

results. In 4 of the present cases, rats alternated between immobile crouching and fast running, and another compound caused prolonged gnawing and grooming.

The effort involved averages about 0.25 man-hours/rat, perhaps 8 min actual observation and 7 min in preparation, data analysis and writing a routine report. This may be more than is justified in an acute screen but seems worthwhile at the subacute stage where early warning and tentative no-effect doses are most useful.

This work was done at Imperial Chemical Industries Ltd., Central Toxicology Laboratory, Alderley Park, Cheshire SK10 4TJ.

References

- SILVERMAN, A.P. (1973). An 'Exploration-Thirst' test of chemical effects on behaviour. *Arch. Pharmac.*, **279**, suppl., p. 25.
SILVERMAN, A.P. & WILLIAMS, H. (1975). Behaviour of rats exposed to trichloroethylene vapour. *Br. J. ind. Med.*, **32**, 308-315.

Comparison of the novel bibenzyl bifluranol with diethylstilboestrol: effect of aromatic fluorine substitution on metabolism

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A novel compound, bifluranol, has been synthesized as part of a programme to examine the effects of various substituents on the endocrinological activity of bibenzyl structures related to hexoestrol (Figure 1). The bibenzyl structure was chosen in preference to the stilbene structure of diethylstilboestrol (DES) due to concern over DES toxicity. Orally bifluranol

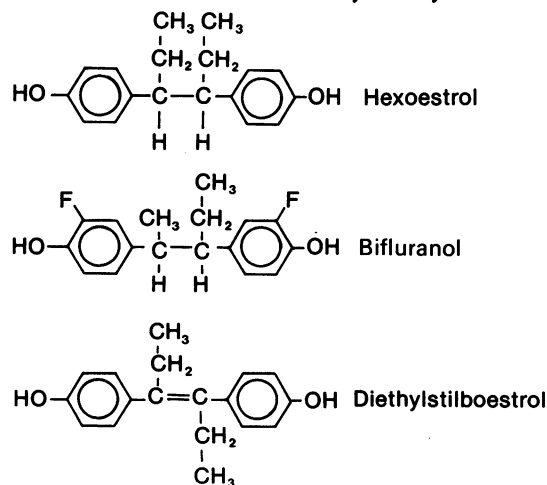


Figure 1 Structures of hexoestrol, bifluranol and diethylstilboestrol.

was shown to be a comparable antiandrogen to DES, but with only one-eighth of the oestrogenic activity, and is currently undergoing clinical evaluation for use in benign prostatic enlargement.

Studies on the fate of [³H]-bifluranol have shown that the disposition in rat is similar to that reported for DES, with the drug being rapidly absorbed, taken up by the liver and eliminated in bile to give predominantly faecal excretion and low urinary excretion. However, the metabolic fate of the two compounds differs significantly. DES has been shown to undergo extensive oxidative metabolism in rat, with up to 30% being converted to a mixture of at least seven products including dienestrol and hydroxy- and methoxy-derivatives of dienestrol and DES (Metzler, 1976). The major biliary conjugate of DES is the monoglucuronide (75%), with small amounts of diglucuronide (10%) and polar products (15%) (Fischer, Millburn, Smith & Williams, 1966). In contrast bifluranol (2 mg/kg) undergoes oxidative metabolism only to the extent of 7% in rat to give a single main oxidation product. Mass spectrometry indicates that this metabolite has an additional hydroxyl group in one of the aromatic rings. The major biliary conjugates also differ, the monoglucuronide (50%) and glucuronide sulphate (42%) being the major products with small amounts of diglucuronide (8%). Further comparative studies are in progress to examine this lower extent of oxidative metabolism and the formation of a mixed double conjugate seen with bifluranol.

References

- FISCHER, L.J., MILLBURN, P., SMITH, R.L. & WILLIAMS, R.T. (1966). The fate of ¹⁴C-stilboestrol in the rat. *Biochem. J.*, **100** (3), 69P.
METZLER, M. (1976). Metabolic activation of carcinogenic diethylstilboestrol in rodents and humans. *J. Toxicol. Env. Hlth. Suppl.*, **1**, 21-35.

tion of expired air demonstrated 5% deacetylation, although desacetylpractolol was not detected in urine, confirming previous observations (Bodem & Chidsey, 1973). Human adverse reactions do not appear to be associated with gross differences in practolol metabolism.

Of eight animal species studied, mouse, rat and dog most closely resemble man in metabolic profile. Minimal deacetylation (5-14% dose) occurs in all species except marmoset (51%). In other species, practolol was recovered largely unchanged in urine, except hamster, which eliminated practolol primarily as 3-hydroxypractolol and its glucuronide (35% dose).

Microsomal studies (Orton & Lowery, 1977) have been extended to show that hepatic enzymes from several species produce intermediary metabolites which bind to proteins. Hamster microsomes produced the highest binding rate although with marked inter-animal variation (0.44-2.44 nmoles bound mg protein⁻¹ 30 min⁻¹). Inhibition and stimulation (53-448% control) were observed in the presence of sodium fluoride, stimulation being the major finding. Tricyclopropene oxide, an epoxide hydratase inhibitor, caused no significant change in binding. *Bis*-[*p*-nitrophenyl] phosphate both inhibited deacetylation *in vitro* and reduced binding. These results suggest, but do not prove, that N-hydroxylation may give rise to the intermediary metabolite(s). Microsomal activation has been utilized immunologically to screen for anti-practolol metabolite antibodies in patients' sera (Amos, Lake & Atkinson, 1977).

The relevance of any hypothesis implicating bound metabolites remains unclear as no toxic signs related to those seen in man were found in albino mouse (18 months, doses up to 100 mg/kg), black (C57 BL/10J) mouse (21 months, 300 mg/kg), rat (24 months, 300 mg/kg), Beagle dog (12 months, 200 mg/kg) and marmoset (6 months, 400 mg/kg). A hamster study reported to us gave rise to no relevant toxic sign. Thus, to date, no animal model for the human adverse reactions is known.

References

- AMOS, H.E., LAKE, B.G. & ATKINSON, H.A.C. (1977). Allergic drug reactions: an *in vitro* model using a mixed function oxidase complex to demonstrate antibodies with specificity for a practolol metabolite. *Clin. Allergy*, **7**, 423-428.
- BODEM, G. & CHIDSEY, C.A. (1973). Pharmacokinetic studies of practolol, a beta adrenergic antagonist, in man. *Clin. Pharmac. Ther.*, **14**, 26-29.
- NICHOLLS, J.T. (1976). Adverse effects of practolol. *Ann. Clin. Res.*, **8**, 229-231.
- ORTON, T.C. & LOWERY, C. (1977). Irreversible protein binding of [¹⁴C]-practolol metabolite(s) to hamster liver microsomes. *Br. J. Pharmac.*, **60**, 319P.
- REEVES, P.R., CASE, D.E., FELIX, R.H., FLUKE, R.W., HOLT, P.J.L., JEPSON, H.T., McCORMICK, D.J., NICHOLLS, J.T. & ZACHARIAS, F.J. (1978). Practolol metabolism: 2. Metabolism in human subjects. *J. Pharmac. exp. Ther.* (in press).

Paraquat-induced formation of hydroperoxide in mouse liver microsomes

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The activation of oxygen is considered to be the underlying mechanism of paraquat (PQ) toxicity. Bus, Aust & Gibson (1974) demonstrated the formation of superoxide anion and an increased peroxidation of microsomal lipids in a system consisting of NADPH-cytochrome c-reductase, microsomal lipids and a NADPH-regenerating system whereas other

authors (Ilett, Stripp, Menard, Reid & Gillette, 1974; Montgomery, 1976) observed a diminished formation of malondialdehyde (MDA).

Mouse liver microsomes were incubated at 37°C in a Soerensen buffer (pH 7.4) with NADPH and a NADPH regenerating system. Oxygen uptake was measured polarographically and MDA formation by the 2-thiobarbitone acid method. Oxygen uptake was increased by PQ in a dose-dependent fashion (K_m 3×10^{-4} M); microsomes from animals pretreated with phenobarbitone had a higher oxygen uptake than microsomes from control animals. MDA formation was decreased by PQ (K_i 6×10^{-5} M) and was not affected by phenobarbitone pretreatment. In the absence of PQ about 1 mole of NADPH was oxidized per mole oxygen. In the presence of PQ (1 mM) the ratio was 2. Addition of NaN_3 (1mM) shifted the NADPH/O₂ ratio towards 1 and increased the speed