PHARMACOLOGY OF THE ADRENOCEPTORS AND CHOLINOCEPTORS OF THE BC₂H1 NONFUSING MUSCLE CELL LINE

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1 We have studied the action of different transmitters on the transmembrane $42K$ or $86Rb$ efflux from tissue cultures of the $BC₃H1$ muscle cell line.

2 The effect of catecholamines and carbachol (CCh) on the isotope efflux rate was measured by addition of the drugs at different times during the washout.

3 Noradrenaline (NA), phenylephrine (Phe), isoprenaline (Iso) and CCh increased ^{42}K and ^{86}Rb efflux rate in a dose-dependent manner.

4 The action of NA seems to be due exclusively to the stimulation of α -receptors, since its effect was blocked by phentolamine but not by propranolol. The effects of Iso on the ⁸⁶Rb efflux were inhibited by propranolol. The β -receptors in the BC₃H1 cells seem to be of the β_2 -type since they are stimulated by Iso and insensitive to NA.

5 The effect of CCh was blocked by (+)-tubocurarine but not by atropine. This result confirms the presence of nicotinic receptors in $BC₃H1$ cells.

Introduction

Schubert, Harris, Devine & Heinemann (1974), separated several cell lines from a brain tumour induced in mice by the injection of nitrosoethylurea (Ivankovic & Druckrey, 1968). One of these cell lines, called $BC₃H1$, defined as a non-fusing muscle cell line has many characteristics of smooth muscle. In the confluent state the cells are elongated, contain thick and thin filaments and their membranes show numerous vesicles closely resembling the stable structures observed by Gabella (1973) in different smooth muscles. The cells have sometimes been observed to contract spontaneously. The cell membrane is excitable, the iontophoretic application of acetylcholine producing ^a rapid depolarization. On the other hand, noradrenaline (NA) has a hyperpolarizing effect. However, impalement of these cultured cells is difficult, making a pharmacological analysis of the membrane response to chemical mediators and drugs rather cumbersome.

The purpose of the present work was to study the membrane response of the $BC₃H1$ cells by a different approach. We have examined the effects of catecholamines, carbachol and some of their antagonists on 86Rb and ⁴²K effluxes. It has been observed previously that molecules which have a depolarizing or a hyperpolarizing effect increase K^+ permeability either primarily or secondarily (Durbin & Jenkinson,

1961; Burgen & Spero, 1968, Worcel & Hamon, 1976). The results obtained indicate that the $BC₃H1$ cells have α - and β -adrenoceptors. The latter seem to correspond to the β_2 -type usually found in mammalian smooth muscle (Lands, Arnold, McAuliff, Luduena & Brown, 1967). On the other hand, there was also a response, unusual in this type of tissue, to the nicotinic cholinoceptor blocker, $(+)$ -tubocurarine. This drug blocked the effect of carbachol on 86Rb and 42K effluxes confirming the presence of nicotinic receptors in these cells.

Methods

Tissue culture

The cloned muscle cell line was grown in Dulbecco modified Eagle's medium (Vogt & Dulbecco, 1963) containing 10% foetal calf serum at 37° C in an atmosphere of 12% CO₂ and 88% air. The line was maintained in the exponential phase of growth by passage every 4 days. The cells were resuspended in 0.25% Viokase in Dulbecco modified Eagle's medium, centrifuged and resuspended in the growth medium. They were plated in plastic tissue culture dishes at 103 cells/cm2. The cells used for efflux studies were grown on 10.5×50 mm glass slides in 90 mm diameter

Figure ¹ Morphological characteristics of the BC₃H1 cells used in this study. (a) Exponential
phase; (b) culture 1 week after reaching confluency. The bar represents 50 μ m.

Figure 2 Schematic representation of the glass apparatus used for the efflux experiments. A slide with the cultured cells loaded with $42K$ or $86Rb$ was placed in the chamber (2 ml volume). The chamber was perfused with either ^a physiological saline solution (PSS) alone (stop-cock in position A) or with the PSS containing the drug tested (stop-cock in position B). The solutions could be changed without any change in the flow, which was maintained constant at 7 mI/min by a roller pump. The effluent was collected at fixed times in test tubes for counting. The chamber and the incoming solutions were maintained at 37'C by circulating water.

dishes. The medium was changed every 2 days. The cells were usually used ¹ week after reaching confluence, except in those cases indicated in the results section (Figure 1).

Measurement of $42K$ and $86Rb$ fluxes

The slide with the cells attached to one side was removed from the dish, rinsed once and incubated for at least 120min in a physiological saline solution (PSS) with the following composition (mM): Na+ 136.9, K⁺ 5.9, Ca²⁺ 2.5, Mg²⁺ 1.2, HCO₃ 15.5, $H_2PO_4^-$ 1.2, Cl^- 133.5 and glucose 11.5. The solutions contained 10 to 20 μ Ci/ml of ⁴²K or ⁸⁶Rb. When the latter isotope was used, all K^+ in the loading and perfusing solutions was replaced by $Rb⁺$. During the loading period and the washout, PSS was gassed with a 95% O_2 and 5% CO_2 mixture. After loading, the slide was placed in a chamber maintained at a constant temperature (Figure 2) after rinsing in PSS for not more than 60s in order to reduce the amount of isotope sticking to the surface. The slide was then perfused continuously with the PSS at a constant flow of 7 ml/min with a roller pump. The volume of the chamber being 2 ml, the complete replacement of the medium around the cells was achieved within one minute. A four way stop-cock allowed solutions to be changed without interruption of the perfusion at any time. The entire effluent was collected every minute or every 2 min and counted in a well γ -counter. At the end of the experiment, the radioactivity remaining in the cells was counted. A desaturation curve was calculated by adding in a reverse order the washout curve to the radioactivity remaining in the cells at the end of the experiment (Hamon, Papadimitriou & Worcel, 1976).

Drugs

All salts were reagent grade. $(-)$ -Phenylephrine hydrochloride, $(-)$ -noradrenaline hydrochloride, $(-)$ -isoprenaline bitartrate, (\pm) -propranolol hydrochloride, carbamylcholine chloride and (+)-tubocurarine chloride were purchased from Sigma and phentolamine hydrochloride from Ciba-Geigy; Viokase, Dulbecco modified Eagle's medium and foetal calf serum from Gibco.

Statistical methods

Means and standard error of the mean have been calculated assuming a normal distribution and allowing for the small size of the samples (Schwartz, 1963).

Results

 $42K$ and $86Rb$ turnovers were analysed in terms of

Figure 3 Desaturation curves for $86Rb$ and $42K$ from BC₃H1 cultures in the confluent state. After a fast initial washout, the desaturation of both isotopes resolves in a single exponential. Single experiments are represented for clarity. The difference between the mean efflux rate (see text) is not significant: $r_{42K} = 0.0083 \pm 0.0006$ /min (n = 7) and $r_{48B} = 0.0092 \pm 0.0009$ /min (n = 18). The ordinate scale represents the percentage counts left in the tissue at a given time after washout. The abscissa scale represents the time of washout.

the rate coefficient which is calculated by the expression:

$$
r = \frac{A_1 - A_2}{\frac{A_1 + A_2}{2}(t_2 - t_1)}
$$

where r is the rate coefficient, A_1 and A_2 are two points in the desaturation curve at time t_1 and t_2 . The rate coefficient decreased rapidly and continuously during the first 5 min, as a consequence of the washout of the isotope adsorbed on to the surface of cells and slides and to diffusion from the extracellular space. After 5 min, the rate became constant and probably represented the membrane-limited efflux from the cells (Worcel & Hamon, 1976).

Preliminary experiments were peformed with ⁴²K as an isotope, but the determination of the efflux curves was complicated by the fact that as a consequence of the short half-life of $42K$, the radioactivity in the washout and slides was very low, especially after prolonged experiments. As a result $42K$ was replaced by ⁸⁶Rb, since the longer half-life of ⁸⁶Rb makes its handling easier. We have compared the efflux rates of the BC_3H1 cells for ⁴²K and ⁸⁶Rb (Figure 3). At the end of the exponential growth phase, the mean efflux rate coefficients for both isotopes were similar: $0.0083 + 0.0006/\text{min}$ (n = 7) for ⁴²K and 0.0092 \pm 0.0009/min (n = 18) for ⁸⁶Rb. It has been shown that $Rb⁺$ behaves like $K⁺$ with respect to ionic channels and the $Na⁺-K⁺$ pump. Widdicombe (1977) observed that $Rb⁺$ can inhibit the ouabain-sensitive uptake of K^+ in the guinea-pig taenia coli smooth muscle. The pump has the same affinity for both Rb^+ and K^+ . Similarly, Rb^+ reduces

the K^+ passive uptake indicating that these ions pass through the same channel in the cell membrane. Similar interactions between Rb^+ and K^+ fluxes have been reported in red blood cells (Sachs & Welt, 1967).

Effects of noradrenaline on the efflux rate

The amine produced an increase in the ⁸⁶Rb efflux rate at concentrations higher than 10^{-7} M. In preliminary experiments, we determined the times of exposure necessary to attain the maximal effects of a given concentration and the intervals required to reduce tachyphylaxis to NA to ^a minimum. The doses of NA were administered over ⁶ min and the slides washed for 20 min before being superfused with another dose of the drug. Increasing concentrations of NA applied under these conditions produced ^a dose-dependent enhancement of 86Rb efflux rates (Figure 4). The maximum effects were attained only after at least ³ min exposure to the drug. This delay seemed to be due at least partly, to diffusion delay in the perfusion chamber since it was longer than the time needed to obtain a hyperpolarization following iontophoretic application (Schubert et al., 1974). In all cases, the effects of NA were faster at higher concentrations, the time to peak being shorter at these levels.

Dose-response curves to NA were obtained at the exponential and confluent stages of growth of the cultures. The results obtained were similar, the ED_{50} value for NA during the exponential phase (before confluence) was 3.6×10^{-7} M. One or two weeks later, the ED_{50} was still 3.8×10^{-7} M (Figure 5). These results indicate that there was no variation in

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Figure 4 (a) Effect of noradrenaline (NA) on the 86 Rb efflux rate from BC₃H1 cells in the confluent state. The arrows at the bottom indicate the beginning of the perfusion of the slide with the physiological salt solution (PSS) containing NA. W indicates the return to the PSS alone. Increasing concentrations of NA enhanced the efflux rate coefficient in ^a dose-dependent manner. A plateau was attained with concentrations higher than 2.5 \times 10⁻⁶ M NA. A single experiment is shown for clarity; 6 similar experiments were performed. The average curve is represented in Figure 5. (b) Effect of repeated exposure of the $BC₃H1$ to 10⁻⁶ M NA, a concentration that produces a submaximal increase in the ⁸⁶Rb efflux. At this concentration and when the intervals between administration are longer than 20 min, no desensitization occurred. ^r is the rate coefficient (see text). Time zero indicates the beginning of the washout.

the response to NA during the differentiation of the cells. All the following experiments were performed with cultures one or two weeks after confluence.

Characterization of adrenoceptors

The action of NA was tested in the presence of different concentrations of phentolamine. In the following experiments the antagonists were added to the PSS. At a concentration of 10^{-6} M phentolamine displaced the NA dose-response curves to the right (Figure 6a). The difference in the maximum response should be disregarded given the variability of the results. With phenylephrine (Phe), a specific α -agonist, it was observed that phentolamine 10^{-6} M still produced the same qualitative and quantitative effects as NA (Figure 6b). These findings suggested that both Phe and even NA act exclusively on α -adrenoceptors. The latter possibility was confirmed further by the action of higher doses of phentolamine which at 5×10^{-5} M practically suppressed the effects of NA (Figure 6a). The preferential stimulation of α -adrenoceptors by NA was confirmed by the experiments performed in the presence of propranolol. This antagonist did not affect NA dose-response curves at concentrations of 10^{-6} M and 5×10^{-5} M.

The preceding experiments indicated that the β -adrenoceptors in BC₃H1 cells are not stimulated by NA. The β -agonist isoprenaline increased $86Rb$ efflux in a dose-dependent manner. This effect was blocked by 10^{-6} M propranolol. At this concentration, the antagonist shifted the curves to the right with ^a dose-ratio of ¹⁰ (Figure 7). On the other hand, phentolamine at 10^{-6} M did not modify the response to isoprenaline. This confirmed the presence, in these cells of β -adrenoceptors that were activated by isoprenaline but not by NA. Consequently these receptors would correspond to the β_2 -type as described by Lands et al. (1967), found especially in smooth muscle cells, and activated by the following series of agonists: isoprenaline $>$ adrenaline \ge noradrenaline, as opposed to β_1 -receptors chiefly found in cardiac sites, activated by the following series: isoprena $line >$ adrenaline = noradrenaline.

Effects of carbamylcholine (CCh) on the $86Rb$ efflux rate

lontophoretic application of acetylcholine to the $BC₃H1$ cells results in membrane depolarization (Schubert et al., 1974). This confirms the results of Patrick, McMillan, Wolfson & O'Brien (1977) who have shown that α -bungarotoxin binds specifically to $BC₃H1$ cell membranes. We first tested the action of CCh on the 86Rb efflux in the same clone used for all the previous experiments. Surprisingly the agonist

Figure 5 Log dose-effect curves of noradrenaline (NA) on cells 1 week after reaching confluence $($ or in the logarithmic phase of growth (\triangle) r is the efflux rate coefficient of ⁸⁶Rb. Each point is the mean of at least 4 experiments. Vertical bars represent s.e. means.

did not modify the efflux rates even at concentrations as high as 10^{-4} M. Positive results were obtained only with the $BC₃H1$ clone used by Patrick et al. (1977). CCh increased the efflux rate constant of ⁸⁶Rb from these cells. The response was more rapid than with NA, phenylephrine or isoprenaline, the maximum 86Rb efflux rate being attained within the first 2 min of exposure to the drug. CCh produced a dose-dependent increase in ${}^{\circ}$ Rb efflux with an ED₅₀ of 4×10^{-6} M. This effect was not blocked by 10^{-6} M atropine. The same concentration of $(+)$ -tubocurarine had a non-competitive blocking effect of CCh action (Figure 8). Our finding confirmed previous results indicating that this clone of the $BC₃H1$ cell line possesses nicotinic cholinoceptors.

Discussion

The purpose of this work was to examine the membrane response of the $BC₃H1$ cells to different putative transmitters and some of their analogues and antagonists. To gauge the membrane effects we have

studied the action of these substances on ⁸⁶Rb and 42K efflux rates. Depolarization of the cell membrane activates an electrically dependent K^+ conductance in smooth, striated and cardiac muscles, as well as in nervous tissue (Ginsborg, 1967). Furthermore the hyperpolarization induced by NA in other excitable tissues appears to be at least partly due to a primary increase in the membrane conductance to K^+ as shown by Bulbring & Tomita (1969) in guinea-pig taenia coli. Direct evidence for such an action has been previously obtained by Jenkinson & Morton (1965; 1967) who found that NA doubled the rate of 42K loss from depolarized taenia coli. Consequently, it might be expected that cell depolarization would result in an increase in K^+ or Rb^+ efflux rates and that hyperpolarization would be frequently, but not always, associated with a primary enhancement of these rates. However, our experiments were designed to quantify the membrane effects of agonists and antagonists using 86Rb and 42K effluxes as tools.

The present results show that $BC₃H1$ cells possess a-adrenoceptors that are activated by NA and Phe and inhibited by phentolamine. Isoprenaline stimulated ⁸⁶Rb efflux from these cells, this effect being blocked by propranolol. Conversely, the NA effect on 86Rb efflux was not modified by propranolol. These findings suggest that the β -receptors present in the $BC₃H1$ line are probably of the β_2 -type (Lands *et al.*, 1967) since they are stimulated by isoprenaline and apparently unaffected by NA up to ^a concentration of 10^{-5} M. Schubert, Tarikas & La Corbiere (1976) studied the effects of NA at a concentration of 10^{-4} M in the presence of 10^{-3} M theophylline on the cyclic AMP levels of fusing and non-fusing muscle cell lines. At these high concentrations NA has ^a very moderate effect on $BC₃H1$ cells, much smaller than the stimulatory action observed in skeletal muscle myoblasts and myotubes.

Responses to cholinoceptor agonists were obtained only in the clone of the $BC₃H1$ line used by Patrick, et al. (1977). There is no explanation yet to account for the failure of certain clones of $BC₃H1$ lines to give such responses. 86Rb effluxes were stimulated in a dose-dependent manner by CCh. Since acetylcholine produced a depolarization of these cells, the increase in 86Rb efflux may be related to an increase in membrane permeability to K^+ secondary to the change in membrane potential. The effect of the cholinoceptor agonist on ⁸^oRb efflux was not modified by atropine in concentrations up to 10^{-6} M. On the other hand $(+)$ -tubocurarine produced a clearcut displacement of CCh dose-response curves. Our findings are consistent with the results of Patrick et al. (1977) who showed that these cells bound α -bungarotoxin and that this binding was preferentially displaced by (+)-tubocurarine and only very weakly by atropine, hexamethonium or decamethonium. These results in-

Figure 6 (a) Log dose-effect curves for noradrenaline (NA) on cells in the stationary phase in the absence (\bullet) (4 experiments) or in the presence of 10^{- \bullet} M(O) (3 experiments) or 5×10^{-5} M(\Box) (1 experiment) phentolamine. (b) Log dose-effect curves for phenylephrine on cells in the stationary phase in the absence (\bullet) (4 experiments) or in the presence of 10^{-6} M phentolamine (\circ) (1 experiment). r is the efflux rate coefficient of ⁸⁶Rb. Vertical bars represent s.e. means.

Figure 7 Log dose-effect curves for isoprenaline on cells in the stationary phase in the absence $(①)$ or in the presence of 10^{-6} M (O) or 5×10^{-5} M ([]) propranolol. ^r is the efflux rate coefficient of ⁸⁶Rb. Each point is the mean of 2 experiments. Since propranolol did not have any effect on NA dose-effect curves, this result suggests that these cells have β_2 -receptors (see text).

dicate that the cholinoceptors of the $BC₃H1$ line are nicotinic.

The $BC₃H1$ cell line was originally described as a smooth muscle cell line (Schubert et al., 1974) on the grounds of morphological, physiological and biochemical properties. These cells resemble in their differentiated state other smooth muscle cells in culture

Figure 8 Log dose-effect curve for carbamylcholine on cells in the stationary phase in the absence (\bullet) and presence of 10⁻⁶ M (+)-tubocurarine (\circ). r is the efflux rate coefficient of ⁸⁶Rb. Each point is the mean of 3 experiments. Vertical bars represent s.e. means. The $BC₃H1$ cells have nicotinic cholinoceptors (see text).

as described previously by Chamley, Campbell & Burnstock (1974), Campbell, Chamley & Burnstock (1974) and Mauger, Worcel, Tassin & Courtois (1975). They are elongated, disposed in parallel bundles running in perpendicular directions when in multilayers. Electronmicrographs shown a few caveolae in the cell membrane (Schubert et al., 1974), thick and thin filaments and no sarcomeres or Z lines. However, these excitable non-fusing muscle cells do not behave pharmacologically as typical smooth muscle cells. On one hand α - and β -adrenoceptors have been described in both vascular and visceral smooth muscles. On the other hand, the presence of nicotinic receptors makes them rather atypical since to our knowledge only muscarinic receptors have been observed in mammalian smooth muscles. In any case, the $BC₃H1$ cell

References

- BULBRING, E. & TOMITA, T. (1969). Increase of membrane conductance by adrenaline in the smooth muscle of guinea-pig taenia coli. Proc. Roy. Soc. B., 172, 89-102.
- BURGEN, A.S.V. & SPERO, L. (1968). The action of acetylcholine and other drugs on the efflux of potassium and rubidium from smooth muscle of guinea-pig intestine. Br. J. Pharmac., 34, 99-115.
- CAMPBELL, G.R., CHAMLEY, J.H. & BURNSTOCK, G. (1974). Development of smooth muscle cells in tissue culture. J. Anat., 117, 295-312.
- CHAMLEY, J.H., CAMPBELL, G.R. & BURNSTOCK, G. (1974). Dedifferentiation, redifferentiation and bundle formation of smooth muscle cells in tissue culture: the influence of cell number and nerve fibres. J. Embryol. exp. Morph., 32, 297-323.
- DURBIN, R.P. & JENKINSON, D.H. (1961). The effect of carbachol on the permeability of depolarized smooth muscle to inorganic ions. J. Physiol., 157, 74-89.
- GABELLA, G. (1973). Fine structure of smooth muscles. Phil. Trans. R. Soc. B., 265, 7-16.
- GINSBORG, B.L. (1967). Ion movements in junctional transmission. Pharmac. Rev., 19, 289-316.
- HAMON, G., PAPADIMITRIOU, A. & WORCEL, M. (1976). Ionic fluxes in rat uterine smooth muscle. J. Physiol., 254, 229-243.
- IVANKOVIC, S. & DRUKREY, H. (1968). Transplazentare erzeugung maligner Tumoren des Nerven Systems. I. Athyl-nitroso-Harnstoff (ANH) an BDIX Ratten. Z. Krebsforsh., 71, 320.
- JENKINSON, G.H. & MORTON, I.K.M. (1965). Effects of noradrenaline and isoprenaline on the permeability of depolarized intestinal smooth muscle to inorganic ions. Nature, Lond., 205, 505-506.
- JENKINSON, D.H. & MORTON, I.K.M. (1967). The effect of noradrenaline on the permeability of depolarized intestinal smooth muscle to inorganic ions. J. Physiol., 188, 373-386.

line may be a suitable model to study interactions between adrenoceptors and cholinoceptors in different stages of cell differentiation.

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- LANDS, A.M., ARNOLD, A., McAULIFF, J.P., LUDUENA, F.P. & BROWN, T.G. Jr. (1967). Differentiation of receptor systems activated by sympathomimetic amines. Nature, Lond., 214, 597-598.
- MAUGER, J.P., WORCEL, M., TASSIN, J. & COURTOIS, Y. (1975). Contractility of smooth muscle cells of rabbit aorta in tissue culture. Nature, Lond., 255, 337-338.
- PATRICK, J., McMILLAN, J., WOLFSON, H. & O'BRIEN, J.C. (1977). Acetylcholine receptor metabolism in a non-fusing muscle cell line. J. biol. Chem., 252, 2143-2153.
- SACHS, J.R. & WELT, L.G. (1967). The concentration dependence of active potassium transport in the human red blood cell. J. clin. Invest., 46, 65-76.
- SCHUBERT, D., HARRIS, A.J., DEVINE, C.E. & HEINEMANN, S. (1974). Characterization of a unique muscle cell line. J. cell Biol., 61, 398-413.
- SCHUBERT, D., TARIKAS, H. & LA CORBIERE, M. (1976). Neurotransmitter regulation of adenosine 3',5'-monophosphate in clonal nerve, glia and muscle cell lines. Science, N.Y. 192, 471-472.
- SCHWARTZ, D. (1963). Les petits échantillons. In Méthodes Statistiques à l'Usage des Médecins et Biologistes. pp. 151-162. Paris: Editions Medicales Flammarion.
- VOGT, M. & DULBECCO, R. (1963). Steps in the neoplastic transformation of hamster embryo cells by polyoma virus. Proc. natn. Acad. Sci. U.S.A., 49, 171-182.
- WIDDICOMBE, J.H. (1977). Ouabain-sensitive ion fluxes in the smooth muscle of the guinea-pig's taenia coli. J. Physiol., 266, 235-254.
- WORCEL, M. & HAMON, G. (1976). Changes in ionic fluxes in uterine smooth muscle induced by carbachol. Physiology of Smooth Muscle. ed. Biilbring, E. & Shuba, M.F. pp. 339-345. New York: Raven Press.

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