# Cortical integration in the visual system of the macaque monkey: large-scale morphological differences in the pyramidal neurons in the occipital, parietal and temporal lobes

## Guy N. Elston<sup>\*</sup>, Rowan Tweedale and Marcello G. P. Rosa

Vision, Touch and Hearing Research Centre, Department of Physiology and Pharmacology, The University of Queensland, Queensland 4072, Australia

Layer III pyramidal neurons were injected with Lucifer yellow in tangential cortical slices taken from the inferior temporal cortex (area TE) and the superior temporal polysensory (STP) area of the macaque monkey. Basal dendritic field areas of layer III pyramidal neurons in area STP are significantly larger, and their dendritic arborizations more complex, than those of cells in area TE. Moreover, the dendritic fields of layer III pyramidal neurons in both STP and TE are many times larger and more complex than those in areas forming 'lower' stages in cortical visual processing, such as the first (V1), second (V2), fourth (V4) and middle temporal (MT) visual areas. By combining data on spine density with those of Sholl analyses, we were able to estimate the average number of spines in the basal dendritic field of layer III pyramidal neurons in each area. These calculations revealed a 13-fold difference in the number of spines in the basal dendritic field between areas STP and V1 in animals of similar age. The large differences in complexity of the same kind of neuron in different visual areas go against arguments for isopotentiality of different cortical regions and provide a basis that allows pyramidal neurons in temporal areas TE and STP to integrate more inputs than neurons in more caudal visual areas.

Keywords: intracellular injection; extrastriate cortex; temporal lobe; dendritic spines; Lucifer yellow

## 1. INTRODUCTION

As in other primates, the cortex of the macaque monkey includes a number of visual areas in the occipital, parietal and temporal lobes (e.g. Rosa 1997). Based on their different functional characteristics and connections, it has been suggested that these areas are organized into processing 'streams' (Ungerleider & Mishkin 1982) that reflect, to some extent, a hierarchical organization (for reviews, see Maunsell & Newsome 1987; Weller 1988; Felleman & Van Essen 1991; Gross *et al.* 1993; Graziano & Gross 1997). Both the 'dorsal' and 'ventral' streams originate in the first and second visual areas (V1 and V2), but they involve different areas of the parietal and temporal lobes.

As well as differences in connectivity and response properties, a number of studies have shown marked variation in the morphology of neurons in striate and extrastriate areas (Lund *et al.* 1993; Elston *et al.* 1996; Fujita & Fujita 1996; Elston & Rosa 1997, 1998*a,b,c*; Peters *et al.* 1997; Jelinek *et al.* 1999). These morphological differences have been implicated in models of surround inhibition (Lund *et al.* 1993) and cortical integration (Amir *et al.* 1993; Elston & Rosa 1997, 1998*a,b*). The present study was conducted to determine the morphology of layer III pyramidal neurons in two cortical areas of the temporal lobe, cytoarchitectural area TE and the superior temporal polysensory (STP) area. The response properties and connections of neurons in areas TE and STP suggest that cells in these areas perform sophisticated analyses. Neurons in area TE, a 'high-order' visual area, have exquisite selectivities for object shape, colour and texture, while those in area STP are involved in the integration of inputs from a number of sensory modalities (for reviews, see Perrett et al. 1990; Gross et al. 1993; Tanaka 1996; Cusick 1997; Yukie 1997). Thus, we were interested in establishing whether these functional differences between temporal areas are paralleled by differences in cell morphology. Moreover, we were interested in comparing the morphology of temporal neurons with those in other visual areas, particularly in view of the persistent belief that neurons in different visual areas are homogenous in their morphology (e.g. Hendry & Calkins 1998).

#### 2. MATERIAL AND METHODS

Tissue was obtained from two adult male macaque monkeys (*Macaca fascicularis*), cases DM4 and MF2 (18 months and 11 years old, respectively). Animals were administered an overdose of barbiturate (sodium pentobarbitone) and perfused with physiological saline, followed by 4% paraformaldehyde (in  $0.1 \text{ mol}^{-1}$  phosphate buffer). Tissue for cell injection was taken from the rostral third of the ventral bank of the superior temporal sulcus (cytoarchitectural area TEa of Seltzer & Pandya (1978) and TEad(s) of Yukie (1997)) and from the

 $<sup>\</sup>label{eq:author} \ensuremath{^*\!Author}\ for\ correspondence\ (g.elston@vthrc.uq.edu.au).$ 



Figure 1. Photomicrographs of layer III pyramidal neurons in area STP that were injected with Lucifer yellow and processed for a light-stable diaminobenzidine reaction product, revealing the cellular anatomy in fine detail. Scale bar,  $100 \,\mu$ m.

middle third of the dorsal bank of the superior temporal sulcus (area STP of Bruce *et al.* (1981), rSTP of Hikosaka *et al.* (1988), STPp of Felleman & Van Essen (1991) and TPOr of Cusick *et al.* (1995)).

The methods of cell injection and morphological analysis have been detailed previously (Elston & Rosa 1997; Elston *et al.* 1997; see also Buhl & Schlote 1987; Einstein 1988; Buhl & Singer 1989). Briefly, 250 µm thick cortical slices cut tangential to the cortical surface were labelled with 4,6 diamidino-2phenylindole (DAPI, Sigma D9542), which enabled the distinction to be made between the larger, less densely spaced cell bodies of layer III pyramidal cells and the small, densely packed cell bodies of spiny multipolar cells in layer IV (see fig. 2 of Elston & Rosa (1997)). Neurons were injected with Lucifer yellow (Sigma L 0259) by continuous hyperpolarizing current (up to 100 nA). The tissue was processed with an antibody to

Proc. R. Soc. Lond. B (1999)

Lucifer yellow (raised by Dr D. Pow), biotinylated and processed for a light-stable reaction product (3,3'-diaminobenzidine, Sigma D8001; figure 1).

In order to be included in the analysis, neurons had to be of the pyramidal type, had to be completely filled and had to have their complete basal dendritic arbor contained within the section (for details see Elston *et al.* 1996). All neurons with an unambiguous apical dendrite were analysed, including modified pyramidal neurons (e.g. DeFelipe & Fariñas 1992). Neurons were drawn with the aid of a camera lucida, scanned and analysed using standard features of NIH-IMAGE software (NIH Research Services, Bethesda). Sholl (1953) analysis was performed in the tangential plane to study the complexity of dendritic fields at different distances from the cell body (Elston & Rosa 1997). Spine densities were determined by counting the number of spines per 10  $\mu$ m of dendrite (as seen with a ×100 Zeiss oil-immersion objective) of different layer III pyramidal neurons. Cell bodies were also drawn at this magnification to test for differences between cross-sectional areas and for correlation between cell body and basal dendritic field areas. Statistical comparisons were made using Statview SE software (Abacus Concepts, Berkeley, CA) for the Macintosh.

## 3. RESULTS

Two hundred and fifty-five neurons were injected in layer III of areas TE and STP of the temporal lobe. One hundred and twenty-seven of these neurons were deemed to be well-filled pyramidal neurons and were the subject of further analyses. Neurons were sampled in areas TE and STP in both cases (for case MF2, area TE n=29 and area STP n=38, and for case DM4, area TE n=21 and area STP n=39). The results were consistent in the two cases, revealing that layer III pyramidal cells in areas STP have larger basal dendritic fields and branch more profusely than those in area TE. Moreover, cells in areas STP and TE are much larger, more complex and spine dense than those previously studied in visual areas of the macaque monkey.

#### (a) Basal dendritic field morphology

For both cases MF2 and DM4, two-tailed unpaired t-tests revealed that neurons in area STP had significantly larger basal dendritic fields than those in area TE  $(t_{65} = 3.13, p < 0.05 \text{ and } t_{58} = 5.52, p < 0.05, \text{ respectively};$ figure 2a). Moreover, ANOVAS (two-way repeated measures,  $2 \times 10$  design) on the results of Sholl (1953) analyses in cases MF2 and DM4, revealed that the number of branches in the basal dendritic fields of layer III pyramidal neurons in area STP was significantly greater than that in area TE  $(F_{679} = 211.9 \text{ and } p < 0.01, \text{ and } F_{601} = 46.5$ and p < 0.01, respectively). Comparison with data from visual areas of the occipital, parietal and temporal lobes (Elston & Rosa 1997, 1998a) revealed that layer III neurons in areas TE and STP have much larger basal dendritic fields with many more bifurcations than those in areas V1, V2, MT or V4 (figures 2 and 3). The extent of these differences is made clear by comparing, for example, the neuron with the smallest basal dendritic field in areas STP  $(1.09 \times 10^5 \,\mu\text{m}^2)$  and TE  $(0.82 \times 10^5 \,\mu\text{m}^2)$  with the largest neuron sampled in the corresponding layer in area V1  $(0.51 \times 10^5 \,\mu m^2)$ .

#### (b) Spine density

Spine densities were calculated for layer III pyramidal neurons sampled in case DM4, an 18-month-old monkey, to allow comparison with previously published data in other visual areas collected from animals of 14-28 months (Elston & Rosa 1997, 1998a). This choice was made in view of the well-established decrease in spine density with ageing (e.g. Lund et al. 1977). The basal dendrites of layer III pyramidal neurons in areas TE and STP are clearly more spinous than those in other visual areas previously studied (figure 4). The peak spine density in areas STP and TE was of the order of 23 spines per 10 µm of dendrite, at approximately one-third the distance from the soma to the distal tips, much greater than that of neurons in areas V1 (seven spines per  $10\,\mu m$ ), V2 (seven spines per  $10 \,\mu$ m), MT (eight spines per  $10 \,\mu$ m) and V4 (13 spines per  $10 \,\mu m$ ; figure 2).

## (c) Total number of dendritic spines

By combining data from the Sholl (1953) analyses and spine densities obtained from the 18-month-old animal, we were able to estimate the number of spines on the basal dendritic field of the 'average' layer III pyramidal neuron in each area. These analyses revealed that layer III pyramidal neurons in area STP had, on average, 8337 spines on their basal dendritic fields, more than the corresponding estimate for area TE (7260 spines). These estimates are considerably higher than those obtained for layer III pyramidal neurons in other visual areas (Elston & Rosa 1997, 1998*a*) such as areas V1 (643; average of the number of spines on the 'average' layer III pyramidal neurons in the blobs and interblobs), V2 (1139), MT (2077) and V4 (2429). It should be noted, however, that these estimates are a best approximation and, therefore, the absolute values should be viewed cautiously as they are most likely to be underestimates. Nonetheless, as a relative measure, these estimations provide a sound indication of differences in the numbers of spines of layer III pyramidal neurons in cortical areas involved in visual processing.

#### (d) Somal areas

We also observed a significant difference in crosssectional area of cell bodies of layer III pyramidal neurons between areas STP and TE (for case MF2  $t_{65}=2.67$ , p < 0.05, and for case DM4,  $t_{58}=3.6$ , p < 0.05). The cell bodies of neurons in area STP tended to be larger than those of neurons in area TE. Moreover, the cross-sectional area of the cell bodies of layer III pyramidal neurons in areas TE and STP tended to be larger than those in other visual areas previously studied in the occipital, parietal and temporal lobes (figure 2*d*; see also Elston & Rosa 1997, 1998*a*).

#### 4. DISCUSSION

The temporal cortex is involved in various aspects of visual function, including object recognition and memory, and the response characteristics of temporal cortex neurons are strongly influenced by behavioural state, including attention (Gross et al. 1969, 1972; Desimone & Gross 1979; Desimone et al. 1984; Rolls 1992; Miyashita et al. 1993a, b). The present study is restricted to two areas of the temporal cortex, areas TE and STP. Area TE receives its main 'feedforward' projections from areas such as V4 and TEO and projects to, among other targets, memoryrelated areas of the medial temporal and parahippocampal cortices. Based on connections and response properties, area TE is considered to be a high-order area of the occipitotemporal stream (for reviews see Maunsell & Newsome 1987; Felleman & Van Essen 1991; Gross et al. 1993; Tanaka 1996; Yukie 1997). The response properties and connections of neurons in area STP suggest that cells in this area perform analyses based on more diverse sets of inputs: not only do they receive projections from visual areas, they also integrate somatosensory and auditory information (for reviews, see Gross et al. 1993; Cusick 1997; Graziano & Gross 1997). Despite differences in opinion on boundaries, function and connectivity of different areas in the superior temporal sulcus, most agree that neurons in these areas are involved in 'global'



Figure 2. Graphs of (*a*) basal dendritic field areas, (*b*) Sholl (1953) analyses, (*c*) spine densities, and (*d*) somal areas for neurons in areas V1, V2, MT, V4, TE and STP. Data for area V1 represent cells found in the blobs of the middle and upper layer III of Hassler (1966). Values illustrated for areas TE and STP represent a combined data set for both animals studied, except in the case of spine densities (*c*) which were determined (per 10  $\mu$ m of dendrite) in an 18-month-old animal. Note that layer III pyramidal neurons in areas TE and STP have larger basal dendritic fields (*a*) which have more branches (*b*) and greater spine density (*c*) than those in areas V1, V2, MT and V4. The degree of variation in the size of their cell bodies (*d*) is less marked amongst visual areas of the parietal and temporal lobes, as compared to areas V1 and V2. Error bars are standard deviations.

Proc. R. Soc. Lond. B (1999)



Figure 3. Drawings of layer III pyramidal neurons in visual areas V1, V2, MT, V4, TE and STP, as seen in the plane of section tangential to the cortical layers. Illustrated cells were selected for having a basal dendritic field area close to the mean for each visual area. Even in this small selection of cells it can be seen that there are differences in the number of bifurcations in the basal dendritic fields of cells in the different areas. Statistical analyses of the results of Sholl (1953) analyses for the entire sample of cells in each area revealed these differences to be significant. Cells are skeletonized images and spines are not illustrated. The insert shows the parts of the brain from which blocks of tissue were excised.

visual analyses, as opposed to local feature extraction, and are strongly influenced by the animal's behavioural state, including attention.

Conventional wisdom based on cytoarchitectural studies using the Nissl stain has assumed that neurons in different extrastriate areas are similar in their morphologies and the differences in their neuronal response properties have largely been attributed to different patterns of connections (Zeki 1978). However, recent studies have revealed large-scale variation in cell morphology between different cortical areas involved in visual processing (Lund *et al.* 1993; Elston *et al.* 1996; Elston & Rosa 1997, 1998*a*,*b*,*c*; Peters *et al.* 1997; Jelinek *et al.* 1999). The present results reveal marked variation in the morphology of neurons in visual areas of the superior

eurons in<br/>norpholo-<br/>nse prop-<br/>a patterns(see Elston & Rosa 1997; figure 3). Furthermore, the<br/>basal dendritic fields of layer III pyramidal neurons in<br/>area TE have, on average, 1.5 times the number of spines<br/>found in those of cells in area TEO, another temporal<br/>area involved in the object recognition pathway (see<br/>Elston & Rosa 1998a). Indeed, comparison of the present<br/>data with those obtained in our previous studies of<br/>neurons in visual areas of the occipital lobe reveals that<br/>pyramidal cell morphology varies to an even greater<br/>extent than previously reported. For example, the basal<br/>dendritic fields of layer III pyramidal neurons in area

temporal sulcus. For example, the 'average' layer III

pyramidal neuron in area STP has fourfold more spines

on its basal dendritic field than that in area MT and 3.2-

fold more spines than that in cytoarchitectural area 7a



Figure 4. High-power photomicrographs of horizontally projecting basal dendrites of layer III pyramidal neurons in areas V1 (left column) and TE (right column). In all examples the cell body is located to the left and the distal tips of the dendrites to the right. Scale bar,  $50 \,\mu$ m.

STP cover, on average, an area more than six times that of neurons in area V1 and have, on average, four times more branches. The differences in the size and complexity of the basal dendritic fields, in conjunction with differences in the spine density, result in spectacular differences in the total number of dendritic spines in the basal dendritic field of the 'average' neuron in the different visual areas (e.g. 13 times more spines in the basal dendritic fields of area STP cells as compared with area V1).

Following Gray's (1959) hallmark discovery that synapses could be divided into two morphological types, various groups have shown that dendritic spines are the principal site for asymmetrical synapses in the neocortex (e.g. Jones & Powell 1969), the presynaptic terminals of which have since been shown to contain the excitatory transmitter glutamate (DeFelipe et al. 1988; Kharazia & Weinberg 1993). Moreover, although variation has been reported, it is generally accepted that, in the visual cortex, each cortical dendritic spine receives one excitatory input and models of cortical processing make this assumption (e.g. DeFelipe 1997). The variation in the total number of basal dendritic spines of layer III pyramidal neurons in different cortical visual areas, in conjunction with the difference in the extent of intrinsic connectivity (Amir et al. 1993; Lund et al. 1993; Fujita & Fujita 1996) and the number of sources of corticocortical inputs (e.g. Felleman & Van Essen 1991), suggest that the number and diversity of excitatory inputs which can be integrated by single neurons differs markedly between areas. Recent observations which revealed a marked increase in the density of supragranular glutamate (GluR2/3) receptor subunits with rostral progression through visual areas of the occipitotemporal pathway (Xu et al. 1997) support this idea. Furthermore, the majority of GABAergic inhibitory inputs in the sensory cortex are found on dendrites and dendritic spines (Beaulieu & Colonnier 1985; Beaulieu & Somogyi 1990; Beaulieu *et al.* 1992). Therefore, differences in the total dendritic length and number of dendritic spines of layer III pyramidal neurons in the different cortical areas would most likely result in the integration of different numbers of inhibitory inputs. The greater number and diversity of inputs to neurons in areas TE and STP may be important for the generation of complex stimulus selectivities and the context dependency reported for cells in these areas (e.g. Perrett *et al.* 1982, 1985*a,b*, 1990; Desimone *et al.* 1984; Mistlin & Perrett 1990; Desimone 1991; Hietanen & Perrett 1993; Oram *et al.* 1993; Oram & Perrett 1996; Baker *et al.* 1988).

We would like to thank Dr David Vaney for generously allowing us access to his laboratory, Dr Soumya Ghosh for providing some of the tissue samples and Dr David Pow for providing his superb antibody to Lucifer yellow. Supported by project grants 961144 and 971113 from the National Health and Medical Research Council of Australia.

#### REFERENCES

- Amir, Y., Harel, M. & Malach, R. 1993 Cortical hierarchy reflected in the organization of intrinsic connections in macaque monkey visual cortex. *J. Comp. Neurol.* 334, 19–64.
- Baker, C. I., Keysers, C. & Perrett, D. I. 1998 Temporal cortex and object permanence: cells responsive to people occluded from sight. *Eur. J. Neurosci.* 10, 235.
- Beaulieu, C. & Colonnier, M. 1985 A laminar analysis of the number of round-asymmetrical and flat-symmetrical synapses on spines, dendritic trunks, and cell bodies in area 17 of the cat. *J. Comp. Neurol.* 231, 180–189.

- Beaulieu, C. & Somogyi, P. 1990 Targets and quantitative distribution of GABAergic synapses in the visual cortex of the cat. *Eur. J. Neurosci.* 2, 296–303.
- Beaulieu, C., Kisvárday, Z., Somogyi, P., Cynader, M. & Cowey, A. 1992 Quantitative distribution of GABA-immunopositive and -immunonegative neurons and synapses in the monkey striate cortex (area 17). *Cerebr. Cortex* 2, 295–309.
- Bruce, C. J., Desimone, R. & Gross, C. G. 1981 Visual properties of neurons in a polysensory area in superior temporal sulcus of the macaque. *J. Neurophysiol.* 46, 369–384.
- Buhl, E. H. & Schlote, W. 1987 Intracellular Lucifer yellow staining and electronmicroscopy of neurones in slices of fixed epitumourous human cortical tissue. *Acta Neuropathol.* 75, 140–146.
- Buhl, E. H. & Singer, W. 1989 The callosal projection in cat visual cortex as revealed by a combination of retrograde tracing and intracellular injection. *Exp. Brain Res.* **75**, 470–476.
- Cusick, C. G. 1997 The superior temporal polysensory region in monkeys. In *Cerebral cortex. 12. Extrastriate cortex in primates* (ed. K. Rockland, J. H. Kaas & A. Peters), pp. 435–468. New York: Plenum.
- Cusick, C. G., Seltzer, B., Cola, M. & Griggs, E. 1995 Chemoarchitectonics and corticocortical terminations within the superior temporal sulcus of the rhesus monkey: evidence for subdivisions of superior temporal polysensory cortex. *J. Comp. Neurol.* **360**, 513–535.
- DeFelipe, J. 1997 Microcircuits of the brain. In *Biological and artificial computation: from neuroscience to technology. Lecture notes in computer science* (ed. J. Mira, R. Moreno-Diaz & J. Cabestany), pp. 1–14. Berlin: Springer.
- DeFelipe, J. & Fariñas, I. 1992 The pyramidal neuron of the cerebral cortex: morphological and chemical characteristics of the synaptic inputs. *Prog. Neurobiol.* 39, 563–607.
- DeFelipe, J., Conti, F., Van Eyck, S. L. & Manzoni, T. 1988 Demonstration of glutamate-positive axon terminals forming asymmetric synapses in cat neocortex. *Brain Res.* 455, 162–165.
- Desimone, R. 1991 Face-selective cells in the temporal cortex of monkeys. J. Cogn. Neurosci. 3, 1–8.
- Desimone, R. & Gross, C. G. 1979 Visual areas in the temporal cortex of the macaque. *Brain Res.* 178, 363–380.
- Desimone, R., Albright, T. D., Gross, C. G. & Bruce, C. 1984 Stimulus-selective properties of inferior temporal neurons in the macaque. *J. Neurosci.* 4, 2051–2062.
- Einstein, G. 1988 Intracellular injection of Lucifer yellow into cortical neurons in lightly fixed sections and its application to human autopsy material. *J. Neurosci. Meth.* **26**, 95–103.
- Elston, G. N. & Rosa, M. G. P. 1997 The occipitoparietal pathway of the macaque monkey: comparison of pyramidal cell morphology in layer III of functionally related cortical visual areas. *Cerebr. Cortex* **7**, 432–452.
- Elston, G. N. & Rosa, M. G. P. 1998a Morphological variation of layer III pyramidal neurones in the occipitotemporal pathway of the macaque monkey visual cortex. *Cerebr. Cortex* 8, 278–294.
- Elston, G. N. & Rosa, M. G. P. 1998b Complex dendritic fields of pyramidal cells in the frontal eye field of the macaque monkey: comparison with parietal areas 7a and LIP. *NeuroReport* **9**, 127–131.
- Elston, G. N. & Rosa, M. G. P. 1998c Morphological variation of layer V pyramidal neurones in visual areas of the temporal lobe of the macaque monkey. *Proc. Aust. Neurosci. Soc. Abstr.* 9, 25.
- Elston, G., Rosa, M. G. P. & Calford, M. B. 1996 Comparison of dendritic fields of layer III pyramidal neurones in striate and extrastriate visual areas of the marmoset: a Lucifer yellow intracellular injection study. *Cerebr. Cortex* 6, 807–813.
- Elston, G. N., Pow, D. V. & Calford, M. B. 1997 Neuronal composition and morphology in layer IV of two vibrissal barrel subfields of rat cortex. *Cerebr. Cortex* **7**, 422–431.

- Felleman, D. J. & Van Essen, D. C. 1991 Distributed hierarchical processing in primate cerebral cortex. *Cerebr. Cortex* 1, 1–47.
- Fujita, I. & Fujita, T. 1996 Intrinsic connections in the macaque inferior temporal cortex. *J. Comp. Neurol.* 368, 467–486.
- Gray, E. G. 1959 Axo-somatic and axo-dendritic synapses of the cerebral cortex; an electron microscope study. *J. Anat.* 93, 420–433.
- Graziano, M. S. A. & Gross, C. G. 1997 Vision, movement, and the monkey brain. In *The association cortex: structure and function* (ed. H. Sakata, A. Mikami & J. Fuster), pp. 219–232. Amsterdam: Harwood Academic Publishers.
- Gross, C. G., Bender, D. B. & Rocha-Miranda, C. E. 1969 Visual receptive fields of neurons in inferotemporal cortex of the monkey. *Science* 166, 1303–1306.
- Gross, C. G., Rocha-Miranda, C. E. & Bender, D. B. 1972 Visual properties of neurones in inferotemporal cortex of the monkey. *J. Neurophysiol.* 35, 96–111.
- Gross, C. G., Rodman, H. R., Gochin, P. M. & Colombo, M. W. 1993 Inferior temporal cortex as a pattern recognition device. In *Computational learning and recognition: Proceedings of the 3rd NEC Research Symposium* (ed. E. Baum), pp. 44–73. Philadelphia, PA: Society for Industrial and Applied Mathematics.
- Hassler, R. 1966 Comparative anatomy of the central visual system in day- and night-active primates. In *Evolution of the forebrain* (ed. R. Hassler & H. Stephen), pp. 419–434. Stuttgart: Thieme.
- Hendry, S. H. C. & Calkins, D. J. 1998 Neuronal chemistry and functional organization in the primate visual system. *Trends Neurosci.* 21, 344–349.
- Hietanen, J. K. & Perrett, D. I. 1993 Motion sensitive cells in the macaque superior temporal polysensory area. I. Lack of response to the sight of the animal's own limb movement. *Exp. Brain Res.* 93, 117–128.
- Hikosaka, K., Iwai, E., Saito, H. & Tanaka, K. 1988 Polysensory properties of neurones in the anterior bank of the caudal superior temporal sulcus of the macaque monkey. *J. Neurophysiol.* **60**, 1615–1637.
- Jelinek, H. F., Elston, G. N. & Rosa, M. G. P. 1999 Fractal analyses of pyramidal neurones in macaque visual cortex. *Proc. Aust. Neurosci. Soc. Abstr.* 10, 186.
- Jones, E. G. & Powell, T. P. S. 1969 Morphological variations in the dendritic spines of the neocortex. *J. Cell Sci.* 5, 509–529.
- Kharazia, V. N. & Weinberg, R. J. 1993 Glutamate in terminals of the thalamocortical fibers in rat somatic sensory cortex. *Neurosci. Lett.* 157, 162–166.
- Lund, J. S., Boothe, R. G. & Lund, R. D. 1977 Development of neurons in the visual cortex (area 17) of the monkey (*Macaca nemistrema*): a Golgi study from fetal day 127 to postnatal maturity. *J. Comp. Neurol.* **176**, 149–188.
- Lund, J., Yoshioka, T. & Levitt, J. B. 1993 Comparison of intrinsic connectivity in different areas of macaque monkey cerebral cortex. *Cerebr. Cortex* 3, 148–162.
- Maunsell, J. H. R. & Newsome, W. T. 1987 Visual processing in monkey extrastriate cortex. A. Rev. Neurosci. 10, 363–401.
- Mistlin, A. J. & Perrett, D. I. 1990 Visual and somatosensory processing in the macaque temporal cortex: the role of 'expectation'. *Exp. Brain Res.* **82**, 437–450.
- Miyashita, Y., Okuno, H. & Hasegawa, I. 1993*a* Tuning and association—neural memory mechanisms of complex visual forms in monkey temporal cortex. *Biomed. Res.* 14, 89–94.
- Miyashita, Y., Date, A. & Okuno, H. 1993b Configurational encoding of complex visual forms by single neurons of monkey temporal cortex. *Neuropsychologia* 31, 1119–1131.
- Oram, M. W. & Perrett, D. I. 1996 Integration of form in the anterior superior temporal polysensory area (STPa) of the macaque monkey. *J. Neurophysiol.* 76, 109–129.
- Oram, M. W., Perrett, D. I. & Hietanen, J. K. 1993 Directional tuning of motion sensitive cells in the anterior superior

temporal polysensory area of the macaque. *Exp. Brain Res.* 97, 274–294.

- Perrett, D. I., Rolls, E. T. & Caan, W. 1982 Visual neurons responsive to faces in the monkey temporal cortex. *Exp. Brain Res.* 47, 329–342.
- Perrett, D. I., Smith, P. A. J., Potter, D. D., Mistlin, A. J., Head, A. S., Milner, A. D. & Jeeves, M. A. 1985a Visual cells in the temporal cortex sensitive to face view and gaze direction. *Proc. R. Soc. Lond.* B 223, 293–317.
- Perrett, D. I., Smith, P. A. J., Mistlin, A. J., Chitty, A. J., Head, A. S., Potter, D. D., Broennimann, R., Milner, A. D. & Jeeves, M. A. 1985b Visual analyses of body movements by neurons in the temporal cortex of the macaque monkey: a preliminary report. *Behav. Brain Res.* 16, 153–170.
- Perrett, D. I., Harries, M. H., Mistlin, A. J., Hietanen, J. K., Benson, P. J., Bevan, R., Thomas, S., Oram, M. W., Ortega, J. & Brierley, K. 1990 Social signals analyzed at the single cell level: someone is looking at me, something touched me, something moved! *Int. J. Comp. Psychol.* 4, 25–55.
- Peters, A., Cifuentes, J. & Sethares, C. 1997 The organization of pyramidal cells in area 18 of the rhesus monkey. *Cerebr. Cortex* 7, 405–421.
- Rolls, E. T. 1992 Neurophysiological mechanisms underlying face processing within and beyond the temporal cortical visual areas. *Phil. Trans. R. Soc. Lond.* B **335**, 11–21.
- Rosa, M. G. P. 1997 Visuotopic organization of primate extrastriate cortex. In *Cerebral cortex. 12. Extrastriate cortex in primates* (ed. K. Rockland, J. H. Kaas & A. Peters), pp. 127–204. New York: Plenum.

- Seltzer, B. & Pandya, D. N. 1978 Afferent cortical connections of the superior temporal sulcus and surrounding cortex in the rhesus monkey. *Brain Res.* 149, 1–24.
- Sholl, D. A. 1953 Dendritic organization in the neurons of the visual and motor cortices of the cat. *J. Anat.* 87, 387–406.
- Tanaka, K. 1996 Inferotemporal cortex and object vision. A. Rev. Neurosci. 19, 109–139.
- Ungerleider, L. G. & Mishkin, M. 1982 Two cortical systems. In Analysis of visual behavior (ed. D. J. Ingle, M. A. Goodale & R. J. W. Mansfield), pp. 549–586. Cambridge, MA: MIT.
- Weller, R. E. 1988 Two cortical visual systems in Old World and New World primates. In *Progress in brain research* (ed. T. P. Hicks & G. Benedek), pp. 293–306. Oxford: Elsevier.
- Xu, L.-H., Tanigawa, H. & Fujita, I. 1997 Distribution of AMPA-type glutamate receptors along the ventral visual cortical pathway in the macaque: a gradient reflecting the cortical hierarchy. *Soc. Neurosci. Abstr.* 23, 2062.
- Yukie, M. 1997 Organization of visual afferent connections to inferior temporal cortex, area TE, in the macaque monkey. In *The association cortex: structure and function* (ed. H. Sakata, A. Mikami & J. Fuster), pp. 247–258. Amsterdam: Harwood Academic Publishers.
- Zeki, S. M. 1978 Uniformity and diversity of structure and function in rhesus monkey prestriate cortex. *J. Physiol. (Lond.)* 277, 273–290.
- As this paper exceeds the maximum length normally permitted, the authors have agreed to contribute to production costs.