

Characterisation of the vasorelaxant properties of the novel endocannabinoid *N*-arachidonoyl-dopamine (NADA)

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1 We have investigated the vascular effects of *N*-arachidonoyl-dopamine (NADA), a novel endocannabinoid/vanilloid. NADA caused vasorelaxant effects comparable to those of anandamide in small mesenteric vessels (G3), the superior mesenteric artery (G0) and in the aorta.

2 In G3, addition of *N*^G-nitro-L-arginine methyl ester (300 μ M) or the dopamine (D₁) receptor antagonist (SCH23390, 1 μ M) did not affect responses to NADA. In the presence of 60 mM KCl, after de-endothelialisation, or after K⁺ channel inhibition with charybdotoxin (100 nM) and apamin (500 nM), relaxant responses to NADA were inhibited.

3 In G3, pretreatment with the vanilloid receptor (VR) agonist capsaicin (10 μ M) or the VR antagonist capsazepine (10 μ M) reduced vasorelaxation to NADA.

4 In G3, application of the CB₁ antagonist SR141716A at 1 μ M but not 100 nM reduced the potency of NADA. Another CB₁ antagonist, AM251 (100 nM and 1 μ M), did not affect vasorelaxation to NADA. After endothelial denudation, SR141716A (1 μ M) did not reduce the responses further. A combination of capsaicin and SR141716A (1 μ M) reduced vasorelaxation to NADA further than with capsaicin pretreatment alone. The novel endothelial cannabinoid (CB) receptor antagonist O-1918 opposed vasorelaxation to NADA in G3.

5 In the superior mesenteric artery (G0), vasorelaxation to NADA was not dependent on an intact endothelium and was not sensitive to O-1918, but was sensitive to capsaicin and SR141716A or AM251 (both 100 nM).

6 The results of the present study demonstrate for the first time that NADA is a potent vasorelaxant. In G3, the effects of NADA are mediated by stimulation of the VR and the novel endothelial CB receptor, while in G0, vasorelaxation is mediated through VR₁ and CB₁ receptors.

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Abbreviations: AM251, *N*-(piperidin-1-yl)-5-(4-iodophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1*H*-pyrazole-3-carboxamide; CB, cannabinoid; ChTX, charybdotoxin; EDHF, endothelium-derived hyperpolarising factor; G3, third-order branch of the superior mesenteric artery; G0, the superior mesenteric artery; L-NAME, *N*^G-nitro-L-arginine methyl ester; NADA, *N*-arachidonoyl-dopamine; SR141716A, *N*-(piperidin-1-yl)-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1*H*-pyrazole-3-carboxamide; VR, vanilloid receptor

Introduction

Since the identification of specific cannabinoid (CB) receptors (Matsuda *et al.*, 1990), several natural ligands for the receptors have been found. One such compound is the novel endogenous substance *N*-arachidonoyl-dopamine (NADA), which has recently been identified in rat and bovine nervous tissues (Bisogno *et al.*, 2000; Huang *et al.*, 2002). Structurally, NADA has a vanillyl-amine moiety and a long-chain unsaturated fatty acid common to CBs. It is reported to bind to vanilloid receptors (VRs) with similar potency to capsaicin in human embryonic kidney cells (Huang *et al.*, 2002), and has similar efficacy to anandamide at the CB₁ receptors in rat brain cells (Bisogno *et al.*, 2000). Very recent data have identified NADA as a central endovanilloid, which can induce hyperalgesia (Chu *et al.*, 2003), and a peripheral endovanilloid, which can cause

smooth muscle contraction in the guinea-pig bronchi and bladder (Harrison *et al.*, 2003).

To date, the cardiovascular actions of NADA have not been examined; however, the effects of the prototypic endocannabinoid anandamide have been widely studied (for a review, see Randall *et al.*, 2002). Several putative mechanisms have been put forward to explain the vasorelaxant effects of anandamide; however, many discrepancies still occur in the literature. Early reports indicated that anandamide acted *via* the release of nitric oxide (Deutsch *et al.*, 1997). However, in rat mesenteric vessels, vasorelaxant effects of anandamide are largely insensitive to inhibition of nitric oxide synthase (Randall *et al.*, 1996; White & Hiley, 1997; Járai *et al.*, 1999). Although nitric oxide does not appear to mediate responses to anandamide, the vasorelaxation is partly endothelium-dependent, in some, but not all preparations (White & Hiley, 1997; Chaytor *et al.*, 1999; Járai *et al.*, 1999). It is therefore likely that anandamide stimulates the release of the

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endothelium-derived hyperpolarising factor (EDHF), which is communicated to smooth muscle cells *via* myoendothelial gap junctions (Chaytor *et al.*, 1999; Harris *et al.*, 2002).

The involvement of currently recognised CB receptors in vasorelaxation to endocannabinoids is also unclear. Some previous studies have implicated the CB₁ receptor on the basis of inhibition by SR141716A, a selective CB₁ receptor antagonist. However, this is complicated by the wide-ranging actions of SR141716A at various concentrations. For example, SR141716A can inhibit myoendothelial gap junctions (Chaytor *et al.*, 1999), which themselves have been implicated in the actions of anandamide *via* EDHF (Chaytor *et al.*, 1999; Harris *et al.*, 2002). SR141716A may also antagonise the novel non-CB₁ endothelial CB receptor proposed by Kunos and colleagues (Járai *et al.*, 1999; Offertáler *et al.*, 2003). Indeed, Offertáler *et al.* (2003) have reported that these responses are sensitive to a novel antagonist, O-1918. SR141716A may also have actions on the VR (De Petrocellis *et al.*, 2001; Millns *et al.*, unpublished data).

One of the most interesting proposals has been that anandamide stimulates VRs on perivascular sensory nerves, and acts *via* the release of the vasodilator neurotransmitter calcitonin gene-related peptide (CGRP; Zygmunt *et al.*, 1999; Ralevic *et al.*, 2000). Whether this action can account for the entire endocannabinoid vasorelaxant response has yet to be clarified, as further studies have not found that the entire response to anandamide is mediated *via* sensory nerves (Ralevic *et al.*, 2000; Harris *et al.*, 2002).

Given the agonist effects of the novel compound NADA on CB and VRs, the possibility exists that NADA may act as a vasorelaxant in a similar manner to anandamide. In this study, vasorelaxant actions of NADA have been identified and characterised in large and small rat arteries. Specifically, the roles of CB receptors, VRs and sensory nerves, the endothelium and EDHF in the vascular effects of NADA have been investigated.

Methods

Blood vessel preparation

Male Wistar rats (250–350 g) were stunned by a blow to the back of the head and killed by cervical dislocation. The aorta, superior mesenteric artery and mesenteric arterial bed were removed rapidly and placed into cold Krebs–Henseleit buffer (composition, mM: NaCl 118, KCl 4.7, MgSO₄ 1.2, KH₂PO₄ 1.2, NaHCO₃ 25, CaCl₂ 2, D-glucose 10). From the mesenteric arterial bed, 2 mm segments of third-order branches of the superior mesenteric artery (G3) were dissected free of adherent connective and adipose tissue. G3 vessels were mounted on fine tungsten wires (40 µm diameter) on a Mulvany–Halpern myograph (Myo-Interface Model 410A, Danish Myo Technology, Denmark) (Mulvany & Halpern, 1977). The superior mesenteric artery (G0; 3–4 mm in length) was also cleaned of adherent tissue, and was mounted on fixed-segment support pins using the Multi Myograph system (Model 610M, Danish Myo Technology, Denmark). The thoracic aorta was dissected free of adherent connective and adipose tissue, and cut into rings 6–8 mm long (Tep-areenan *et al.*, 2003). The aortic rings were placed in 50 ml organ baths containing Krebs–Henseleit buffer, mounted between two stainless steel hooks and

attached by thread to an isometric force displacement transducer (LETICA 210). Tension was measured and was recorded on a MacLab 4e recording system (ADInstruments, U.K.).

Once mounted, all vessels were kept at 37°C in Krebs–Henseleit buffer and gassed with 5% CO₂ in O₂. The mesenteric vessels were stretched to an optimal passive tension of 5 mN and the thoracic aorta to 10 mN tension. All vessels were allowed to equilibrate, and the contractile integrity of each was tested by its ability to contract to 60 mM KCl by at least 5 mN.

In some mesenteric preparations, the endothelium was removed by abrasion with a human hair. Preparations (both G3 and G0) were considered denuded when relaxation to 10 µM carbachol after precontraction was less than 20% relaxation (see White & Hiley, 1997). All other G3 and G0 preparations were endothelium-intact.

Experimental protocol

Viable vessels were contracted with U46619 (10–300 nM), a thromboxane mimetic, to increase tension by at least 5 mN. The average increase in tension for G3 vessels was 10.4 ± 0.4 mN (*n* = 121), for the superior mesenteric artery it was 19.8 ± 0.6 mN (*n* = 60), and for the aorta it was 9.0 ± 0.7 mN (*n* = 16). Once a stable contraction was achieved, the vasorelaxant effects of NADA were assessed as cumulative concentration–response curves by addition of NADA to the buffer. The steady-state response to NADA was taken at each concentration and expressed as a percentage relaxation of the imposed U46619 contraction.

To assess the involvement of dopamine receptors in vasorelaxation to NADA, the dopamine receptor antagonist SCH23390 (1 µM, Martin & Broadley, 1995) was added to preparations 20 min before pre-contraction. The involvement of nitric oxide was investigated using the nitric oxide synthase inhibitor, *N*^G-nitro-L-arginine methyl ester (L-NAME, 300 µM, Randall & Griffith, 1991). In these experiments, L-NAME was present in the buffer throughout the whole experiment. To assess the role of K⁺ channels, vessels were depolarised and contracted with 60 mM KCl (Adeagbo & Triggle, 1993) and concentration–response curves constructed.

To investigate the role of the endothelium in NADA-mediated vasorelaxation, the endothelium was removed using a human hair (White & Hiley, 1997). The role of the EDHF was also assessed using a number of K⁺ channel inhibitors in the presence of both nitric oxide synthase and cyclooxygenase inhibitors (McCulloch *et al.*, 1997). In these experiments, L-NAME (300 µM) and indomethacin (10 µM) were present in the KREBS solution throughout the experiments to inhibit nitric oxide and prostanoid synthesis, respectively. Preparations were then incubated with charybdotoxin (ChTX) to block large calcium-activated K⁺ channels and voltage-sensitive K⁺ channels (ChTX, 100 nM) and apamin to block small calcium-activated K⁺ channels (500 nM), as this combination inhibits EDHF responses (Randall & Kendall, 1998; Zygmunt *et al.*, 1997). Concentration–response curves were then constructed to NADA.

To assess the involvement of VRs, capsazepine (10 µM, a VR antagonist) was added to the buffer 20 min before precontraction (Zygmunt *et al.*, 1999). Also, some vessels were incubated for 1 h with the vanilloid agonist capsaicin (10 µM) to deplete

the sensory nerves of neurotransmitters. This was followed by a 20-min washout period (Zygmunt *et al.*, 1999; Harris *et al.*, 2002).

To assess the involvement of CB receptors, either SR141716A (100 nM and 1 μ M, a CB₁ receptor antagonist, Rinaldi-Carmona *et al.*, 1994) or AM251 (100 nM and 1 μ M, a CB₁ receptor antagonist; Gardiner *et al.*, 2002) were added to the preparations 20 min before precontraction. The effects of SR141716A were also examined in conjunction with endothelial denudation and after capsaicin pretreatment to deplete sensory nerves. In addition, endothelial denudation was combined with capsaicin pretreatment alone, or in combination with SR141716A at 1 μ M.

To establish whether NADA acts at the novel endothelial CB receptor, some vessels were treated with O-1918 (1 μ M), a silent antagonist of the receptor at which abnormal cannabidiol acts, which is neither CB₁ nor CB₂. This receptor is proposed to be a novel endothelial CB receptor (Offertaler *et al.*, 2003).

As a comparison to G3 vessels, the vascular effects of NADA were also investigated in the superior mesenteric artery (G0) to establish any possible regional differences between resistance (G3) and conduit (G0) vessels. Specifically, the effects of L-NAME, de-endothelialisation, capsaicin treatment and the antagonists SR141716A, AM251 and O-1918 were examined.

Statistical analysis

The concentration of vasorelaxant giving the half-maximal response (EC₅₀) was obtained from the concentration–response curve fitted to a sigmoidal logistic equation, with the minimum vasorelaxation set to zero using the GraphPad Prism package (Tep-areenan *et al.*, 2003). Maximal responses and pEC₅₀ (negative logarithm of the EC₅₀) values are expressed as mean \pm s.e.m. The number of animals in each group is represented by *n*. Data were compared, as appropriate, by Student's unpaired *t*-test or by analysis of variance (ANOVA) with statistical significance between manipulations and controls determined by Dunnett's *post hoc* test.

Drugs

All drugs were supplied by Sigma Chemical Co. (Poole, U.K.), except where stated. Carbachol and L-NAME were dissolved in Krebs–Henseleit solution. NADA, anandamide, SCH23390, capsaicin, capsazepine and SR141716A were dissolved in ethanol at 10 mM, with further dilutions made in distilled water. Anandamide and NADA were obtained as ethanolic solutions from Tocris (U.K.). SR141716A was supplied by Research Biochemicals International as part of the Chemical Synthesis Programme of the National Institute of Mental Health contract (NOIMH3003). SCH23390 was obtained from the Schering Corporation (U.S.A.). AM251 was a kind gift from Professor Sheila Gardiner and was initially dissolved in DMSO to 10 mM, with further dilutions in distilled water. O-1918 was generously donated by Dr George Kunos. O-1918 was dissolved in ethanol at 10 mM, with further dilutions made in distilled water.

Results

NADA caused vasorelaxation in all of the precontracted blood vessels tested. The small resistance arteries (G3) were significantly more responsive to NADA than the superior mesenteric artery (G0) or the aorta (Figure 1a; Table 1). An original representative trace of the effects of NADA in precontracted G3 vessels is presented in Figure 2. In G3, the vasorelaxant effects of NADA were significantly more potent than anandamide ($P < 0.05$), with a slightly lower maximal vasorelaxant effect ($P < 0.05$). In the superior mesenteric artery, NADA was significantly more effective than anandamide ($P < 0.05$). The vasorelaxant effects of NADA were comparable to those of anandamide in the aorta (Table 1; Figure 1b–d).

Isolated mesenteric resistance arteries (G3)

Characterisation of vasorelaxant response to NADA

When vessels were precontracted with 60 mM KCl, the maximal relaxant response to NADA was significantly inhibited (Table 2; Figure 3a). In preparations contracted with U46619, the dopamine receptor antagonist SCH23390 (1 μ M) had a small significant inhibitory response on the fitted maximal vasorelaxation response to NADA (Table 2; Figure 3b). However, the actual mean values obtained at the maximal concentration of NADA were not different from that seen in control arteries (control, $91.6 \pm 3.5\%$ relaxation; SCH23390, $82.3 \pm 6.7\%$ relaxation). Addition of L-NAME (300 μ M) did not affect vasorelaxation to NADA (Table 2; Figure 3c).

Endothelium-dependence and the role of EDHF

Removal of the endothelium resulted in a significant reduction in the relaxant responses to NADA (Table 2; Figure 4a). In endothelium-intact vessels, and in the presence of L-NAME (300 μ M) and indomethacin (10 μ M), the combination of 100 nM ChTX and 500 nM apamin produced comparable inhibition of NADA-induced vasorelaxation in G3 vessels (Table 2; Figure 4b).

Involvement of VRs Pretreatment with capsazepine (10 μ M) caused a 10.5-fold shift in the concentration–response curve to NADA, without affecting the maximal response (Table 2; Figure 5a). Similarly, pretreatment with capsaicin (10 μ M) significantly reduced the relaxant effects of NADA (Figure 5b). In the presence of L-NAME (300 μ M), capsaicin was still effective at reducing the effects of NADA (Figure 5b).

Effects of CB receptor antagonism The CB₁ receptor antagonist SR141716A (1 μ M) caused a 6.8-fold rightward shift in the concentration–response curve to NADA (Table 2; Figure 6a). However, SR141716A at 100 nM did not reduce the vasorelaxant responses to NADA (Figure 6a). A second CB₁ receptor antagonist, AM251, did not affect the responses to NADA at 1 μ M (Figure 6b), but at 100 nM, AM251 had a small inhibitory effect on the maximal response to NADA (Table 2; Figure 6b).

In endothelium-denuded preparations, the addition of SR141716A (1 μ M) did not affect the vasorelaxation to NADA further than was seen following endothelial denudation alone (Table 2; Figure 7a). In endothelium-intact preparations,

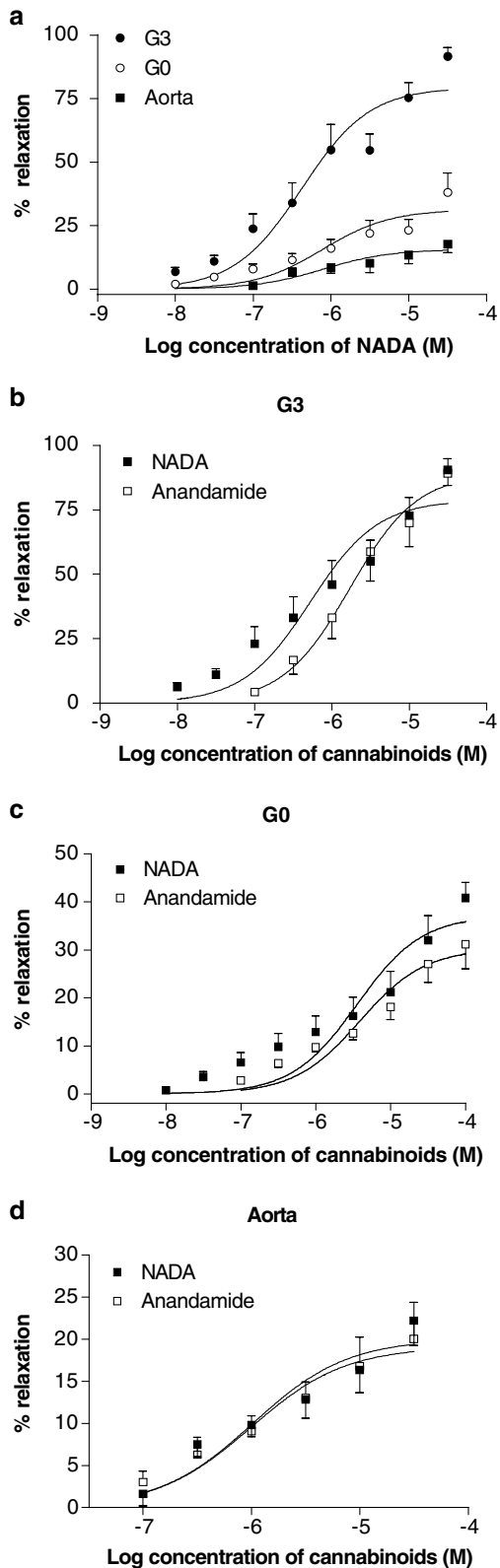


Figure 1 Vasorelaxation to NADA in isolated segments of G0 ($n = 7$) and G3 ($n = 10$) of the mesenteric bed and of the aorta ($n = 8$) compared to that of anandamide (G0 $n = 7$; G3 $n = 9$; aorta $n = 8$). Data are given as means, with error bars representing s.e.m.

capsaicin treatment for 1 h was combined with incubation with SR141716A ($1 \mu\text{M}$). In this experiment, the potency of NADA in G3 vessels was reduced further than that with capsaicin

Table 1 Comparison of the vasorelaxant effects of NADA and anandamide in the third-order branch of the superior mesenteric artery (G3), the superior mesenteric artery (G0) and in the aorta

	NADA			Anandamide		
	pEC_{50}	R_{max}	n	pEC_{50}	R_{max}	n
G3	$6.39 \pm 0.12^*$	$79.1 \pm 4.4^*$	10	5.77 ± 0.14	89.2 ± 7.1	9
G0	5.45 ± 0.15	$37.2 \pm 3.0^*$	7	5.41 ± 0.14	30.2 ± 2.4	7
Aorta	5.99 ± 0.17	20.0 ± 1.8	8	6.00 ± 0.21	19.1 ± 2.2	8

Data are given as mean \pm s.e.m. * denotes a significant difference between anandamide and NADA ($P < 0.05$).

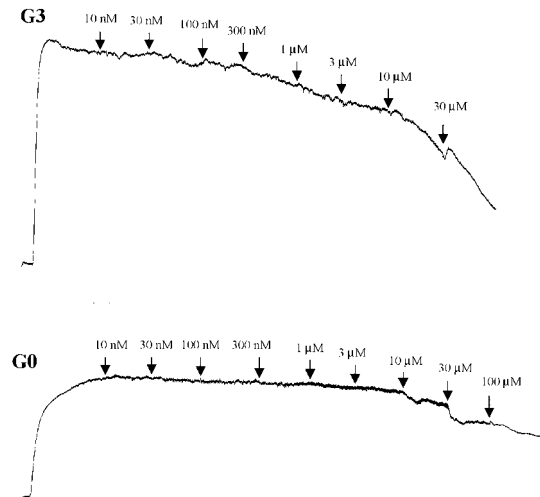


Figure 2 A representative trace of the vasorelaxant effects of NADA in preconstricted G3 and G0 mesenteric arteries.

treatment alone (Figure 7b). When capsaicin treatment was combined with endothelial denudation, it was found that, while the potency of NADA was reduced, there was no reduction in the maximum vasorelaxant effects (Figure 7c; Table 2). With the further addition of SR141716A at $1 \mu\text{M}$, the potency of NADA was reduced somewhat further, with no change in the maximal vasorelaxant response to NADA (Figure 7d; Table 2).

Addition of O-1918 ($1 \mu\text{M}$) caused a 13.5-fold rightward shift in the concentration–response curve to NADA (Table 2, Figure 9a).

Superior mesenteric artery (G0)

In the superior mesenteric artery, the presence of L-NAME ($300 \mu\text{M}$) had a modest inhibitory effect on the maximum vasorelaxant response to NADA (Table 3; Figure 8a). In contrast to the G3 vessels, de-endothelialisation in the superior mesenteric artery (G0) did not reduce vasorelaxation to NADA (Table 3; Figure 8c). Pretreatment with capsaicin substantially reduced the effects of NADA in the superior mesenteric artery (Table 3; Figure 8b), and the inhibitory effect of capsaicin on responses to NADA was not protected by the presence of $300 \mu\text{M}$ L-NAME (Figure 8c).

Unlike G3 vessels, pretreatment with SR141716A at 100 nM reduced the maximal vasorelaxant response to NADA (Table 3; Figure 8d). Similarly, AM251 (100 nM) also caused a reduction in the maximal response to NADA (Table 3; Figure 8d). The vasorelaxant responses to NADA were not

Table 2 Effects of various treatments on the vasorelaxation responses to NADA in third-order branches (G3) of the rat superior mesenteric artery

	pEC_{50}	R_{max}	n
Control	6.39 ± 0.12	79.1 ± 4.4	10
KCl contracted vessels	6.37 ± 0.16	28.1 ± 2.1	4
SCH23390 (1 µM)	6.34 ± 0.22	65.8 ± 6.9**	7
L-NAME (300 µM)	6.45 ± 0.14	85.3 ± 5.4	6
Endothelium denuded	5.70 ± 0.29**	39.1 ± 4.7**	6
ChTx (100 nM) & apamin (500 nM) with L-NAME (300 µM) & indomethacin (10 µM)	5.73 ± 0.29**	40.0 ± 7.0**	5
Capsazepine (10 µM)	5.37 ± 0.11**	78.7 ± 6.3	5
Capsaicin pretreatment (10 µM)	6.05 ± 0.17**	60.3 ± 6.4**	10
Capsaicin pretreatment (10 µM) & L-NAME (300 µM)	5.51 ± 0.16**	62.8 ± 5.5**	6
SR141716A (100 nM)	6.73 ± 0.21	75.3 ± 6.8	5
SR141716A (1 µM)	5.56 ± 0.17**	63.8 ± 7.0**	6
Endothelium denuded & SR141716A (1 µM)	5.78 ± 0.28**	47.1 ± 7.8**	6
Endothelium denuded & capsaicin pretreatment (10 µM)	5.17 ± 0.14**	90.0 ± 10.4	6
Endothelium denuded & capsaicin pretreatment (10 µM) & SR141716A	4.91 ± 0.14**	87.9 ± 11.5	6
Capsaicin pretreatment (10 µM) & SR141716A (1 µM)	5.02 ± 0.34**	54.1 ± 16.2**	6
AM251 (100 nM)	6.86 ± 0.30	65.8 ± 8.3**	6
AM251 (1 µM)	6.73 ± 0.21	69.8 ± 6.4	6
O-1918 (1 µM)	5.26 ± 0.20**	69.8 ± 10.8	6

Data are given as means ± s.e.m. Significant inhibitory effects compared with control vasorelaxant responses are indicated by *. * $P < 0.05$, ** $P < 0.01$, as determined by ANOVA.

inhibited by O-1918 in the superior mesentery artery (Table 3; Figure 9b).

Discussion

The aim of the present study was to characterise the vasorelaxant responses of the novel endogenous CB/vanilloid NADA. The results of this study show for the first time that NADA is a potent vasorelaxant in a number of blood vessels. The mechanisms of action of NADA and anandamide appear to be similar in small resistance vessels (G3), involving stimulation of VRs and the novel endothelial receptor, which may be coupled to the release of EDHF. Regional differences in the actions of NADA were also observed, such that, in larger mesenteric vessels (G0), relaxation to NADA was mediated through stimulation of the VR and the CB₁ receptor. There was no endothelial component to the responses to NADA in G0, and the novel endothelial receptor antagonised by O-1918 does not appear to be functional in these blood vessels.

It was hypothesised that NADA would have a vasorelaxant effect similar to anandamide, based on its structure and common pharmacology with anandamide (Bisogno *et al.*, 2000; Huang *et al.*, 2002). The initial findings of the present study identified NADA as a vasorelaxant, with similar potency to that of anandamide. More specifically, vasorelaxant activity was found in small resistance and larger conduit vessels, although this was more marked in the small vessels. Little is currently known about the synthesis, source or physiological relevance of NADA, but, given its commonality with anandamide, it might be speculated that NADA is of some pathophysiological importance (Wagner *et al.*, 1997).

To exclude the possibility that NADA acts at dopamine receptors, experiments were carried out in the presence of

SCH23390, an antagonist of the D₁ receptor, as stimulation of the D₁ receptor is known to cause vasodilatation (Goldberg, 1985). It was found that the effects of NADA were not through stimulation of this receptor. This is in agreement with Bisogno *et al.* (2000), who showed that NADA has little affinity for either D₁ or D₂ receptors in rat brain membranes.

In order to investigate the mechanisms of vasorelaxation, initial experiments showed that NADA was sensitive to a high potassium depolarising buffer. This is consistent with the notion that CBs may exert their effects through direct activation of potassium channels (Randall & Kendall, 1998) or through mediation *via* EDHF (Chaytor *et al.*, 1999). Alternatively, the high potassium solution may act on sensory nerves either by rapidly depleting or inhibiting the release of neurotransmitters *via* a depolarisation-clamp mechanism.

In the present study, the vasorelaxant responses to NADA in G3 vessels were found to be partly endothelium-dependent. This is in agreement with the findings of Chaytor *et al.* (1999) for anandamide in similar isolated arterial preparations, but contrary to those of White & Hiley (1997). However, it is clear that some other CBs such as abnormal cannabidiol are clearly endothelium-dependent (Járai *et al.*, 1999). This might highlight some potential differences in the actions of various CB molecules. To characterise the endothelium-derived vasorelaxant involved in the NADA response, experiments were performed in the presence of the nitric oxide synthase inhibitor L-NAME, and after potassium channel blockade, to inhibit EDHF activity. It was found that NADA does not act *via* the release of NO, which is in agreement with some reports on anandamide in isolated mesenteric vessels (White & Hiley, 1997; Chaytor *et al.*, 1999). However, relaxation to NADA was reduced when potassium channels were inhibited, suggesting that release of EDHF contributes to vasorelaxation to NADA, which is in agreement with findings

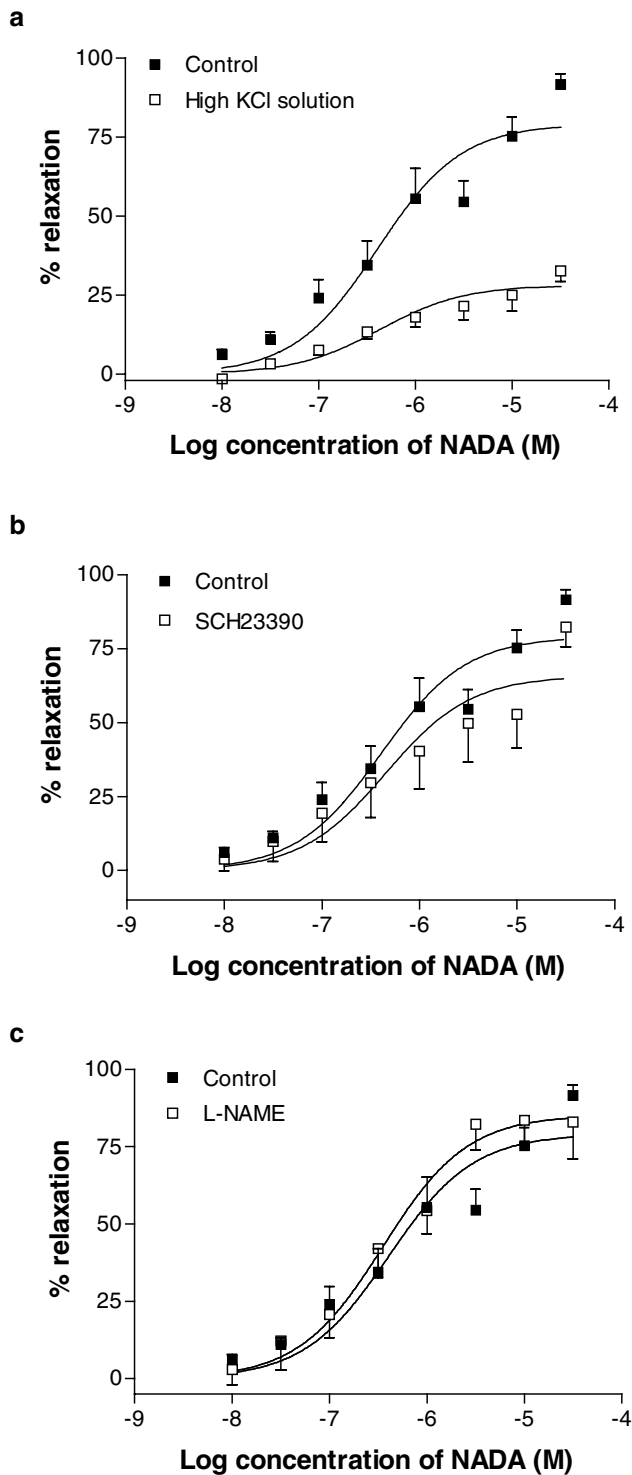


Figure 3 Vasorelaxant responses to NADA in the presence of a high potassium solution ($n=4$, a), the dopamine receptor antagonist SCH23390 ($1 \mu\text{M}$, $n=7$, b), and after nitric oxide synthase inhibition by L-NAME ($300 \mu\text{M}$, $n=6$, c). Data are given as means, with error bars representing s.e.m.

reported for anandamide in rabbit mesenteric vessels (Chaytor *et al.*, 1999). The endothelium-dependent component of vasorelaxation to NADA accounted for about 50% of the response.

In 1999, Zygmunt *et al.* investigated the possibility that anandamide stimulates VRs on the basis that anandamide is

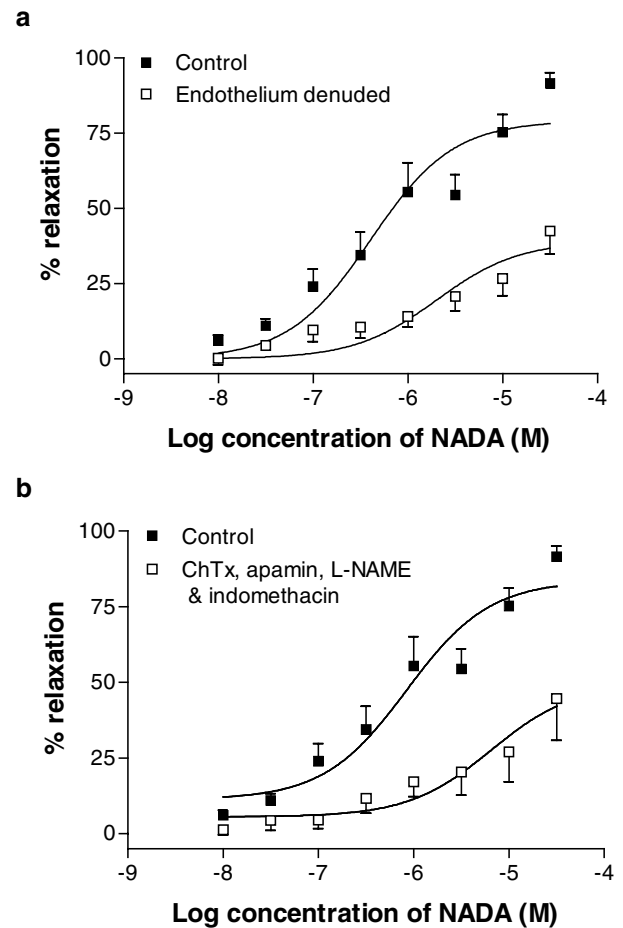


Figure 4 Vasorelaxant responses to NADA in small resistance vessels after removal of the endothelium ($n=6$, a), and after inhibition of EDHF activity with ChTx (100 nM) and apamin (500 nM) in the presence of L-NAME ($300 \mu\text{M}$) and indomethacin ($10 \mu\text{M}$) ($n=5$, b). Data are given as means, with error bars

structurally related to capsaicin. They found that, in isolated mesenteric arteries, vasorelaxation to anandamide was sensitive to the VR antagonist capsazepine, capsaicin pretreatment and CGRP receptor antagonism. A role for sensory nerves in vasorelaxation to CBs has been confirmed since then (Ralevic *et al.*, 2000; Harris *et al.*, 2002), although it is not thought that stimulation of the VR accounts for the entire relaxant response to anandamide (White *et al.*, 2001; Harris *et al.*, 2002; Ho & Hiley, 2003). Since NADA has also been found to stimulate VRs (Huang *et al.*, 2002), their potential involvement in the vasorelaxant response to NADA was investigated. It was found that both pretreatment with capsaicin and antagonism of the VR with capsazepine reduced the effects of NADA, confirming the involvement of VRs in vasorelaxation to NADA. This was true for both the resistance mesenteric artery and the superior mesenteric artery. Although it has been previously shown in the whole mesenteric arterial bed that the inhibitory effects of capsaicin on vasorelaxation to anandamide can be prevented by blocking nitric oxide synthesis (Harris *et al.*, 2002), this was not found to be the case in the present study for NADA.

NADA and anandamide have been shown to have similar potency at the CB₁ receptor; so, the possibility exists that vasorelaxation to NADA may be mediated, in part, through

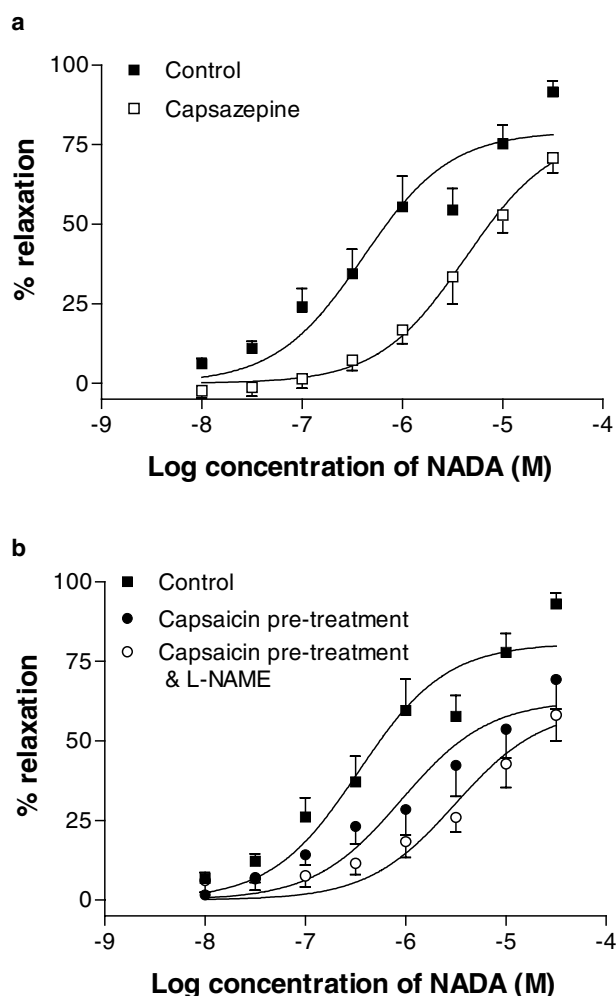


Figure 5 Effects of the VR antagonist capsazepine (10 μM , $n=5$, a), and pretreatment with the VR agonist capsaicin (10 μM , $n=10$, b) on vasorelaxation to NADA in small resistance vessels. Data are given as means with error bars representing s.e.m.

stimulation of this receptor. The involvement of CB receptors was examined using two different CB₁ receptor antagonists, SR141716A and AM251, in endothelium intact preparations. Interestingly, in G3 vessels, SR141716A at 1 μM reduced vasorelaxation to NADA, which may have suggested action *via* the CB₁ receptor, as at 1 μM this antagonist is regarded as being selective (Rinaldi-Carmona *et al.*, 1994). However, in the nanomolar range, SR141716A did not affect responses to NADA, and, furthermore, the structurally similar CB₁ antagonist AM251 had no inhibitory effect on the potency of NADA at either 100 nM or 1 μM . This would appear to rule out the involvement of the CB₁ receptor in vasorelaxation to NADA in G3 vessels, but does not exclude the possibility of NADA acting at a novel CB receptor sensitive to SR141716A at 1 μM (Járai *et al.*, 1999; Offertáler *et al.*, 2003). Interestingly, these findings are similar to work from Harris *et al.* (2002), where vasorelaxation to anandamide was inhibited in the whole mesenteric arterial bed in the presence of SR141716A at 3 or 10 μM , but not 1 μM , and was not affected by another CB₁ receptor antagonist, LY320135, at 10 μM . Accordingly, it was suggested that SR141716A at 3 μM or 10 μM might be acting at a novel CB receptor with low affinity for SR141716A. However, it cannot be ruled out that SR141716A may be

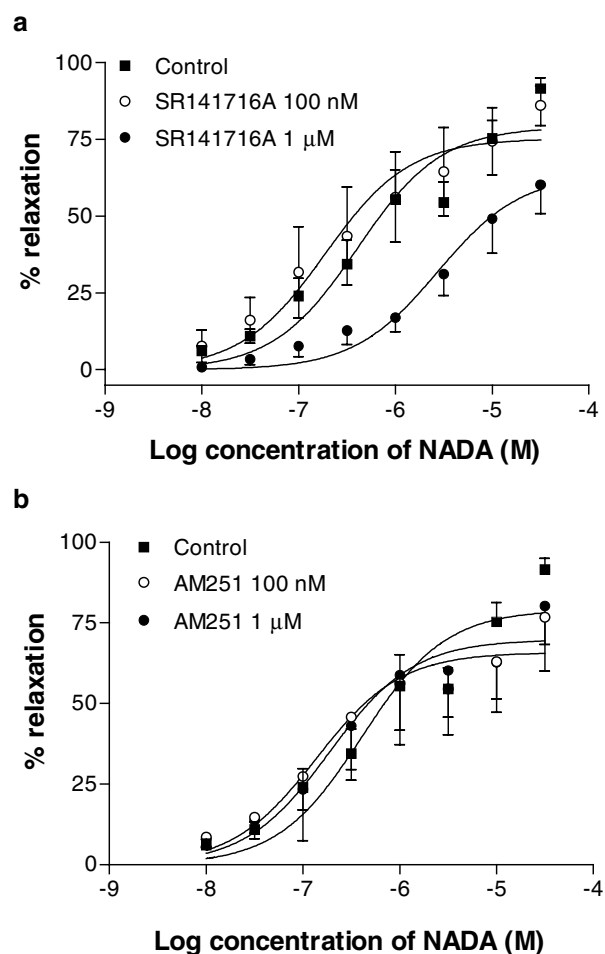


Figure 6 Effects of the CB₁ receptor antagonists SR141716A at 100 nM ($n=5$) and 1 μM ($n=6$, a) and AM251 at 100 nM ($n=6$) and at 1 μM ($n=6$, b) on vasorelaxation to NADA in small resistance vessels. Data are given as means, with error bars representing s.e.m.

opposing the EDHF-mediated response *via* inhibition of myoendothelial gap junctions, which has been observed by Chaytor *et al.* (1999) at a concentration of 10 μM .

After endothelial denudation, SR141716A at 1 μM did not inhibit the vasorelaxant effect of NADA in G3 further than was seen following endothelial denudation alone, suggesting that SR141716A acts at an endothelial site. Járai *et al.* (1999) reported a similar finding using anandamide, and this group postulated that a novel endothelial receptor exists that is sensitive to SR141716A at 1 μM . To further investigate this, a chemically modified analogue of cannabidiol was used in the present study. This compound binds to the site of action of abnormal cannabidiol, which has been shown to be neither the CB₁ nor the CB₂ receptor (Járai *et al.*, 1999; Offertáler *et al.*, 2003). In the present study, it was found that O-1918 antagonised the actions of NADA in a similar fashion to SR141716A at 1 μM , confirming that NADA acts at this proposed novel endothelial receptor, and suggesting also that SR141716A at 1 μM antagonises this receptor.

Since the endothelium-derived relaxing factor stimulated by NADA has been shown in the present study to be sensitive to a combination of drugs that are believed to inhibit EDHF activity (Randall & Kendall, 1998; Zygmunt *et al.*, 1997), stimulation of the novel endothelial receptor may be coupled

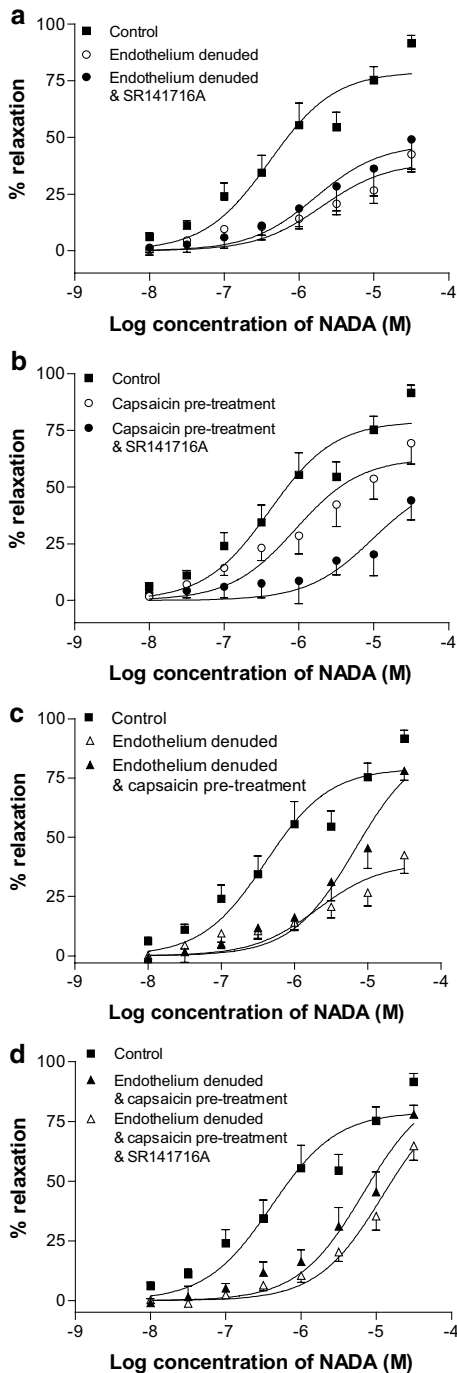


Figure 7 Effects of the CB₁ receptor antagonist SR141716A (1 μ M) on vasorelaxation to NADA in small resistance vessels after endothelial denudation ($n=6$, a) and after pretreatment of vessels with capsaicin (10 μ M, $n=6$, b) and after all the three manipulations ($n=6$, d). Data are given as means, with error bars representing s.e.m.

to the release of EDHF. It was also shown in this study that this pathway is independent of stimulation of the VR, since SR141716A at 1 μ M in combination with CGRP depletion by capsaicin reduced vasorelaxation to NADA to a greater extent than was seen with either manipulation alone. This is in agreement with previous work, which has shown that vasorelaxation to CBs is not mediated entirely by sensory nerves (Ralevic *et al.*, 2000; Harris *et al.*, 2002; Ho & Hiley, 2003).

Table 3 Effects of various treatments on the vasorelaxation responses to NADA in the rat superior mesenteric artery (G0)

	pEC_{50}	R_{max}	n
Control	5.45 ± 0.15	37.2 ± 3.0	7
L-NAME	5.63 ± 0.15	$23.4 \pm 3.0^*$	8
Endothelium denuded	5.29 ± 0.17	48.7 ± 5.0	7
Capsaicin pretreatment (10 μ M)	$3.75 \pm 0.92^{**}$	$17.6 \pm 6.4^{**}$	6
Capsaicin and L-NAME (300 μ M)	5.10 ± 0.74	$9.7 \pm 4.5^{**}$	6
SR141716A (100 nM)	6.21 ± 0.11	$16.3 \pm 2.3^{**}$	7
AM251 (100 nM)	$6.36 \pm 0.51^{**}$	$10.7 \pm 2.3^{**}$	6
O-1918 (1 μ M)	5.11 ± 0.14	56.1 ± 5.1	6

Data are given as mean \pm s.e.m. * denotes a significant inhibitory effect compared with control vasorelaxant responses (* $P < 0.05$, ** $P < 0.01$).

There are different mechanisms of CB-mediated vasorelaxation depending on the vascular bed studied (see Randall *et al.*, 2002). For example, vasorelaxation to anandamide in the rat coronary vessels does not involve sensory nerves (White *et al.*, 2001), and Andersson *et al.* (2002) showed that, while anandamide is a full agonist of the VR in guinea-pig mesenteric arteries, it is a weak agonist of this receptor in main bronchi. This might suggest that the actions of CBs are dependent on receptor distribution and density, and perivascular nerve density in a given tissue. Indeed, there have been reports of regional differences in anandamide-induced membrane potential changes between the main and daughter branches of the mesenteric artery (Vanheel & van de Voorde, 2001). In the present study, a number of experiments were performed in the superior mesenteric artery (G0) to compare with results found in the small resistance vessels (G3). It was found that there are both differences and similarities between the two vessel types. The main difference is that, in the superior mesenteric artery, the vasorelaxant response to NADA was not sensitive to removal of the endothelium, suggesting that the novel endothelial CB receptor is not expressed in this larger conduit vessel. This is supported by the finding that the cannabidiol analogue O-1918 did not reduce the vasorelaxant response to NADA in the superior mesenteric artery. It is of note that the responses of the endothelium-denuded G3 vessels were identical to those of the control G0 vessels, which could suggest that the endothelial pathway is the only difference between the artery types (see Figure 10).

Unlike G3 vessels, vasorelaxation to NADA in the superior mesenteric artery (G0) was sensitive to both SR141716A and AM 251 at 100 nM, suggesting that stimulation of the CB₁ receptor plays a role in the vasorelaxation of larger blood vessels to NADA. Given that both manipulations caused substantial inhibitory effects on vasorelaxation to NADA in G0, this might suggest that there is a degree of synergy between the sensory nerve-mediated vasodilation and the CB₁ receptor-mediated effect. Accordingly, in G0, there is clear evidence for participation of both sensory nerves and the CB₁ receptor; however, in G3, there is no evidence that CB₁ receptors participate in the response to NADA. Indeed, the residual relaxation to NADA in G3 in the presence of SR141716A and following both endothelial denudation and treatment with capsaicin (Figure 7d) points to an additional, unidentified mechanism of action.

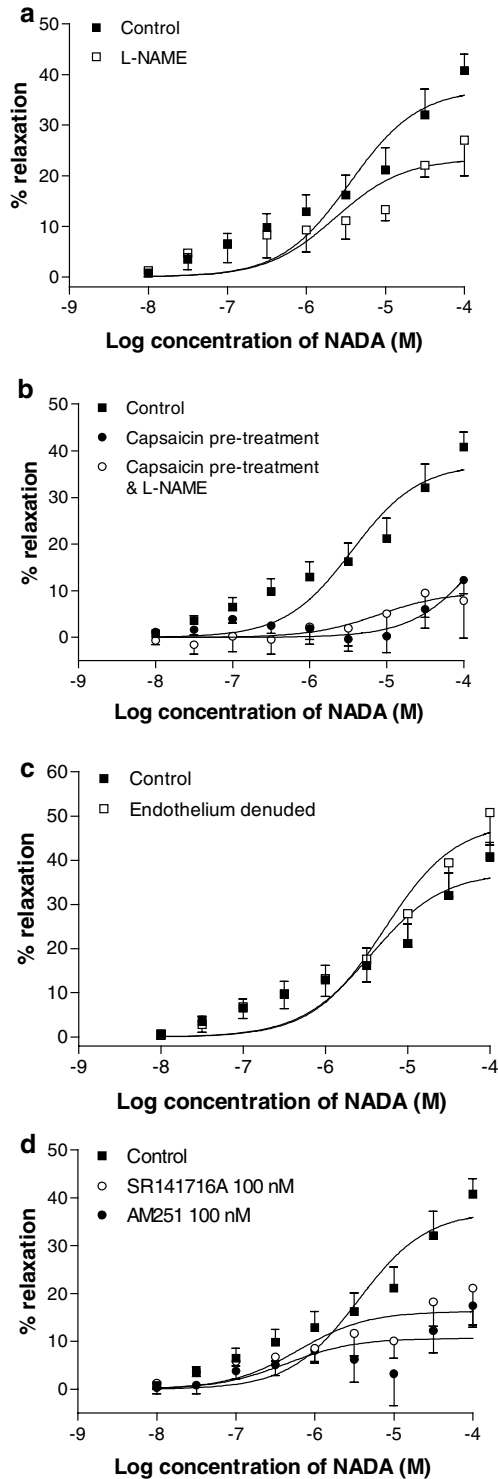


Figure 8 Vasorelaxant responses to NADA in the superior mesenteric artery after nitric oxide synthase inhibition by L-NAME (300 μ M, $n = 8$, a), de-endothelisation ($n = 7$, b), depletion of sensory neurotransmitters by capsaicin (10 μ M, $n = 6$, c), and after CB₁ receptor antagonism with SR141716A ($n = 7$) or AM251 ($n = 6$) at 100 nM (d). Data are given as means with error bars representing s.e.m.

In conclusion, the present study has demonstrated and characterised, for the first time, the vasorelaxant properties of the novel endogenous cannabinoid NADA. Its actions appear to be similar to those of the first identified endocannabinoid

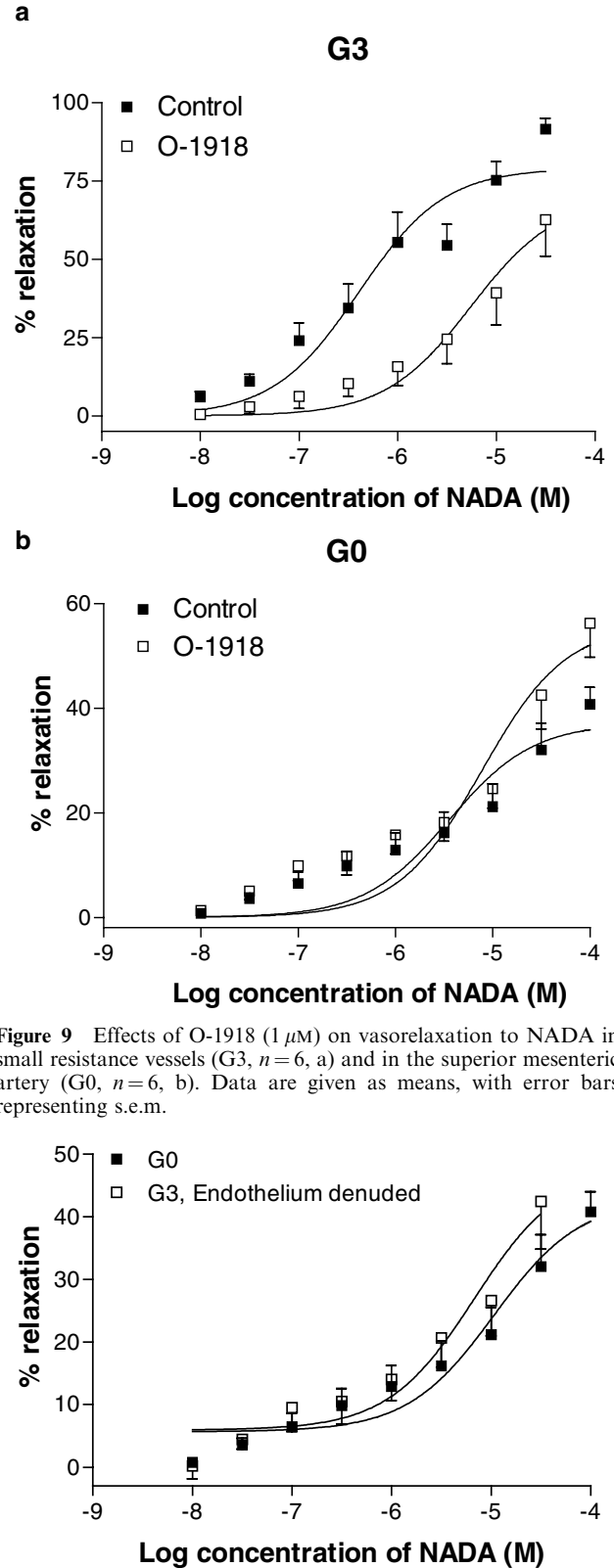


Figure 9 Effects of O-1918 (1 μ M) on vasorelaxation to NADA in small resistance vessels (G3, $n = 6$, a) and in the superior mesenteric artery (G0, $n = 6$, b). Data are given as means, with error bars representing s.e.m.

Figure 10 Vasorelaxant effects of NADA in the superior mesenteric artery compared with G3 vessels with the endothelium removed.

anandamide. In small resistance arteries, NADA acts at the VR and the novel endothelial CB receptor, which appears to be coupled to EDHF release. In the superior mesenteric artery,

NADA acts through stimulation of the vanilloid and CB₁ receptors. The difference in efficacy between the small and large vessels may lie solely in the absence of the endothelial pathway in the superior mesenteric artery (see Figure 10).

References

- ADEAGBO, A.S. & TRIGGLE, C.R. (1993). Varying extracellular [K⁺]: a functional approach to separating EDHF- and EDNO-related mechanisms in perfused rat mesenteric arterial bed. *J. Cardiovasc. Pharmacol.*, **21**, 423–429.
- ANDERSSON, D.A., ADNER, M., HOGESTATT, E.D. & ZYGMUNT, P.M. (2002). Mechanisms underlying tissue selectivity of anandamide and other vanilloid receptor agonists. *Mol. Pharmacol.*, **62**, 705–713.
- BISOGNO, T., MELCK, D., BOBROV, M.Y.U., GRETSKAYA, N.M., BEZUGLOV, V.V., DE PETROCELLIS, L. & DI MARZO, V. (2000). *N*-acyl-dopamines: novel synthetic CB(1) cannabinoid-receptor ligands and inhibitors of anandamide inactivation with cannabimimetic activity *in vitro* and *in vivo*. *Biochem. J.*, **351**, 817–824.
- CHAYTOR, A.T., MARTIN, P.E., EVANS, W.H., RANDALL, M.D. & GRIFFITH, T.M. (1999). The endothelial component of cannabinoid-induced relaxation in rabbit mesenteric artery depends on gap junctional communication. *J. Physiol.*, **520**, 539–550.
- CHU, C.J., HUANG, S.M., DE PETROCELLIS, L., BISOGNO, T., EWING, S.A., MILLER, J.D., ZIPKIN, R.E., DADDARIO, N., APPENDINO, G., DI MARZO, V. & WALKER, J.M. (2003). *N*-oleoyldopamine, a novel endogenous capsaicin-like lipid that produces hyperalgesia. *J. Biol. Chem.*, **278**, 13633–13639.
- DE PETROCELLIS, L., BISOGNO, T., MACCARRONE, M., DAVIS, J.B., FINAZZI-AGRO, A. & DI MARZO, V. (2001). The activity of anandamide at vanilloid VR1 receptors requires facilitated transport across the cell membrane and is limited by intracellular metabolism. *J. Biol. Chem.*, **276**, 12856–12863.
- DEUTSCH, D.G., GOLIGORSKY, M.S., SCHMID, P.G., KREBSBACH, R.J., SCHMID, R.J., DAS, S.K., DEY, S.K., ARREAZA, G., THORUP, C., STEFANO, G. & MOORE, L.C. (1997). Production and physiological actions of anandamide in the vasculature of the rat kidney. *J. Clin. Invest.*, **100**, 1538–1546.
- GARDINER, S.M., MARCH, J.E., KEMP, P.A. & BENNETT, T. (2002). Influence of the CB₁ receptor antagonist, AM 251, on the regional haemodynamic effects of WIN-55212-2 or HU 210 in conscious rats. *Br. J. Pharmacol.*, **136**, 581–587.
- GOLDBERG, L.I. (1985). Dopamine: receptors and clinical applications. *Clin. Physiol. Biochem.*, **3**, 120–126.
- HARRIS, D., MCCULLOCH, A.I., KENDALL, D.A. & RANDALL, M.D. (2002). Characterisation of vasorelaxant responses to anandamide in the rat mesenteric arterial bed. *J. Physiol.*, **539**, 893–902.
- HARRISON, S., DE PETROCELLIS, L., TREVISANI, M., BENVENUTI, F., BIFULCO, M., GEPPETTI, P. & DI MARZO, V. (2003). Capsaicin-like effects of *N*-arachidonoyl-dopamine in the isolated guinea pig bronchi and urinary bladder. *Eur. J. Pharmacol.*, **475**, 107–114.
- HO, W.S. & HILEY, C.R. (2003). Endothelium-independent relaxation to cannabinoids in rat-isolated mesenteric artery and role of Ca²⁺ influx. *Br. J. Pharmacol.*, **139**, 585–597.
- HUANG, S.M., BISOGNO, T., TREVISANI, M., AL-HAYANI, A., DE PETROCELLIS, L., FEZZA, F., TOGNETTO, M., PETROS, T.J., KREY, J.F., CHU, C.J., MILLER, J.D., DAVIES, S.N., GEPPETTI, P., WALKER, J.M. & DI MARZO, V. (2002). An endogenous capsaicin-like substance with high potency at recombinant and native vanilloid VR1 receptors. *Proc. Natl. Acad. Sci. U.S.A.*, **99**, 8400–8405.
- JÁRAI, Z., WAGNER, J.A., VARGA, K., LAKE, K.D., COMPTON, D.R., MARTIN, B.R., ZIMMER, A.M., BONNER, T.I., BUCKLEY, N.E., MEZEY, E., RAZDAN, R.K., ZIMMER, A. & KUNOS, G. (1999). Cannabinoid-induced mesenteric vasodilation through an endothelial site distinct from CB₁ or CB₂ receptors. *Proc. Natl. Acad. Sci. U.S.A.*, **96**, 14136–14141.
- MARTIN, S.W. & BROADLEY, K.J. (1995). Atypical antagonism of D1-receptor-mediated vasodilator response in the perfused kidney by SCH23390. *Pharmacol. Res.*, **31**, 289–297.
- MATSUDA, L.A., LOLAIT, S.J., BROWNSTEIN, M.J., YOUNG, A.C. & BONNER, T.L. (1990). Structure of a cannabinoid receptor and functional expression of the cloned cDNA. *Nature*, **346**, 561–564.
- MCCULLOCH, A.I., BOTTRILL, F.E., RANDALL, M.D. & HILEY, C.R. (1997). Characterisation and modulation of EDHF-mediated relaxations in the rat isolated superior mesenteric arterial bed. *Br. J. Pharmacol.*, **120**, 1431–1438.
- MULVANY, M.J. & HALPERN, W. (1977). Contractile properties of small arterial resistance vessels in spontaneously hypertensive and normotensive rats. *Circ. Res.*, **41**, 19–26.
- OFFERTÁLER, L., MO, F.M., BATKAI, S., LIU, J., BEGG, M., RAZDAN, R.K., MARTIN, B.R., BUKOSKI, R.D. & KUNOS, G. (2003). Selective ligands and cellular effectors of a G protein-coupled endothelial cannabinoid receptor. *Mol. Pharmacol.*, **63**, 699–705.
- RALEVIC, V., KENDALL, D.A., RANDALL, M.D., ZYGMUNT, P.M., MOVAHED, P. & HÖGESTÄTT, E.D. (2000). Vanilloid receptors on capsaicin-sensitive sensory nerves mediate relaxation to methanandamide in the rat isolated mesenteric arterial bed and small mesenteric arteries. *Br. J. Pharmacol.*, **130**, 1483–1488.
- RANDALL, M.D., ALEXANDER, S.P.H., BENNETT, T., BOYD, E.A., FRY, J.R., GARDINER, S.M., KEMP, P.A., MCCULLOCH, A.I. & KENDALL, D.A. (1996). An endogenous cannabinoid as an endothelium-derived vasorelaxant. *Biochem. Biophys. Res. Commun.*, **229**, 114–120.
- RANDALL, M.D. & GRIFFITH, T.M. (1991). Differential effects of L-arginine on the inhibition by *N*^G-nitro-L-arginine methyl ester of basal and agonist-stimulated EDRF activity. *Br. J. Pharmacol.*, **104**, 743–749.
- RANDALL, M.D., HARRIS, D., KENDALL, D. & RALEVIC, V. (2002). Cardiovascular effects of cannabinoids. *Pharm. Ther.*, **95**, 191–202.
- RANDALL, M.D. & KENDALL, D.A. (1998). Anandamide and endothelium-derived hyperpolarising factor act *via* a common vasorelaxant mechanism in rat mesentery. *Eur. J. Pharmacol.*, **346**, 51–53.
- RINALDI-CARMONA, M., BARTH, F., HEAULME, M., SHIRE, D., CALANDRA, B., CONGY, C., MARTINEZ, S., MARUANI, J., NELIAT, G., CAPUT, D., FERRARA, P., SOUBRIÉ, P., BRELIÈRE, J.C. & LE FUR, G. (1994). SR141716A, a potent and selective antagonist of the brain cannabinoid receptor. *FEBS Lett.*, **350**, 240–244.
- TEP-AREENAN, P., KENDALL, D.A. & RANDALL, M.D. (2003). Mechanisms of vasorelaxation to testosterone in the rat aorta. *Eur. J. Pharmacol.*, **465**, 125–132.
- VANHEEL, B. & VAN DE VOORDE, J. (2001). Regional differences in anandamide- and methanandamide-induced membrane potential changes in rat mesenteric arteries. *J. Pharmacol. Exp. Ther.*, **296**, 322–328.
- WAGNER, J.A., VARGA, K., ELLIS, E.F., RZIGALINSKI, B.A., MARTIN, B.R. & KUNOS, G. (1997). Activation of peripheral CB₁ cannabinoid receptors in haemorrhagic shock. *Nature*, **390**, 518–521.
- WHITE, R. & HILEY, C.R. (1997). A comparison of EDHF-mediated and anandamide-induced relaxations in the rat isolated mesenteric artery. *Br. J. Pharmacol.*, **122**, 1573–1584.
- WHITE, R., HO, W.S., BOTTRILL, F.E., FORD, W.R. & HILEY, C.R. (2001). Mechanisms of anandamide-induced vasorelaxation in rat isolated coronary arteries. *Br. J. Pharmacol.*, **134**, 921–929.
- ZYGMUNT, P.M., HOGESTATT, E.D., WALDECK, K., EDWARDS, G., KIRKUP, A.J. & WESTON, A.H. (1997). Studies on the effects of anandamide in rat hepatic artery. *Br. J. Pharmacol.*, **122**, 1679–1686.
- ZYGMUNT, P.M., PETERSSON, J., ANDERSSON, D.A., CHUANG, H.H., SORGARD, M., DI MARZO, V., JULIUS, D. & HOGESTATT, E.D. (1999). Vanilloid receptors on sensory nerves mediate the vasodilator action of anandamide. *Nature*, **400**, 452–457.

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