



The role of parasite-induced immunodepression, rank and social environment in the modulation of behaviour and hormone concentration in male laboratory mice (*Mus musculus*)

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Peripheral immune responsiveness in male laboratory mice was reduced by infection with the trichostrongyloid nematode *Heligmosomoides polygyrus*. Responsiveness was also lower among high-ranking (aggressive) males regardless of infection status. Reduced responsiveness in both infected animals and high rankers was associated with elevated serum corticosterone concentration (a potential immunodepressant) and was compounded among high-ranking males by subsequent high aggressiveness. As in previous experiments, only low rankers modulated testosterone secretion in relation to current immunocompetence and corticosterone concentration. The lack of any downregulation of aggression in response to parasite-induced immunodepression contrasted with previous results using antithymocyte serum and may be due to the more localized nature of immunodepression during *H. polygyrus* infection. However, the additional increase in corticosterone concentration resulting from exposure to female odour and destabilized aggressive social relationships did result in downregulation of aggression among high rankers and of testosterone among mice generally, suggesting that modulation rules of thumb are at least partly dependent on the proximate cues associated with immunodepression.

Keywords: immunocompetence; parasite; mice; modulation; aggression; social rank; testosterone; corticosterone

1. INTRODUCTION

The idea that interrelationships between behaviour, immune function and other physiological factors may reflect adaptive life history trade-offs has recently been gaining theoretical and empirical support (e.g. Folstad & Karter 1992; Wedekind & Folstad 1994; Sheldon & Verhulst 1996; Barnard & Hurst 1996; Barnard *et al.* 1996a,b, 1997a). Although interest has mainly focused on sexual selection and the effect on secondary sexual characters of modulating immunodepressive sex steroids to protect immunity (e.g. Folstad & Karter 1992; Owens & Short 1995; Hillgarth & Wingfield 1997), current immunocompetence may act more generally as a constraint on behavioural and physiological decisions where these risk imposing a burden on immune function.

In a series of experiments with male laboratory mice (*Mus musculus*), we have shown that behaviour and serum hormone (testosterone and corticosterone) and immunoglobulin concentrations covary in a rank-dependent fashion that is consistent with the adaptive modulation hypothesis and differences in life history strategy between

individuals (Barnard *et al.* 1994, 1996a,b; see also Klein *et al.* (1997) for evidence from other rodent species). Moreover, the covariation can be manipulated predictably by increasing social stress or depressing immune function experimentally (Barnard *et al.* 1996b, 1997a,b).

However, although these results provide strong support for the adaptive modulation hypothesis, experimental manipulation of immunocompetence has so far been limited to the use of antithymocyte serum (ATS) (to depress thymus-mediated immunity (Doenhoff & Leuchars 1977; Levey & Medawar 1966; Barnard *et al.* 1997a,b)). While ATS is relatively benign in terms of cytotoxic and other unwelcome side-effects (Levey & Medawar 1966; Lance *et al.* 1973), it is nevertheless an artificial, and fairly extreme, means of compromising immunity. If modulation of behaviour and physiology in relation to immune function is a general feature of decision-making in mice, similar responses should be expected following other means of inducing immunodepression (e.g. direct immune component or pathogen product administration (Friedman *et al.* 1996; Bluthé *et al.* 1997)), particularly those mimicking more natural causes of depressed immunity such as the immunodepressive effects of certain parasitic infections (Playfair 1982; Urban *et al.* 1992).

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Many parasites appear to enhance their survival in the host through a range of immunodepressive strategies (Raybourne *et al.* 1983; Mazingue *et al.* 1983; Liew *et al.* 1987; Cross & Klesius 1989). One such species is the trichostrongyloid nematode *Heligmosomoides polygyrus bakeri*, a natural intestinal parasite of house mice. *H. polygyrus* causes chronic infections (Robinson *et al.* 1989) by downregulating local inflammatory responses in the intestine (Behnke *et al.* 1993), as well as systemic antibody production (Ali & Behnke 1983), probably through interfering with T-cell function (Behnke *et al.* 1993). Although the major impact on host response is locally in the intestine, where mastocytosis is severely depressed (Behnke & Wakelin 1977) and the local mucosal immune response against other intestinal nematodes greatly weakened (Behnke *et al.* 1978), this downregulation is not amenable to intercurrent monitoring. However, systemic downregulation, although weaker, provides a convenient intercurrent yardstick of overall impact. As well as downregulating immune function, low doses of *H. polygyrus* are also known to affect the development of social status (Freeland 1981) and various behavioural attributes (e.g. spatial learning and predator aversion (Kavaliers & Colwell 1995; Kavaliers *et al.* 1998)) in mice. In the present experiment, we repeated Barnard *et al.*'s (1997*a,b*) experimental design, but replaced ATS treatment with infection by *H. polygyrus*. As in the ATS experiments, infected and uninfected mice were divided into two treatments on the basis of exposure or otherwise to female odours (and thus potential reproductive opportunity). Following Barnard *et al.* (1997*a,b*), we expected infected mice to downregulate testosterone and aggressive behaviour, but maintain or increase time spent sleeping, relative to controls, and for these differences between infected and uninfected mice to disappear when animals were exposed to female odours. Although corticosterone has shown no evidence of modulation in relation to immunocompetence in our previous experiments (see Barnard *et al.* 1996*a*, 1997*a*; Smith *et al.* 1996), glucocorticoids have an inhibitory effect on the secretory function of Th2 cells which characterize the immune response to helminth infections generally (Grencis *et al.* 1991; Finkelman *et al.* 1995; Padgett *et al.* 1995; but see Golding *et al.* 1994) and are known to be effective in protective immunity to *H. polygyrus* (Urban *et al.* 1992). Some reduction in host-induced corticosterone levels might thus be expected among infected animals.

2. METHODS

(a) Pre-experimental procedure

The subjects were two batches of 64 male laboratory mice of the randomly bred CFLP strain (see Barnard *et al.* 1993) purchased from Bantin and Kingman Ltd, Hull, UK, at 42 days of age. Twenty-three female CFLP mice of the same age as the males were purchased from the same suppliers at the same time as the second batch and immediately established in groups of 7–8 animals in large polypropylene cages (48 cm × 28 cm × 13 cm) in a separate room from the males.

The experiment followed Barnard *et al.* (1997*a,b*) in being carried out in two parts: a 'no female odour' treatment (batch 1) followed by a 'female odour' treatment (batch 2). This prevented exposure of the former to ambient female odours while maintaining both treatments in the same physical environment (see

discussion and references in Barnard *et al.* (1997*b*)). The same parasite stock was used to infect mice in both batches (see below), and any chance batch effects in other variables were taken into account in subsequent analyses (see Barnard *et al.* 1997*b*).

(b) Pre-isolation/infection period

Prior to the experiment, animals were maintained, individually marked with black hair dye and blood sampled (figure 1, sample 1 (88 µl from the tail)) exactly as described by Barnard *et al.* (1997*b*). Three days after the pre-experimental blood samples, the males in each batch were re-allocated arbitrarily to 16 groups of four previously unfamiliar individuals (32 groups in total see Barnard *et al.* 1997*b*) and their behaviour recorded over; the next 8 d following the comprehensive methods of Hurst *et al.* (1996) and Barnard *et al.* (1997*a,b*). A full list of behaviour categories and definitions is given in Table 1 of Barnard *et al.* (1997*a,b*).

(c) Isolation/infection period

At the end of the pre-infection period, males were weighed again and a second 88 µl blood sample taken from the tail (figure 1, sample 2). Groups were then allocated randomly into either infected or sham-infected control treatments (eight groups per batch each), and mice were separated and housed singly in the same sized cages as pre-infection groups.

On the day after separation, groups in the infection treatment were infected by gavage with 150 L3 larvae of *H. polygyrus* in 0.2 ml of distilled water, while control groups were given 0.2 ml of distilled water only. This dose was chosen because it is well tolerated by mice and is in the low dose range for studies claiming effects of subclinical infections of *H. polygyrus* on behaviour and underlying physiological mechanisms (e.g. Freeland 1981; Kavaliers & Colwell 1995). Moreover, it is within the range causing non-specific systemic depression of antibody (Ali & Behnke 1983) and cell-mediated responses (Ali & Behnke 1984). Mice then remained singly housed for 14 d to allow the worms to develop to the adult immunodepressive stage. On day 13 of separation, mice were weighed for a third time and an 88 µl blood sample taken retro-orbitally (figure 1, sample 3). To confirm that non-specific systemic immunodepression had been induced by infection with *H. polygyrus*, each individual was given an intraperitoneal injection on day 14 of 0.2 ml of a sheep erythrocyte suspension (SRBC) containing 25×10^7 SRBC ml⁻¹, each mouse receiving 5×10^7 SRBC (Barnard *et al.* 1997*a,b*; figure 1).

(d) Post-isolation/infection period

The day after injection with SRBC, mice were re-established in their pre-isolation/infection groups. 'No odour' groups (batch 1) were re-established on clean sawdust, whereas 'female odour' groups (batch 2) were given soiled sawdust from the cages of two arbitrarily allocated, singly housed females following the procedure of Barnard *et al.* (1997*b*). Behavioural observations were then taken for 8 d exactly as during the pre-isolation/infection period.

(e) Organ weights, worm burdens and blood assays

At the end of the post-infection observation period, males were weighed for the final time, killed using chloroform and exsanguinated. The kidneys, adrenal glands, spleen, thymus gland, testes, preputial glands, seminal vesicles, heart and mesenteric lymph nodes (MLN) of each individual were carefully dissected out and weighed. The intestinal worm burden was

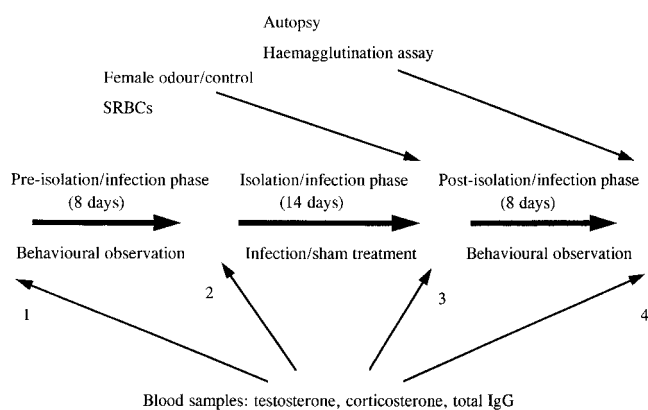


Figure 1. Flow diagram of experimental design (see § 2).

recovered, and worms sexed and counted within 48 h, following the established methods of Jenkins & Behnke (1977). SRBC haemagglutination titres, packed cell volume (PCV) and serum concentrations of testosterone, corticosterone and total IgG (a convenient bystander measure of peripheral immune responsiveness) were assayed using the standard techniques in Barnard *et al.* (1994, 1997*a,b*).

(f) Statistical analyses

All analyses were done using Statgraphics Plus v.7 (Manugistics Ltd, Maryland, USA). Parametric analyses were used throughout (data were \log_{10} or square root transformed as necessary and tested for normality using a Kolmogorov–Smirnov one-sample test). Wherever there were *a priori* reasons for expecting trends or differences in a particular direction, probabilities associated with significance tests are indicated as one-tailed.

3. RESULTS

(a) Social rank

Following Barnard *et al.* (1993, 1994, 1996*a,b*, 1997*a,b*), high and low rank categories within groups were defined on the basis of the ratio of attacks initiated and received by each male during the pre-treatment period of grouping, and high-ranking males initiated disproportionately more attacks than low rankers over the pre-treatment period ($t_{52}=4.95$, $p<0.0001$). Five groups, in which the incidence of aggression was very low, were omitted from later analyses because it was not possible to allocate males to rank categories. All other groups comprised either one or two high-ranking males and two or three low-ranking males. All analyses relating to social status were based on high and low rank categories and, where there was more than one individual per category within cages, data were averaged to control for non-independence (Barnard *et al.* 1996*a,b*, 1997*a,b*).

(b) Pre-isolation/infection differences

Three-way analysis of variance (ANOVA) revealed no significant chance pre-treatment biases in any measure of hormone concentration, immunocompetence or body weight with respect to subsequent infection status or odour treatment. Because female odour treatments applied only during the post-isolation/infection phase, results for hormone and IgG concentrations during the

period of isolation/infection were analysed with subsequent odour treatments combined.

(c) Effects of infection, rank and odour treatment on immunocompetence

(i) Infection status

As expected (Ali & Behnke 1983), three-way ANOVA showed that infection with *H. polygyrus* reduced peripheral immune responsiveness as measured by terminal haemagglutination titre ($F_{1,46}=3.87$, one-tailed $p<0.05$, figure 2*a*). Also, as expected with the introduction of foreign antigen and the known IgG1 hypergammaglobulinaemia associated with *H. polygyrus* infection (Chapman *et al.* 1979; Williams & Behnke 1983), total serum IgG concentration increased during the period of isolation/infection (change between sample points 2 and 3 in figure 1) in those animals treated with *H. polygyrus* (two-way ANOVA, $F_{1,49}=93.40$, $p<0.0001$). During the post-isolation/infection phase, and following SRBC challenge, however, further increase in IgG concentration among infected mice was damped compared with the now increased response in control animals (three-way ANOVA, $F_{1,45}=11.98$, $p<0.002$, figure 2*b*). Within infected groups, partial regression analysis revealed a significant negative relationship between worm burden at autopsy and change in IgG concentration over the period of isolation/infection ($t_{20}=-2.23$, $p<0.05$), so that burdens were greater the smaller the IgG response. There was no independent relationship with haemagglutination titre.

Relative (% terminal body weight) MLN ($F_{1,46}=56.66$, $p<0.0001$) and spleen ($F_{1,46}=14.13$, $p<0.001$) weights showed a significant increase among infected animals, but there was no effect of infection on thymus weight.

There was no significant impact of infection on change in body weight or PCV over the period of the experiment, supporting our assumption that our low infection dose did not have deleterious clinical consequences.

(ii) Rank

High rankers had significantly lower haemagglutination titres than low rankers (three-way ANOVA, $F_{1,46}=10.54$, $p<0.005$, figure 3) and there was a pronounced, but just non-significant ($F_{1,46}=3.72$, $p=0.059$), interaction between rank and infection status, with infected low rankers showing a much reduced titre compared with uninfected controls (mean \pm s.e. titre (1/dilution) in infected groups = 52.61 ± 8.00 , $n=12$; in uninfected groups = 88.89 ± 9.36 , $n=15$) but high rankers showing similarly low titres in both (mean \pm s.e. in infected groups = 40.08 ± 5.69 , $n=13$; in uninfected groups = 40.53 ± 11.53 , $n=15$). There were no significant effects of rank and no interaction for changes in IgG concentration during either the isolation/infection or post-isolation/infection phases, and no difference between rank categories in the regression relationship between worm burden and IgG (see above). Rank showed no effects (main or interaction) on worm burden, or weight of MLN, spleen or thymus.

(iii) Odour treatment

There were no significant biases in IgG measures with respect to post-isolation/infection odour treatment during the isolation/infection period, and no significant effect of odour treatment on post-isolation/infection measures of

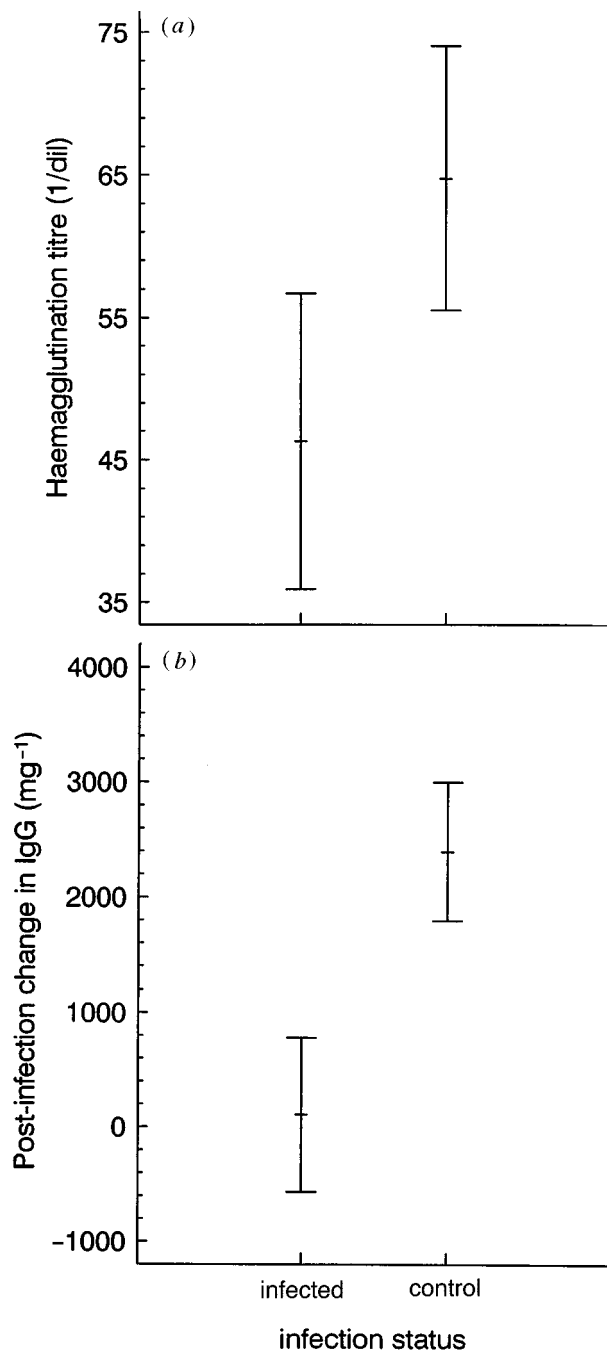


Figure 2. Effect of infection status on (a) haemagglutination titre and (b) post-isolation/infection phase change in total IgG concentration (means \pm least squares deviations (l.s.d.) from ANOVA). See text.

IgG or on haemagglutination titre. However, terminal worm burden among infected animals was significantly lower following exposure to female odour (two-way ANOVA by odour treatment and rank, $F_{1,19}=45.30$, $p<0.0001$), with no interaction between odour treatment and rank. There were no effects of odour on MLN, spleen or thymus weight, but, in keeping with the reduced worm burden, there was a significant interaction between infection status and odour treatment on spleen weight ($F_{1,46}=5.64$, $p<0.05$), with spleen weight being reduced among infected animals following the presence of female odour.

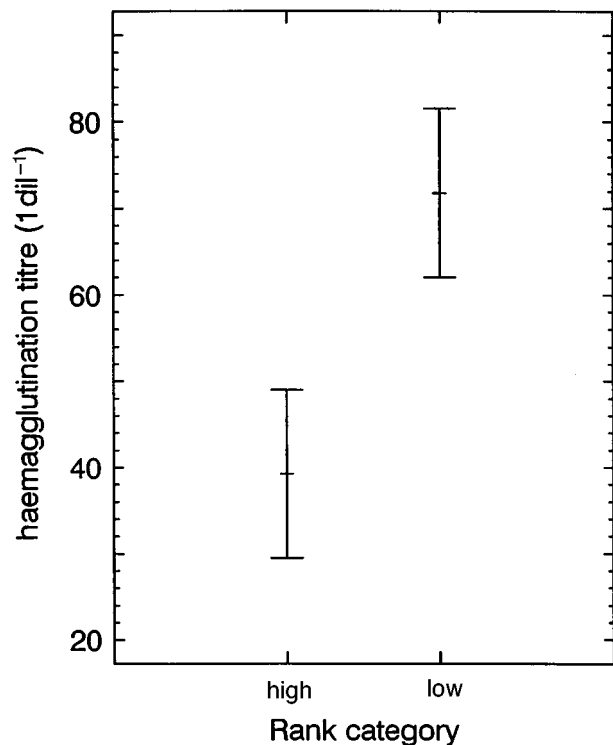


Figure 3. Effect of rank category on haemagglutination titre (means \pm l.s.d. from ANOVA). See text.

(d) *Immunocompetence and hormone modulation*

(i) *Isolation/infection phase*

Two-way ANOVA of changes in hormone concentrations over the period of isolation and infection showed no significant main effects of infection status or rank for testosterone. However, the interaction between infection status and rank was significant at the 10% level, with high rankers tending to reduce testosterone relative to low rankers in infected groups (from 10.11 ± 2.34 to 7.21 ± 3.50 (mean \pm s.e.) ng ml⁻¹ in high rankers but 6.64 ± 1.8 to 9.10 ± 1.59 ng ml⁻¹ in low rankers) but to increase it in control groups (7.36 ± 2.27 to 12.69 ± 3.58 ng ml⁻¹ in high rankers compared with 6.53 ± 1.04 to 4.47 ± 1.26 ng ml⁻¹ in low rankers) ($F_{1,40}=3.03$, $p=0.089$). As before (Barnard *et al.* 1994, 1996*a,b*), partial regression analysis revealed a rank difference in the change in testosterone relative to measures of immunocompetence, testosterone over the period of infection showing a significant increase with IgG among low rankers ($t_{24}=2.53$, $p<0.05$), but not among high rankers ($t_{22}=-0.39$, n.s.).

In contrast, two-way ANOVA of change in serum corticosterone concentration over the period of infection showed strongly significant differences with both infection status ($F_{1,45}=7.25$, $p<0.01$) and rank ($F_{1,45}=6.21$, $p<0.02$). Corticosterone levels declined sharply over the period in uninfected groups (from 118.22 ± 18.50 to 55.63 ± 4.12 ng ml⁻¹ versus 88.38 ± 6.96 to 77.82 ± 19.70 ng ml⁻¹ in infected groups) and showed a sharper decline among low rankers (from 124.52 ± 19.65 to 56.45 ± 3.96 ng ml⁻¹ compared with 84.22 ± 6.18 to 74.42 ± 17.64 among high rankers). However, the decline among low rankers was reduced in infected groups while infection resulted in an increase among high rankers (interaction in figure 4). Again as previously (Barnard *et*

al. 1994, 1996a,b; Smith *et al.* 1996), corticosterone showed no significant change relative to immunocompetence measures. Partial regression analysis showed that worm burden increased significantly with increasing corticosterone concentration ($t_{20}=2.89$, $p<0.01$), but there was no effect of change in testosterone and no difference in either relationship between rank categories.

(ii) *Post-isolation/infection phase*

Taking odour treatment into account, post-isolation/infection phase testosterone showed a precipitate drop (from 8.15 ± 1.62 to 2.10 ± 0.55 ng ml⁻¹), rather than the expected increase, in mice exposed to female odour (three-way ANOVA, $F_{1,41}=11.01$, $p<0.01$), but with no independent effects of infection status or rank and no interactions. Corticosterone concentration, however, increased, with the increase being greater among infected individuals (77.83 ± 19.70 to 440.80 ± 38.69 ng ml⁻¹ compared with 55.63 ± 4.12 to 308.85 ± 33.77 ng ml⁻¹ in uninfected controls; $F_{1,44}=9.90$, $p<0.005$) and, as expected (Smith *et al.* 1996), those exposed to female odours (79.59 ± 16.48 to 409.10 ± 39.83 ng ml⁻¹ compared with 50.39 ± 4.90 to 329.85 ± 33.77 ng ml⁻¹ in those not exposed to odours; $F_{1,44}=3.93$, one-tailed $p<0.05$). The greater increase in corticosterone among mice exposed to female odour was reflected in significantly greater relative adrenal weights in these individuals ($F_{1,46}=4.78$, $p<0.05$).

Partial regression analyses by rank category once again showed no significant association between change in testosterone levels and immunocompetence measures among high rankers, but a significant positive association with haemagglutination titre ($t_{25}=3.48$, $p<0.01$) among low rankers. Testosterone among low rankers also decreased with increasing corticosterone concentration ($t_{25}=-2.65$, $p<0.02$). Change in corticosterone showed no significant relationships with measures of immunocompetence and no significant relationships emerged between changes in hormone or IgG concentrations and eventual worm burden.

(e) *Effects of treatment on behaviour*

ANOVA revealed no significant biases with respect to subsequent infection or odour treatment in any behaviour category showing treatment effects after infection. During the post-isolation/infection phase, however, the highly significant difference between rank categories in the amount of aggression initiated remained ($F_{1,45}=13.50$, $p<0.001$) and there was a strong significant interaction between rank and odour treatment ($F_{1,45}=19.28$, $p<0.001$) with high rankers showing less aggression and low rankers more when female odours were present (figure 5). As expected following a period of isolation (Cairns *et al.* 1985, Hurst *et al.* 1994), low rankers showed a greater tendency to challenge high rankers in post-isolation/infection groups, with the number of attacks received by high rankers increasing significantly over pre-isolation/infection phase levels ($F_{1,45}=5.60$, $p<0.05$). Again, there was a suggestive interaction between rank and odour treatment ($F_{1,45}=3.41$, $p=0.073$), high rankers showing a greater increase in attacks received, and low rankers a marked reduction, when female odours were present. There were no significant main or interaction effects of

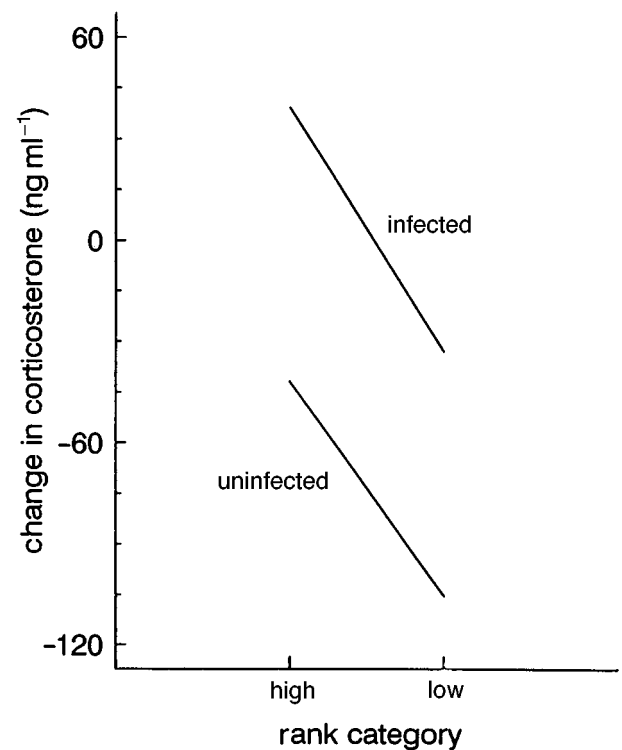


Figure 4. Effects of rank category and infection status on change in corticosterone concentration during the isolation/infection phase (interaction plot from ANOVA, error bars omitted for clarity). See text.

infection status on aggression initiated or received during the post-isolation/infection phase.

The only other behaviour showing a significant effect of experimental treatments was mounting, the frequency of mounts initiated during the post-isolation/infection phase increasing when female odours were presented ($F_{1,46}=13.50$, $p<0.001$; see also Barnard *et al.* 1997b). There were no effects of infection status and no interaction with either infection status or rank.

(f) *Behaviour, hormone concentrations and immune function*

Partial regression analysis was used to investigate the effect of post-isolation/infection phase behaviour on hormone concentrations. Rank categories were analysed separately with infection status and odour treatment included as dummy variables and post-isolation/infection phase body weight and testosterone concentration as additional independent variables. Analyses revealed a significant positive effect of aggression initiated on change in corticosterone concentration during the post-isolation/infection phase among high rankers ($t_{23}=2.19$, $p<0.05$), but not among low rankers. There were no significant effects of aggression received or any other behaviours included simultaneously as independent variables. There were no significant effects of any behaviour category on post-isolation/infection phase measures of testosterone concentration. However, inclusion of the change in corticosterone concentration over the post-isolation/infection period showed a significant negative correlation with the change in testosterone in both rank categories when female odour was present ($t_{10}=-2.32$, one-tailed $p<0.05$ for high rankers; $t_{12}=-2.38$, one-tailed $p<0.01$ for low

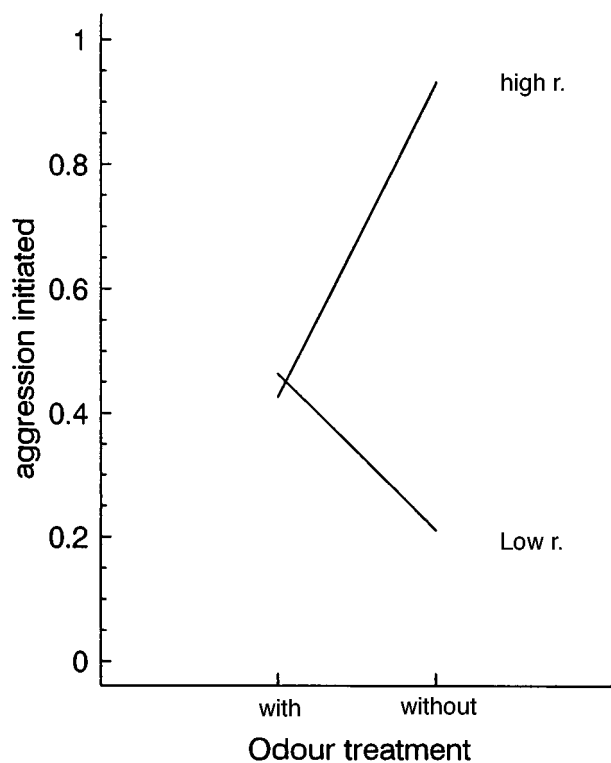


Figure 5. Effects of odour treatment and rank category on the ($\log_{10}+1$) number of aggressive acts initiated during the post-isolation/infection phase (interaction plot from ANOVA, error bars omitted for clarity). See text.

rankers), but only among low rankers ($t_{11} = -3.69$, one-tailed $p < 0.001$) when odour was not present. Analyses of the effects of behaviour on haemagglutination titre and (in infected animals only) worm burden showed a significant reduction in haemagglutination response with increased aggression received ($t_{25} = -2.35$, one-tailed $p < 0.01$) and an increase in worm burden with increased aggression initiated ($t_{10} = 2.26$, one-tailed $p < 0.05$, figure 6) but, again, only among high rankers. No significant effects of any other behaviour on measures of immune response or worm burden emerged in either rank category.

4. DISCUSSION

The results showed the expected reduction in immune responsiveness among animals infected with *H. polygyrus*. However, associations between infection status and changes in behaviour were less direct than those resulting from ATS-induced immunodepression (Barnard *et al.* 1997a) and appeared to be mediated by rank-related changes in corticosterone levels. As found by Smith *et al.* (1996), changes in corticosterone also appeared to underlie behavioural responses to the substrate odours of females. However, as Barnard *et al.* (1997a) have stressed, these apparent effects of hormones do not necessarily reflect direct causal relationships at the level of measurement (in this case, total serum concentrations), but instead may correlate with associated underlying metabolic pathways and mechanisms (e.g. the activity of precursors, metabolites and the modulation of receptors and binding globulins) (see, for example, Kotani *et al.* 1974; Grossman & Roselle 1986; Folstad & Karter 1992; Roberts *et al.* 1996).

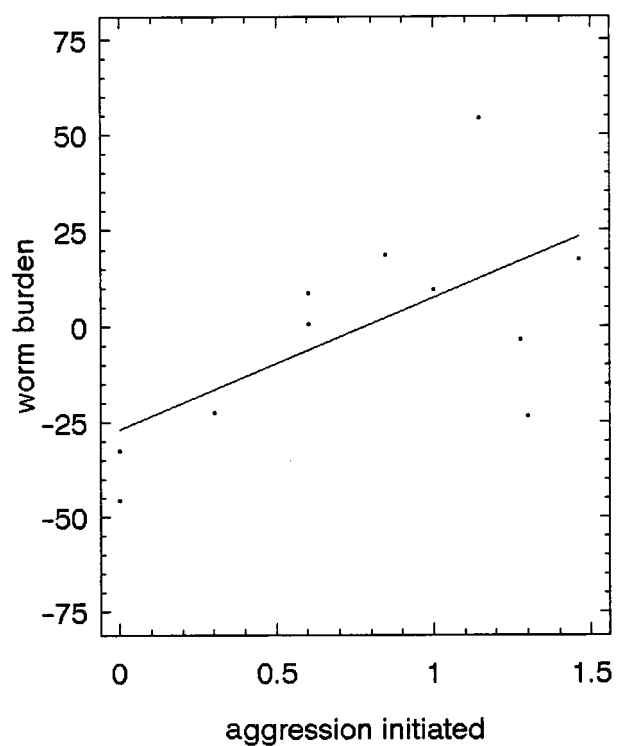


Figure 6. Component plot from partial regression analysis of the relationship between the ($\log_{10}+1$) number of aggressive acts initiated by high-ranking males in the post-isolation/infection phase and terminal worm burden. Regression equation: $y = 126.60 + 34.50(\text{agg})$, where $\text{agg} = (\log_{10}+1)$ no. aggressive acts initiated; no other independent variables entered the equation. See text.

Overall, high rankers showed reduced immune responsiveness compared with low rankers, with the main effects of infection with *H. polygyrus* appearing to be due to a reduction in responsiveness among low rankers. The difference between ranks may be explicable in terms of the interrelationship between aggression, corticosterone concentration, immunocompetence and resistance to *H. polygyrus* among high rankers in which post-infection aggressive behaviour correlated positively with corticosterone concentration and negatively with haemagglutination titre and resistance to *H. polygyrus* (measured as terminal worm burden). There was no evidence that infected animals downregulated corticosterone concentration. Instead, the post-infection phase relationships compounded a tendency for high rankers and infected mice generally to maintain corticosterone concentration during the period of isolation and infection, when concentrations dropped among low rankers and uninfected controls. The difference between ranks is consistent with a previously reported link between elevated corticosterone levels and reduced resistance among high-ranking males (Smith *et al.* 1996). The maintenance of corticosterone levels among infected animals, however, could reflect a confounding effect of parasite-induced glucocorticoid secretion (Hillgarth & Wingfield 1997) as a manipulative strategy to reduce the efficacy of the host's immune response (see also Behnke *et al.* 1992). Although glucocorticoids have a stimulatory effect on the initial cell proliferation phase of T lymphocytes, and thus

some elevation might have been expected on this account (e.g. Maier *et al.* 1994; Padgett *et al.* 1995), the change in corticosterone concentration during the infection phase was the best hormone-measure predictor of eventual worm burden. The negative relationship between the two is more in keeping with the later impact of glucocorticoids on the secretion of Th2 cytokines (Padgett *et al.* 1995; Rook & Zumia 1997) and thus depression of the Th2 arm of the immune response. This is consistent with effects of glucocorticoid drugs in prolonging intestinal nematode infections (Wakelin & Selby 1974), increasing the susceptibility of rodent hosts to *H. polygyrus* (Quinnell *et al.* 1991) and depressing the expression of acquired resistance to *H. polygyrus* (Behnke & Parish 1979).

In contrast to corticosterone, testosterone once again showed rank-related modulation, with correlations between hormone concentration and measures of immunocompetence being evident only among low rankers. However, unlike the situation with *Babesia* infections (Barnard *et al.* 1994, 1996*a,b*), there was no testosterone-dependent increase in parasite burden among high rankers in the present experiment, perhaps because resistance to the two parasites relies on a different emphasis on the Th1 and Th2 arms of the acquired immune response. Androgens tend to modulate Th1 responses, which are broadly associated with combating intracellular parasites such as *Babesia* (but see Allen & Maizels 1997).

These interrelationships suggest that, as before (e.g. Barnard *et al.* 1996*a,b*, 1997*a,b*), aggression was costly in terms of reduced immunocompetence, but, unlike the experiments in which immunocompetence was reduced by administering ATS (Barnard *et al.* 1997*a*), mice immunocompromised by *H. polygyrus* did not downregulate aggressive behaviour. This may reflect a difference in the modes of immunodepression by *H. polygyrus* and ATS treatment, the major impact of *H. polygyrus* on host immune response being locally in the intestine, with a weaker reduction in peripheral responsiveness compared with ATS treatment (cf. haemagglutination titres in figure 2*a* and Barnard *et al.* (1997*a,b*)). Such localized immunodepression may be less likely to trigger compensatory responses than systemic downregulation (see below).

The main effect of introducing female odour on the behaviour of males was to increase the amount of aggression initiated by low-ranking individuals against high rankers, which showed a marked reduction in aggressiveness as a result. The resulting destabilization of competitive social relationship (see Smith *et al.* (1996) for a fuller discussion) caused a general elevation of corticosterone levels and adrenal weights, to which the reduction in aggression by high rankers, and the general downregulation of testosterone concentration, may have been compensatory responses (Barnard *et al.* 1996*a*). Interestingly, partial regression by rank and odour treatment revealed that testosterone was downregulated in relation to corticosterone concentration rather than aggression and that downregulation occurred in high rankers only when female odours were present. We have shown previously (Barnard *et al.* 1996*a*) that testosterone concentration in male CFLPs of both ranks drops sharply in situations of increased social conflict. Increased aggression

in groups may also account for the discrepancy with some other studies, including our own, that have shown an increase in testosterone in male mice exposed to the odours of females (Macrides *et al.* 1975; Batty 1978; Smith *et al.* 1996), but in single rather than grouped animals.

Taken together, the results support our earlier conclusions (Barnard *et al.* 1994, 1996*a,b*) that high- and low-ranking males differ in their tendency to modulate potential immunodepressants (in this case testosterone) and trade-off future susceptibility to infection. However, the results also suggest that the difference may vary with the proximate cues relating to immunodepression. Thus, socially salient cues such as increased rate of being attacked, which are associated with elevated levels of immunodepressive corticosterone, reduced peripheral immune responsiveness and short-term changes in reproductive potential, may have acted as a stronger selection pressure for modulatory rules of thumb than helminth infections, which tend to be chronic and ubiquitous, with a more localized and perhaps manipulative impact on host immune function. The study therefore points to the potential importance of life history considerations and proximate rules of thumb in decisions relating to the conservation of immune function.

We thank Francis Gilbert and two anonymous referees for helpful comments and discussion, Ian Davies, Charlotte Nevison and Jill Brown for assistance during autopsies, and David Fox for Animal House facilities. The work was supported by a research grant from the Biotechnology and Biological Sciences Research Council to C.J.B. and J.M.B. and carried out under Home Office licence 40/1086.

REFERENCES

- Ali, N. M. H. & Behnke, J. M. 1983 *Nematospiroides dubius*: factors affecting the primary response to SRBC in infected mice. *J. Helminthol.* **57**, 343–353.
- Ali, N. M. H. & Behnke, J. M. 1984 Non-specific immunodepression by *Nematospiroides dubius*: of concurrent responses to oxazolone and lipopolysaccharide. *J. Helminthol.* **58**, 301–311.
- Allen, J. E. & Maizels, R. M. 1997 Th1-Th2: reliable paradigm or dangerous dogma? *Immunol. Today* **18**, 387–393.
- Barnard, C. J. & Hurst, J. L. 1996 Welfare by design: the natural selection of welfare criteria. *Anim. Welfare* **5**, 405–433.
- Barnard, C. J., Behnke, J. M. & Sewell, J. 1993 Social behaviour, stress and susceptibility to infection in house mice (*Mus musculus*): effects of duration of grouping and aggressive behaviour prior to infection on susceptibility to *Babesia microti*. *Parasitology* **107**, 183–192.
- Barnard, C. J., Behnke, J. M. & Sewell, J. 1994 Social behaviour and susceptibility to infection in house mice (*Mus musculus*): effects of group size, aggressive behaviour and status-related hormonal responses prior to infection on resistance to *Babesia microti*. *Parasitology* **108**, 487–496.
- Barnard, C. J., Behnke, J. M. & Sewell, J. 1996*a* Social status and resistance to disease in house mice (*Mus musculus*): status-related modulation of hormonal responses in relation to immunity costs in different social and physical environments. *Ethology* **102**, 63–84.
- Barnard, C. J., Behnke, J. M. & Sewell, J. 1996*b* Environmental enrichment, immunocompetence and resistance to *Babesia microti* in male laboratory mice. *Physiol. Behav.* **60**, 1223–1231.
- Barnard, C. J., Behnke, J. M., Gage, A. R., Brown, H. & Smithurst, P. R. 1997*a* Modulation of behaviour and

- testosterone concentration in immunodepressed male laboratory mice (*Mus musculus*). *Physiol. Behav.* **61**, 907–917.
- Barnard, C. J., Behnke, J. M., Gage, A. R., Brown, H. & Smithurst, P. R. 1997b Immunity costs and behavioural modulation in male laboratory mice (*Mus musculus*) exposed to the odour of females. *Physiol. Behav.* **62**, 857–866.
- Batty, J. 1978 Acute changes in plasma testosterone levels and their relation to measures of sexual behaviour in the male house mouse (*Mus musculus*). *Anim. Behav.* **26**, 349–357.
- Behnke, J. M. 1987 Evasion of immunity by nematode parasites causing chronic infections. *Adv. Parasitol.* **26**, 1–71.
- Behnke, J. M. & Parish, H. A. 1979 *Nematospiroides dubius*: arrested development of larvae in immune mice. *Expl Parasitol.* **47**, 116–127.
- Behnke, J. M. & Wakelin, D. 1977 *Nematospiroides dubius*: stimulation of acquired immunity in inbred strains of mice. *J. Helminthol.* **57**, 167–176.
- Behnke, J. M., Barnard, C. J. & Wakelin, D. 1992 Understanding chronic nematode infections: evolutionary considerations, current hypotheses and the way forward. *Int. J. Parasitol.* **22**, 861–907.
- Behnke, J. M., Wakelin, D. & Wilson, M. M. 1978 *Trichinella spiralis*: delayed rejection in mice concurrently infected with *Nematospiroides dubius*. *Exp. Parasitol.* **46**, 121–130.
- Behnke, J. M., Wahid, F. N., Grecnis, R. K., Else, K. J., Ben-Smith, A. W. & Goyal, P. K. 1993 Immunological relationships during primary infection with *Heligmosomoides polygyrus* (*Nematospiroides dubius*): down regulation of specific cytokine secretion (IL-9 and IL-10) correlates with poor mastocytosis and chronic survival of adult worms. *Parasitol. Immunol.* **15**, 415–412.
- Bluthe, R. M., Dantzer, R. & Kelley, K. W. 1997 Central mediation of the effects of interleukin 1 on social exploration and body weight in mice. *Psychoneuroimm* **22**, 1–11.
- Cairns, R. B., Hood, K. E. & Midlairn, J. 1985 On fighting in mice: is there a sensitive period for isolation? *Anim. Behav.* **33**, 166–180.
- Chapman, C. B., Knopf, P. M., Anders, R. F. & Mitchell, G. F. 1979 IgG1 hypergammaglobulinaemia in chronic parasitic infections in mice: magnitude of the response in mice infected with various parasites. *Aust. J. Expl Biol. Med. Sci.* **57**, 369–87.
- Cross, D. A. & Klesius, P. H. 1989. Soluble extracts from larval *Ostertagia ostertagia* modulating immune function. *Int. J. Parasitol.* **19**, 57–61.
- Doenhoff, M. J. & Leuchars, E. 1977 Effects of irradiation, antithymocyte serum and corticosteroids on PHA and LPS responsive cells of the mouse. *Int. Arch. Allergy Appl. Immunol.* **53**, 505–514.
- Finkelman, F. D., Gause, W. C. & Urban, J. F. Jr 1995 Cytokine control of protective immunity against nematode infections. In *Molecular approaches to parasitology* (ed. J. C. Boothroyd & R. Komuniecki), pp. 467–476. New York: Wiley-Liss Inc.
- Folstad, I. & Karter, A. J. 1992 Parasites, bright males and the immunocompetence handicap. *Am. Nat.* **139**, 603–622.
- Freeland, W. J. 1981 Parasitism and behavioral dominance among male mice. *Science* **213**, 461–462.
- Friedman, E. M., Reyes, T. M. & Coe, C. L. 1996 Context-dependent behavioral effects of interleukin 1 in the rhesus monkey (*Macaca mulatta*). *Psychoneuroimm* **21**, 455–468.
- Golding, B., Zaitseva, M. & Golding, H. 1994 The potential for recruiting immune responses toward Type 1 or Type 2 T cell help. *Am. J. Trop. Med.* **50**(4) Suppl., 33–40.
- Grecnis, R. K., Hulter, L. & Else, K. J. 1991 Host protective immunity to *Trichinella spiralis* in mice: activation of Th cell subsets and lymphokine secretion in mice expressing different response phenotypes. *Immunology* **74**, 329–332.
- Grossman, C. J. & Roselle, G. A. 1986 The control of immune response by endocrine factors and the clinical significance of such regulation. *Progr. Clin. Biochem. Med.* **4**, 9–56.
- Hillgarth, N. & Wingfield, J. C. 1997 Testosterone and immunosuppression in vertebrates: implications for parasite-mediated sexual selection. In *Parasites & pathogens; effects on host hormones and behavior* (ed. N. E. Beckage), pp. 143–155. London: Chapman and Hall.
- Hurst, H. L. 1993 The priming effects of urine substrate marks on interactions between male mouse mice, *Mus musculus domesticus* Shwarz and Schwarz. *Anim. Behav.* **45**, 55–81.
- Hurst, J. L., Fang, J. & Barnard, C. J. 1994 The role of substrate odours in maintaining social tolerance between male house mice, *Mus musculus domesticus*: relatedness, incidental kinship effects and the establishment of social status. *Anim. Behav.* **48**, 157–167.
- Hurst, J. L., Barnard, C. J., Hare, R., Wheeldon, E. B. & West, C. D. 1996 Housing and welfare in laboratory rats: status-dependent time-budgeting and pathophysiology in single sex groups maintained in open rooms. *Anim. Behav.* **52**, 335–360.
- Jenkins, S. N. & Behnke, J. M. 1977 Impairment of primary expulsion of *Trichuris muris* in mice concurrently infected with *Nematospiroides dubius*. *Parasitology* **75**, 71–78.
- Kavaliers, M. & Colwell, D. D. 1995 Reduced spatial learning in mice infected with the nematode *Heligmosomoides polygyrus*. *Parasitology* **110**, 591–597.
- Kavaliers, M., Colwell, D. D. & Perrot-Sinal, T. S. 1998 Opioid and non-opioid NMDA mediated predator-induced analgesia in mice and the effects of parasitic infection. *Brain Res.* (In the press.)
- Klein, S. L., Hairston, J. E., DeVries, A. C. & Nelson, R. J. 1997 Social environment and steroid hormones affect species and sex differences in immune function in voles. *Horm. Behav.* **32**, 30–39.
- Kotani, M., Korenaga, M., Nawa, Y. & Kotani, M. 1974 Inhibition by testosterone of immune reactivity and of lymphoid regeneration in irradiated and marrow reconstituted mice. *Experientia* **34**, 1343–1345.
- Lance, E. M., Medawar, P. B. & Taub, R. N. 1973 Antilymphocyte serum. *Adv. Immunol.* **17**, 2–92.
- Levey, R. H. & Medawar, P. B. 1966 Some experiments on the action of antilymphoid sera. *Ann. N.Y. Acad. Sci.* **129**, 164–177.
- Liew, F. Y., Scott, M. T., Lim, D. S. & Croft, S. L. 1987 Suppressive substance produced by T-cells from mice chronically infected with *Trypanosoma cruzi*. *J. Immunol.* **139**, 2452–2457.
- Macrides, F., Bartke, A. & Dalterio, S. 1975 Strange females increase plasma testosterone levels in male mice. *Science* **189**, 1104–1106.
- Maier, S. F., Watkins, L. R. & Fleshner, M. 1994 Psychoneuroimmunology: the interface between behaviour, brain and immunity. *Am. Psychol.* **49**, 1004–1017.
- Mazingue, C., Dessaint, J. P., Schmitt-Verhulst, A. M., Cerottini, J. C. & Capron, A. 1983 Inhibiting cytotoxic T lymphocytes by a schistosome-derived inhibitory factor is dependent on an inhibition of the production of interleukin. *Int. Arch. Allergy Appl. Immunol.* **72**, 22–29.
- Owens, I. P. F. & Short, R. V. 1995 Hormonal basis of sexual dimorphism in birds: implications for new theories of sexual selection. *Trends Ecol. Evol.* **10**, 44–47.
- Padgett, D. A., Sheridan, J. F. & Loria, R. 1995 Steroid hormone regulation of a polyclonal Th2 immune response. *Ann. N.Y. Acad. Sci.* **774**, 323–325.
- Playfair, J. H. L. 1982 Workshop report: suppressor cells in infectious disease. *Parasite Immunol.* **4**, 299–304.
- Quinnell, R. J., Behnke, J. M. & Keymer, A. E. 1991 Host specificity of and cross-immunity between two strains of *Heligmosomoides polygyrus*. *Parasitology* **102**, 419–427.
- Raybourne, R., Desowitz, R. S., Kliks, M. M. & Deardoff, T. L. 1983 *Anisakis simplex* and *Terranova* spp.: inhibition of larval

- excretory-secretory products of mitogen-induced rodent lymphoblast proliferation. *Expl Parasitol.* **55**, 238–298.
- Roberts, C. W., Satsokar, A. & Alexander, J. 1996 Sex steroids, pregnancy-associated hormones and immunity to parasitic infection. *Parasitol. Today* **12**, 382–388.
- Robinson, M., Wahid, F. N., Behnke, J. M. & Gilbert, F. S. 1989 Immunological relationships during primary infection with *Heligmosomoides polygyrus* (*Nematospiroides dubius*): dose-dependent expulsion of adult worms. *Parasitology* **98**, 115.
- Rook, G. A. W. & Zumia, A. 1997 Gulf War syndrome: is it due to a systemic shift in cytokine balance towards a Th2 profile? *Lancet* **349**, 1831–1833.
- Sheldon, B. C. & Verhulst, S. 1996 Ecological immunology: costly parasite defences and trade-offs in evolutionary ecology. *Trends Ecol. Evol.* **11**, 317–321.
- Smith, F. V. 1996 Behaviour and immune function in laboratory mice (*Mus musculus*). Unpublished Ph.D. thesis, University of Nottingham, UK.
- Smith, F. V., Barnard, C. J. & Behnke, J. M. 1996 Social odours, hormone modulation and resistance to disease in male laboratory mice (*Mus musculus*). *Anim. Behav.* **52**, 141–153.
- Telford, G., Wheeler, D. J., Appleby, P., Bowen, J. G. & Pritchard, D. I. 1998 *Heligmosomoides polygyrus* immunomodulatory factor (IMF) targets T-lymphocytes. *Infect. Immun.* (In the press.)
- Urban, J. F., Madden, K. B., Svetic, A., Cheever, A., Trotta, P. P., Gause, W. C., Katona, I. M. & Finkelman, F. D. 1992 The importance of Th2 cytokines in protective immunity to nematodes. *Immunol. Rev.* **127**, 205–220.
- Wakelin, D. & Selby, G. R. 1974 The induction of immunological tolerance to the parasitic nematode *Trichuris muris* is cortisone-treated mice. *Immunology* **26**, 1–10.
- Wedekind, C. & Folstad, I. 1994 Adaptive or nonadaptive immunosuppression by sex hormones? *Am. Nat.* **143**, 936–938.
- Williams, D. J. & Behnke, J. M. 1983 Host protective antibodies and serum immunoglobulin isotypes in mice chronically infected or repeatedly immunized with the nematode parasite *Nematospiroides dubius*. *Immunology* **48**, 37–47.

As this paper exceeds the maximum length normally permitted, the authors have agreed to contribute towards production costs.

