

## Genetic Polymorphism and Molecular Epidemiology of *Leishmania (Viannia) braziliensis* from Different Hosts and Geographic Areas in Brazil

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**Numerical zymotaxonomy and variability of the internal transcribed spacers (ITS) between the small and large subunits of the rRNA genes were used to examine strain variation and relationships in natural populations of *Leishmania (Viannia) braziliensis*. A total of 101 strains from distinct hosts and Brazilian geographic regions were assigned to 15 zymodemes clustered in two major genetic groups. The great number of isolates (48.5%) placed in zymodeme IOC/Z-27 were collected on the Atlantic coast. The high molecular diversity found in populations in the Amazon Basin was related to the great number of sandfly vector(s) in that region. The results of the restriction fragment length polymorphism analysis of the ITS depicted considerable intraspecific variation. Genotypic groups A, B, and C contained 39, 40, and 22 isolates, which were divided into 16, 10, and 15 genotypes, respectively. The genetic polymorphism observed demonstrates the degree of diversity of *L. (V.) braziliensis* strains from different regions where they are endemic. The results reinforce the clonal theory for *Leishmania* parasites showing the genetic diversity of this pathogen and an association of *L. (V.) braziliensis* genotypes with specific transmission cycles, probably reflecting an adaptation of different clones to the vector species involved.**

Infections with the parasitic protozoan *Leishmania (Viannia) braziliensis* Vianna 1911 (Kinetoplastida: Trypanosomatidae) or strain variants are recognized as causing human illness in many areas of (sub)tropical America (at least 15 countries), where it constitutes a significant public health problem. Many of these parasites seem to have a unique life cycle, with different phlebotomine sandfly (Diptera: Psychodidae) vectors and/or animal reservoirs and a different geographic distribution (13). This pathogen is capable of producing a variety of clinical manifestations, such as (i) cutaneous leishmaniasis (CL), which may heal spontaneously; (ii) mucosal leishmaniasis (ML), a hyperergic invasive ulcerative form that progresses in the absence of any apparent cellular defect (15); and (iii) disseminated CL (4).

Most of the environmental factors affecting the epidemiology of the various leishmaniasis are still poorly understood. Wild mammals serve as reservoirs for most of the New World *Leishmania* spp. (21), but there is increasing evidence that some of the human pathogenic *Leishmania* strains can be maintained in both sylvan and urban cycles. In the case of *L. (V.) braziliensis*, the principal vertebrate hosts in the sylvan cycle have not been identified, but there is evidence that dogs, horses, and donkeys may serve as reservoir hosts of this para-

site (15). The existence of an urban cycle involving peridomestic sandfly species for *L. (V.) braziliensis* reflects the ability of these parasites and their vectors to adapt to changes in their original forested habitats with important public health implications. Studies using molecular techniques to characterize *L. (V.) braziliensis* populations from different regions have shown a relationship between level of similarity among the parasite populations (12, 24) and their geographic range, but recent data have also indicated that the considerable variability detected among these parasites is more probably related to the sandfly vector(s) and/or animal reservoir(s) involved in the transmission cycles (18).

Pathogens that produce many different genetic variants are more prone to infect multiple hosts (37). Although several studies have discussed the polymorphism observed in natural populations of different *Leishmania* species (8, 25, 34), until now there has been little information available about the genetic variability of the parasites and the correlation with eco-epidemiological features of the disease (16). The risk factors for clinical leishmaniasis are still poorly understood and probably are influenced by host and parasite features.

Considering the public health importance of leishmaniasis caused by *L. (V.) braziliensis* in Brazil and the role of the genetic polymorphism of the parasites in the epidemiology of the disease, we applied multilocus enzyme electrophoresis (MLEE) and the analysis of the restriction fragment length polymorphism (RFLP) of the internal transcribed spacers (ITS) of the rRNA genes as typing methods to determine the level of genetic variation in natural population of *L. (V.) bra-*

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TABLE 1. Results of molecular typing and provenance details of 101 *L. (V.) braziliensis* strains collected over a 23-year period that were studied for their genetic relatedness

Zymodeme <sup>a</sup>	Genotypic type <sup>b</sup> (no. of stocks)	Geographic origin <sup>c</sup>	Clinical form <sup>d</sup>
Z-27 (MHOM/BR/66/M2903)	B6(1)	RJ, Angra dos Reis	CL
	B6 (1)	RJ, Itacoara	ML
	B6 (1)	RJ, Maricá	ML
	B5 (1), B6 (8), B9 (1)	RJ, Mesquita	CL
	B6 (3)	RJ, Paraty	CL
	B9 (1)	RJ, Quicamã	CL
	B5 (1), B6 (5), B7 (1), B8 (1)	RJ, Rio de Janeiro	CL
	B6 (1)	RJ, São Gonçalo	CL
	B6 (1)	RJ, Teresópolis	CL
	A12 (1), A16 (1)	ES, Afonso Cláudio	CL
	A1 (1)	ES, Água Doce do Norte	CL
	B4 (1)	ES, Aracruz	CL
	A16 (1)	ES, Castelo	CL
	B3 (1)	ES, Fundão	CL
	A9 (1)	ES, Itaguaçu	CL
	A7 (2)	ES, Itarana	CL
	A13 (1), A16 (1)	ES, Muniz Freire	CL
	B2 (4)	ES, Santa Leopoldina	CL
	B10 (2)	ES, Serra	CL
	B1 (5)	ES, Viana	CL, ML
Z-31 (IPAR/BR/80/IM231)	C13 (1)	RO, BR-319Rd, Km866	
Z-32 (ICAR/BR/86/IM2978)	C1 (1)	RO, Cachoeira Samuel	
Z-34 (MHOM/BR/88/IM3476)	C2 (1)	AM, Tapauá River	CL
Z-35 (MHOM/BR/88/IM3483)	C3 (1)	AM, Tapauá River	CL
	C5 (1), C6 (2), C7 (4), C8 (3), C9 (2)	AM, Coari-Urucu River	CL
	C4 (1)	AM, Manaus	ML
Z-45 (MHOM/BR/91/JRS)	A4 (4), A6 (2), A8 (1), A10 (1), A11 (1), A14 (6), A15 (1)	PE, Amaraji	CL
Z-53 (MAGO/BR/92/IN154)	C15 (1)	RO, Porto Velho	
Z-63 (MHOM/BR/91/IM3708)	C10 (1)	AM, Coari-Urucu River	CL
Z-64 (MHOM/BR/91/IM3711)	C11 (1)	AM, Coari-Urucu River	CL
Z-65 (MHOM/BR/91/IM3723)	C12 (1)	AM, Coari-Urucu River	CL
Z-69 (MDID/BR/95/IM4159)	C14 (1)	AM, Barcelos	
Z-72 (MHOM/BR/98/AFS)	A4 (1), A6 (3), A11 (1), A14 (1)	PE, Amaraji	CL
Z-73 (MHOM/BR/98/ASB)	A4 (3), A13 (2)	PE, Amaraji	CL, ML
Z-74 (MHOM/BR/98/EJS)	A3 (1), A5 (1)	PE, Amaraji	CL
Z-75 (MHOM/BR/98/FJS)	A2 (1)	PE, Amaraji	CL

<sup>a</sup> The zymodeme number of the Instituto Oswaldo Cruz (classification was established by numerical zymotaxonomic analysis) is given. The reference strain designation code is given in parentheses as follow host (I for Insecta [CAR, *Psychodopygus carrerai*; PAR, *Psychodopygus paraensis*]; M for Mammalia [AGO, *Agouti paca*; DID, *Didelphis marsupialis*; HOM, *Homo sapiens*])/country of origin/year of isolation/original code.

<sup>b</sup> Denotes the identity between isolates by using all polymorphic rDNA results.

<sup>c</sup> Codes for states in Brazil (AM, Amazonas; ES, Espírito Santo; PE, Pernambuco; RJ, Rio de Janeiro; RO, Rondônia) are given.

<sup>d</sup> Leishmaniasis lesions in humans (CL, cutaneous; ML, mucosal).

*ziliensis* derived from different endemic areas. MLEE are rapidly evolving markers and have been the most universally accepted means of characterizing and differentiating the many diverse types of leishmanial parasite (5, 30). The electrophoretic mobilities and banding patterns of several different enzymes are usually checked and compared to produce enzyme profiles that designate zymodemes. The relationships among the zymodemes are often verified by numerical taxonomic methods (5, 22, 30, 36). Recently, the ITS of the rRNA genes were reported to be useful markers to type *Leishmania* parasites (6, 33, 34). These regions are flanking the ribosomal genes, which are found in all eukaryotic organisms, and are clustered in tandem arrays on different chromosomes. The ITS regions of the rRNA genes evolve more rapidly than the functional domains that they flank, and phylogenetic studies of several organisms have demonstrated that they are useful to infer about the relationship among closely related populations of organisms (17).

The observed genetic variability and the relationships among the isolates were the basis for investigating whether the genetic differences in *L. (V.) braziliensis* reflect distinct ecoepidemiological features of the infection since these strains are endemic in the distinct areas studied, thus bringing to light fundamental information for future control programs of the disease.

#### MATERIALS AND METHODS

**Leishmanial strains.** The parasites used in the present study and the sources of original stocks are listed in Table 1. We analyzed *Leishmania (V.) braziliensis* isolates from four geographic localities in Brazil that are regions where cutaneous leishmaniasis is endemic: Rio de Janeiro, Espírito Santo, and Pernambuco, as well as in the Amazonian region. *Leishmania (V.) braziliensis* reference strains and isolates used in the present study are maintained at DIFIOCRUZ *Leishmania* Type Culture Collection (registration no. 731 [WFCC World Data Center on Microorganisms Directory]).

**MLEE.** The methods used to prepare samples and study the electrophoretic mobility of some enzymes in agarose gels were performed as described elsewhere (5, 19). Allelic variation for the following enzymes was assessed: aconitate hy-

dratase (EC 4.2.1.3), glucose-6-phosphate dehydrogenase (EC 1.1.1.49), glucose phosphate isomerase (EC 5.3.1.9), isocitrate dehydrogenase NAD and NADP (EC 1.1.1.42), malate dehydrogenase (EC 1.1.1.37), malic enzyme (EC 1.1.1.40), mannose phosphate isomerase (EC 5.3.1.8), nucleosidase (EC 3.2.2.1), 6-phosphogluconate dehydrogenase (EC 1.1.1.43), phosphoglucomutase (EC 1.4.1.9), Leu-Pro dipeptidase (EC 3.4.13.9), and Leu-Gly dipeptidase (EC 3.4.11.1). The bands produced on the gels were numbered according to enzymatic mobility, and the enzyme profiles were subjected to numerical analysis and used to group the samples in zymodemes. Leishmanial strains with the same electromorphs were classified into the same zymodeme (5). One strain from each zymodeme was chosen as the reference for that zymodeme (Table 1).

**Intergenic region typing (IRT).** Genomic RFLP analysis of the ITS between the small and large subunits of the ribosomal DNA (rDNA) locus (PCR-RFLP ITSrDNA) of leishmanial parasites used in the present study (Table 1) was done, as described elsewhere (6). The ITS were PCR amplified and digested with restriction enzymes (*AluI*, *BstUI*, *EcoRI*, *FspI*, *HaeIII*, *HhaI*, *RsaI*, and *TaqI*). Digestion products were separated by high-resolution electrophoresis in a 12% polyacrylamide gel by using the Genephor apparatus (Amersham Pharmacia Biotech). The fragment sizes were estimated relative to molecular weight markers and the banding patterns were used to group the isolates in genotypes (each genotype grouped isolates with the same banding pattern for all of the restriction enzyme used) and were also subjected to numerical analysis.

**Numerical analysis of MLEE or PCR-RFLP ITSrDNA data.** For this numerical analysis, a matrix with the presence or absence of the bands was built and analyzed by using the NTSYS software program (version 2.1; Exeter Software). The similarity matrices constructed by using the simple matching coefficient ( $S_{sm} = m/n$ , where  $m$  is the number of matches and  $n$  is the total number of characters) were transformed into phenograms by the UPGMA (unweighted pair-group method with arithmetic averages) algorithm (35). The confidence of the groups was tested applying a bootstrap analysis with 1,000 replicates using Free-Tree software (version 0.9.1.50). The average and standard deviation of the similarities observed among individuals from each area (considering the clusters observed by the phenetic analysis) where the infection is endemic were calculated as a parameter of genetic diversity.

## RESULTS

**MLEE.** Fourteen enzyme loci of 101 stocks of *L. (V.) braziliensis* from different hosts and Brazilian geographic areas were analyzed, and the electromorphic profiles were compared to reference strains revealing 15 zymodemes (Table 1). The most common zymodeme (Z-27) grouped 49 isolates from Espírito Santo and Rio de Janeiro States. In contrast, 31 strains collected in a rural area in Pernambuco State were classified into five zymodemes. Furthermore, most of the 22 stocks from the Amazon and Rondônia States were individually assigned to separate zymodemes. However, the analysis of allelic variation occurring in stocks compared to the reference strain MHOM/BR/1966/M2903 of *L. (V.) braziliensis* revealed that some zymodemes differ from each other in only one enzymatic locus position (data not shown).

Affinities between zymodemes were calculated by using the simple matching similarity coefficient and were transformed into a phenogram by using mean distances between groups (Fig. 1A). The 15 *L. (V.) braziliensis* zymodemes could be clustered into two major groups, with high level of statistic support as demonstrated by the bootstrap value obtained for each group (Fig. 1A). Each group clustered strains collected on either the Atlantic coast or the Amazon region. One group (comprising isolates from CL or ML patients) was less diverse compared to the other group (comprising isolates from humans and various wild animals and sylvan sandflies). Furthermore, the later group contained two parasites (Z-53 and Z-69) that clustered independently of other zymodemes. These two zymodemes were assigned to the second group based on their geographic origin and because they presented a higher level of

similarity with the zymodemes of this group than to the ones of the other group.

**Genotyping.** The IRT allowed the examination of polymorphisms among *L. (V.) braziliensis* zymodemes (Fig. 2). All leishmanial isolates indicated in Table 1 were analyzed. It was possible to characterize 41 genotypes, each representing parasites with unique ITS fragment profiles. By using numerical analysis, the strains were studied for their genetic relatedness (Fig. 1B). Three statistically supported groups were observed. Genotypic groups A, B, and C comprised 39, 40, and 22 isolates, which were divided into 16, 10, and 15 related genotypes, respectively. It was observed that zymodemes into which more than one isolate was assigned are represented by more than one genotype: Z-27 included 49 samples and 16 genotypes, Z-35 included 14 samples and 7 genotypes, Z-45 included 16 samples and 7 genotypes, and Z-72 included 6 samples and 4 genotypes (Table 1). Except for Z-27, all of the other zymodemes representing large clusters included isolates from the same geographic locality.

Isolates characterized as the same zymodeme (Z-27) were clustered into two independent clusters (A and B). Cluster A comprised isolates from rural localities in Pernambuco and Espírito Santo. Cluster B accounted for strains related with domestic animals and peridomestic sandflies that were assigned to the same zymodeme (Z-27). Group C included only isolates from the Amazonian region. This group was more diverse, presenting some genotypes (C12, C13, C14, and C15) that clustered independently from the others, but with more affinity for the genotypes of group C than for the ones of group A or B. A clonal population (genotype B6) classified in this group contained 21 isolates from different geographic locations in the Espírito Santo and Rio de Janeiro states. In contrast, eight isolates characterized as genotype A4 were assigned to separate zymodemes. Furthermore, parasites showing a high degree of genetic similarity were found to cause either CL or ML in humans (Table 1).

**Geographic distribution and genetic diversity of the genotypes.** A geographic structuring was observed for some genotypes but not for others that was related to genotypes from more distant areas where the organisms were endemic (Fig. 1 and 3). The genetic similarity average based on the MLEE and IRT data was calculated for each locality, and it was assumed as a parameter to evaluate the level of genetic diversity. We observed that the isolates from the Amazonian region presented the highest level of genetic diversity and that the isolates from Rio de Janeiro were more homogeneous (Fig. 3). The isolates from Espírito Santo were considered as being involved in two distinct transmission cycles. The Espírito Santo isolates from urban areas grouped together with the isolates from Rio de Janeiro (group B) and those from rural areas clustered with isolates from Pernambuco (group A) (Fig. 3).

## DISCUSSION

Advances in molecular technology are facilitating the study of the ecology of *Leishmania* clonal populations by providing information on (i) sources of infection, (ii) transmission patterns, (iii) response to treatment, and (iv) the importance of immunity in preventing reinfection (27). In the New World, the *Leishmania (Viannia)* parasites (7, 8), the main causal agents

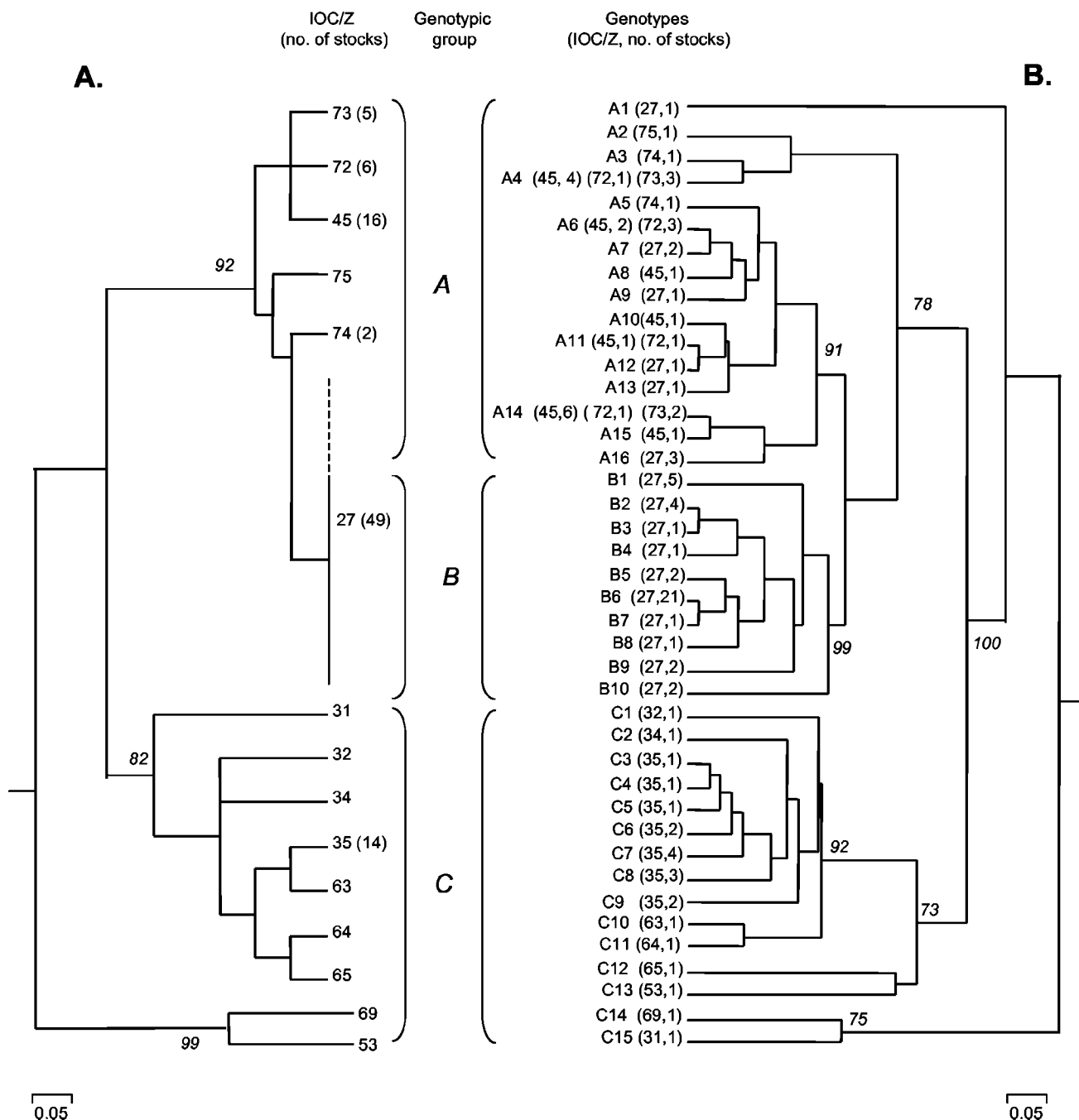


FIG. 1. (A) MLEE phenogram showing the similarity between groups of *L. (V.) braziliensis* zymodemes. Unless indicated otherwise by the number in parentheses, IOC/Z contains a unique isolate. (B) Molecular tree showing the clustering of *L. (V.) braziliensis* genotypes classified in this work (PCR-RFLP ITSrDNA). The bootstrap values (italic numbers) are shown for the principal clusters. Details about strains are given in Table 1. The three main clusters represent parasites collected from different hosts and geographic areas in Brazil (i.e., the Atlantic coast [A and B] or the Amazon region [C]). Isolates characterized as IOC/Z-27 (outlined with continuous and dashed lines) presented genotypes in either group A and B. A, Strains collected in rural localities (Espírito Santo and Pernambuco) where different sandfly and animal species are involved in the transmission cycle; B, strains from urban areas (Espírito Santo and Rio de Janeiro) where transmission is related with a cycle involving domestic animals and preidomestic sandfly species; and C, strains from the Amazon region (in this case, the parasites are maintained in an enzootic cycle involving various wild animals and sylvan sand flies).

of American CL, represent a biologically diverse group of microorganisms showing considerable intraspecies variability (5, 6, 12) that complicates taxonomic classification and epidemiological studies.

The present systematic molecular study revealed *L. (V.) braziliensis* strains to be an extremely diverse population. The multilocus data on a diverse collection of strains from different hosts and ecologic regions in Brazil indicate a clonal popula-



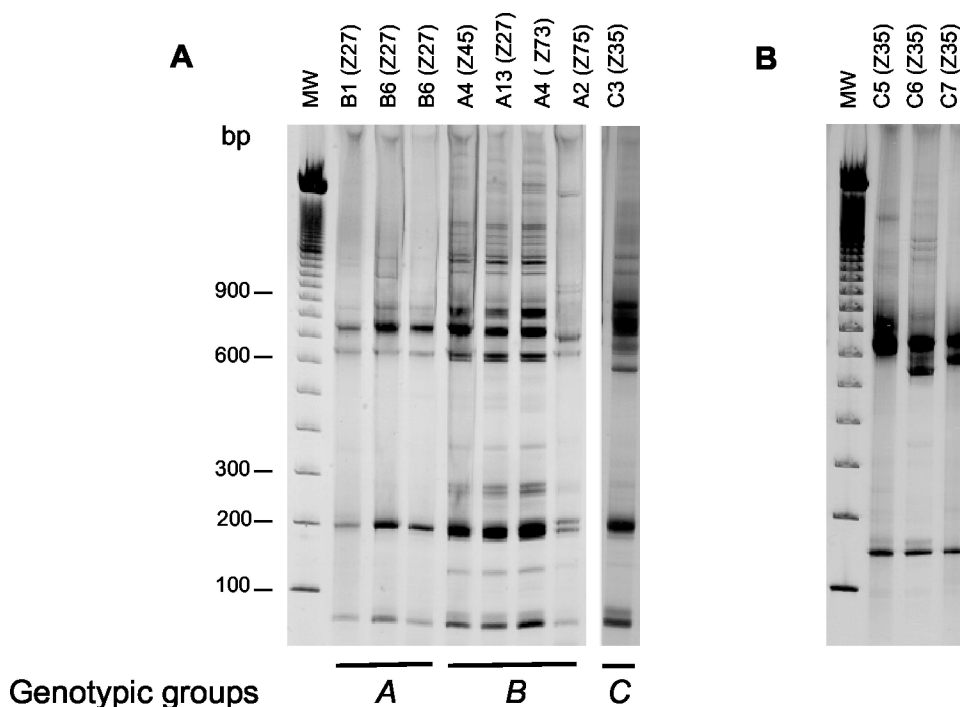


FIG. 2. Acrylamide (12%) gel electrophoresis comparison of ITS fragment patterns generated with the restriction enzymes *Bst*UI (A) and *Hha*I (B), among selected strains of *L. (V.) braziliensis* from Brazil. The codes in each lane represent the genotypes and the respective zymodemes (IOC/Z). The genotypic groups correspond to those described in Fig. 1. (A) Differences among groups A, B, and C; (B) differences among isolates from the same zymodeme.

tion structure, with some zymodemes (e.g., IOC/Z-27) widely distributed and others seemingly unique and localized to a particular endemic focus (e.g., IOC/Z-35 and IOC/Z-45). The IRT analysis resulted in a very high degree of discrimination among isolates from diverse regions, confirming the ability of this technique to discriminate between closely related parasites (6). The three distinct genotypic groups described herein may represent important genetic variation within *L. (V.) braziliensis*, which could explain the plasticity of these parasites and of their ability to adapt to changing ecological conditions. Changes in the ITSrDNA locus, which is a noncoding region, should not reflect the ability to adapt to ecological changes. However, because of the high level of polymorphism in the composition of the ITSrDNA locus, several molecular epidemiological studies have been conducted by using this tool. Analysis of the ITS regions of the nuclear rRNA operon for different parasites has demonstrated the utility of this marker in detect genetic diversity and the association of this with several epidemiological aspects (9, 26)

The present study also indicates that most of the genotypes are specific to some geographic areas (e.g., genotype B1 contains isolates from one geographic location, in Espírito Santo, that were collected over a distinct period of time [between 1982 and 1995]), showing the importance of clonal dissemination versus sexual reproduction for this organism in nature, at least occurring in some regions. The stability of this genotype adapting to some environmental modifications through years in the same area is a clear proof of the aforementioned statement.

*L. (V.) braziliensis* can infect vertebrate hosts of different

species and orders (21) and has been associated with a number of different sandfly species (15). Our analyses indicated that the molecular diversity found in parasites from the Amazon Basin is apparently related to the great number of sandfly vector(s) and/or animal reservoir(s) involved in the transmission cycles (13, 14). In contrast, the *L. (V.) braziliensis* populations circulating in the Brazilian Atlantic coast (in the present study, representing 81.2% of the isolates collected from human beings and dogs in old established communities in nonforested areas) showed a lower level of heterogeneity than did the Amazonian strains and have peridomestic sandfly species such as *Lutzomyia intermedia* and *Lutzomyia whitmani* as the principal suspected vectors (1, 3, 10). The presence of the same genotypes in humans and dogs in the same area suggests that specific transmission cycles could be defined by using the proposed methodologies.

A total of 49% of the parasites collected in areas (Rio de Janeiro or Espírito Santo) where sylvan animals are probably not involved in the transmission cycle were clustered together into the same zymodeme (IOC/Z-27) of the *L. (V.) braziliensis* reference strain (MHOM/BR/1966/M2903), which was isolated from a patient with American CL in the Serra dos Carajás, Pará State. In this case, a sylvan sandfly, *Psychodopygus wellcomei*, has been determined to be the vector (20, 31). In contrast, isolates from Pernambuco, where grass mice (*Bolomys lasiurus*) and black rats (*Rattus rattus*) have been found to be parasite reservoirs (S. P. Brandão-Filho, M. E. F. Brito, F. G. Carvalho, E. A. Ishikawa, E. Cupolillo, and J. J. Shaw, Abstr. Wordleish 2000, abstr. 100, 2000), were grouped into five distinct zymodemes. Factors that predispose pathogens to

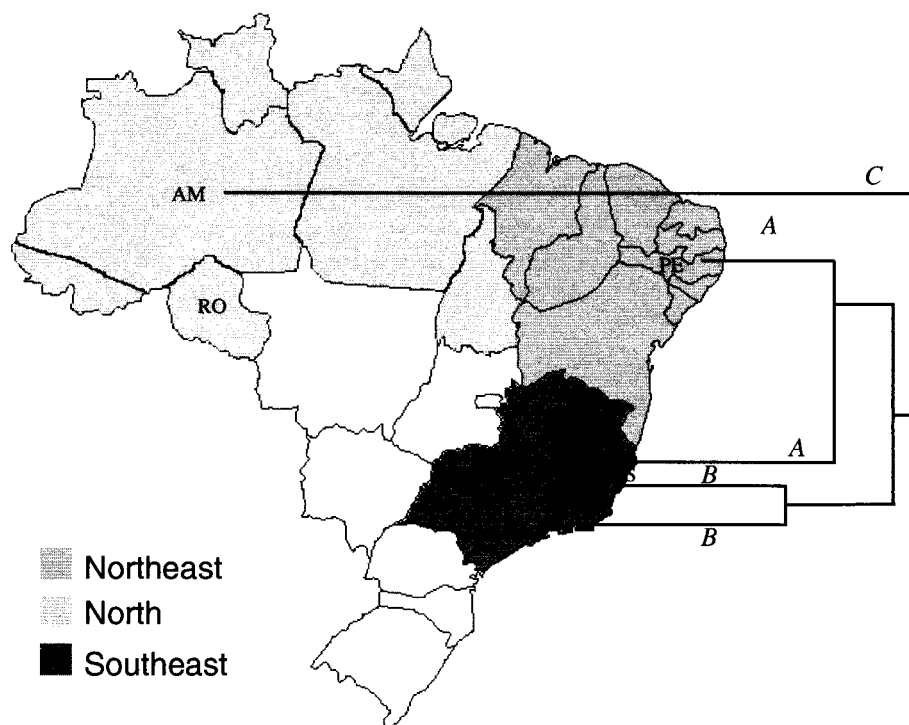


FIG. 3. Map of Brazil showing the areas from which the *L. (V.) braziliensis* isolates originated. A schematic and summarized dendrogram showing the relationship among the genotypic groups (A, B, and C as defined in Fig. 1) was superimposed. The level of genetic diversity found among the isolates from each area where the parasites were endemic was estimated by using the average of similarities as follows (averages  $\pm$  the standard deviations of MLEE and IRT, respectively): Amazonia ( $0.75 \pm 0.082$  and  $0.76 \pm 0.095$ ), Pernambuco ( $0.92 \pm 0.01$  and  $0.83 \pm 0.082$ ), Espírito Santo A ( $1.0$  and  $0.89 \pm 0.05$ ), Espírito Santo B ( $1.0$  and  $0.83 \pm 0.11$ ), and Rio de Janeiro ( $1.0$  and  $0.93 \pm 0.035$ ). The Brazilian states where the isolates were collected included Amazonia, Rondônia, Pernambuco, Espírito Santo, and Rio de Janeiro (see Table 1).

infect multiple hosts include high levels of genetic diversity and opportunities for cross-species transmission (37). The extent of host adaptation is therefore linked to and limited by the genetic variability. Pathogens that contain more accumulating mutations should produce more genetic variants and are more likely to be generalists. An alternative explanation to the extent of host adaptation of some organisms is to consider that the infection of a given host is only related to opportunity; therefore, parasites that infect several different hosts are more likely to be isolated in different transmission cycles.

The RFLP-ITSrDNA data show that parasites from urban areas (Espírito Santo and Rio de Janeiro), where transmission is associated with *Lutzomyia intermedia*, cluster together. Another cluster comprises parasites from rural localities in Espírito Santo and Pernambuco, where *Lutzomyia whitmani* is the principal vector species. Other studies using different molecular markers have demonstrated a geographic structuring of *Leishmania* parasites (2, 16, 23, 33, 34), and populations of *L. (V.) braziliensis* isolates collected from geographically closed areas seem to be more genetically similar (12, 24). However, the present study indicates that the geographic structuring of *L. (V.) braziliensis* reflects the plasticity of these parasites to adapt to different vector species (or populations of this vectors) involved in the transmission cycle. Furthermore, our results indicate an association between the diversity of vectors involved in the transmission of *L. (V.) braziliensis* and the genetic diversity of the parasites. Evaluation of the phospho-

glycan composition of these leishmanial populations is fundamental to an understanding of the vector-parasite interaction (32). Another important approach is to study the population structure of the sandfly vectors in areas where the genetic diversity of *L. (V.) braziliensis* has been determined.

It was recently suggested that the dispersion of *Leishmania* clones is associated with the vector and reservoir movements (18). However, what is important to note is the plasticity of leishmanial parasites, their ability to adapt to changes in ecologic conditions, and a possible evolutionary relationship between them and their vectors as observed in the Old World (28).

The clinical expression of leishmanial infection is dependent on a number of different factors (15). The identification of virulence factors that contribute to the pathogenesis of *L. (V.) braziliensis* infections are poorly understood, mainly because of the lack of good in vivo models. A better understanding of the genetic basis of pathogenesis in ML is needed, as are studies on the role of host factors in immune susceptibility to this form of the disease. Our genotyping studies did not identify a pathogenic clone specifically associated with the development of ML. However, in other analyses we were able to detect distinct genotypic types of a single *L. (V.) braziliensis* isolate from a patient with ML (unpublished data). Competitive interactions among coinfecting clones could affect host immune responses and play a role in determining the immunopathology of the progressive and destructive form of ML. Some underlying

causes of its hyperergic response might include the following factors: (i) the resistance of some parasite clones to elimination (27) and the persistence of "allergic" antigen that evokes hyperergic hypersensitivity inflammatory responses and (ii) autoimmune phenomena related to antigens cross-reactive between leishmanial parasites and host tissues (11, 29).

In conclusion, the combination of MLEE and IRT-ITSrDNA is useful for studies of the interface of population genetic and epidemiology. The application of these molecular techniques could allow us to better determine the genetic diversity of various *Leishmania* parasites circulating in nature. This information is expected to yield new approaches to mitigate parasite transmission and virulence in humans and eventually to improve parasite control.

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