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Renal fibrosis: not just PAI-1 in the sky

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A delicate balance exists between ECM synthesis and degradation such that interruption of the corresponding pathways results in increased plasminogen activator inhibitor-1 (PAI-1), pathological matrix accumulation, and glomerulosclerosis. A new study (see the related article beginning on page 379) demonstrates that therapy with a mutant PAI-1 increases matrix turnover and reduces glomerulosclerosis by competing with endogenous PAI-1, suggesting therapeutic utility in the treatment of fibrotic renal disease.

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Progressive fibrosis leads to end-organ failure in multiple organs, including heart, lung, liver, and kidney. The process of substitution of functional parenchymal cells by fibrotic tissue has been intensively investigated. Past progress over the last few decades has resulted in remarkable gains in slowing the progression of chronic kidney disease. The major emphasis has been on control of blood pressure, both systemi-

cally and within the glomerulus. The efficacy of angiotensin I-converting enzyme inhibitors was linked to control of glomerular pressure beyond systemic pressure effects mediated by dilation of the efferent arterioles of the glomerulus (1). However, normalization of glomerular pressure did not completely halt progression, nor could it regress existing fibrosis. Numerous additional factors thus were implicated in progressive renal injury, including modulation of ECM and parenchymal and infiltrating cell interactions. In addition to angiotensin, TGF- β has been recognized as a key mediator of renal fibrogenesis. Interestingly, both angiotensin and TGF- β induce plasminogen activator inhibitor-1 (PAI-1) (2).

PAI-1 and fibrosis mechanisms

How might PAI-1 affect fibrosis? PAI-1 is the major inhibitor of tissue-type plasminogen activator (t-PA) and

urokinase-type plasminogen activator (u-PA), which activate plasminogen to yield plasmin, which in turn degrades fibrin. PAI-1 not only inhibits fibrinolysis but also has complex interactions with matrix, promoting net proteolysis (3). t-PA, u-PA, and plasmin can degrade a wide range of ECM proteins; they also activate latent MMPs, in particular MMP-1 and MMP-3, and indirectly activate MMP-2. Models of glomerular sclerosis and/or interstitial fibrosis have been developed in both rats and mice to further examine these mechanisms. Noble and colleagues previously demonstrated that direct manipulation of the plasmin/plasminogen activator system with recombinant t-PA treatment decreased glomerular matrix accumulation in the anti-Thy1 model of glomerular matrix expansion (4). This model does not result in progressive renal damage and shows no attendant interstitial fibrosis but allows in vivo determination of mechanisms of glomerular matrix accumulation. Interstitial fibrosis can be induced in vivo in either rats or mice by unilateral ureteral obstruction (UUO). In contrast to the anti-Thy1 model, UUO does not result in significant glomerular lesions but demonstrates early macrophage infiltration and robust interstitial fibrosis, along with tubular injury. The unobstructed contralateral kidney serves as an ideal control. The UUO model thus allows in-depth in vivo study of mechanisms of interstitial fibrosis, and interactions between epithelial and infiltrating cells. The use of diverse models of renal scarring thus allows

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Nonstandard abbreviations used:

plasminogen activator inhibitor-1 (PAI-1); tissue-type plasminogen activator (t-PA); urokinase-type plasminogen activator (u-PA); unilateral ureteral obstruction (UUO); angiotensin type 1 (AT1); epithelial-mesenchymal transition (EMT).

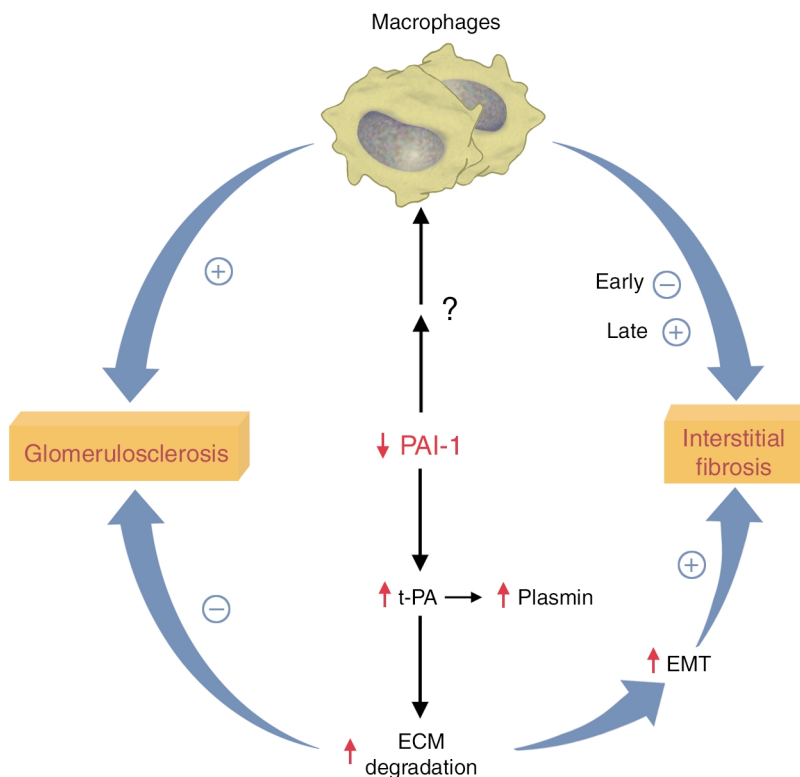


Figure 1

PAI-1 modulates scarring in a complex manner, including effects on cell migration, matrix turnover, and macrophage infiltration. Whereas increased t-PA promotes dissolution of matrix and subsequently lessens glomerulosclerosis, this matrix breakdown facilitates cell migration and epithelial-mesenchymal transition (EMT) and thus contributes to increased interstitial fibrosis. The effects of PAI-1 that facilitate enhanced macrophage infiltration *in vivo* are complex and have not yet been established. Macrophages have beneficial effects early in interstitial fibrosis and promote fibrosis later. The increased number of glomerular macrophages is linked to glomerulosclerosis. u-PA and its receptor and vitronectin also interact with PAI-1 and contribute to its final effects on fibrosis.

dissection of the divergent mechanisms that contribute to glomerular versus interstitial scarring (Figure 1). Thus, in contrast to the effects of t-PA on glomerular matrix accumulation, t-PA^{-/-} mice have *reduced* interstitial fibrosis after injury caused by UOU (see below) (5).

In normal, non-scarred kidneys, there is very little PAI-1 expression, but acute infusion of angiotensin markedly upregulates PAI-1 mRNA and protein via the angiotensin type 1 (AT1) receptor by a non-pressure-dependent mechanism (6). This induction has been shown *in vitro* to be augmented by aldosterone via a putative glucocorticoid-response element in the PAI-1 promoter (7). In fibrotic renal diseases, PAI-1 is also increased and localizes to areas of glomerulosclerosis (8). Conversely,

inhibition of angiotensin or aldosterone decreases PAI-1 and also decreases renal scarring (8, 9). Decreased glomerular PAI-1 may thus lessen severity of glomerulosclerosis by allowing increased proteolysis. PAI-1 also affects interstitial fibrosis. PAI-1^{-/-} mice had attenuated interstitial fibrosis after either UOU or protein overload compared with wild-type but were not completely protected from injury (10). In this issue of the *JCI*, the novel and exciting studies from Noble and colleagues show that inhibition of PAI-1 by a mutant PAI-1, which binds matrix vitronectin but does not inhibit plasminogen activators, results in significant, albeit incomplete, protection against glomerular matrix accumulation (11). Mechanisms demonstrated by the authors include increased

glomerular plasmin with direct degradation of accumulated excess matrix.

However, the results of Noble and colleagues (11) suggest that PAI-1 also has more far-reaching effects, including indirect effects on matrix synthesis. Thus, mRNA expression for matrix molecules was also decreased, perhaps because of the reduced macrophage infiltration observed with PAI-1 inhibition. The mechanisms for decreased macrophage infiltration with PAI-1 inhibition remain to be determined (Figure 1). The macrophage has a particularly critical role after injury. Macrophages produce numerous profibrotic factors, which increase matrix synthesis by resident parenchymal cells. However, in the absence of TGF- β signaling, macrophages may be ineffectual in promoting fibrosis. This has been illustrated experimentally in β_6 integrin knockout mice. The heterodimeric integrin $\alpha_5\beta_6$, expressed in epithelia in the skin, lung, and kidney, is one key local activator of TGF- β . TGF- β circulates in an inactive form, linked to latency-associated peptide (LAP). β_6 integrin binds to this TGF- β -LAP inactive complex, effecting local TGF- β activation. β_6 ^{-/-} mice were resistant to lung fibrosis induced by bleomycin, despite robust numbers of infiltrating macrophages (12). We have shown similar protection in β_6 ^{-/-} mice in the renal fibrosis model of UOU (13). Thus, macrophages may not be profibrotic in the absence of local TGF- β activation.

Macrophages and fibrosis

Of note, glomerular and interstitial macrophage infiltration may not activate identical injury responses. This possibility is supported by our observation of differing phenotype of glomerular versus interstitial macrophages in human diabetic nephropathy; only the latter expressed PPAR- γ (14). It is also possible that macrophage infiltration, especially interstitial, has an early, beneficial role after injury, whereas persistent macrophage infiltration, whether glomerular or interstitial, is profibrotic. Recently, we have further demonstrated that infiltrating macrophages

even play a beneficial role in early injury after UUO, and this salutary effect is dependent on intact AT1 receptor (15). Wild-type mice were radiated and underwent bone marrow reconstitution with *AT1a*^{-/-} marrow. Surprisingly, these mice had more severe fibrosis after UUO than identically treated mice reconstituted with wild-type marrow. These beneficial effects of macrophages are similar to the well-established role of macrophages in early wound healing, in which phagocytosis is a key element in resolution of injury. In Noble and colleagues' study (11), macrophage infiltrate was decreased, which could be causal in decreasing glomerular injury. Whether PAI-1 antibody also decreases interstitial macrophages and thereby contributes to decreased interstitial fibrosis remains to be determined. Even those remaining glomerular macrophages could be inhibited in transducing fibrotic effects, because TGF- β content of glomeruli was also reduced by the mutant PAI-1, thus reducing downstream fibrosis-effector mechanisms. This decrease in TGF- β possibly could result from increased clearance of TGF- β by PAI-1 mutant protein binding to vitronectin.

PAI-1 and cell migration

PAI-1 also importantly modulates cellular adhesion and migration and thus has impact on inflammation, wound healing, angiogenesis and cell migration, and tumor cell metastasis (3). PAI-1 competes with the u-PA receptor (u-PAR) for the NH₂ terminus-binding domain on vitronectin. When PAI-1 binds to vitronectin, u-PAR interaction with the vitronectin and surrounding ECM is prevented, thus inhibiting vitronectin-dependent cellular adhesion and migration. This effect is demonstrated in PAI-1^{-/-} mice, which show enhanced smooth muscle cell migration, while PAI-1 overexpression in vitro inhibits this process (16, 17). However, in vivo PAI-1 enhances cell migration in some settings. Increased PAI-1 is linked to macrophage infiltration, and more aggressive tumor cell metastasis, pointing to complex

interactions with vitronectin and matrix in vivo, and possible direct chemotactic actions of PAI-1 (Figure 1) (3). The mutant PAI-1 used by Noble and colleagues (11) maintains interactions with vitronectin and thus can have cell migration-inhibitory effects independent of its proteolytic activity. This might not only decrease macrophage infiltration but also affect epithelial-mesenchymal transition (EMT). EMT is characterized by migration of epithelial cells into the interstitium, where they become mesenchymal, myofibroblast-type cells. Neilson and colleagues have recently elegantly demonstrated that this process contributes to renal fibrosis in vivo after UUO (18). The importance of EMT in interstitial fibrosis was also supported by a decrease in interstitial collagen after UUO in t-PA^{-/-} mice. This protection was linked to decreased MMP-9 induction and preserved tubular basement membrane integrity, postulated to prevent EMT. These findings contrast with the effects of increased t-PA to protect against glomerular matrix accumulation, discussed above. The mutant PAI-1 antibody used by Noble and colleagues could possibly inhibit this migration by its persistent interaction with vitronectin. However, effects on interstitial fibrosis cannot be extrapolated from their studies in the anti-Thy1 model, which has only mesangial proliferation and matrix expansion and no fibrosis.

In conclusion, the studies of Noble and her colleagues (11) show us that modulating fibrosis is a goal within reach, and inhibition of these mechanisms could confer further benefits beyond existing therapies in treatment of chronic kidney diseases. It is important to note that disease was not completely prevented, indicating that additional targets might yet remain for optimal intervention in fibrosis. Further understanding of the complex interactions of parenchymal cells, infiltrating cells, and matrix will allow optimal targeting and development of novel antifibrotic therapies. Synergistic therapies could halt progression of

lesions and even potentially regress existing scarring (19).

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