosis and Guillain-Barre syndrome, characterized by widespread loss of myelin, may see their neuronal symptoms exacerbated *via* the release of NO that accompanies the severe inflammation in the central and peripheral nervous system occurring in these conditions. NO may also be important in secondary neuronal cell death following trauma. In the spinal cord, nNOS expression precedes the death of motoneurones, which follows avulsion of spinal nerve roots (Wu *et al.*, 1994b), and pre-treatment with nNOS inhibitors substantially increases the number of surviving neurones (Wu & Li, 1993). NOS neurones are resistant to NMDA and NO neurotoxicity although the protective mechanism responsible is still not fully known (Koh & Choi, 1988). Likewise, the excessive release of both glutamate and NO, coupled with mitochondrial dysfunction and oxidative stress, has been implicated in a number of neurodegenerative diseases. This highlights a potential therapeutic role for specific NOS inhibitors in their pharmacological control (Hobbs *et al.*, 1999). For instance, nNOS is induced in various cortical regions following epileptic seizures (Huh *et al.*, 2000). NOS neurones are spared in Alzheimer's disease and NOS inhibitors provide neuroprotection against toxicity elicited by fragments of human  $\text{B-amyloid}$  in primary cortical cultures (Hyman *et al.*, 1992). Inhibition of nNOS markedly reduces the loss of dopamine neurones and clinical symptoms in a baboon model of Parkinson's disease (Hantraye *et al.*, 1996) and inhibition of NOS is protective in models of Huntington's disease (Deckel, 2001; Schultz *et al.*, 1995). Finally, significant changes in nNOS activity in the cerebrum and cerebellum follow the administration of metals such as aluminium and mercury, and suggest an involvement of this mediator in cerebral diseases induced by metals (Cucarella *et al.*, 1998).

The influence of peripheral systemic inflammatory conditions on the expression of central NOS isoforms is still controversial, however, overproduction of NO by iNOS is known to play a pathological role in acute inflammatory disorders of the CNS. There are increased levels of NO production in viral and bacterial infections such as meningitis, and a role for NO has been clearly indicated in the disruption of the blood-brain barrier during inflammatory conditions (Brian *et al.*, 1995; Visser *et al.*, 1994; Zheng *et al.*, 1993). There is little data available as to how inhibitors of iNOS modulate the course of these diseases, but considering

the role played by NO in defences against infection, it is logical to assume that they exacerbate the disease. Furthermore, substantial progress has been made towards identifying NO as a potential toxic mediator in inflammatory encephalitic diseases (Parkinson *et al.*, 1999). NO has been implicated as a potential mediator of microglia-dependent primary demyelination, a hallmark of multiple sclerosis, and iNOS induction has been noted in the brains of patients with this autoimmune disease (Bo *et al.*, 1994). NO may also be involved in the pathogenesis of sporadic amyotrophic lateral sclerosis and that of AIDS dementia. In the latter condition, neurotoxicity induced by certain HIV coat proteins is partially mediated by activation of NOS (Dawson *et al.*, 1993), whereas HIV patients who develop severe dementia exhibit a substantial increase in cortical iNOS (Adamson *et al.*, 1996).

## Nitric oxide in the peripheral nervous system

### Introduction

First debated in the late 1980s (Bowman *et al.*, 1986; Gillespie, 1987; Gillespie *et al.*, 1989; Li & Rand, 1989a), it is now well established that NO has a leading role as an inhibitory neurotransmitter of peripheral non-adrenergic, non-cholinergic (NANC) nerves. Peripheral nitrergic nerves have a widespread distribution, and are particularly important in that they produce relaxation of smooth muscle in the gastrointestinal, respiratory, vascular and urogenital systems. It is generally assumed that free NO is the transmitter substance released by nitrergic nerves (Lilley & Gibson, 1997; Martin *et al.*, 1994; Wood & Garthwaite, 1994). However, several controversial experimental observations (Barbier & Lefebvre, 1992; Gillespie & Sheng, 1990; Hobbs *et al.*, 1991; Rajanayagam *et al.*, 1993) have pointed to the possibility of obtaining a closely related redox product from NO (Stamler *et al.*, 1992), and also suggest that the inhibitory nitrergic transmitter is a NO-releasing molecule (Myers *et al.*, 1990; Thomas & Ramwell, 1989; Vedernikov *et al.*, 1992).

Furthermore, there is evidence that NO is not the only mediator involved in NANC neurotransmission, and that the release of a combination of messengers is involved in inhibitory NANC responses. Vasoactive peptide (VIP) seems to be the most important of these but roles for ATP, neuropeptide Y, enkephalin, peptide histidine isoleucine, carbon monoxide, pituitary adenyl cyclase activating peptide, gastrin-releasing peptide and dynorphin have also been suggested (Furness *et al.*, 1995). There is some evidence for the existence of 'solely nitrergic neurones' (Hohler *et al.*, 1995; Smet *et al.*, 1994), but most functional and morphological studies have demonstrated the existence of neurones and nerve fibres containing various neurotransmitters (D'Amato *et al.*, 1992). The relative contribution of such mediators varies greatly depending on factors such as stimuli and the tissue innervated. There is still discrepancy as to whether neurones release mediators upon stimulation, most likely to be the case because of their co-localization, or whether they are activated to release, preferentially, a specific neurotransmitter (Furness *et al.*, 1995; Schemann *et al.*, 1995; Smet *et al.*, 1994). Considerably more controversial is the

proposal of a sequential link between NO and VIP (Boeckxstaens *et al.*, 1991b; Ellis & Undem, 1992; Grider *et al.*, 1992; Murthy *et al.*, 1995). According to this hypothesis, VIP is the primary NANC neurotransmitter, but its release is dependent on NO synthesis. Furthermore, relaxation by VIP is produced partially *via* activation of adenylate cyclase and partially *via* stimulation of NO production in the smooth muscle. Said NO may relax the smooth muscle through activation of sGC and diffuse back to the nerve terminal to further enhance VIP release (Dick & Lefebvre, 2000; Teng *et al.*, 1988).

### Effects of peripherally released NO

**Gastrointestinal system** In the gastrointestinal tract the majority of NOS positive fibres are intrinsic, with smooth muscle cells containing sCG next to axon varicosities containing NOS (Bredt *et al.*, 1990; Ekblad *et al.*, 1994). This neuronally produced NO is implicated in many physiological and pathophysiological reflexes in which changes in gastrointestinal muscle relaxation are noted (Barrachina *et al.*, 2001; Calatayud *et al.*, 2001). Dysfunction of the inhibitory NANC nerves in the lower oesophageal sphincter results in the motility disorder, achalasia (Mearin *et al.*, 1993), and is probably involved in oesophageal spasms and related primary motor disorders in the oesophageal body (Yamato *et al.*, 1992). Gastric NANC-mediated relaxation following vagal stimulation, food intake or distension of the antrum or duodenum was among the first NANC effects described (Martinson, 1965). Indeed, pyloric hypertrophy and gastric distension are the most prominent abnormalities in nNOS knock out mice (Huang *et al.*, 1993). The importance of such inhibitory NANC neurones in human gastric function is illustrated by the frequent complaint by vagotomized patients of epigastric bloating and discomfort after meals, which has been related to an abnormal gastric receptive relaxation (Koster & Madsen, 1979; Troncon *et al.*, 1995). Infantile hypertrophic pyloric stenosis has been attributed to a lack of NOS-containing nerves at the pylorus (Vanderwinden *et al.*, 1992) while diabetic gastropathy has been linked to a loss of nNOS that can be treated with insulin and the phosphodiesterase-5 inhibitor sildenafil (Watkins *et al.*, 2000). A similar diminution in nNOS expression and activity in the myenteric plexus is associated with the delay in colonic transit appearing with advanced age (Takahashi *et al.*, 2000). In contrast, increases in the activity of NANC nerves have been held responsible for the decreased gastrointestinal motility appearing during pregnancy (Shah *et al.*, 2001). Similarly, the severe constipation characterizing Hirschsprung's disease results from the absence of intramural inhibitory NANC in the smooth muscle cells of the internal anal sphincter (Vanderwinden *et al.*, 1993). Furthermore, *in vivo* topical application of nitroglycerin causes a reduction in human anal pressure, pointing to the use of NO-donors in conditions such as anal fissures, haemorrhoids and proctalgia (Loder *et al.*, 1994).

**Pulmonary system** The density of extrinsic NOS-containing fibres increases progressively from the top of the trachea to the primary bronchi, and then diminishes as the bronchial diameter decreases (Fischer *et al.*, 1993; Fischer & Hoffmann, 1996). Nitrergic nerves are believed to represent the main



nervous bronchodilator pathway in humans, and dysfunction of this system may be implicated in the increased tone and hyper-responsiveness observed in asthma (Belvisi *et al.*, 1995). Furthermore, inhalation of NO has become an important therapeutic tool in the treatment of diseases such as acute respiratory distress syndrome, hypoxic respiratory failure, high pulmonary artery pressure, lung transplantation, sickle cell disease and specifically paediatric conditions such as neonatal pulmonary hypertension (Weinberger *et al.*, 1999; 2001; Weinacker & Vaszar, 2001). Indeed, new approaches to the treatment of some of these diseases involve potentiation of NO responses with inhibitors of phosphodiesterase-5 (Fernandes *et al.*, 1994; Gibson, 2001; Weinberger *et al.*, 2001).

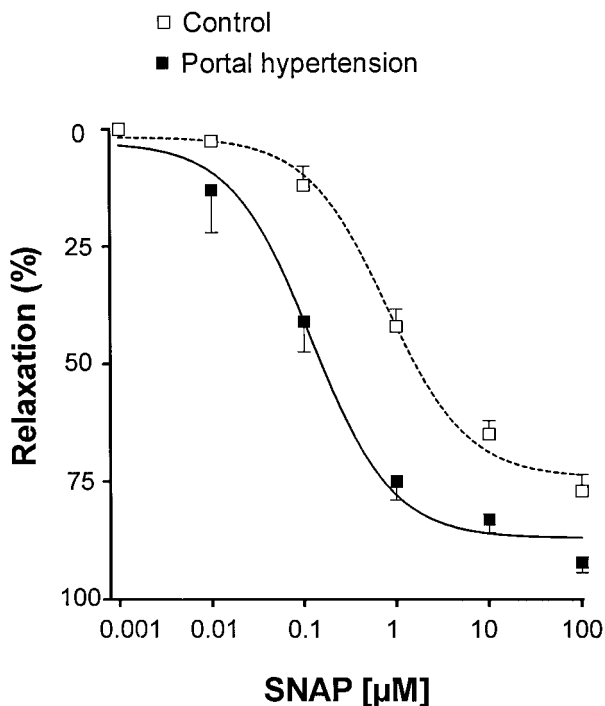
**Vascular system** nNOS is found in the perivascular nerves of various blood vessels and appears to constitute an alternative regional control mechanism for blood flow, independent of eNOS (Bredt *et al.*, 1990; Huang *et al.*, 1999; Martinez Cuesta *et al.*, 1996; Figure 5). This neuronally produced NO seems to be particularly relevant in the regulation of cerebral blood flow (Estrada & Defelipe, 1998; Faraci & Heistad, 1998). High levels of nNOS are present in vasodilator nerves in cerebral blood vessels (Bredt *et al.*, 1990; Thomsen *et al.*, 1993), although, in most cases, nNOS is co-localized with different vasoactive neurotransmitters (Bredt *et al.*, 1990; Estrada & Defelipe, 1998). In the brain, activity-dependent activation of nNOS is associated with a local increase in blood flow, and this response is

prevented by inhibitors of NOS (Iadecola *et al.*, 1993). The initial vascular response to neuronal ischaemia and the implication of nNOS in this condition have been discussed here previously. In addition to this relationship, it has been suggested that blockade of NANC vasodilatation by haemolysate or haemoglobin may contribute to the vaso-spasm observed in haemorrhages (Estrada & Defelipe, 1998). Abnormal dilatation of cerebral vessels appears to mediate vascular headaches. Furthermore, the finding that blockade of NO synthesis aborts acute attacks of migraine points to the use of the pharmacological manipulation of nNOS in the development of anti-migraine compounds (Thomsen & Olesen, 1998).

**Urogenital system** nNOS is most prominent in the parasympathetic postganglionic innervation of the urethra. Likewise, stimulation of bladder afferent nerves leads to the release of NO and chronic irritation of the bladder augments nNOS expression in dorsal root ganglion cells. Finally, bladder hyperactivity provoked by intravesical irritants can be moderated by inhibition of NO synthesis, thus suggesting a role for spinal cord NO in the micturition reflex pathway (de Groat & Yoshimura, 2001).

Recent years have seen a major focus on the pharmacological modulation of the NO released by the endothelium and nitrenergic nerves and which is involved in penile erection. nNOS neurones innervate the corpus cavernosum and blood vessels of the penis and nerve stimulation leads to erection, which involves sGC stimulation and is blocked by NOS inhibitors (Burnett *et al.*, 1992; Holmquist *et al.*, 1992; Rajfer *et al.*, 1992). Nitrenergic neurones are also implicated in the effects of sexual hormones. For instance, nNOS levels in the penis decrease substantially after castration but return to normal levels following testosterone replacement (Penson *et al.*, 1996). Levels of nNOS diminish with age, and this decrease correlates with impaired erectile responses (Carrier *et al.*, 1997). Similarly, impotence occurring with diabetes mellitus, spinal cord injury and treatments for prostate cancer is now related to damage of the nitrenergic structures controlling erection (Goldstein *et al.*, 1998). Phosphodiesterase-5 is the isoenzyme predominantly responsible for cyclic GMP hydrolysis in the corpus cavernosum, and recently, different isoforms of this isozyme have been described (Lin *et al.*, 2000). Selective inhibition with drugs such as zaprinast or sildenafil restores erectile responses, which are linked to prolongation of the NO/sGC/cyclic GMP signalling pathway (Bivalacqua *et al.*, 2000; Gibson, 2001; Saenz de Tejada *et al.*, 1989). However, this mechanism of action is dependent on a level of integrity of the nitrenergic nerves and a pre-activated endogenous NO-cyclic GMP system. This explains the clinical observation that sildenafil does not aid erection in patients with complete loss of sacral nerve activity nor where there is an absence of sexual arousal (Maytom *et al.*, 1999).

Nitrenergic structures also innervate smooth muscle structures in the female urogenital tract, and are particularly abundant in the clitoral corpus cavernosum (Burnett *et al.*, 1997; Papka *et al.*, 1995) where they appear to be responsible for the NANC erectile response of the clitoris (Cellek & Moncada, 1998). There have been few studies of female sexual dysfunction, but existing results suggest that inhibitors of phosphodiesterase-5 may be effective in specific cases, particularly those associated with the use of anti-depressant,



**Figure 5** Selective destruction of nitrenergic nerves in a model of portal hypertension results in supersensitivity of vascular tissue to the effects of exogenously administered NO. The graph shows cumulative concentration-response to SNAP in isolated mesenteric veins from control and portal hypertensive rats. Relaxations induced by SNAP are expressed as % of decrease induced by KCL (30 mM), and each point is the mean  $\pm$  s.e.mean of at least five experiments.

anti-psychotic and anti-anxiety agents (Shen *et al.*, 1999). Encouraging results have also been obtained with sildenafil in other cases of sexual dysfunction (Kaplan *et al.*, 1999; Sipski *et al.*, 2000), but these need confirmation. The use of sildenafil to aid *in vitro* fertilization is also a possibility since its application in the vagina increases both uterine blood flow and the thickness of the endometrium (Chwalisz & Garfield, 2000). Finally, NO appears to be responsible for a tonic inhibition of spontaneous contractile activity in the uterus while there is evidence of increased biosynthesis of NO during pregnancy and a rapid drop in NOS activity preceding delivery. This points to the involvement of nitergic mechanisms during pregnancy, which promote a relaxed state in the uterus, whereas a decrease in responsiveness to NO would appear to be involved in the initiation of labour (Chwalisz & Garfield, 2000; Weiner & Thompson, 1997).

**Skeletal muscle** The high levels of nNOS expressed in skeletal muscle, particularly the muscle-specific splice variant nNOS $\mu$ , tend to be located beneath the sarcolemma of fast twitch fibres, emphasizing the role of NO as a modulator of contractile force (Kobzik *et al.*, 1994; Nakane *et al.*, 1993). NO derived from sarcolemmal nNOS is also implicated in various other physiological functions occurring near the muscle membrane. Myocytes fuse to form muscle myotubes during muscle development, and this process is prevented by inhibition of NO (Lee *et al.*, 1994). In myocyte/motor neuron co-cultures, NO produced at the postsynaptic muscle membrane functions as a retrograde messenger, regulating myotube innervation (Wang *et al.*, 1995). In mature muscle fibres, NOS modulates glucose uptake across the sarcolemma. Although glucose uptake in skeletal muscle is regulated by both rigorous exercise and insulin, inhibition of NO synthesis has a selective action on glucose uptake in the former (Roberts *et al.*, 1997). Interestingly, regular exercise increases nNOS protein expression in the muscle and this has long-lasting enhancing effects on glucose transport in the muscle (Roberts *et al.*, 1997). Finally, both eNOS and iNOS isoforms are also present in the skeletal muscle, the former mostly related with the control of skeletal blood flow and the latter with inflammatory conditions and responses elicited by cytokines or lipopolysaccharides (LPS) (Stamler & Meissner, 2001).

Several muscular diseases have been linked to a dystrophin deficiency, and although the specific cause is unconfirmed, perturbed NO signalling would seem to be responsible (Brenman *et al.*, 1995). A mutation in the rod-like domain of dystrophin causes Becker's dystrophy and results in a loss of sarcolemmal nNOS, while other components of the dystrophin complex are preserved (Chao *et al.*, 1996).

## References

- ADAMSON, D.C., WILDEMANN, B., SASSAKI, M., GLASS, J.D., MCARTHUR, J.C., CHRISTOV, V.I., DAWSON, T.M. & DAWSON, V.L. (1996). Immunologic NO synthase: elevation in severe AIDS dementia and induction by HIV-1 gp41. *Science*, **274**, 1917–1921.
- AGUILLA, M.C. (1994). Growth hormone-releasing factor increases somatostatin release and mRNA levels in the rat paraventricular nucleus via nitric oxide activation of guanylate cyclase. *Proc. Natl. Acad. Sci. U.S.A.*, **91**, 782–786.
- ALDERTON, W.K., COOPER, C.E. & KNOWLES, R.G. (2001). Nitric oxide synthases: structure, function and inhibition. *Biochem. J.*, **357**, 593–615.
- ALMEIDA, A., ALMEIDA, J., BOLAÑOS, J.P. & MONCADA, S. (2001). Different responses of astrocytes and neurons to nitric oxide: The role of glycolytically generated ATP in astrocyte protection. *Proc. Natl. Acad. Sci. U.S.A.*, **98**, 15294–15299.

Similarly, patients with Duchenne muscular dystrophy and *mdx* mice that lack dystrophin both exhibit a reduction of nNOS in the sarcolemma (Brenman *et al.*, 1995; Chao *et al.*, 1996). This diminishing of dystrophin disrupts the normal link between the extracellular matrix and myofiber cytoskeleton (Campbell, 1995), which results in sarcolemmal damage and myofiber necrosis. Sarcolemmal instability in Duchenne dystrophy leads to a repeated cycle of myofibre degeneration and subsequent regeneration. Redistribution of nNOS from sarcolemma to cytosol is thought to be involved in myofibre necrosis, whereas the involvement of NO in myofibre differentiation suggests that an altered sarcolemmal nNOS signalling contributes to failed muscle regeneration in Duchenne dystrophy.

## Concluding remarks

The last few years have seen the publication of a plethora of information concerning the multiple roles played by NO in the nervous system. Although we now know that NO is involved in many aspects of CNS function, we are still far away from the pharmacological breakthrough which could have a clinical impact in CNS related diseases other than stroke. This is not the case in the peripheral nervous system where the introduction of sildenafil has represented a major breakthrough in a condition for which, until now, there existed complicated and only partially effective drug treatment or simply a psychological approach. However, one should bear in mind the many advances in our knowledge in this field, and how our insight has improved regarding diseases about which we knew relatively little 10 years ago. The field continues to grow and recent important findings include the role of NO in mitochondrial function, clarification of signalling pathways, targeting of diseases associated with organ-specific changes in nitergic activity, development of more selective NOS inhibitors and the identification of different isoforms of phosphodiesterase-5. These discoveries raise the prospect of future therapeutic leads, thus achieving the pharmacological goal of linking physiological knowledge with drug development.

The author would like to thank Prof Salvador Moncada for his insight on the topics discussed. The help of Annie Higgs, Prof Arnold Herman, Selim Cellek, Maria Dolores Barrachina, Dora Martí and Brian Normanly is also very much appreciated. The writing of this manuscript has been supported by CICYT (Comisión Interministerial de Ciencia y Tecnología) grants IFD97-1029, SAF98-0118 and SAF2001-0763.

- BAGETTA, G., CORASANITI, M.T., MELINO, G., PAOLETTI, A.M., FINAZZI-AGRO, A. & NISTICO, G. (1993). Lithium and tacrine increase the expression of nitric oxide synthase mRNA in the hippocampus of rat. *Biophys. Res. Commun.*, **197**, 1132–1139.
- BARBIER, A.J. & LEFEBVRE, R.A. (1992). Effect of LY 83583 on relaxation induced by non-adrenergic non-cholinergic nerve stimulation and exogenous nitric oxide in the rat gastric fundus. *Eur. J. Pharmacol.*, **219**, 331–334.
- BARBIER, A.J. & LEFEBVRE, R.A. (1995). Relaxant influence of phosphodiesterase inhibitors in the cat gastric fundus. *Eur. J. Pharmacol.*, **276**, 41–47.
- BARNETTE, M.S., GROUS, M., MANNING, C.D., CALLAHAM, J.F. & BARONE, F.C. (1990). Inhibition of neurally induced relaxation of canine lower esophageal sphincter by opioid peptides. *Eur. J. Pharmacol.*, **182**, 363–368.
- BARRACHINA, M.D., PANES, J. & ESPLUGUES, J.V. (2001). Role of nitric oxide in gastrointestinal inflammatory and ulcerative diseases: perspective for drugs development. *Curr. Pharm. Des.*, **7**, 31–48.
- BAYGUINOV, O. & SANDERS, K.M. (1993). Role of nitric oxide as an inhibitory neurotransmitter in the canine pyloric sphincter. *Am. J. Physiol.*, **264**, G975–G983.
- BELTRAN, B., BARRACHINA, M.D., MENEZ, A., QUINTERO, E. & ESPLUGUES, J.V. (1999). Synthesis of nitric oxide in the dorsal nucleus of the vagus mediates the inhibition of gastric acid secretion by central bombesin. *Br. J. Pharmacol.*, **127**, 1603–1610.
- BELTRÁN, B., MATHUR, A., DUCHEN, M., ERUSALIMSKY, J.D. & MONCADA, S. (2000). The effect of nitric oxide on cell respiration: a key to understanding its role in cell survival or death. *Proc. Natl. Acad. Sci. U.S.A.*, **26**, 14602–14607.
- BELVISI, M.G., WARD, J.K., MITCHELL, J.A. & BARNES, P.J. (1995). Nitric oxide as a neurotransmitter in human airways. *Arch. Int. Pharmacodyn. Ther.*, **329**, 97–110.
- BENDER, A.T., SILVERSTEIN, A.M., DEMADY, D.R., KANELAKIS, K.C., NOGUCHI, S., PRATT, W.B. & OSAWA, Y. (1999). Neuronal nitric-oxide synthase is regulated by the Hsp90-based chaperone system in vivo. *J. Biol. Chem.*, **274**, 1472–1478.
- BIVALACQUA, T.J., CHAMPION, H.C., HELSTROM, W.J. & KADOWITZ, P.J. (2000). Pharmacotherapy for erectile dysfunction. *Trends Pharmacol. Sci.*, **21**, 484–489.
- BO, L., DAWSON, T.M., WESSELINGH, S., MORK, S., CHOI, S., KONG, P.A., HANLEY, D. & TRAPP, B.D. (1994). Induction of nitric oxide synthase in demyelinating regions of multiple sclerosis brains. *Ann. Neurol.*, **36**, 778–786.
- BOECKXSTAENS, G.E., DE MAN, J.G., PELCKMANS, P.A., HERMAN, A.G. & VAN MAERCKE, Y.M. (1993). Alpha 2-adrenoceptor-mediated modulation of the nitric innervation of the canine isolated ileocolonic junction. *Br. J. Pharmacol.*, **109**, 1079–1084.
- BOECKXSTAENS, G.E., PELCKMANS, P.A., BULT, H., DE MAN, J.G., HERMAN, A.G. & VAN MAERCKE, Y.M. (1991a). Evidence for nitric oxide as mediator of non-adrenergic non-cholinergic relaxations induced by ATP and GABA in the canine gut. *Br. J. Pharmacol.*, **102**, 434–438.
- BOECKXSTAENS, G.E., PELCKMANS, P.A., RAMPART, M., RUYTJENS, I.F., VERBEUREN, T.J., HERMAN, A.G. & VAN MAERCKE, Y.M. (1990). GABA receptor-mediated stimulation of non-adrenergic non-cholinergic neurones in the dog ileocolonic junction. *Br. J. Pharmacol.*, **101**, 460–464.
- BOECKXSTAENS, G.E., PELCKMANS, P.A., RUYTJENS, I.F., BULT, H., DE MAN, J.G., HERMAN, A.G. & VAN MAERCKE, Y.M. (1991b). Bioassay of nitric oxide released upon stimulation of non-adrenergic non-cholinergic nerves in the canine ileocolonic junction. *Br. J. Pharmacol.*, **103**, 1085–1091.
- BOGERS, J.J., PELCKMANS, P.A., BOECKXSTAENS, G.E., DE MAN, J.G., HERMAN, A.G. & VAN MAERCKE, Y.M. (1991). The role of nitric oxide in serotonin-induced relaxations in the canine terminal ileum and ileocolonic junction. *Naunyn Schmiedeberg's Arch. Pharmacol.*, **344**, 716–719.
- BOHME, G.A., BON, C., LEMAIRE, M., REIBAUD, M., PIOT, O., STUTZMANN, J.M., DOBLE, A. & BLANCHARD, J.C. (1993). Altered synaptic plasticity and memory formation in nitric oxide synthase inhibitor-treated rats. *Proc. Natl. Acad. Sci. U.S.A.*, **90**, 9191–9194.
- BOHME, G.A., BON, C., STUTZMANN, J.M., DOBLE, A. & BLANCHARD, J.C. (1991). Possible involvement of nitric oxide in long-term potentiation. *Eur. J. Pharmacol.*, **199**, 379–381.
- BOJE, K.M. & AKORA, P.K. (1992). Microglial-produced nitric oxide synthase and reactive nitrogen oxides mediate neuronal cell death. *Brain Res.*, **587**, 250–256.
- BOLAÑOS, J.P. & ALMEIDA, A. (1999). Roles of nitric oxide in brain hypoxia-ischemia. *Biochim. Biophys. Acta*, **1411**, 415–436.
- BOLAÑOS, J.P., HEALES, S.J., LAND, J.M. & CLARK, J.B. (1995). Effect of peroxynitrite on the mitochondrial respiratory chain: differential susceptibility of neurones and astrocytes in primary culture. *J. Neurochem.*, **64**, 1965–1972.
- BOLAÑOS, J.P., PEUCHEN, S., HEALES, S.J., LAND, J.M. & CLARK, J.B. (1994). Nitric oxide-mediated inhibition of the mitochondrial respiratory chain in cultured astrocytes. *J. Neurochem.*, **63**, 910–916.
- BOWMAN, A., GILLESPIE, J.S. & SOARES DA SILVA, P. (1986). A comparison of the action of the endothelium-derived relaxant factor and the inhibitory factor from bovine retractor penis on rabbit aortic smooth muscle. *Br. J. Pharmacol.*, **87**, 175–181.
- BRAISSANT, O., GOTOH, T., LOUP, M., MORI, M. & BACHMANN, C. (1999). L-arginine uptake, the citrulline-NO cycle and arginase II in the rat brain: an in situ hybridization study. *Mol. Brain Res.*, **70**, 231–241.
- BREDT, D.S., FERRIS, C.D. & SNYDER, S.H. (1992). Nitric oxide synthase regulatory sites: phosphorylation by cyclic AMP dependent protein kinases, protein kinase C, calcium/calmodulin protein kinase; identification of flavin and calmodulin binding sites. *J. Biol. Chem.*, **267**, 10976–10981.
- BREDT, D.S., HWANG, P.M. & SNYDER, S.H. (1990). Localization of nitric oxide synthase indicating a neural role for nitric oxide. *Nature*, **347**, 768–770.
- BREDT, D.S., HWANG, P.M., GLATT, C.E., LOWENSTEIN, C., REED, R.R. & SNYDER, S.H. (1991). Cloned and expressed nitric oxide synthase structurally resembles cytochrome P-450 reductase. *Nature*, **351**, 714–718.
- BREDT, D.S. & SNYDER, S.H. (1989). Nitric oxide mediates glutamate-linked enhancement of cGMP levels in the cerebellum. *Proc. Natl. Acad. Sci. U.S.A.*, **86**, 9030–9033.
- BRENMAN, J.E., CHAO, D.S., GEE, S.H., MCGEE, A.W., CRAVEN, S.E., SANTILLANO, D.R., WU, Z., HUANG, F., XIA, H., PETERS, M.F., FROEHNER, S.C. & BREDT, D.S. (1996). Interaction of nitric oxide synthase with the postsynaptic density protein PSD-95 and alpha1-syntrophin mediated by PDZ domains. *Cell*, **84**, 757–767.
- BRENMAN, J.E., CHAO, D.S., XIA, H., ALDAPE, K. & BREDT, D.S. (1995). Nitric oxide synthase complexed with dystrophin and absent from skeletal muscle sarcolemma in Duchenne muscular dystrophy. *Cell*, **82**, 743–752.
- BRIAN, J.E., HEISTAD, D.D. & FARACI, F.M. (1995). Dilatation of cerebral arterioles in response to lipopolysaccharide in vivo. *Stroke*, **26**, 277–280.
- BROWN, G.C. (2000). Nitric oxide as a competitive inhibitor of oxygen consumption in the mitochondrial respiratory chain. *Acta Physiol. Scand.*, **168**, 667–674.
- BRÜNE, B. & LAPETINA, E.G. (1989). Activation of cytosolic ADP-ribosyltransferase by nitric oxide generating agents. *J. Biol. Chem.*, **264**, 8455–8458.
- BRÜNE, B. & LAPETINA, E.G. (1991). Phosphorylation of nitric oxide synthase by protein kinase A. *Biochem. Biophys. Res. Commun.*, **181**, 921–926.
- BURNETT, A.L., CALVIN, D.C., SILVER, R.I., PEPPAS, D.S. & DOCIMO, S.G. (1997). Immunohistochemical description of nitric oxide isoforms in human clitoris. *J. Urol.*, **158**, 75–78.
- BURNETT, A.L., LOWENSTEIN, C.J., BREDT, D.S., CHANG, T.S. & SNYDER, S.H. (1992). Nitric oxide: a physiologic mediator of penile erection. *Science*, **257**, 401–403.
- CALAPAI, G., SQUADRITO, F., ALTAVILLA, D., ZINGARELLI, B., CAMPO, G.M., CILIA, M. & CAPUTI, A.P. (1992). Evidence that nitric oxide modulates drinking behaviour. *Neuropharmacology*, **31**, 761–764.
- CALATAYUD, S., BARRACHINA, M.D. & ESPLUGUES, J.V. (2001). Nitric oxide – relation to integrity, injury and healing of the gastric mucosa. *Microsc. Res. Tech.*, **53**, 325–335.

- CAMPBELL, K.P. (1995). Three muscular dystrophies: loss of cytoskeleton-extracellular matrix linkage. *Cell*, **80**, 675–679.
- CARRIER, S., NAGARAJU, P., MORGAN, D.M., BABA, K., NUNES, L. & LUE, T.F. (1997). Age decreases nitric oxide-containing nerve fibers in the rat penis. *J. Urol.*, **157**, 1088–1092.
- CAZAL, L., GIARDINO, L. & CECCATELLI, S. (1993). NOS mRNA in the paraventricular nucleus of young and old rats after immobilization stress. *NeuroReport*, **4**, 627–630.
- CECCATELLI, S. & ERIKSSON, M. (1993). The effect of lactation on nitric synthase gene expression. *Brain Res.*, **625**, 177–179.
- CECCATELLI, S., HULTING, A.L., ZHANG, X., GUSTAFSSON, L., VILLAR, M. & HOKFELT, T. (1993). Nitric oxide synthase in the rat anterior pituitary gland and the role of nitric oxide in regulation of luteinizing hormone secretion. *Proc. Natl. Acad. Sci. U.S.A.*, **90**, 11292–11296.
- CELLEK, S. & MONCADA, S. (1998). Nitroergic neurotransmission mediates the non-adrenergic non-cholinergic responses in the clitoral corpus cavernosum of the rabbit. *Br. J. Pharmacol.*, **125**, 1627–1629.
- CHAO, D.S., GOROSPE, J.R., BRENNAN, J.E., RAFAEL, J.A., PETERS, M.F., FROEHLER, S.C., HOFFMAN, E.P., CHAMBERLAIN, J.S. & BREDDT, D.S. (1996). Selective loss of sarcolemmal nitric oxide in Becker muscular dystrophy. *J. Exp. Med.*, **184**, 609–618.
- CHAO, C.C., HU, S., MOLITOR, T.W., SHASKAN, E.G. & PETERSON, P.K. (1992). Activated microglia mediate neuronal cell injury via a nitric oxide mechanism. *J. Immunol.*, **149**, 2736–2741.
- CHOI, Y.B., TENNETI, L., LE, D.A., ORTIZ, J., BAI, G., CHEN, H.S. & LIPTON, S.A. (2000). Molecular basis for NMDA receptor-coupled ion channel modulation by S-nitrosylation. *Nat. Neurosci.*, **3**, 15–21.
- CHWALISZ, K. & GARFIELD, R.E. (2000). Role of nitric oxide in implantation and menstruation. *Hum. Reprod.*, **15**(Suppl. 3): 96–111.
- CLEMENTI, E., BROWN, G.C., FEELISCH, M. & MONCADA, S. (1998). Persistent inhibition of cell respiration by nitric oxide: crucial role of S-nitrosylation of mitochondrial complex I and protective action of glutathione. *Proc. Natl. Acad. Sci. U.S.A.*, **95**, 7631–7636.
- CONTESTABILE, A. (2000). Roles of NMDA receptor activity and nitric oxide production in brain development. *Brain Res. Rev.*, **32**, 476–509.
- COSTA, A., TRAINER, P., BESSER, M. & GROSSMAN, A. (1993). Nitric oxide modulates the release of corticotropin-releasing hormone from the rat hypothalamus in vitro. *Brain Res.*, **605**, 187–192.
- COUET, J., LI, S., OKAMOTO, T., IKEZU, T. & LISANTI, M.P. (1997). Identification of peptide and protein ligands for the scaffolding domain. Implications for the interaction of caveolin with caveolae-associated proteins. *J. Biol. Chem.*, **272**, 6525–6533.
- CROSBY, G., MAROTA, J.J. & HUANG, P.L. (1995). Intact nociception-induced neuroplasticity in transgenic mice deficient in neuronal nitric oxide synthase. *Neuroscience*, **69**, 1013–1017.
- CUCARELLA, C., MONTOLIU, C., HERMENEGILDO, C., SAEZ, R., MANZO, L., MIÑANA, M.D. & FELIPO, V. (1998). Chronic exposure to aluminum impairs glutamate-nitric oxide-cyclic GMP pathway. *J. Neurochem.*, **70**, 1609–1614.
- CUDEIRO, J. & RIVADULLA, C. (1999). Sight and insight - on the physiological role of nitric oxide in the visual system. *Trends Neurosci.*, **22**, 109–116.
- D'AMATO, M., CURRO, D., MONTUSCHI, P. & CURRO, D. (1992). Evidence for dual components in the non-adrenergic non-cholinergic relaxation in the rat gastric fundus: role of endogenous nitric oxide and vasoactive intestinal polypeptide. *J. Auton. Nerv. Syst.*, **37**, 175–186.
- DANIEL, H., HEMART, N., JAILLARD, D. & CREPEL, F. (1993). Long-term depression requires nitric oxide and guanosine 3':5' cyclic monophosphate production in rat cerebellar Purkinje cells. *Eur. J. Neurosci.*, **5**, 1079–1082.
- DAWSON, V.L., DAWSON, T.M., LONDON, E.D., BREDDT, D.S. & SNYDER, S.H. (1991). Nitric oxide mediates glutamate neurotoxicity in primary cortical cultures. *Proc. Natl. Acad. Sci. U.S.A.*, **88**, 6368–6371.
- DAWSON, V.L., DAWSON, T.M., UHL, G.R. & SNYDER, S.H. (1993). Human immunodeficiency virus type I coat protein neurotoxicity mediated by nitric oxide in primary cortical cultures. *Proc. Natl. Acad. Sci. U.S.A.*, **90**, 3256–3259.
- DAWSON, V.L., KIZUSHI, V., HUANG, P.L., SNYDER, S.H. & DAWSON, T.M. (1996). Resistance to neurotoxicity in cortical cultures from neuronal nitric oxide synthase-deficient mice. *J. Neurosci.*, **16**, 2479–2487.
- DECKEL, A.W. (2001). Nitric oxide and nitric oxide synthase in Huntington's disease. *J. Neurosci. Res.*, **64**, 99–107.
- DE GROAT, W.C. & YOSHIMURA, N. (2001). Pharmacology of the lower urinary tract. *Annu. Rev. Pharmacol. Toxicol.*, **41**, 691–721.
- DE LUCA, B., MONDA, M. & SULLO, A. (1995). Changes in eating behavior and thermogenic activity following inhibition of nitric oxide formation. *Am. J. Physiol.*, **268**, R1533–R1538.
- DICK, J.M. & LEFEBVRE, R.A. (2000). Interplay between nitric oxide and vasoactive intestinal polypeptide in the pig gastric fundus smooth muscle. *Eur. J. Pharmacol.*, **397**, 389–397.
- DINERMAN, J.L., DAWSON, T.M., SCHELL, M.J., SNOWMAN, A. & SNYDER, S.H. (1994). Endothelial nitric oxide synthase localized to hippocampal pyramidal cells: implications for synaptic plasticity. *Proc. Natl. Acad. Sci. U.S.A.*, **91**, 4214–4218.
- DRAPIER, J.C. & BOUTON, C. (1996). Modulation by nitric oxide of metalloprotein regulatory activities. *Bioessays*, **18**, 549–556.
- EKBLAD, E., ALM, P. & SUNDLER, F. (1994). Distribution, origin and projections of nitric oxide synthase-containing neurons in gut and pancreas. *Neuroscience*, **63**, 233–248.
- ELIASSON, M.J.L., SAMPEI, K., MANDIR, A.S., HURN, P.D., TRAYSTMAN, R.J., BAO, J., PIEPER, A., WANG, Z.Q., DAWSON, T.M., SNYDER, S.H. & DAWSON, V.L. (1997). Poly(ADP-ribose) polymerase gene disruption renders mice resistant to cerebral ischemia. *Nat. Med.*, **3**, 1089–1095.
- ELLIS, J.L. & UNDEM, B.J. (1992). Inhibition by L-NG-nitro-L-arginine of nonadrenergic-noncholinergic-mediated relaxations of human isolated central and peripheral airway. *Am. Rev. Respir. Dis.*, **146**, 1543–1547.
- ENDO, M., MAIESE, K. & WAGNER, J.A. (1994). Expression of the inducible form of nitric oxide synthase by reactive astrocytes after transient global ischemia. *Brain Res.*, **651**, 92–100.
- ESPLUGUES, J.V., BARRACHINA, M.D., BELTRAN, B., CALATAYUD, S., WHITTLE, B.J.R. & MONCADA, S. (1996). Inhibition of gastric acid secretion by stress: a protective reflex mediated by cerebral nitric oxide. *Proc. Natl. Acad. Sci. U.S.A.*, **93**, 14839–14844.
- ESTRADA, C. & DEFELIPE, J. (1998). Nitric oxide-producing neurons in the neocortex: morphological and functional relationship with intraparenchymal microvasculature. *Cerebral Cortex*, **8**, 193–203.
- FANG, M., JAFFREY, S.R., SAWA, A., YE, K., LUO, X. & SNYDER, S.H. (2000). Dexas 1: a G protein specifically coupled to neuronal nitric oxide synthase via CAPON. *Neuron*, **28**, 183–193.
- FARACI, F.M. & HEISTAD, D.D. (1998). Regulation of the cerebral circulation: role of the endothelium and potassium channels. *Physiol. Rev.*, **75**, 53–97.
- FERGUS, A. & LEE, K.S. (1997). Regulation of cerebral microvessels by glutamatergic mechanisms. *Brain Res.*, **754**, 35–45.
- FERNANDES, L.B., ELLIS, J.L. & UNDEM, B.J. (1994). Potentiation of nonadrenergic noncholinergic relaxation of human isolated bronchus by selective inhibitors of phosphodiesterase isozymes. *Am. J. Respir. Crit. Care Med.*, **150**, 1384–1390.
- FISCHER, A. & HOFFMANN, B. (1996). Nitric oxide synthase in neurons and nerve fibers of lower airways and in vagal sensory ganglia of man: Correlation with neuropeptides. *Am. J. Respir. Crit. Care Med.*, **154**, 209–216.
- FISCHER, A., MUNDEL, P., MAYERM, B., PREISSLER, U., PHILIPPIN, B. & KUMMER, W. (1993). Nitric oxide synthase in guinea pig lower airway innervation. *Neurosci. Lett.*, **149**, 157–160.
- FUKUYAMA, N., TAKIZAWA, S., ISHIDA, H., HOSHIAI, K., SHINOHARA, Y. & NAKAZAWA, H. (1998). Peroxynitrite formation in focal cerebral ischemia-reperfusion in rats occurs predominantly in the peri-infarct region. *J. Cereb. Blood Flow Metab.*, **18**, 123–129.
- FURNESS, J.B., YOUNG, H.M., POMPOLO, S., BORNSTEIN, J.C., KUNZE, W.A.A. & MCCONALOGUE, K. (1995). Plurichemical transmission and chemical coding of neurons in the digestive tract. *Gastroenterology*, **108**, 554–563.

- GARCÍA-CERDEÑA, G., MARTASEK, P., MASTERS, B.S.S., SKIDD, P.M., COUET, J., LI, S., LISANTI, M.P. & SESSA, W.C. (1997). Dissecting the interaction between nitric oxide synthase and caveolin. Functional significance of the NOS caveolin binding domain in vivo. *J. Biol. Chem.*, **272**, 25437–25440.
- GARCÍA-ZARAGOZA, E., BARRACHINA, M.D., MORENO, L. & ESPLUGUES, J.V. (2000). Role of central glutamate receptors, nitric oxide and soluble guanylyl cyclase in the inhibition by endotoxin of rat gastric acid secretion. *Br. J. Pharmacol.*, **130**, 1283–1288.
- GARTHWAITE, J. & BOULTON, C.L. (1995). Nitric oxide signaling in the central nervous system. *Annu. Rev. Physiol.*, **57**, 683–706.
- GARTHWAITE, J., CHARLES, S.L. & CHESS-WILLIAMS, R. (1988). Endothelium derived relaxing factor release on activation of NMDA receptors suggests a role as intercellular messenger in the brain. *Nature*, **336**, 385–388.
- GARTHWAITE, J., GARTHWAITE, G., PALMER, R.M. & MONCADA, S. (1989). NMDA receptor activation induces nitric oxide synthesis from arginine in rat brain slices. *Eur. J. Pharmacol.*, **172**, 413–416.
- GIBSON, A. (2001). Phosphodiesterase 5 inhibitors and nitrergic transmission: from zaprinast to sildenafil. *Eur. J. Pharmacol.*, **411**, 1–10.
- GIBSON, A. & MIRZAZADEH, S. (1989). N-methylhydroxylamine inhibits and M&B 22948 potentiates relaxations of the mouse anococcygeus to non-adrenergic, non-cholinergic field stimulation and to nitrovasodilator drugs. *Br. J. Pharmacol.*, **96**, 637–644.
- GILLESPIE, J.S. (1987). Searching for the non-adrenergic non-cholinergic autonomic transmitter. In *Pharmacology*, ed. Rand, M.J., Raper, C. pp. 161–170. Amsterdam: Excerta Medica.
- GILLESPIE, J.S., LIU, X.R., MARTIN, W. & LIU, X. (1989). The effects of L-arginine and NG-monomethyl L-arginine on the response of the rat anococcygeus muscle to NANC nerve stimulation. *Br. J. Pharmacol.*, **98**, 1080–1082.
- GILLESPIE, J.S. & SHENG, H. (1990). The effects of pyrogallol and hydroquinone on the response to NANC nerve stimulation in the rat anococcygeus and the bovine retractor penis muscles. *Br. J. Pharmacol.*, **99**, 194–196.
- GOLDSTEIN, I., LUE, T.F., PADMA-NATHAN, H., ROSEN, R.C., STEERS, W.D. & WICKER, P.A. (1998). Oral sildenafil in the treatment of erectile dysfunction. *N. Engl. J. Med.*, **338**, 1397–1404.
- GRIDER, J.R., MURTHY, K.S., JIN, J.G. & MAKHLOUF, G.M. (1992). Stimulation of nitric oxide from muscle cells by VIP: prejunctional enhancement of VIP release. *Am. J. Physiol.*, **262**, G774–G778.
- GROLL-KNAPP, E., HAIDER, M., KIENZL, K., HANDLER, A. & TRIMMEL, M. (1988). Changes in discrimination learning and brain activity (ERPs) due to combined exposure to NO and CO in rats. *Toxicology*, **49**, 441–447.
- GUO, Y., WARD, M.E., BEASJOURS, S., MORI, M. & HUSSAIN, S.N.A. (1997). Regulation of cerebral nitric oxide production in response to prolonged in vivo hypoxia. *Neurosci. Res.*, **49**, 89–97.
- GUSTAFSSON, L.E., WIKLUND, C.U., WIKLUND, N.P., PERSSON, M.G. & MONCADA, S. (1990). Modulation of autonomic neuroeffector transmission by nitric oxide in guinea pig ileum. *Biochem. Biophys. Res. Commun.*, **173**, 106–110.
- HALEY, J.E., WILCOX, G.L. & CHAPMAN, P.F. (1992). The role of nitric oxide in hippocampal long-term potentiation. *Neuron*, **8**, 211–216.
- HAMMER, B., PARKER JR., W.D. & BENNETT JR., J.P. (1993). NMDA receptors increase OH radicals in vivo by using nitric oxide synthase and protein kinase C. *NeuroReport*, **5**, 72–74.
- HANBAUER, I., WINK, D., OSAWA, Y., EDELMAN, G.M. & GALLY, J.A. (1992). Role of nitric oxide in NMDA-evoked release of [<sup>3</sup>H]-dopamine from striatal slices. *NeuroReport*, **3**, 409–412.
- HANTRAYE, P., BROUILLET, E., FERRANTE, R., PALFI, S., DOLAN, R., MATTHEWS, R.T. & BEAL, M.F. (1996). Inhibition of neuronal nitric oxide synthase prevents MPTP-induced parkinsonism in baboons. *Nat. Med.*, **2**, 1017–1021.
- HAYASHI, Y., NISHIO, M., NAITO, Y., YOKOKURA, H., NIMURA, Y., HIDAKA, H. & WATANABE, Y. (1999). Regulation of neuronal nitric-oxide synthase by calmodulin kinases. *J. Biol. Chem.*, **274**, 20597–20602.
- HEMMENS, B., WOSCHITZ, S., PITTERS, E., KLOSCH, B., VOLKER, C., SCHMIDT, K. & MAYER, B. (1998). The protein inhibitor of neuronal nitric oxide synthase (PIN): characterization of its actions on pure nitric oxide synthases. *FEBS Lett.*, **430**, 397–400.
- HESS, D.T., PATTERSON, S.I., SMITH, D.S. & SKENE, J.H.P. (1993). Neuronal growth cone collapse and inhibition of protein fatty acylation by nitric oxide. *Nature*, **366**, 562–565.
- HOBBS, A.J., TUCKER, J.F. & GIBSON, A. (1991). Differentiation by hydroquinone of relaxations induced by exogenous and endogenous nitrates in non-vascular smooth muscle: role of superoxide anions. *Br. J. Pharmacol.*, **104**, 645–650.
- HOBBS, A.J., HIGGS, A. & MONCADA, S. (1999). Inhibition of nitric oxide synthase as a potential therapeutic target. *Annu. Rev. Pharmacol. Toxicol.*, **39**, 191–220.
- HOHLER, B., OLRY, R., MAYER, B. & KUMMER, W. (1995). Nitric oxide synthase in guinea pig sympathetic ganglia: correlation with tyrosine hydroxylase and neuropeptides. *Histochem. Cell Biol.*, **104**, 21–28.
- HOLMQUIST, F., HEDLUNG, H. & ANDERSSON, K. (1992). Characterization of inhibitory neurotransmission in the isolated corpus cavernosum from rabbit and man. *J. Physiol.*, **449**, 295–311.
- HOLSCHER, C. & ROSE, S.P. (1992). An inhibitor of nitric oxide synthesis prevents memory formation in the chick. *Neurosci. Lett.*, **145**, 165–167.
- HOYT, K.R., TANG, L.H., AIZENMAN, E. & REYNOLDS, I.J. (1992). Nitric oxide modulates NMDA-induced increases in intracellular Ca<sup>2+</sup> in cultured rat forebrain neurons. *Brain Res.*, **592**, 310–316.
- HUANG, P.L., DAWSON, T.M., BRETT, D.S., SNYDER, S.H. & FISHMAN, M.C. (1993). Targeted disruption of the neuronal nitric oxide synthase gene. *Cell*, **75**, 1273–1286.
- HUANG, P.L., GYURKO, R. & ZHANG, L. (1999). Cardiovascular effects of nitric oxide: lessons learned from endothelial nitric oxide synthase knockout mice. In *Endothelium, nitric oxide and atherosclerosis*, ed. Panza, J.A. & Cannon III, R.O. pp. 37–48. Armonk, NY: Futura Publishing Co.
- HUANG, Z., HUANG, P.L., MA, J., MENG, W., AYATA, C., FISHMAN, M.C. & MOSKOWITZ, M.A. (1996). Enlarged infarcts in endothelial nitric oxide synthase knockout mice are attenuated by nitro-L-arginine. *J. Cereb. Blood Flow Metab.*, **16**, 981–987.
- HUANG, Z., HUANG, P.L., PANAHIAN, N., DALKARA, T., FISHMAN, M.C. & MOSKOWITZ, M.A. (1994). Effects of cerebral ischemia in mice deficient in neuronal nitric oxide synthase. *Science*, **265**, 1883–1885.
- HUH, Y., HEO, K., PARK, C. & AHN, H. (2000). Transient induction of neuronal nitric oxide synthase in neurons of the rat cerebral cortex after status epilepticus. *Neurosci. Lett.*, **281**, 49–52.
- HUNTER, T. (2000). Signaling - 2000 and beyond. *Cell*, **100**, 113–127.
- HUSI, H., WARD, M.A., CHOUDHARY, J.S., BLACKSTOCK, W.P. & GRANT, S.G.N. (2000). Proteomic analysis of NMDA receptor-adhesion protein signaling complexes. *Nat. Neurosci.*, **3**, 661–669.
- HYMAN, B.T., MARZLOFF, K., WENNIGER, J.J., DAWSON, T.M., BRETT, D.S. & SNYDER, S.H. (1992). Relative sparing of nitric oxide synthase-containing neurons in the hippocampal formation in Alzheimer's disease. *Ann. Neurol.*, **32**, 818–820.
- IADECOLA, C. (1997). Bright and dark sides of nitric oxide in ischemic brain injury. *Trends Neurosci.*, **20**, 132–139.
- IADECOLA, C., XU, X., ZHANG, F., EL-FAKAHANY, E.E. & ROSS, M.E. (1995a). Marked induction of calcium-independent nitric oxide synthase activity after focal ischemia. *J. Cereb. Blood Flow Metab.*, **15**, 52–59.
- IADECOLA, C., ZHANG, F. & XU, X. (1993). Role of nitric oxide synthase-containing vascular nerves in cerebrovasodilation elicited from cerebellum. *Am. J. Physiol.*, **264**, R738–R746.
- IADECOLA, C., ZHANG, F., XU, S., CASEY, R. & ROSS, M.E. (1995b). Inducible nitric oxide synthase gene expression in brain following cerebral ischemia. *J. Cereb. Blood Flow Metab.*, **15**, 378–384.
- IWASE, K., IYAMA, K.-I., AKAGI, K., YANO, S., FUKUNAGA, K., MIYAMOTO, E., MORI, M. & TAKIGUCHI, M. (1998). Precise distribution of neuronal nitric oxide synthase mRNA in the rat brain revealed by non-radioisotopic in situ hybridisation. *Mol. Brain Res.*, **53**, 1–12.

- JAFFREY, S.R., ERDJUMENT-BROMAGE, H., FERRIS, C.D., TEMPST, P. & SNYDER, S.H. (2001). Protein S-nitrosylation: a physiological signal for neuronal nitric oxide. *Nat. Cell Biol.*, **3**, 193–197.
- JAFFREY, S.R., SNOWMAN, A.M., ELIASSON, M.J.L., COHEN, N.A. & SNYDER, S.H. (1998). CAPON: a protein associated with neuronal nitric oxide synthase that regulates its interaction with PSD95. *Neuron*, **20**, 115–124.
- JAFFREY, S.R. & SNYDER, S.H. (1996). PIN: an associated protein inhibitor of neuronal nitric oxide synthase. *Science*, **274**, 774–777.
- KADER, A., FRAZZINI, V.I., SOLOMON, R.A. & TRIFILETTI, R.R. (1993). Nitric oxide production during focal cerebral ischemia in rats. *Stroke*, **24**, 1709–1716.
- KAPAS, L., SHIBATA, M., KIMURA, M. & KRUEGER, J.M. (1994). Inhibition of nitric oxide synthesis suppresses sleep in rabbits. *Am. J. Physiol.*, **266**, R151–R157.
- KAPLAN, S.A., REIS, R.B., KOHN, I.J., IKEGUCHI, E.F., LAOR, E., TE, A.E. & MARTINS, A.C.P. (1999). Safety and efficacy of sildenafil in postmenopausal women with sexual dysfunction. *Urology*, **53**, 481–486.
- KELLER, J.N., KINDY, M.S., HOLTSBERG, F.W., ST CLAIR, D.K., YEN, H.C., GERMEYER, A., STEINER S.M., BRUCE KELLER, A.J., HUTCHINS, J.B. & MATTSON, M.P. (1998). Mitochondrial manganese superoxide dismutase prevents neural apoptosis and reduces ischemic brain injury: suppression of peroxynitrite production, lipid peroxidation, and mitochondrial dysfunction. *J. Neurosci.*, **18**, 687–697.
- KENDRICK, K.M., GUEVARA GUZMAN, R., ZORRILLA, J., HINTON, M.R., BROAD, K.D., MIMMACK, M. & OHKURA, S. (1997). Formation of olfactory memories mediated by nitric oxide. *Nature*, **388**, 670–674.
- KISS, J.P. & VIZI, E.S. (2001). Nitric oxide: a novel link between synaptic and nonsynaptic transmission. *Trends Neurosci.*, **24**, 211–215.
- KITCHENER, P.D., VAN DER ZEDE, C.E.E.M. & DIAMOND, J. (1993). Lesion induced NADPH-diaphorase reactivity in neocortical pyramidal neurones. *NeuroReport*, **4**, 487–490.
- KITTO, K.F., HALEY, J.E. & WILCOX, G.L. (1992). Involvement of nitric oxide in spinally mediated hyperalgesia in the mouse. *Neurosci. Lett.*, **148**, 1–5.
- KLATT, P., SCHMIDT, K. & MAYER, B. (1992). Brain nitric oxide synthase is a haemoprotein. *Biochem. J.*, **288**, 15–17.
- KNOWLES, R.G., PALACIOS, M., PALMER, R.M. & MONCADA, S. (1989). Formation of nitric oxide from L-arginine in the central nervous system: a transduction mechanism for stimulation of the soluble guanylate cyclase. *Proc. Natl. Acad. Sci. U.S.A.*, **86**, 5159–5162.
- KOBIK, L., REID, M.B., BRETT, D.S. & STAMLER, J.S. (1994). Nitric oxide in skeletal muscle. *Nature*, **372**, 546–548.
- KOH, J.Y. & CHOI, D. (1988). Vulnerability of cultured cortical neurons to damage by excitotoxins: differential susceptibility of neurons containing NADP-diaphorase. *J. Neurosci.*, **8**, 2153–2163.
- KOLESNIKOV, Y.A., PAN, Y.X., BABEY, A.M., JAIN, S., WILSON, R. & PASTERNAK, G.W. (1997). Functionally differentiating two neuronal nitric oxide isoforms through antisense mapping: evidence for opposing NO actions on morphine analgesia and tolerance. *Proc. Natl. Acad. Sci. U.S.A.*, **94**, 8220–8225.
- KOLLEGER, H., MCBEAN, G.J. & TIPTON, K.F. (1993). Reduction of striatal N-methyl-D-aspartate toxicity by inhibition of nitric oxide synthase. *Biochem. Pharmacol.*, **45**, 260–264.
- KOMEIMA, K., HAYASHI, Y., NAITO, Y. & WATANABE, Y. (2000). Inhibition of neuronal nitric-oxide synthase by calcium/calmodulin-dependent protein kinase II through Ser847 phosphorylation in NG108-NG115 neuronal cells. *J. Biol. Chem.*, **275**, 28139–28143.
- KORNAU, H.C., SCHENKER, L.T., KENNEDY, M.B. & SEEBURG, P.H. (1995). Domain interaction between NMDA receptor subunits and the postsynaptic density protein PSD-95. *Science*, **269**, 1737–1740.
- KOSTER, N. & MADSEN, P. (1979). The intragastric pressure before and immediately after truncal vagotomy. *Scand. J. Gastroenterol.*, **5**, 381–383.
- KURIYAMA, K. & OHKUMA, S. (1995). Role of nitric oxide in central synaptic transmission: effects on neurotransmitter release. *Jpn. J. Pharmacol.*, **69**, 1–8.
- LAFON-CAZAL, M., PIETRI, S., CULCASI, M. & BOCKAERT, J. (1993). NMDA-dependent superoxide production and neurotoxicity. *Nature*, **364**, 535–537.
- LANE, P., HAO, G. & GROSS, S.S. (2001). S-nitrosylation is emerging as a specific and fundamental posttranslational protein modification: head to head comparison with O-phosphorylation. *Sci SKTE*, **86**, 1–9.
- LEE, K.H., BAEK, M.Y., MOON, K.Y., SONG, W.G., CHUNG, C.H., HA, D.B. & KANG, M.S. (1994). Nitric oxide as a messenger molecule for myoblast fusion. *J. Biol. Chem.*, **269**, 14371–14374.
- LI, C.G. & RAND, M.J. (1989a). Evidence for a role of nitric oxide in the neurotransmitter system mediating relaxation of the rat anococcygeus muscle. *Clin. Exp. Pharmacol. Physiol.*, **16**, 933–938.
- LI, C.G. & RAND, M.J. (1989b). Prejunctional inhibition of non-adrenergic non-cholinergic transmission in the rat anococcygeus muscle. *Eur. J. Pharmacol.*, **168**, 107–110.
- LILLEY, E. & GIBSON, A. (1997). Release of the antioxidant ascorbate and urate from nitrergically-innervated smooth muscle. *Br. J. Pharmacol.*, **122**, 1746–1752.
- LIN, C.S., LAU, A., TU, R. & LUE, T.F. (2000). Expression of three isoforms of cGMP-binding cGMP-specific phosphodiesterase (PDE5) in human penile cavernosum. *Biochem. Biophys. Res. Commun.*, **268**, 628–635.
- LINCOLN, T.M., DEY, N. & SELLAK, H. (2001). cGMP-dependent protein kinase signaling mechanisms in smooth muscle: from the regulation of tone to gene expression. *J. Appl. Physiol.*, **91**, 1421–1430.
- LING, L., KARIUS, D.R., FISCUS, R.R. & SPECK, D.F. (1992). Endogenous nitric oxide required for an integrative respiratory function in the cat brain. *J. Neurophysiol.*, **68**, 1910–1912.
- LODER, P.B., KAMM, M.A., NICHOLLS, R.J. & PHILLIPS, R.K. (1994). 'Reversible chemical sphincterectomy' by local application of glyceryl trinitrate. *Br. J. Surg.*, **81**, 1386–1389.
- MALINSKI, T., BAILEY, F., ZHANG, Z.G. & CHOPP, M. (1993). Nitric oxide measured by porphyrinic microsensor in rat brain after transient middle cerebral artery occlusion. *J. Cereb. Blood Flow Metab.*, **13**, 355–358.
- MALMBERG, A.B. & YAKSH, T.L. (1993). Spinal nitric oxide synthesis inhibition blocks NMDA-induced thermal hyperalgesia and produces antinociception in the formalin test in rats. *Pain*, **54**, 291–300.
- MANI, S.K., ALLEN, J.M., RETTORI, V., MCCANN, S.M., O'MALLEY, B.W. & CLARK, J.H. (1994). Nitric oxide mediates sexual behavior in female rats. *Proc. Natl. Acad. Sci. U.S.A.*, **91**, 6468–6472.
- MAO, J. (1999). NMDA and opioid receptors: their interactions in antinociception, tolerance and neuroplasticity. *Brain Res. Rev.*, **30**, 289–304.
- MARKS, K.A., MALLARD, C.E., ROBERTS, I., WILLIAMS, C.E., GLUCKMAN, P.D. & EDWARDS, A.D. (1996). Nitric oxide synthase inhibition attenuates delayed vasodilation and increases injury after cerebral ischemia in fetal sheep. *Pediatr. Res.*, **40**, 185–191.
- MARTIN, W., MCALLISTER, K.H. & PAISLEY, K. (1994). NANC neurotransmission in the bovine retractor penis muscle is blocked by superoxide anion following inhibition of superoxide dismutase with diethylthiocarbamate. *Neuropharmacology*, **33**, 1293–1301.
- MARTINSON, J. (1965). Vagal relaxation of the stomach. Experimental re-investigation of the concept of transmission mechanism. *Acta Physiol. Scand.*, **64**, 453–462.
- MARTINEZ CUESTA, M.A., MORENO, L., PIQUE, J.M., BOSCH, J., RODRIGO, J. & ESPLUGUES, J.V. (1996). Nitric oxide-mediated  $\beta_2$ -adrenoceptor relaxation is impaired in mesenteric veins from portal-hypertensive rats. *Gastroenterology*, **111**, 727–735.
- MAYER, B., KLATT, P., BOHME, E. & SCHMIDT, K. (1992). Regulation of neuronal nitric oxide and cyclic GMP formation by  $Ca^{2+}$ . *J. Neurochem.*, **59**, 2024–2029.
- MAYER, D.J., MAO, J., HOLT, J. & PRICE, D.D. (1999). Cellular interactions of neuropathic pain, morphine tolerance, and their interactions. *Proc. Natl. Acad. Sci. U.S.A.*, **96**, 7731–7736.
- MAYTOM, M.C., DERRY, F.A., DINSMORE, W.W., GLASS, C.A., SMITH, M.D., ORR, M. & OSTERLOH, I.H. (1999). A two-part pilot study of sildenafil (Viagra<sup>®</sup>) in men with erectile dysfunction caused by spinal cord injury. *Spinal Cord*, **37**, 110–116.

- MEARIN, F., MOURELLE, M., GUARNER, F., SALAS, A., RIVEROS MORENO, V., MONCADA, S. & MALAGELADA, J.R. (1993). Patients with achalasia lack nitric oxide synthase in the gastro-oesophageal junction. *Eur. J. Clin. Invest.*, **23**, 724–728.
- MEFFERT, M.K., CALAKOS, N.C., SCHELLER, R.H. & SCHULMAN, H. (1996). Nitric oxide modulates synaptic vesicle docking fusion reactions. *Neuron*, **16**, 1229–1236.
- MELIS, M.R., STANCAMPIANO, R. & ARGIOLOS, A. (1994). Prevention by N<sup>G</sup>-nitro-L-arginine methyl ester of apomorphine and oxytocin induced penile erection and yawning: site of action in the brain. *Pharmacol. Biochem. Behav.*, **48**, 799–804.
- MELIS, M.R., STANCAMPIANO, R. & ARGIOLOS, A. (1995). Role of nitric oxide in penile erection and yawning induced by 5-HT<sub>1c</sub> receptor agonists in male rats. *Naunyn Schmiedeberg's Arch. Pharmacol.*, **351**, 439–445.
- MIGAUD, M., CHARLESWORTH, P., DEMPSTER, M., WEBSTER, L.C., WATABE, A.M., MAKHINSON, M., HE, Y., RAMSAY, M.F., MORRIS, R.G., MORRISON, J.J., O'DELL, T.J. & GRANT, S.G. (1998). Enhanced long-term potentiation and impaired learning in mice with mutant postsynaptic density-95 protein. *Nature*, **396**, 433–439.
- MINAMI, T., NISHIHARA, I., ITO, S., SAKAMOTO, K., HYODO, M. & HAYASHI, O. (1995). Nitric oxide mediates allodynia induced by intrathecal administration of prostaglandin E<sub>2</sub> or prostaglandin F<sub>2</sub>. *Pain*, **61**, 285–290.
- MOLLACE, V., RODINI, P., MASSOUD, R., ROTIROTI, D. & NIATICO, G. (1995). Age-dependent changes of NO synthase activity in the rat brain. *Biochem. Biophys. Res. Commun.*, **215**, 822–827.
- MONCADA, S. & ERUSALIMSKY, J.D. (2002). Nitric oxide and cell respiration: physiology and pathophysiology or just pharmacology? *Nature* (in press).
- MONCADA, S., HIGGS, A. & FURCHGOTT, R. (1997). International Union of Pharmacology nomenclature in nitric oxide research. *Pharmacol. Rev.*, **49**, 137–142.
- MONTAGUE, P.R., GANCAICO, C.D., WINN, M.J., MARCHASE, R.B. & FRIEDLANDER, M.J. (1994). Role of NO production in NMDA receptor-mediated neurotransmitter release in cerebral cortex. *Science*, **263**, 973–977.
- MOORE, P.K., OLUYOMI, A.O., BABBEDGE, R.C., WALLACE, P. & HART, S.L. (1991). L-NG-nitro arginine methyl ester exhibits antinociceptive activity in the mouse. *Br. J. Pharmacol.*, **102**, 198–202.
- MOORE, P.K., WALLACE, P., GAFFEN, Z., HART, S.L. & BABBEDGE, R.C. (1993). Characterization of the novel nitric oxide synthase inhibitor 7-nitro indazole and related indazoles: antinociceptive and cardiovascular effects. *Br. J. Pharmacol.*, **110**, 219–224.
- MORLEY, J.E. & FLOOD, J.F. (1992). Evidence that nitric oxide modulates food intake in mice. *Life Sci.*, **49**, 707–711.
- MORRIS, B.J. (1995). Stimulation of immediate early gene expression in striatal neurons by nitric oxide. *J. Biol. Chem.*, **270**, 24740–24744.
- MURPHY, S. (2000). Production of nitric oxide by glial cells: regulation and potential roles in the CNS. *Glia*, **29**, 1–14.
- MURTHY, K.S., GRIDER, J.R., JIN, J.G. & MAKHLOUF, G.M. (1995). Interplay of VIP and nitric oxide in the regulation of neuromuscular activity in the gut. *Arch. Int. Pharmacodyn. Ther.*, **329**, 27–38.
- MYERS, P.R., MINOR JR., R.L., GUERRA JR., R., BATES, J.N. & HARRISON, D.G. (1990). Vasorelaxant properties of the endothelium-derived relaxing factor more closely resemble S-nitrosocysteine than nitric oxide. *Nature*, **345**, 161–163.
- NAGAFUJI, T., MATSUI, T., KOIDE, T. & ASANO, T. (1992). Blockade of nitric oxide formation by N-nitro-L-arginine mitigates ischemic brain edema and subsequent cerebral infarction in rats. *Neurosci. Lett.*, **147**, 159–162.
- NAKANE, M., MITCHELL, J., FÖRSTERMANN, U. & MURAD, F. (1991). Phosphorylation by calcium calmodulin-dependent protein kinase II and protein kinase C modulates the activity of nitric oxide synthase. *Biochem. Biophys. Res. Commun.*, **180**, 1396–1402.
- NAKANE, M., SCHMIDT, H.H., POLLOCK, J.S., FORSTERMANN, U. & MURAD, F. (1993). Cloned human brain nitric oxide synthase is highly expressed in skeletal muscle. *FEBS Lett.*, **316**, 175–180.
- NELSON, R.J., DEMAS, G.E., HUANG, P.L., FISHMAN, M.C., DAWSON, V.L., DAWSON, T.M. & SNYDER, S.H. (1995). Behavioural abnormalities in male mice lacking neuronal nitric oxide synthase. *Nature*, **378**, 383–386.
- NOWICKI, J.P., DUVAL, D., POIGNET, H. & SCATTON, B. (1991). Nitric oxide mediates neuronal death after focal cerebral ischemia in the mouse. *Eur. J. Pharmacol.*, **204**, 339–340.
- O'DELL, T.J., HAWKINS, R.D., KANDEL, E.R. & ARANCIO, O. (1991). Tests of the roles of two diffusible substances in long-term potentiation: evidence for nitric oxide as a possible early retrograde messenger. *Proc. Natl. Acad. Sci. U.S.A.*, **88**, 11285–11289.
- O'DELL, T.J., HUANG, P.L., DAWSON, T.M., DINERMAN, J.L., SNYDER, S.H., KANDEL, E.R. & FISHMAN, M.C. (1994). Endothelial NOS and the blockade of LTP by NOS inhibitors in mice lacking neuronal NOS. *Science*, **265**, 542–546.
- OKERE, C.O. & KABA, H. (2000). Increased expression of neuronal nitric oxide synthase mRNA in the accessory olfactory bulb during the formation of olfactory recognition memory in mice. *Eur. J. Neurosci.*, **12**, 4552–4556.
- PAPKA, R.E., MCCURDY, J.R., WILLIAMS, S.J., MAYER, B., MARSON, L. & PLATT, K.B. (1995). Parasympathetic preganglionic neurons in the spinal cord involved in uterine innervation are cholinergic and nitric oxide-containing. *Anat. Rec.*, **241**, 554–562.
- PARKINSON, J.F., MITROVIC, B. & MERRILL, J.E. (1999). The role of nitric oxide in multiple sclerosis. *J. Mol. Med.*, **75**, 174–186.
- PENSON, D.F., NG, C., CAI, L., RAJFER, J. & GONZALEZ CADAVID, N.F. (1996). Androgen and pituitary control of penile nitric oxide synthase and erectile function in the rat. *Biol. Reprod.*, **55**, 567–574.
- PEUNOVA, N. & ENIKOLOPOV, G. (1993). Amplification of calcium-induced gene transcription by nitric oxide in neuronal cells. *Nature*, **364**, 450–453.
- PEUNOVA, N. & ENIKOLOPOV, G. (1995). Nitric oxide triggers a switch to growth arrest during differentiation of neuronal cells. *Nature*, **375**, 68–73.
- PIEPER, A.A., BLACKSHAW, S., CLEMENTS, E.E., BRAT, D.J., KRUG, D.K., WHITE, A.J., PINTO-GARCIA, P., FAVIT, A., CONOVER, J.R., SNYDER, S.H. & VERMA, A. (2000). Poly(ADP-ribosylation) basally activated by DNA strand breaks reflects glutamate-nitric oxide neurotransmission. *Proc. Natl. Acad. Sci. U.S.A.*, **97**, 1845–1850.
- PIEPER, A.A., VERMA, A., ZHANG, J. & SNYDER, S.H. (1999). Poly(ADP-ribose)polymerase, nitric oxide and cell death. *Trends Pharmacol. Sci.*, **20**, 171–181.
- QUINTANA, E., GARCIA-ZARAGOZA, E., MARTINEZ CUESTA, M.A., CALATAYUD, S., ESPLUGUES, J.V. & BARRACHINA, M.D. (2001). A cerebral nitric oxide pathway modulates endotoxin-induced changes in gastric motility. *Br. J. Pharmacol.*, **134**, 325–332.
- RAJANAYAGAM, M.A., LI, C.G. & RAND, M.J. (1993). Differential aspects of hydroxocobalamin on NO-mediated relaxations in rat aorta and anococcygeus muscle. *Br. J. Pharmacol.*, **108**, 3–5.
- RAJFER, J., ARONSON, W.J., BUSH, P.A., DOREY, F.J. & IGNARRO, L.J. (1992). Nitric oxide as a mediator of relaxation of the corpus cavernosum in response to nonadrenergic, noncholinergic neurotransmission. *N. Engl. J. Med.*, **326**, 90–94.
- REGIDOR, J., MONTESDEOCA, J., RAMIREZ-GONZALEZ, J.A., HERNANDEZ-URQUIA, C.M. & DIVAC, I. (1993). Bilateral induction of NADPH-diaphorase activity in neocortical and hippocampal neurons by unilateral injury. *Brain Res.*, **631**, 171–174.
- REISER, G. (1990a). Endothelin and a Ca<sup>2+</sup> ionophore raise cyclic GMP levels in a neuronal line via formation of nitric oxide. *Br. J. Pharmacol.*, **101**, 722–726.
- REISER, G. (1990b). Mechanisms of stimulation of cyclic-GMP level in a neuronal cell line mediated by serotonin (5-HT<sub>3</sub>) receptors. Involvement of nitric oxide, arachidonic acid metabolism and cytosolic Ca<sup>2+</sup>. *Eur. J. Biochem.*, **189**, 547–552.
- RIVIER, C. & SHEN, G.H. (1994). In the rat, endogenous nitric oxide modulates the response of the hypothalamic-pituitary-adrenal axis to interleukin-1 beta, vasopressin, and oxytocin. *J. Neurosci.*, **14**, 1985–1993.

- ROBERTS, C.K., BARNARD, R.J., SCHECK, S.H. & BALON, T.W. (1997). Exercise-stimulated glucose transport in skeletal muscle is nitric oxide dependent. *Am. J. Physiol.*, **273**, E220–E225.
- RODRIGUEZ-CRESPO, I., STRAUB, W., GAVILANES, F. & ORTIZ DE MONTELLANO, P.R. (1998). Binding of dynein light chain (PIN) to neuronal nitric oxide synthase in the absence of inhibition. *Arch. Biochem. Biophys.*, **359**, 297–304.
- ROGERS, N.E. & IGNARRO, L.J. (1992). Constitutive nitric oxide synthase from cerebellum is reversibly inhibited by nitric oxide formed from L-arginine. *Biochem. Biophys. Res. Commun.*, **189**, 242–249.
- SAENZ DE TEJADA, I., GOLDSTEIN, I., AZADZOL, K., KRANE, R.J. & COHEN, R.A. (1989). Impaired neurogenic and endothelium-mediated relaxation of penile smooth muscle from diabetic men with impotence. *N. Engl. J. Med.*, **320**, 1025–1030.
- SAKUMA, I., TOGASHI, H., YOSHIOKA, M., SAITO, H., YANAGIDA, M., TAMURA, M., KOBAYASHI, T., YASUDA, H., GROSS, S.S. & LEVI, R. (1992). NG-methyl-L-arginine, an inhibitor of L-arginine-derived nitric oxide synthesis, stimulates renal sympathetic nerve activity in vivo. A role for nitric oxide in the central regulation of sympathetic tone? *Circ. Res.*, **70**, 607–611.
- SALTER, M., KNOWLES, R.G. & MONCADA, S. (1991). Widespread tissue distribution species distribution and changes in activity of Ca<sup>2+</sup>-dependent and Ca<sup>2+</sup>-independent nitric oxide synthase. *FEBS Lett.*, **291**, 145–149.
- SAMDANI, A.F., DAWSON, T.M. & DAWSON, V.L. (1997). Nitric oxide synthase in models of focal ischemia. *Stroke*, **28**, 1283–1288.
- SANDOR, N.T., BRASSAI, A., PUSKAS, A. & LENDVAI, B. (1995). Role of nitric oxide in modulating neurotransmitter release from rat striatum. *Brain Res. Bull.*, **36**, 483–486.
- SCHAAD, N.C., VANECEK, J. & SCHULZ, P.E. (1994). Photoneuronal regulation of rat pineal nitric oxide synthase. *J. Neurochem.*, **62**, 2496–2499.
- SCHEMANN, M., SCHAAF, C. & MADER, M. (1995). Neurochemical coding of enteric neurons in the guinea pig stomach. *J. Comp. Neurol.*, **353**, 161–178.
- SCHMIDT, H.H., GAGNE, G.D., NAKANE, M., POLLOCK, J.S., MILLER, M.F. & MURAD, F. (1992a). Mapping of neural nitric oxide synthase in the rat suggests frequent co-localization with NADPH diaphorase but not with soluble guanylyl cyclase, and novel paraneural functions for nitrinergic signal transduction. *J. Histochem. Cytochem.*, **40**, 1439–1456.
- SCHMIDT, H.H.W., POLLOCK, J.S., NAKANE, M., FÖRSTERMANN, U. & MURAD, F. (1992b). Ca<sup>2+</sup>/calmodulin-regulated nitric oxide synthases. *Cell Calcium*, **13**, 427–434.
- SCHULTZ, J.B., MATTHEWS, R.T., MUQIT, M.M.K., BROWNE, S.E. & BEAL, M.F. (1995). Inhibition of neuronal nitric oxide synthase by 7-nitroimidazole protects against MPTP induced neurotoxicity in mice. *J. Neurochem.*, **64**, 936–939.
- SCHUMAN, E.M. & MADISON, D.V. (1991). A requirement for the intercellular messenger nitric oxide in long-term potentiation. *Science*, **254**, 1503–1506.
- SHAH, S., NATHAN, L., SINGH, R., FU, Y.S. & CHAUDHURI, G. (2001). E<sub>2</sub> and not P<sub>4</sub> increases NO release from NANC nerves of the gastrointestinal tract: implications in pregnancy. *Am. J. Physiol.*, **280**, R1546–R1554.
- SHEN, W.W., UROSEVICH, Z. & CLAYTON, D.O. (1999). Sildenafil in the treatment of female sexual dysfunction induced by selective serotonin reuptake inhibitors. *J. Reprod. Med.*, **44**, 535–542.
- SHENG, H., SCHMIDT, H.H., NAKANE, M., MITCHELL, J.A., POLLOCK, J.S., FÖRSTERMANN, U., MURAD, F. & SCHMIDT, H.H.W. (1992). Characterization and localization of nitric oxide synthase in non-adrenergic non-cholinergic nerves from bovine retractor penis muscles. *Br. J. Pharmacol.*, **106**, 768–773.
- SHIBUKI, K. & OKADA, D. (1991). Endogenous nitric oxide release required for long-term synaptic depression in the cerebellum. *Nature*, **349**, 326–328.
- SILVAGNO, F., XIA, H. & BREDDT, D.S. (1996). Neuronal nitric-oxide synthase- $\mu$ , an alternatively spliced isoform expressed in differentiated skeletal muscle. *J. Biol. Chem.*, **271**, 11204–11208.
- SIPSKI, M.L., ROSEN, R.C., ALEXANDER, C.J. & HAMER, R.M. (2000). Sildenafil effects on sexual and cardiovascular responses in women with spinal cord injury. *Urology*, **55**, 812–815.
- SMET, P.J., EDYVANE, K.A., JONAVICIUS, J. & MARSHALL, V.R. (1994). Colocalization of nitric oxide synthase with vasoactive intestinal peptide, neuropeptide Y, and tyrosine hydroxylase in nerves supplying the human ureter. *J. Urol.*, **152**, 1292–1296.
- SODERLING, T.R. (2000). CaM-kinases: modulators of synaptic plasticity. *Curr. Opin. Neurobiol.*, **10**, 375–380.
- SON, H., HAWKINS, R.D., MARTIN, K., KIEBLER, M., HUANG, P.L., FISHMAN, M.C. & KANDEL, E.R. (1996). Long-term potentiation is reduced in mice that are doubly mutant in endothelial and neuronal nitric oxide synthase. *Cell*, **87**, 1015–1023.
- SORKIN, L.S. (1993). NMDA evokes an L-NAME sensitive spinal release of glutamate and citrulline. *NeuroReport*, **4**, 479–482.
- SOUTHAM, E. & GARTHWAITE, J. (1993). The nitric oxide-cyclic GMP signalling pathway in rat brain. *Neuropharmacology*, **32**, 1267–1277.
- STAGLIANO, N.E., DIETRICH, W.D., PRADO, R., GREEN, E.J. & BUSTO, R. (1997). The role of nitric oxide in the pathophysiology of thromboembolic stroke in the rat. *Brain Res.*, **759**, 32–40.
- STAMLER, J.S. & MEISSNER, G. (2001). Physiology of nitric oxide in skeletal muscle. *Physiol. Rev.*, **81**, 209–237.
- STAMLER, J.S., SINGEL, D.J. & LOSCALZO, J. (1992). Biochemistry of nitric oxide and its redox-activated forms. *Science*, **258**, 1898–1902.
- STEEL, J.H., TEREINGHI, G., CHUNG, J.M., NA, H.S., CARLTON, S.M. & POLAK, J.M. (1994). Increased nitric oxide synthase immunoreactivity in rat dorsal root ganglia in a neuropathic pain model. *Neurosci. Lett.*, **169**, 81–84.
- TAKAHASHI, T., QOUBAITARY, A., OWYANG, C. & WILEY, J.W. (2000). Decreased expression of nitric oxide synthase in the colonic myenteric plexus of aged rats. *Brain Res.*, **883**, 15–21.
- TANAKA, K., SHIRAI, T., NAGATA, E., DEMBO, T. & FUKUUCHI, Y. (1997). Immunohistochemical detection of nitrotyrosine in postischemic cerebral cortex in gerbil. *Neurosci. Lett.*, **235**, 85–88.
- TENG, B.Q., MURTHY, K.S., KEUMMERLE, J.F., GRIDER, J.R., SASE, K., MICHEL, T. & MAKHLOUF, G.M. (1988). Expression of endothelial nitric oxide synthase in human and rabbit gastrointestinal smooth muscle cells. *Am. J. Physiol.*, **275**, G342–G351.
- THOMAS, G. & RAMWELL, P.W. (1989). Vascular relaxation mediated by hydroxylamines and oximes: their conversion to nitrites and mechanism of endothelium dependent vascular relaxation. *Biochem. Biophys. Res. Commun.*, **164**, 889–893.
- THOMSEN, L.L., IVERSEN, H.K., BRINCK, T.A. & OLESEN, J. (1993). Arterial supersensitivity to nitric oxide (nitroglycerin) in migraine sufferers. *Cephalalgia*, **13**, 395–399.
- THOMSEN, L.L. & OLESEN, J. (1998). Nitric oxide theory of migraine. *Clin. Neurosci.*, **5**, 28–33.
- TIDBALL, J.G., LAVERGNE, E., LAU, K.S., SPENCER, J.J., STULL, J.T. & WEHLING, M. (1998). Mechanical loading regulates NOS expression and activity in developing and adult skeletal muscle. *Am. J. Physiol.*, **275**, C260–C266.
- TOGASHI, H., SAKUMA, I., YOSHIOKA, M., KOBAYASHI, T., YASUDA, H., KITABAKE, A., SAITO, H., GROSS, S.S. & LEVI, R.A. (1992). A central nervous system action of nitric oxide in blood pressure regulation. *J. Pharmacol. Exp. Ther.*, **262**, 343–347.
- TOMINAGA, T., SATO, S., OHNISHI, T. & OHNISHI, S.T. (1993). Potentiation of nitric oxide formation following bilateral carotid occlusion and focal cerebral ischemia in the rat: in vivo detection of the nitric oxide radical by electron paramagnetic resonance spin trapping. *Brain Res.*, **614**, 342–346.
- TOMITA, S., NICOLL, R.A. & BREDDT, D.S. (2001). PDZ protein interactions regulating glutamate receptor function and plasticity. *J. Cell. Biol.*, **153**, F19–F24.
- TORPHY, T.J., FINE, C.F., BURMAN, M., BARNETTE, M.S. & ORMSBEE III, H.S. (1986). Lower esophageal sphincter relaxation is associated with increased cyclic nucleotide content. *Am. J. Physiol.*, **251**, G786–G793.
- TRONCON, L.E.A., THOMPSON, D.G., AHLUWALIA, N.K., BARLOW, J. & HEGGIE, L. (1995). Relations between upper abdominal symptoms and gastric distension abnormalities in dysmotility like functional dyspepsia and after vagotomy. *Gut*, **37**, 17–22.



- UTEPBERGENOV, D.I., MERTSCH, K., SPORBERT, A., TENZ, K., PAUL, M., HASELOFF, R.F. & BLASIG, I.E. (1998). Nitric oxide protects blood-brain barrier in vitro from hypoxia/reoxygenation-mediated injury. *FEBS Lett.*, **424**, 197–201.
- VANDERWINDEN, J.M., DE LAET, M.H., SCHIFFMANN, S.N., MAILLEUX, P., LOWENSTEIN, C.J. & SNYDER, S.H. (1993). Nitric oxide synthase distribution in the enteric nervous system of Hirschsprung's disease. *Gastroenterology*, **105**, 969–973.
- VANDERWINDEN, J.M., MAILLEUX, P., SCHIFFMANN, S.N., VANDERHAEGHEN, J.J. & DE LAET, M.H. (1992). Nitric oxide synthase activity in infantile hypertrophic pyloric stenosis. *N. Engl. J. Med.*, **327**, 511–515.
- VEDERNIKOV, Y.P., MORDVINCEV, P.I., MALENKOVA, I.V. & VANIN, A.F. (1992). Similarity between the vasorelaxing activity of dinitrosyl iron cysteine complexes and endothelium-derived relaxing factor. *Eur. J. Pharmacol.*, **211**, 313–317.
- VENEMA, V.J., JU, H., ZOU, R. & VENEMA, R.C. (1997). Interaction of neuronal nitric-oxide synthase with caveolin-3 in skeletal muscle. Identification of a novel caveolin scaffolding/inhibitory domain. *J. Biol. Chem.*, **272**, 28187–28190.
- VERGE, V.M.K., XU, Z., XU, X., WIESENFELD HALLIN, Z. & HOKFELT, T. (1992). Marked increase in nitric oxide synthase mRNA in rat dorsal root ganglia after peripheral axotomy: in situ hybridization and functional studies. *Proc. Natl. Acad. Sci. U.S.A.*, **89**, 11617–11621.
- VINCENT, S.R. (1995). Localization of nitric oxide neurons in the central nervous system. In *Nitric oxide in the nervous system*. ed Vincent, S.R. pp. 83. London: Academic Press.
- VISSER, J.J., SCHOLTEN, R.J. & HOEKMAN, K. (1994). Nitric oxide synthesis in meningococcal meningitis [letter]. *Ann. Intern. Med.*, **120**, 345–346.
- WALLIS, R.A., PANIZZON, K.L., HENRY, D. & WASTERLAIN, C.G. (1993). Neuroprotection against nitric oxide injury with inhibitors of ADP-ribosylation. *NeuroReport*, **5**, 245–248.
- WANG, T., XIE, Z. & LU, B. (1995). Nitric oxide mediates activity-dependent synaptic suppression at developing neuromuscular synapses. *Nature*, **374**, 262–266.
- WATANABE, A., ONO, M., SHIBATA, S. & WATANABE, S. (1995). Effect of a nitric oxide synthase inhibitor, N-nitro-L-arginine methyl ester, on light-induced phase delay of circadian rhythm of wheel-running activity in golden hamsters. *Neurosci. Lett.*, **192**, 25–28.
- WATKINS, C.C., SAWA, A., JAFFREY, S., BLACKSHAW, S., BARROW, R.K., SNYDER, S.H. & FERRIS, C.D. (2000). Insulin restores neuronal nitric oxide synthase expression and function that is lost in diabetic gastropathy. *J. Clin. Invest.*, **106**, 373–384.
- WEINACKER, A.B. & VASZAR, L.T. (2001). Acute respiratory distress syndrome: physiology and new management strategies. *Annu. Rev. Med.*, **52**, 221–237.
- WEINBERGER, B., HECK, D.E., LASKIN, D.L. & LASKIN, J.D. (1999). Nitric oxide in the lung: therapeutic and cellular mechanisms of action. *Pharmacol. Ther.*, **84**, 401–411.
- WEINBERGER, B., WEISS, K., HECK, D.E., LASKIN, D.L. & LASKIN, J.D. (2001). Pharmacologic therapy of persistent pulmonary hypertension of the newborn. *Pharmacol. Ther.*, **89**, 67–79.
- WIENCKEN, A.E. & CASAGRANDE, V.A. (1999). Endothelial nitric oxide synthetase (eNOS) in astrocytes: another source of nitric oxide in neocortex. *Glia*, **26**, 280–290.
- WEINER, C.P. & THOMPSON, L.P. (1997). Nitric oxide and pregnancy. *Semin. Perinatol.*, **21**, 367–380.
- WILDERMAN, M.J. & ARMSTEAD, W.M. (1997). Role of neuronal NO synthase in relationship between NO and opioids in hypoxia-induced pial artery dilation. *Am. J. Physiol.*, **273**, H1807–H1815.
- WINK, D.A., HANBAUER, I., KRISHNA, M.C., DEGRAFF, W., GAMSON, J. & MITCHELL, J.B. (1993). Nitric oxide protects against cellular damage and cytotoxicity from reactive oxygen species. *Proc. Natl. Acad. Sci. U.S.A.*, **90**, 9813–9817.
- WOLF, G. (1997). Nitric oxide and nitric oxide synthase: biology, pathology, localization. *Histol. Histopathol.*, **12**, 251–261.
- WOOD, J. & GARTHWAITE, J. (1994). Models of the diffusional spread of nitric oxide: implications for neural oxide signalling and its pharmacological properties. *Neuropharmacology*, **33**, 1235–1244.
- WU, H.H., WILLIAMS, C.V. & MCLOON, S.C. (1994a). Involvement of nitric oxide in the elimination of a transient retinotectal projection in development. *Science*, **265**, 1593–1596.
- WU, W. & LI, L. (1993). Inhibition of nitric oxide synthase reduces motoneuron death due to spinal root avulsion. *Neurosci. Lett.*, **153**, 121–124.
- WU, W., LIUZZI, F.J., SCHINCO, F.P., DEPTO, A.S., LI, Y., MONG, J.A., DAWSON, T.M. & SNYDER, S.H. (1994b). Neuronal nitric oxide synthase is induced in spinal neurons by traumatic injury. *Neuroscience*, **61**, 719–726.
- XU, J. & TSENG, L.F. (1995). Nitric oxide/cyclic guanosine monophosphate system in the spinal cord differentially modulates intracerebroventricularly administered morphine and B-endorphin induced antinociception in the mouse. *J. Pharmacol. Exp. Ther.*, **274**, 8–16.
- XUE, L., FARRUGIA, G., MILLER, S.M., FERRIS, C.D., SNYDER, S.H. & SZRUSZEWSKI, J.H. (2000). Carbon monoxide and nitric oxide as coneurotransmitters in the enteric nervous system: evidence from genomic deletion of biosynthetic enzymes. *Proc. Natl. Acad. Sci. U.S.A.*, **97**, 1851–1855.
- YAMADA, K., NODA, Y., NAKAYAMA, S., KOMORI, Y., SUGIHARA, H., HASEGAWA, I. & NABESHIMA, T. (1995). Role of nitric oxide in learning and memory and in monoamine metabolism in the rat brain. *Br. J. Pharmacol.*, **115**, 852–858.
- YAMATO, S., SPECHLER, S.J. & GOYAL, R.K. (1992). Role of nitric oxide in esophageal peristalsis in the opossum. *Gastroenterology*, **103**, 197–204.
- YANG, G., CHAN, P.H., CHEN, J., CARLSON, E., CHEN, S.F., WEINSTEIN, P., EPSTEIN, C.J. & KAMII, H. (1994). Human copper-zinc superoxide dismutase transgenic mice are highly resistant to reperfusion injury after focal cerebral ischemia. *Stroke*, **25**, 165–170.
- YOSHIDA, T., LIMMROTH, V., IRIKURA, K. & MOSKOWITZ, M.A. (1994). The NOS inhibitor, 7-nitroindazole, decreases focal infarct volume but not the response to topical acetylcholine in pial vessels. *J. Cereb. Blood Flow Metab.*, **14**, 924–929.
- YOUNG, H.M., MCCONALOGUE, K., FURNESS, J.B. & DE VENTE, J. (1993). Nitric oxide targets in the guinea-pig intestine identified by induction of cyclic GMP immunoreactivity. *Neuroscience*, **55**, 583–596.
- ZHANG, J., DAWSON, V.L., DAWSON, T.M. & SNYDER, S.H. (1994). Nitric oxide activation of poly(ADP-ribose) synthetase in neurotoxicity. *Science*, **263**, 687–689.
- ZHANG, J. & SNYDER, S.H. (1992). Nitric oxide stimulates auto-ADP-ribosylation of glyceraldehyde-3-phosphate dehydrogenase. *Proc. Natl. Acad. Sci. U.S.A.*, **89**, 9382–9385.
- ZHANG, Z.G., REIF, D., MACDONALD, J., TANG, W.X., KAMP, D.K., GENTILE, R.J., SHAKESPEARE, W.C., MURRAY, R.J. & CHOPP, M. (1996). ARL 17477, a potent and selective neuronal NOS inhibitor decreases infarct volume after transient middle cerebral artery occlusion in rats. *J. Cereb. Blood Flow Metab.*, **16**, 599–604.
- ZHENG, Y.M., SCHAFFER, M.K., WEIHE, E., SHENG, H., CORISDEO, S., FU, Z.F., KOPROWSKI, H. & DIETZSCHOLD, B. (1993). Severity of neurological signs and degree of inflammatory lesions in the brains of rats with Borna disease correlate with the induction of nitric oxide synthase. *J. Virol.*, **67**, 5786–5791.
- ZHU, H. & BARR, G.A. (2001). Opiate withdrawal during development: are NMDA receptors indispensable? *Trends. Pharmacol. Sci.*, **22**, 404–408.
- ZHUO, M., MELLER, S.T. & GEBHART, G.F. (1993). Endogenous nitric oxide is required for tonic cholinergic inhibition of spinal mechanical transmission. *Pain*, **54**, 71–78.

(Received September 25, 2001

Revised December 19, 2001

Accepted December 20, 2001)