

## Unexpected Link between Iron and Drug Resistance of *Candida* spp.: Iron Depletion Enhances Membrane Fluidity and Drug Diffusion, Leading to Drug-Susceptible Cells<sup>∇</sup>

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Received 30 May 2006/Returned for modification 15 July 2006/Accepted 30 August 2006

**In this study, we show that iron depletion in *Candida albicans* with bathophenanthroline disulfonic acid and ferrozine as chelators enhanced its sensitivity to several drugs, including the most common antifungal, fluconazole (FLC). Several other species of *Candida* also displayed increased sensitivity to FLC because of iron restriction. Iron uptake mutations, namely,  $\Delta$ *ptr1* and  $\Delta$ *ptr2*, as well as the copper transporter mutation  $\Delta$ *ccc2*, which affects high-affinity iron uptake in *Candida*, produced increased sensitivity to FLC compared to that of the wild type. The effect of iron depletion on drug sensitivity appeared to be independent of the efflux pump proteins Cdr1p and Cdr2p. We found that iron deprivation led to lowering of membrane ergosterol by 15 to 30%. Subsequently, fluorescence polarization measurements also revealed that iron-restricted *Candida* cells displayed a 29 to 40% increase in membrane fluidity, resulting in enhanced passive diffusion of the drugs. Northern blot assays revealed that the *ERG11* gene was considerably down regulated in iron-deprived cells, which might account for the lowered ergosterol content. Our results show a close relationship between cellular iron and drug susceptibilities of *C. albicans*. Considering that multidrug resistance is a manifestation of multifactorial phenomena, the influence of cellular iron on the drug susceptibilities of *Candida* suggests iron as yet another novel determinant of multidrug resistance.**

Iron plays a key role in providing natural resistance to infections in humans (5). Interestingly, recent studies suggest that there could be a correlation between intracellular iron concentration and the multidrug resistance (MDR) phenomenon in mammalian cells (14). Iron depletion in mammalian cells with iron chelators is known to activate hypoxia-inducible factor 1 (HIF-1) (6–9, 31). Cellular iron status-dependent activation of HIF-1 results in the activation of its target gene, *MDR1* (7). Ambient hypoxia also causes resistance to chemotherapy by the induction of human *MDR1* in growing tumors via activation of HIF-1 (7–9, 31).

Recent studies of the role of iron in recurrent vulvovaginal candidosis have revealed that this element is important not only for the normal function of host immunity but also for pathogenic *Candida* owing to the fact that absence of this metal resulted in reduced virulence and hence reduced yeast invasion of the host epithelium (40). Availability of iron has been found to play a critical role in different clinical infections, and this represents a challenge to investigate the role of iron more closely (5). Kuipers et al. (25–27) have shown that lactoferrin, an iron-binding glycoprotein, is synergistic with antifungals against different *Candida* species. However, the effect

of iron on the drug susceptibility of *Candida* cells has yet to be demonstrated experimentally.

Iron is usually present in its ferric form complexed to environmental ligands (3, 5, 11, 21, 23). The insoluble ferric form cannot be taken up by *Candida* directly and has to be first solubilized by conversion to the ferrous form by the cell surface ferric reductase encoded by *CaCFL1* (3, 11, 18, 21). In abundance, ferrous iron is taken up by an iron uptake system comprising the iron transporter *FTR2*. In contrast, when the availability of iron is low in the environment, *Candida* uses the high-affinity iron uptake system, which comprises a membrane permease (encoded by *CaFTR1*) (36) and a multicopper oxidase (encoded by *CaFET3*) (12, 13). Since *CaFET3* has an essential requirement for copper (13, 42), reductive iron uptake in *Candida* also depends on copper availability, which is mediated by the intracellular copper transporter encoded by *CCC2* (23). An additional mechanism for iron uptake, a siderophore transporter (encoded by *SITI*) also exists in *Candida*, which makes use of the low-molecular-mass organic molecule siderophore to bind extracellular iron (20, 22).

The focus of this study was to find out whether the availability of iron could have an impact on the susceptibility of *Candida* to antifungal drugs. We observed that iron deprivation is a mechanism by which to enhance drug susceptibility in *Candida* cells. Our results suggest that iron depletion further introduces an increase in membrane fluidity, which in turn leads to enhanced passive diffusion of drugs, thereby resulting in enhanced drug susceptibility. We could also link changes in membrane fluidity to lowered ergosterol levels found in iron-deprived *Candida* cells probably because of down regulation of *ERG11*.

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∇ Published ahead of print on 5 September 2006.

TABLE 1. Strains used in this study

Strain	Genotype	Source or reference
<i>C. albicans</i> strains		
CAF2-1	<i>URA3/Δura3::imm434</i>	15
CAI4	<i>Δura3::imm434/Δura3::imm434</i>	15
C4-SHB1.1- <i>Δsit1</i>	<i>Δura3::imm434/Δura3::imm434 Δsit1/Δsit1::hisG-URA3-hisG</i>	20
<i>Δftr1</i>	<i>Δura3::imm434/Δura3::imm434 Δftr1::hisG/Δftr1::hisG</i>	36
<i>Δftr2</i>	<i>Δura3::imm434/Δura3::imm434 Δftr2::hisG/Δftr2::hisG</i>	36
<i>Δftr1 Δftr2</i>	<i>Δura3::imm434/Δura3::imm434 Δftr1::hisG/Δftr1::hisG Δftr2::hisG/Δftr2::hisG</i>	36
<i>Δccc2</i>	<i>Δura3::imm434/Δura3::imm434 Δccc2::hisG/Δccc2::hisG</i>	42
DSY449	<i>URA3/Δura3::imm434 Δcdr1::hisG/Δcdr1::hisG</i>	39
DSY1025	<i>Δura3::imm434/Δura3::imm434 Δcdr1::hisG/Δcdr1::hisG Δcdr2::hisG/Δcdr2::hisG</i>	39
DSY1024(II)	<i>Δura3::imm434/Δura3::imm434 Δcdr1::hisG/Δcdr1::hisG Δcdr2::hisG/Δcdr2::hisG Δcamdr1::hisG/Δcamdr1::hisG Δftu1::hisG/Δftu1::hisG</i>	29
Non- <i>C. albicans</i> strains		
<i>Candida krusei</i> ATCC <sup>a</sup> 6258		Ranbaxy Laboratories, India
<i>Candida tropicalis</i> ATCC 750		Ranbaxy Laboratories, India
<i>Candida parapsilosis</i> ATCC 22019		Ranbaxy Laboratories, India
<i>Candida kefyr</i> ATCC 2512		Ranbaxy Laboratories, India
<i>Candida dubliniensis</i> ATCC 33		Ranbaxy Laboratories, India
<i>Candida glabrata</i> ATCC 90030		Ranbaxy Laboratories, India

<sup>a</sup> ATCC, American Type Culture Collection.

## MATERIALS AND METHODS

**Materials.** Medium chemicals were obtained from HiMedia (Mumbai, India) and Difco (Detroit, Mich.). Rhodamine 6G (R6G), nystatin, cycloheximide, anisomycin, bathophenanthroline disulfonic acid (BPS), bathocuproine disulfonate (BCS), ferrozine, ferrous ammonium sulfate (FAS), ferric chloride (FeCl<sub>3</sub>), 2-deoxy-D-glucose (DOG), dinitrophenol (DNP), and 1,6-diphenyl-1,3,5-hexatriene were obtained from Sigma Chemical Co. (St. Louis, MO). FLC was kindly provided by Ranbaxy Laboratories, India. All of the chemicals used in this study were of analytical grade.

**Growth media.** Strains were routinely grown in YPD broth (1% yeast extract, 2% peptone, 2% dextrose), which served as iron-sufficient medium (ISM). Two percent (wt/vol) Bacto Agar (Difco, BD Biosciences, NJ) was added to YPD broth to make solid medium. For *Ura*<sup>-</sup> strains, uridine (Sisco Research Laboratory, Mumbai, India) at a concentration of 100 μg/ml was added to the growth medium. Iron-poor medium (IPM) was prepared by adding 200 μM BPS or 200 μM ferrozine to YPD or otherwise as mentioned in the figure legends (see Fig. 1, 2, and 4). Copper was chelated from YPD by adding 500 μM BCS. Iron-rich medium was prepared by adding either 200 μM FAS or 100 μM FeCl<sub>3</sub> to YPD.

**Strains used.** The *Candida albicans* strains and various *Candida* spp. used in this study are listed in Table 1. All of the strains were stored in 15% (vol/vol) glycerol stock at -80°C. Before each experiment, the cells were freshly revived on YPD plates from this stock. For all of these studies, *Candida* cells were maintained on YPD at 37°C. To begin with, all of the experiments were done with iron-replete cells. Cells suffered iron stress only during the experiments.

**Growth curve.** Cells were resuspended to an optical density at 600 nm (OD<sub>600</sub>) of 0.1 in YPD broth with or without BPS and were incubated in a shaking water bath at 37°C. OD<sub>600</sub> readings were taken at intervals of 1 h, and then a line graph was plotted to show differences in growth.

**Drug susceptibility tests.** Drug susceptibilities were measured by both broth microdilution and spot assays. MICs for the strains were determined by the broth microdilution method as described previously (32, 33, 35). The following stock solutions were used (the solvent used is given in parenthesis): FLC, 1 mg/ml (water); cycloheximide, 20 mg/ml (water); anisomycin, 10 mg/ml (water); nystatin, 10 mg/ml (water). The final drug concentrations used in this study are given in the figure legends. Susceptibilities to various drugs were checked in YPD. In order to have iron-limited conditions for the cells, BPS (a ferrous iron chelator) or ferrozine (a ferrous iron chelator) was added at the concentrations indicated in the figure legends to deplete the YPD of iron. Iron mutants were rescued by addition of either 200 μM FAS or 100 μM FeCl<sub>3</sub> to YPD containing FLC.

**Drug diffusion and efflux assay.** (i) **Passive diffusion.** Passive diffusion of the fluorescent compound R6G was determined by a protocol described previously (32, 33, 35). In a typical diffusion assay, done as described earlier (32, 33, 35), to

achieve de-energization of exponentially grown *Candida* cells for depletion of intracellular ATP, cells were resuspended in de-energization buffer (phosphate-buffered saline without glucose) with 5 mM DOG and 5 mM DNP at a cell density of 10<sup>8</sup> ml<sup>-1</sup>. R6G was then added to the de-energized cell suspension to a final concentration of 10 μM and incubated for 40 min, following which the cells were rapidly centrifuged and the extracellular concentration of R6G in the supernatant was determined spectrophotometrically at 527 nm.

(ii) **R6G efflux.** The functionality of Cdr1p was checked by assaying the energy-dependent efflux of R6G, a known substrate of this drug extrusion pump. The protocol of the efflux assay has been described in our earlier publications (24, 32, 33, 35).

**Measurement of fluorescence polarization.** Steady-state fluorescence polarization studies were done with *Candida* cells with the fluorescent probe 1,6-diphenyl-1,3,5-hexatriene at excitation and emission wavelengths of 360 and 426 nm, respectively (32, 33, 35).

**Quantitation of ergosterol.** Sterols were extracted by the alcoholic KOH method and the percentage of ergosterol was calculated as described previously (32, 33, 35). The extracted sterols indicated a four-peak spectral absorption pattern produced by ergosterol and 24(28)-dehydroergosterol [24(28)-DHE] contents. Both ergosterol and 24(28)-DHE absorb at 281.5 nm, whereas only 24(28)-DHE absorbs at 230 nm. Ergosterol content is determined by subtracting the amount of 24(28)-DHE (calculated from the *A*<sub>230</sub>) from the total ergosterol-plus-24(28)-DHE content (calculated from the *A*<sub>281.5</sub>). Ergosterol content was calculated as a percentage of the wet weight of the cells with the following equations: % ergosterol + % 24(28)-DHE = [(*A*<sub>281.5/290</sub>) × *F*]/pellet weight, % 24(28)-DHE = [(*A*<sub>230/518</sub>) × *F*]/pellet weight, and % ergosterol = [% ergosterol + % 24(28)-DHE] - % 24(28)-DHE, where *F* is the factor for dilution in petroleum ether and 290 and 518 are the *E* values (in percent per centimeter) determined for crystalline ergosterol and 24(28)-DHE, respectively.

**RNA isolation and hybridization.** Northern blot analyses were carried out essentially by standard protocols as described before (35, 38). Equal loading of RNA was checked by rRNA bands. RNA was electrophoresed on a denaturing formaldehyde gel (1.2%) and blotted and UV cross-linked onto Hybond-N<sup>+</sup> nylon (Amersham Biosciences) membranes. Membrane-bound RNA was stained with methylene blue before hybridization to check rRNA bands for equal loading. Relative intensities of *ERG11* mRNA and *ERG3* mRNA signals in Northern hybridizations were quantitated by exposure of the hybridized membrane in a Fuji FLA5000 phosphorimager. *ERG11* and *ERG3* probes were made by PCR amplification with primers *ERG11F* (5'-ATACATGAATTCTACTGCTGCTGCCAAAGC-3'), *ERG11R* (5'-ATACATAAGCTTCCCAAATGATTTCTGCTG-3'), *ERG3F* (5'-ATACATGAATTCTTCACTTTTCCACCGATTG-3'), and *ERG3R* (5'-ATACATAAGCTTATCATCTGGTCTTCTGTA-3').

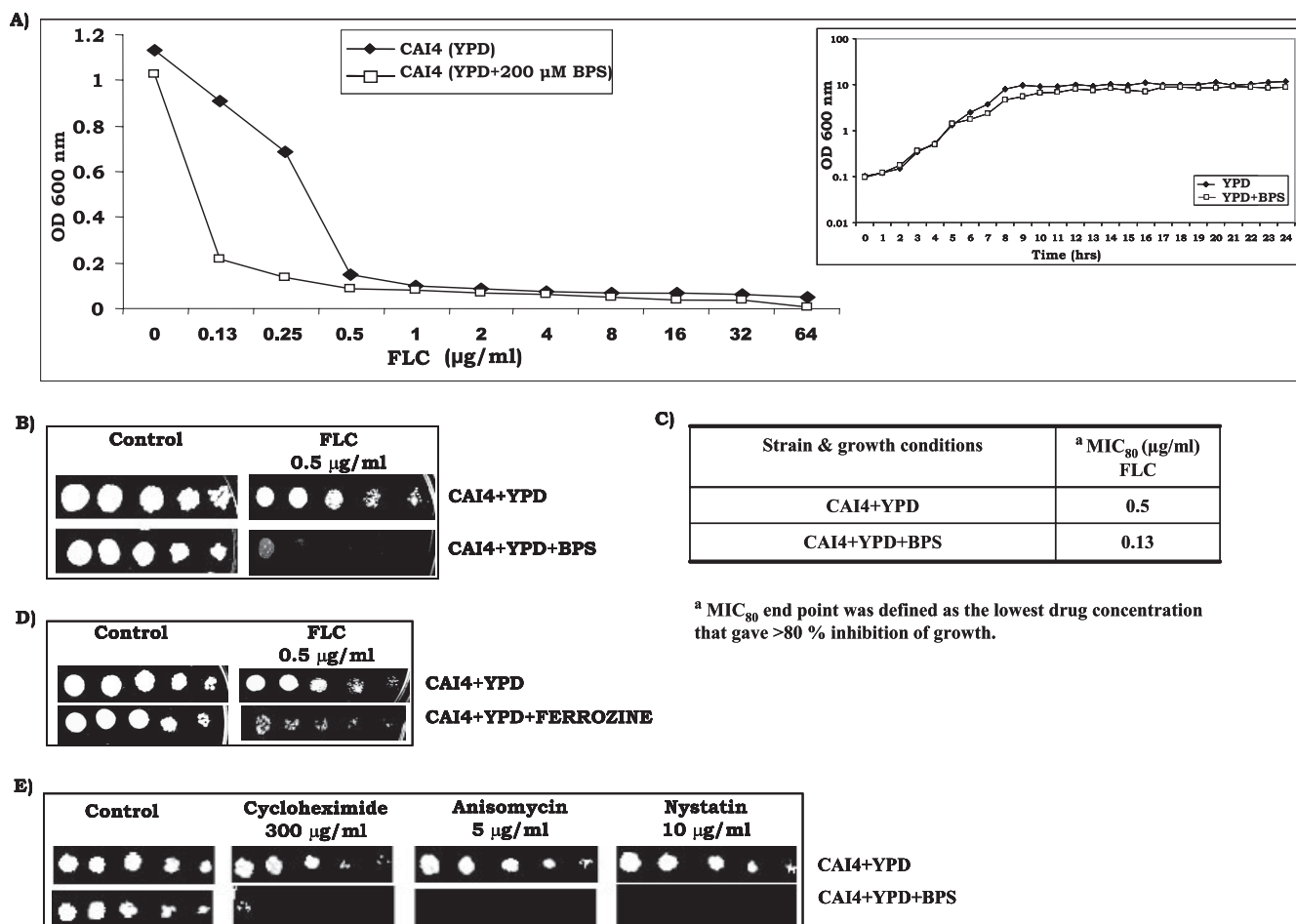


FIG. 1. (A) Determination of dose-ranging inhibition by broth microdilution assay of CAI4 with FLC at concentrations varying from 0.13 to 64 µg/ml in the absence (◆) or presence (□) of 200 µM BPS. Growth of cells was evaluated both visually and by reading the  $A_{600}$  in a microplate reader as described earlier (32, 33, 35). The inset shows the growth curve of CAI4 cells as a semilogarithmic plot of  $OD_{600}$  as a function of time (in hours) in the absence (◆) or presence (□) of 200 µM BPS. (B) Drug resistance profile of CAI4 cells as determined by spot assay for FLC in the absence or presence of 200 µM BPS. For spot assays, 5-µl volumes of fivefold serial dilutions of each yeast culture ( $A_{600}$ , 0.1) were spotted onto YPD plates in the absence (control) or presence of FLC (0.5 µg/ml). Growth differences were evaluated with drug-free controls following incubation of the plates for 48 h as described elsewhere (32, 33, 35). Growth was not affected by the presence of the solvents used for the drugs (data not shown). (C) MICs of FLC for CAI4 cells in the absence or presence of BPS. The lowest drug concentration that gave >80% inhibition of growth compared to the drug-free control was determined by broth microdilution assay and evaluated both visually and by reading the  $A_{600}$  in a microtiter plate reader as described earlier (32, 33, 35). (D) Drug susceptibility tests for CAI4 cells by spot assay in the absence or presence of 200 µM ferrozine. FLC was used at a concentration of 0.5 µg/ml. (E) Spot assays of CAI4 cells in the absence or presence of 200 µM BPS for cycloheximide (300 µg/ml), anisomycin (5 µg/ml), and nystatin (10 µg/ml). Growth differences were recorded as described above. Growth was not affected by the presence of the solvents used for the drugs (data not shown).

## RESULTS

**Iron starvation makes *C. albicans* susceptible to FLC.** It is known that iron deprivation affects the growth of several organisms (3, 11, 21, 34). Therefore, we first assessed if the growth of *C. albicans* cells is affected by BPS at the concentration used in this study. The inset in Fig. 1A illustrates that the growth of CAI4 was not affected when cells were grown in 200 µM BPS. It should be pointed out that concentrations of BPS higher than 200 µM caused growth inhibition and hence were not used. The chelation of  $Fe^{2+}$  (ferrous) by BPS was confirmed by the assay for ferroxidase activity and was performed as described elsewhere (12). The enzyme ferroxidase converts  $Fe^{2+}$  (ferrous) to  $Fe^{3+}$  (ferric), and the remaining  $Fe^{2+}$  (ferrous) complexes with ferrozine to give a purple color. The

reduced intensity of the color of the ferrozine- $Fe^{2+}$  (ferrous) complex, as indicated by the decrease in absorbance at 570 nm, implies less availability of  $Fe^{2+}$  (ferrous). At 570 nm, the number of arbitrary units per 200 µg protein of the ferrozine- $Fe^{2+}$  (ferrous) complex was 0.5 in untreated CAI4 cells, which was reduced to 0.38 in BPS-treated cells. These results clearly showed that BPS at 200 µM was able to chelate iron in the medium without affecting the growth of the cells.

Two independent methods, namely, broth microdilution and spot assays, were used to find out whether iron depletion causes any change in drug susceptibility of *Candida* cells. The dose-ranging inhibition curve depicted in Fig. 1A confirmed that even though the growth of BPS-treated and untreated *Candida* cells in drug-free medium was comparable; however,

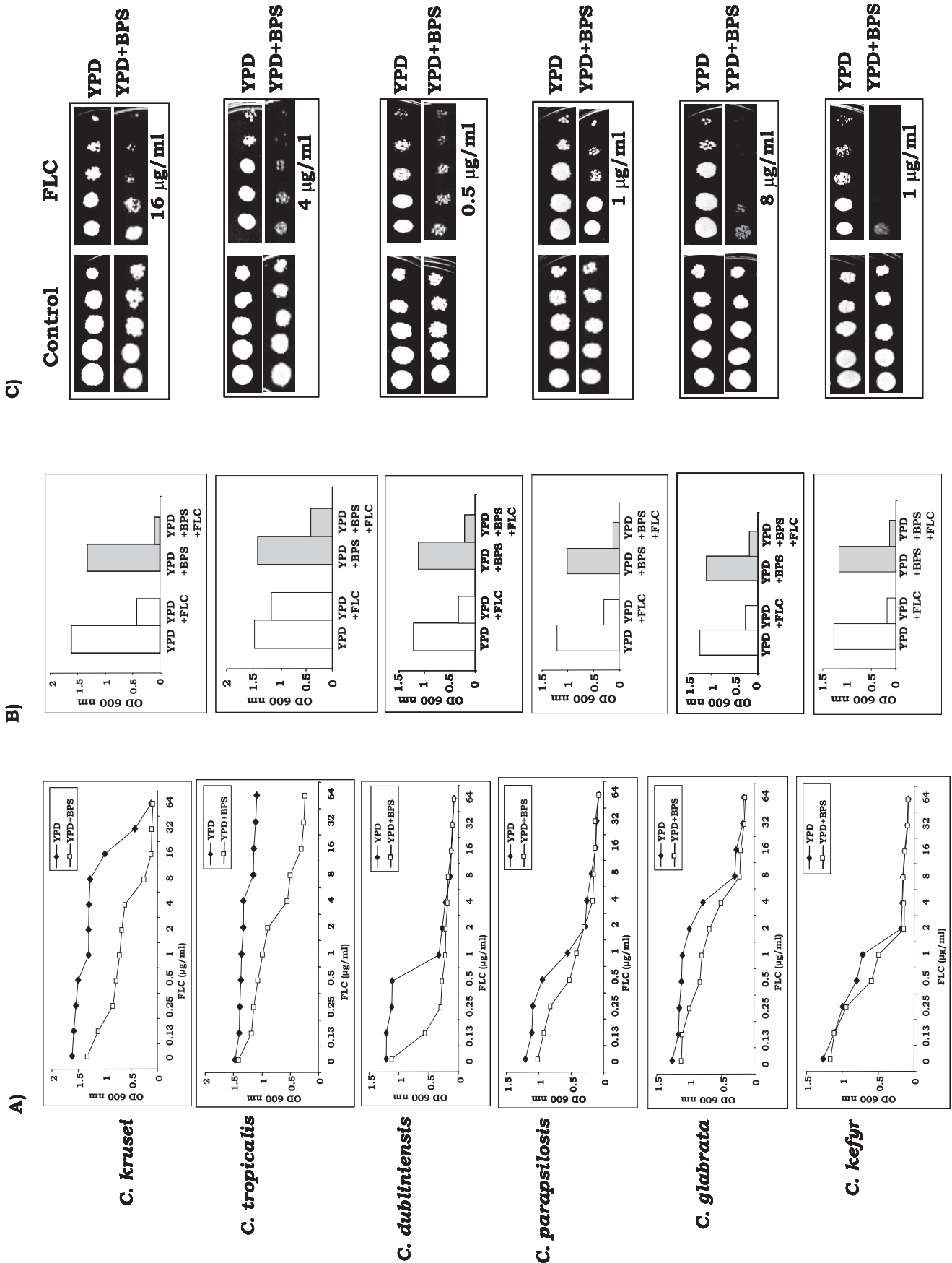


FIG. 2. Drug resistance profiles of non-*C. albicans* species as determined by dose-ranging inhibition curves (A) for non-*C. albicans* species of *Candida* with FLC at concentrations varying from 0.13 to 64  $\mu\text{g/ml}$  in the absence (◆) or presence (□) of BPS (100  $\mu\text{M}$  for all non-*C. albicans* spp. except 200  $\mu\text{M}$  for *C. tropicalis*). (B) MICs of FLC for CAI4 cells in the absence or presence of BPS are presented as histograms, and the  $\text{OD}_{600}$  of the cells is plotted on the y axis. The lowest drug concentration that gave >80% inhibition of growth compared to the drug-free control was determined by broth microdilution assay and evaluated both visually and by reading the  $A_{600}$  in a microtiter plate reader as described earlier (32, 33, 35). Clear bars represent control without BPS, and shaded bars represent cells grown in the presence of BPS. BPS was used at a concentration of 100  $\mu\text{M}$  for all of the species except *C. tropicalis*, for which the concentration used was 200  $\mu\text{M}$ . The lowest FLC concentration that gave >80% inhibition of growth compared to the drug-free control in the absence of BPS was 32  $\mu\text{g/ml}$  (*C. krusei*), 8  $\mu\text{g/ml}$  (*C. tropicalis*), 1  $\mu\text{g/ml}$  (*C. dubliniensis*), 4  $\mu\text{g/ml}$  (*C. parapsilosis*), 16  $\mu\text{g/ml}$  (*C. glabrata*), or 2  $\mu\text{g/ml}$  (*C. kefyr*) and is represented by the second open bar in each graph in panel B. The second shaded bar in each graph in panel B depicts the growth of cells in the presence of the above-mentioned concentration of BPS and that of FLC, which was 32  $\mu\text{g/ml}$  (*C. krusei*), 8  $\mu\text{g/ml}$  (*C. tropicalis*), 1  $\mu\text{g/ml}$  (*C. dubliniensis*), 4  $\mu\text{g/ml}$  (*C. parapsilosis*), 16  $\mu\text{g/ml}$  (*C. glabrata*), or 2  $\mu\text{g/ml}$  (*C. kefyr*). (C) Spot assay for FLC in the absence or presence of 100  $\mu\text{M}$  BPS for all other species except *C. tropicalis*, for which the concentration used was 200  $\mu\text{M}$ .

the cells growing in the presence of BPS were distinctly more susceptible to FLC compared to those growing under iron-sufficient conditions. Spot assay data in Fig. 1B revealed that cells growing in the presence of BPS showed increased sensitivity to FLC compared to that of those growing under iron-sufficient conditions. Spot assays (Fig. 1B) also confirmed the MIC results (Fig. 1C).

**Iron depletion with other chelators also increases the drug sensitivity of *C. albicans* cells to FLC.** BPS has been used in studies including all of the classical iron-copper biology experiments (2, 4, 10, 30, 31). Our objective in this study was to find out whether the drug susceptibility of *Candida* cells is also affected when the cellular iron level is compromised by other chelators. To validate the results obtained with BPS, we used ferrozine ( $\text{Fe}^{2+}$  chelator). Figure 1D reveals that ferrozine-treated cells also showed increased susceptibility to FLC. Similar results were obtained with another  $\text{Fe}^{3+}$  chelator, desferrioxamine (data not shown).

**Iron deprivation enhances the susceptibility of *C. albicans* cells to other drugs.** To check if an increase in the susceptibility of *C. albicans* due to iron deprivation is limited to FLC, we performed spot assays with cycloheximide (300  $\mu\text{g/ml}$ ), nystatin (5  $\mu\text{g/ml}$ ), and anisomycin (10  $\mu\text{g/ml}$ ). We observed that CAI4 cells under IPM conditions displayed enhanced susceptibility to the other tested drugs as well (Fig. 1E). Thus, it is apparent that iron depletion led to enhanced susceptibility of *C. albicans* cells to a variety of drugs.

**Iron depletion results in an increase in the drug sensitivity of other *Candida* spp.** Both MIC and spot tests were performed for six different *Candida* species with FLC as a test drug under ISM and IPM conditions (Fig. 2). On the basis of broth microdilution assay data depicted in Fig. 2A as dose-ranging inhibition curves, it became apparent that, similar to *C. albicans*, all of the non-*C. albicans* species became sensitive to FLC under iron-deprived conditions. To reconfirm broth microdilution assay results, the lowest FLC concentration that showed a marked difference in growth between ISM and IPM media was selected for each species and spot tests were performed. The spot assay results depicted in Fig. 2C largely confirmed the MIC results shown in Fig. 2B. The extent of sensitivity of non-*C. albicans* species to FLC was variable for different species and was in the following order: *C. kefyr* > *C. glabrata* > *C. tropicalis* > *C. krusei* > *C. dubliniensis* > *C. parapsilosis*.

***Candida* mutants defective in iron and intracellular copper transport show increased susceptibility to FLC.** In order to confirm the role of iron in the drug susceptibility of *Candida* cells, we used different iron acquisition-defective *Candida* mutants in the following experiment. All of the null mutants and wild-type cells showed similar growth in YPD, but when their growth was challenged with FLC in the spot assay, null mutants defective in iron uptake, namely,  $\Delta\text{ftr1}$  (defective in high-affinity iron uptake) and  $\Delta\text{ftr1} \Delta\text{ftr2}$  (null mutant of both the high-affinity iron transporters), showed increased susceptibility to FLC in the complete absence of any iron chelator. Mutant strain  $\Delta\text{ftr2}$  (devoid of another iron transporter) only showed a marginal increase in susceptibility, whereas  $\Delta\text{sit1}$  (defective in siderophore transport) showed no change in its sensitivity to FLC (Fig. 3A). Interestingly,  $\Delta\text{ccc2}$  cells (defective in copper transport) also showed enhanced sensitivity to FLC in the absence of any chelator (Fig. 3A). Of note here is that copper

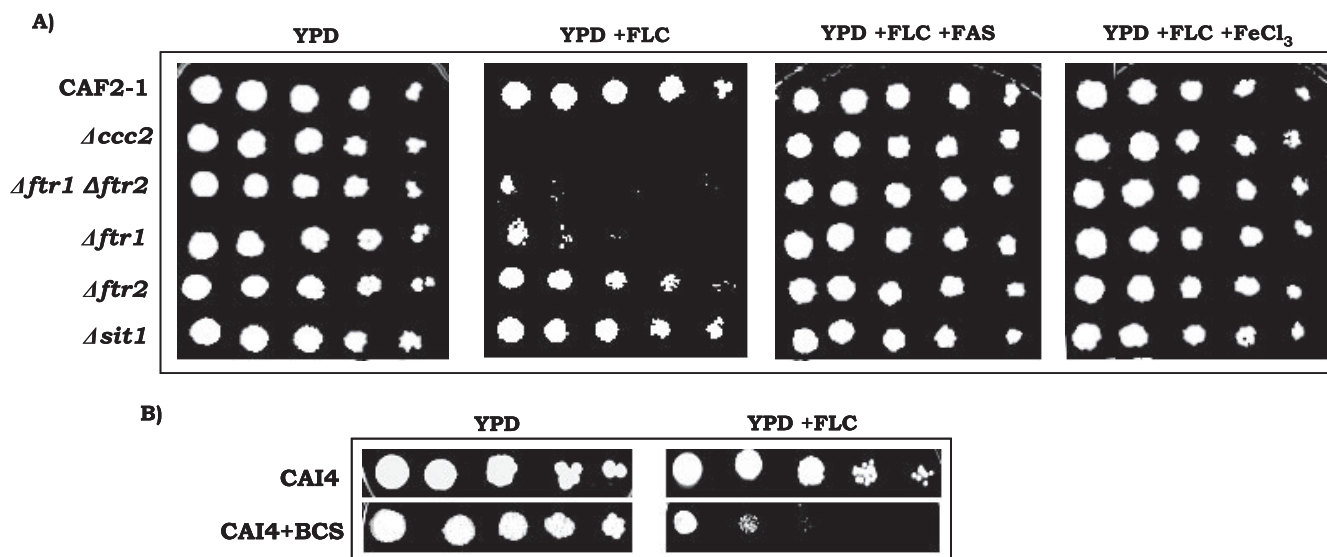


FIG. 3. (A) Drug resistance profiles of iron mutants as determined by spot assay in the absence (control) or presence of FLC (0.5  $\mu\text{g/ml}$ ) and reversal of FLC susceptibility by rescuing the growth defect of iron acquisition-defective mutant cells by addition of 200  $\mu\text{M}$  FAS or 100  $\mu\text{M}$  ferric chloride to a YPD plate containing FLC. (B) Spot assay for untreated CAI4 cells and CAI4 cells treated with 500  $\mu\text{M}$  BCS in the absence (control) or presence of FLC (0.5  $\mu\text{g/ml}$ ).

is an essential component of the multicopper oxidase (FET3) responsible for high-affinity iron uptake. Chelation of copper from the medium by BCS resulted in cells with increased sensitivity to FLC (Fig. 3B). These results indicate that chelation of copper, which affects high-affinity iron transport in *Candida*, enhances the drug sensitivity of *Candida* cells even under iron-rich medium conditions.

**Supplementation of the medium with iron reverses the enhanced susceptibility of *Candida* to FLC.** To confirm the iron and copper chelation effects on the drug sensitivity of *Candida* cells, we tested if supplementation of the medium with iron salts could reverse the effect. For this, we used  $\Delta ftr1$ ,  $\Delta ftr2$ ,  $\Delta ftr1 \Delta ftr2$ , and  $\Delta ccc2$  mutant cells and assessed their susceptibilities to FLC under iron-rich medium conditions. Figure 3A shows that when these cells were grown in the presence of 200  $\mu\text{M}$  FAS or 100  $\mu\text{M}$   $FeCl_3$ , the inhibitory effect of FLC on growth was reversed.

**Synergism between iron depletion and sensitivity to FLC is not dependent on multidrug efflux transporter activity.** Overexpression of drug efflux pumps is one of the well-known mechanisms for the development of FLC resistance in *Candida* (16, 19). To examine the role, if any, of major ABC efflux pumps, namely, *CDR1*, *CDR2*, and MFS pump member *CaMDR1*, in synergism with iron deprivation and drug sensitivity, we did an experiment with efflux pump-encoding gene null mutants DSY449 ( $\Delta cdr1$ ), DSY1024II ( $\Delta cdr1 \Delta cdr2 \Delta camdr1 \Delta flu1$ ), and DSY1025 ( $\Delta cdr1 \Delta cdr2$ ). Spot assays were done to investigate how the sensitivities of drug efflux pump mutants DSY449 ( $\Delta cdr1$ ), DSY1024II ( $\Delta cdr1 \Delta cdr2 \Delta camdr1 \Delta flu1$ ), and DSY1025 ( $\Delta cdr1 \Delta cdr2$ ) were affected when they were exposed to FLC under iron-limiting conditions. Both MIC and spot test data clearly demonstrate that there was no difference in sensitivity to FLC under ISM and IPM conditions between null mutants and the wild type (Fig. 4A and B).

The involvement of drug efflux pumps in influencing the

drug sensitivity of *Candida* cells under IPM conditions was further ruled out by examining and comparing R6G (a fluorescent substrate of Cdr1p) efflux rates. In a typical experiment, *Candida* cells grown under IPM and ISM conditions were de-energized by exposure to DOG and DNP as described in our previous publications (32, 33, 35). R6G was added to a final concentration of 10  $\mu\text{M}$  to the de-energized cells resuspended in phosphate-buffered saline at a cell density  $10^8 \text{ ml}^{-1}$  and incubated to attain steady intracellular accumulation. The efflux of equilibrated fluorescent R6G was initiated by the addition of 1 mole of glucose. The extracellular concentration of R6G was monitored by measuring its  $A_{527}$ . Our results show that there was no difference in the extracellular concentration of R6G between the cells grown under IPM and ISM conditions (data not shown). This suggests that the efflux rates and levels of R6G remained unchanged irrespective of the intracellular iron status. Thus, any contribution of efflux pump protein activity in enhancing the drug susceptibilities of *Candida* cells upon iron deprivation was excluded from these experiments.

**Membrane ergosterol level is altered under iron-depleted conditions.** Recent microarray-based experiments suggested that cellular iron regulates a host of genes, including those involved in membrane homeostasis (28). We explored the effect of iron deprivation on membrane lipid composition, which in turn may affect the ability of the drug to permeate the cell membrane. In order to examine if iron affects the level of ergosterol, one of the main constituents of the cell membrane, we checked the ergosterol contents of iron-depleted cells and iron mutant cells. There was a marked reduction in ergosterol content in CAI4 cells under IPM conditions, which ranged from 10% for BCS-treated cells to 32.5% for BPS-treated cells (Table 2). Of note here is that the decrease in ergosterol levels could be reversed by iron supplementation (data not shown). Interestingly, iron mutant cells ( $\Delta ftr1$  and  $\Delta ccc2$ ) also showed

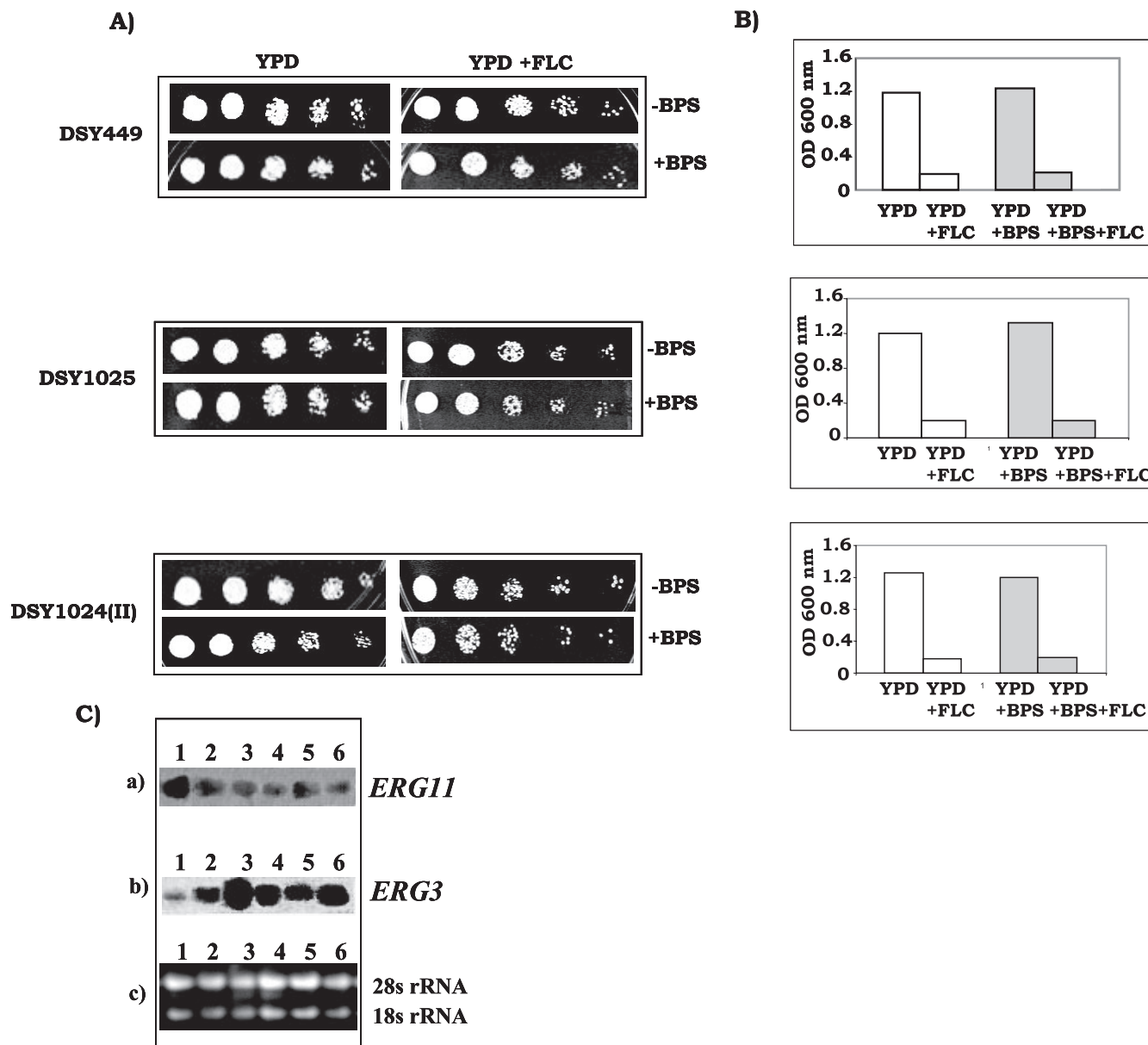


FIG. 4. (A) Drug susceptibilities of drug efflux pump mutants DSY449, DSY1025, and DSY1024(II) as determined by spot assays in the absence or presence of 100 μM BPS. To detect any effect of BPS on growth, a low concentration of FLC (0.125 μg/ml), at which mutant cells could grow, was used. (B) MICs of FLC for drug efflux pump mutants in the absence or presence of BPS are represented as histograms, and the OD<sub>600</sub> of the cells is plotted on the y axis. Clear bars represent cells grown without BPS, and shaded bars represent cells grown in the presence of 100 μM BPS. The lowest FLC concentration that gave >80% inhibition of growth compared to the drug-free control was 0.125 μg/ml for all of the mutant cells grown with or without BPS. (C) Northern blot analyses of *ERG11* and *ERG3*. (a) *ERG11* transcript level. (b) *ERG3* transcript levels in strain CAI4 (lane 1), strain CAI4 grown in the presence of 200 μM BPS (lane 2), strain  $\Delta ccc2$  (lane 3), strain  $\Delta fir1$  (lane 4), strain  $\Delta fir2$  (lane 5), and strain  $\Delta fir1 \Delta fir2$  (lane 6). (c) Loading controls for verifying equal gel loading of total RNA.

reduced ergosterol contents compared to wild-type cells (Table 2). No other major change in membrane lipids was observed under IPM conditions (data not shown).

**Iron starvation leads to a more fluid membrane.** We had earlier observed an intricate relationship between the (i) membrane fluidity and lipid composition and (ii) drug susceptibility of *Candida* cells (32, 33, 35). We had proposed that both entry and extrusion of drugs may be affected if membrane lipids are altered. In view of the reduction in ergosterol levels under IPM

conditions, this aspect was explored. We used steady-state fluorescence polarization (measured as a “p” value) to examine the physical state of the membranes of cells grown under IPM and iron-rich medium conditions. A decrease in the p value implies a decrease in membrane order or an increase in membrane fluidity. The CAI4 strain of *C. albicans* showed a reduced p value under iron depletion conditions and under copper depletion conditions as well (Table 2). A direct link between membrane fluidity and intracellular iron levels was

TABLE 2. Ergosterol contents, p values, and passive diffusion of R6G in untreated and BPS- and BCS-treated wild-type cells and mutant cells with defective iron uptake

Strain (condition for growth)	% Ergosterol <sup>a</sup>	p value <sup>b</sup>	OD <sub>527</sub> <sup>c</sup>
CAI4	1.22 ± 0.05	0.171 ± 0.001	0.309 ± 0.023
CAI4 (200 μM BPS)	0.81 ± 0.02	0.08 ± 0.01	0.209 ± 0.02
CAI4 (500 μM BCS)	1.08 ± 0.04	0.113 ± 0.008	0.202 ± 0.033
$\Delta$ <i>ftr1</i>	0.97 ± 0.02	0.103 ± 0.01	0.237 ± 0.019
$\Delta$ <i>ccc2</i>	1.03 ± 0.033	0.114 ± 0.01	0.202 ± 0.02

<sup>a</sup> The mean ergosterol content of cells is expressed as a percentage of the wet weight of cells ± the standard deviation of three sets of experiments (33).

<sup>b</sup> The mean p value of cells (inversely proportional to membrane fluidity) ± the standard deviation of three sets of experiments is shown (32, 33, 35).

<sup>c</sup> The mean OD<sub>527</sub> of supernatant, for checking passive diffusion of R6G inside cells (a lower OD implies a higher concentration of R6G inside the cells), ± the standard deviation of three sets of experiments is shown (32, 33, 35).

established when mutants defective in iron transport, viz.,  $\Delta$ *ftr1*, and copper transport, viz.,  $\Delta$ *ccc2*, were analyzed. These mutant cells possessed increased membrane fluidity compared to the wild-type strain (Table 2), which could be reversed by addition of iron (data not shown).

**Iron depletion leads to an increase in passive diffusion of drugs.** To test the effect of passive influx of the drug, we first blocked the contribution of the efflux pumps by de-energizing the cells in order to deplete the intracellular ATP. R6G diffusion was then monitored in de-energized cells as described in Materials and Methods. Table 2 depicts extracellular levels of R6G in IPM compared to CAI4 cells grown under ISM conditions. It is clear that depriving cells of either iron or copper resulted in enhanced passive diffusion, as was evident from a decreased extracellular concentration of R6G. The mutants defective in high-affinity iron ( $\Delta$ *ftr1*) and copper ( $\Delta$ *ccc2*) transport also showed enhanced membrane fluidity and diffusion of R6G (Table 2). The enhanced passive diffusion under iron-starved conditions could be reversed by addition of iron to the medium (data not shown). Similarly, with  $\Delta$ *ftr1* and  $\Delta$ *ccc2* cells, enhanced R6G diffusion could be brought back to normal levels by supplementation of the medium with iron salts (data not shown).

**Ergosterol biosynthetic genes are regulated by iron.** Recent microarray data showed that among all of the ergosterol biosynthetic genes, iron deprivation results in down regulation of *ERG11* and up regulation of *ERG3* (both genes are known to have a reverse relationship) (28). Since we observed in our study that iron deprivation leads to lowering of ergosterol content, we wanted to correlate it with the transcript level of these genes. To find out whether expression of any of these two genes is affected by iron depletion, we checked *ERG3* and *ERG11* transcript levels by Northern blot analyses with iron-starved and iron acquisition-defective mutant strains of *C. albicans*. We observed that iron deprivation led to the down regulation of *ERG11*, which encodes lanosterol 14- $\alpha$  demethylase, a key enzyme of ergosterol biosynthesis (Fig. 4C). Northern blot assays of iron-deprived cells, as well as of iron transport-defective mutants ( $\Delta$ *ftr1*,  $\Delta$ *ftr2*,  $\Delta$ *ftr1*  $\Delta$ *ftr2*, and  $\Delta$ *ccc2*), showed considerable down regulation of *ERG11* transcript. *ERG3* acts downstream of *ERG11* in the ergosterol biosynthesis pathway and encodes the  $\Delta$ 5,6-desaturase. In contrast to *ERG11*, *ERG3* was up regulated under IPM conditions (Fig.

4C). In an azole-inhibited pathway, Erg3p is responsible for converting the nontoxic 14-methyl intermediates (1) which accumulate because of azole inhibition of lanosterol 14- $\alpha$  demethylase into the toxic sterol 14-methylergosta-8,24(28)-dien-3,6-diol. *ERG3*, being up regulated under iron-depleted conditions, becomes synergistic with azoles because of greater accumulation of the toxic intermediate.

## DISCUSSION

Iron is an absolute requirement for most organisms and assumes an important role in host innate immunity (5, 17, 40). It has also been established that in *Candida*, tight regulatory specialized iron acquisition tactics constitute virulence factors (20, 36). However, the requirement of the high-affinity iron permease for infection in a mouse model (36), the requirement of a siderophore transporter for epithelial invasion, and the iron-dependent endothelial injury caused by *C. albicans* have established a role for iron in systemic infections (17). In the present study, we explored if iron availability affects the drug susceptibilities of *Candida* cells. For the first time, our results demonstrate that alterations in the intracellular iron concentration could also play a role in the defense mechanism of *Candida* against drugs. We have demonstrated that deprivation of iron results in increased sensitivity of *C. albicans* cells to a host of drugs. Since all of the drugs tested have different mechanisms of action, with varied targets within the cell, this implies that iron starvation affects the cell in a broad manner. Other non-*C. albicans* species of *Candida* tested, e.g., *C. krusei*, *C. tropicalis*, *C. dubliniensis*, *C. kefyr*, *C. parapsilosis*, and *C. glabrata*, also turned sensitive to drugs under iron-depleted conditions, which confirmed that the effect of iron is not limited to a single *Candida* species and may represent another mechanism of regulation of drug resistance in these opportunistic fungi. There have been a few earlier reports suggesting the possibility of a link between intracellular iron levels and drug resistance. For example, lactoferrin, an iron-binding glycoprotein, was found to be synergistic with antifungals against *C. albicans*, *C. glabrata*, and *C. tropicalis* (25–27).

The possibility of iron depletion resulting in a growth defect in *C. albicans* cells and hence an increase in drug sensitivity was ruled out. We alleviated such concerns by monitoring the growth of CAI4 cells and demonstrated that while BPS was sufficient to chelate iron at the concentration used in this study, it did not affect the growth of the cells (Fig. 1A, inset). This was further confirmed when we used various iron uptake mutants ( $\Delta$ *ftr1*,  $\Delta$ *ftr2*, and double mutant  $\Delta$ *ftr1*  $\Delta$ *ftr2*), as well as an intracellular copper transporter mutant ( $\Delta$ *ccc2*). Interestingly, all iron transport-defective mutants showed comparable growth and were found to be inherently sensitive to FLC compared to the wild type without iron chelation (Fig. 3A). However,  $\Delta$ *sit1*, a siderophore uptake mutant, did not show a similar increase in susceptibility to FLC compared to wild-type cells (Fig. 3A). This could be explained by the fact that since both the high- and low-affinity iron uptake machinery of  $\Delta$ *sit1* mutant cells is still intact, cells do not need to resort to siderophore-mediated iron uptake to satisfy their needs. A direct link between iron levels and drug resistance was further established when the drug-sensitive phenotype was found to be reversed upon supplementation of the growth medium with



extra iron salts (ferrous ammonium sulfate or ferric chloride) (Fig. 3A). Taken together, our results confirmed that *Candida* cells become relatively more sensitive to drugs when they are deficient in iron, either because of chelation of the metal or because of their iron acquisition defect mutations ( $\Delta$ *ftr1*,  $\Delta$ *ftr2*,  $\Delta$ *ftr1*  $\Delta$ *ftr2*, and  $\Delta$ *ccc2*).

Since overexpression of the drug efflux pump-encoding genes is one of the predominant mechanisms responsible for conferring a drug resistance phenotype on *C. albicans*, we first hypothesized that the observed changes in drug susceptibility due to iron deprivation could be the result of reduced drug efflux. However, when we used various efflux pump null mutants, viz.,  $\Delta$ *cdr1*,  $\Delta$ *cdr1*  $\Delta$ *cdr2*, and  $\Delta$ *cdr1*  $\Delta$ *cdr2*  $\Delta$ *camd1*  $\Delta$ *flu1*, and compared their sensitivities to FLC under IPM and ISM conditions, none of the mutants showed any further increase in sensitivity to FLC (Fig. 4A and B). This implies that the synergism between antifungals and iron starvation is not directly related to the activity of the efflux pump proteins. The issue that iron levels exert their influence on drug susceptibilities of *Candida* cells by an independent mechanism was finally settled when we observed that the efflux of the fluorescent substrate R6G mediated by the Cdr1p and Cdr2p efflux proteins remained the same, irrespective of the iron levels in *Candida* cells.

A recent genome-wide study of gene expression as a function of alterations in environmental concentrations of iron revealed that a host of genes are highly expressed at low iron concentrations while several transcripts are up regulated under high-iron conditions (28). Among the iron-regulated genes, cytochrome (*ERG11*, *ERG3*) and fatty acid metabolism (*OLE1*) genes were also found to be affected by the iron status of cells. Since we observed the ergosterol content of cells in IPM to be lower than that of those grown under ISM conditions, we hypothesized that changes in *ERG* genes under iron deprivation conditions could affect ergosterol levels, which could explain the higher membrane fluidity and enhanced passive drug entry observed. We observed that *ERG11* was considerably down regulated under iron-restricted conditions, while *ERG3* showed the opposite effect under similar conditions. Interestingly, CAI4 cells grown in IPM showed a substantial reversible decrease in fluorescence anisotropy (p value [membrane order]), implying that iron deprivation indeed results in ergosterol-dependent membrane fluidization. It was further evident that iron deprivation fluidizes the membrane when the iron uptake mutants  $\Delta$ *ftr1*,  $\Delta$ *ftr2*, and  $\Delta$ *ftr1*  $\Delta$ *ftr2* and the intracellular copper transporter mutant  $\Delta$ *ccc2* showed inherently lower ergosterol content and enhanced membrane fluidity (data not shown). This increase in membrane fluidity directly results in increased passive diffusion of drugs and sensitization of *Candida* cells. The fact that CAI4 cells under iron deprivation, iron acquisition-defective mutant  $\Delta$ *ftr1*, and intracellular copper transporter mutant  $\Delta$ *ccc2* showed increased R6G passive diffusion and *ERG11* gene down regulation supports our conclusions.

Taken together, our results demonstrate that iron deprivation of *Candida* cells regulates ergosterol synthesis genes, resulting in lower levels of this very important constituent of the membrane. Ergosterol-depleted cellular membrane, in turn, becomes more fluid, presumably allowing faster passive entry of drugs and thus increasing the drug sensitivity of the cells. A

recent demonstration by Raymond's group that Upc2p, a zinc cluster factor that regulates *ERG* genes, also affects iron acquisition genes, points to a close relationship between iron and ergosterol metabolism in *Candida* (37). The possibility of co-regulation of MDR, lipid biosynthesis, and iron acquisition genes through common regulators also exists, as has already been observed in several instances. On the one hand, *PDR1* and *PDR3*, well-known Zn(II)-Cys<sub>6</sub>-Zn(II) transcription factors which regulate pleiotropic drug resistance in *Saccharomyces cerevisiae*, target the sphingolipid biosynthetic gene *IPT1*, and on the other hand, these key regulators also affect the efflux and storage of cellular iron (41). In conclusion, changes in the drug susceptibility of *Candida* due to iron represent a well-regulated new defense mechanism that merits a closer look.

#### ACKNOWLEDGMENTS

The work presented in this paper has been supported in part by grants to R.P. from the Department of Biotechnology, India [BT/PR3825/MED/14/488(a)/2003 and BT/PR4862/BRB/10/360/2004], and support to C.K.M. in the form of a Wellcome Trust senior research fellowship. T.P. and A.C. thank the Council of Scientific and Industrial Research and the Indian Council of Medical Research, respectively, for senior research fellowships.

We also thank D. Sanglard, Switzerland, for generously providing drug efflux pump knockout strains of *C. albicans*. J. Ernst, Y. Wang, and D. Kornitzer are also acknowledged for providing iron mutant strains. We thank Ranbaxy Laboratories (New Delhi, India) for providing FLC and clinical isolates of other non-*C. albicans* *Candida* spp.

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