

# Fruits and vegetables that are sources for lutein and zeaxanthin: the macular pigment in human eyes

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## Abstract

**Background**—It has been suggested that eating green leafy vegetables, which are rich in lutein and zeaxanthin, may decrease the risk for age related macular degeneration. The goal of this study was to analyse various fruits and vegetables to establish which ones contain lutein and/or zeaxanthin and can serve as possible dietary supplements for these carotenoids.

**Methods**—Homogenates of 33 fruits and vegetables, two fruit juices, and egg yolk were used for extraction of the carotenoids with hexane. Measurement of the different carotenoids and their isomers was carried out by high performance liquid chromatography using a single column with an isocratic run, and a diode array detector. **Results**—Egg yolk and maize (corn) contained the highest mole percentage (% of total) of lutein and zeaxanthin (more than 85% of the total carotenoids). Maize was the vegetable with the highest quantity of lutein (60% of total) and orange pepper was the vegetable with the highest amount of zeaxanthin (37% of total). Substantial amounts of lutein and zeaxanthin (30–50%) were also present in kiwi fruit, grapes, spinach, orange juice, zucchini (or vegetable marrow), and different kinds of squash. The results show that there are fruits and vegetables of various colours with a relatively high content of lutein and zeaxanthin.

**Conclusions**—Most of the dark green leafy vegetables, previously recommended for a higher intake of lutein and zeaxanthin, have 15–47% of lutein, but a very low content (0–3%) of zeaxanthin. Our study shows that fruits and vegetables of various colours can be consumed to increase dietary intake of lutein and zeaxanthin.

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filter, to protect the underlying tissues from phototoxic damage.<sup>4–5</sup> This has been proposed as a factor, especially in the pathophysiology of age related macular degeneration (ARMD).<sup>6</sup> Hammond *et al*<sup>7</sup> found a correlation between macular pigment density and iris colour, and suggested a correlation between dietary carotenoid intake and macular pigment density. Such a correlation was demonstrated in two subjects by Landrum *et al*<sup>8</sup> and in a larger group by Hammond *et al*.<sup>9</sup>

In recent years several epidemiological studies reported the inverse correlation between micronutrients in plasma and the risk of certain eye disorders thought to be related to free radical generation. Jacques *et al*<sup>10</sup> suggested a relation between intake of antioxidant vitamins and carotenoids and the prevention of cataract. In 1993 the Case Control Study Group for Eye Diseases of the United States evaluated the relation between a decreased plasma level of lutein and zeaxanthin and the risk of the exudative form of neovascular ARMD.<sup>11</sup> Seddon *et al* reported the results of further investigations from the Case Control Study Group for Eye Diseases, which showed that a high dietary intake of carotenoids, in particularly those in dark green leafy vegetables, was associated with a 43% lower risk for ARMD.<sup>12</sup> West *et al*<sup>13</sup> suggested a protective effect of high plasma  $\alpha$  tocopherol values of ARMD, and demonstrated a correlation between ARMD and an antioxidant index consisting of ascorbic acid,  $\alpha$  tocopherol, and  $\beta$  carotene. In 1995 Mares-Perlman *et al*<sup>14</sup> reported that a low plasma level of lycopene was associated with ARMD. The role of carotenoids in protection from ARMD was discussed in reviews by Snodderly<sup>15</sup> and Landrum *et al*.<sup>16</sup>

Fruits and vegetables are the most important source of carotenoids in the human diet, and knowledge about this is important for preventive medicine. Several publications report the qualitative and quantitative content of carotenoids in different fruits and vegetables.<sup>17–21</sup> However, many of the procedures used did not allow the separation of lutein and zeaxanthin, and they are typically reported as a single group. To date, there is no comprehensive information on the separate content of lutein and zeaxanthin in fruits and vegetables. Since the macular pigments are not evenly distributed throughout the human retina,<sup>1,2</sup> knowledge about this may be valuable. The goal of our study was to analyse a number of fruits and vegetables for the content of each macular pigment. This may help to give guidelines as to

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The two major carotenoids in the human macula and retina are lutein and zeaxanthin,<sup>1,2</sup> and they are often referred to as xanthophylls, or macular pigment. Handelman *et al*<sup>1</sup> found a fivefold higher content of these carotenoids in the macula compared with the peripheral retina. Snodderly *et al*<sup>3</sup> reported that macaque monkeys, which have foveal cone types like the human retina, have a consistent pattern of more zeaxanthin than lutein in the foveal centre. In the periphery zeaxanthin declines more rapidly than lutein, so that lutein becomes dominant.<sup>2</sup> The carotenoids are thought to function as antioxidants and/or as a blue light

which food products can help decrease the risk of cataract and ARMD.

## Methods

### SOURCES OF CAROTENOID STANDARDS

Apo-10'-carotenal and zeaxanthin were a generous gift by Hoffmann-La Roche, Nutley, NJ, USA. Lutein,  $\beta$  carotene, and *O*-ethyl-hydroxylamine were purchased from Fluka, Ronkonkoma, NY, USA. Apo-10'-carotenal-methyl oxime was used as internal standard, and was synthesised as described elsewhere.<sup>22</sup>

### SAMPLE PREPARATION

For the extraction procedure a 5 g piece of fruit or vegetable was used. The tissue was homogenised with a pestle in a mortar containing 1 ml of phosphate buffered saline (PBS) (10 mM  $\text{NaH}_2\text{PO}_4$ , water with 0.15 N NaCl titrated to pH 4.7). Sodium sulphate and sodium chloride were from Fisher Scientific, Fair Lawn, NJ, USA. The homogenate was transferred to a 7 ml vial and 1 ml of methanol, containing butylated hydroxytoluene (0.5 mg/ml) (BHT Sigma Chemical Co, St Louis, MO, USA). BHT was added before mixing the sample for 30 seconds. Ethanol was from Midwest Grain Products Co, Weston, MI, USA. For fruit juices and egg yolk, 2 ml was mixed with 1 ml methanol BHT (0.5 mg/ml). To extract the carotenoids from the samples, 2 ml of hexane and 1 ml of hexane containing the internal standard was added to the samples, and they were vortexed for 3 minutes, and centrifuged at  $1800 \times g$  for 3 minutes. The upper layer of the

sample containing the carotenoids was collected and passed through a Pasteur pipette, prepared with ~150 g of anhydrous sodium sulphate to remove any traces of moisture. The hexane extraction was repeated once with 2 ml of hexane. The extracted carotenoids in hexane are stable for several weeks at  $-20^\circ\text{C}$ . Since the amount of extracted carotenoids was different for each of the fruits and vegetables the absorbance of 1 ml of the obtained hexane phase was measured in a Shimadzu Spectrophotometer UV-2101PC. To detect the carotenoids by high performance liquid chromatography (HPLC), a certain volume of the hexane phase (100  $\mu\text{l}$ –1 ml), corresponding to an optical density of 0.3, was evaporated under a steam of argon. The residue was redissolved in 60  $\mu\text{l}$  methanol and 20  $\mu\text{l}$  was injected on the HPLC.

### HIGH PERFORMANCE LIQUID CHROMATOGRAPHY

The HPLC consisted of a Beckman System Gold Programmable Solvent Module 125 (Beckman Instruments Inc, Palo Alto, CA, USA), a Beckman System Gold Diode Array Detector Module 168, and a Rheodyne 7725i Injector valve with a 20  $\mu\text{l}$  injection loop (Rheodyne Inc, Cotati, CA, USA). The diode array detector was set in channel A at wavelength 450 nm to detect the carotenoids. The second channel B was set at 300 nm to detect retinols and tocopherols. A reverse phase 5  $\mu\text{m}$   $\text{C}_{18}$  column, 250  $\times$  4.6 mm (VYDAC, 218TP54, Hesperia, CA, USA), with a precolumn (40  $\times$  4.6 mm) was used. For

Table 1 Carotenoids in fruits and vegetables

	Neoxanthins and violaxanthins	Lutein and zeaxanthin	Lutein	Zeaxanthin	Cryptoxanthins	Lycopenes	$\alpha$ carotene	$\beta$ carotene
Egg yolk	8	89	54	35	4	0	0	0
Maize (corn)	9	86	60	25	5	0	0	0
Kiwi	38	54	54	0	0	0	0	8
Red seedless grapes	23	53	43	10	4	5	3	16
Zucchini squash	19	52	47	5	24	0	0	5
Pumpkin	30	49	49	0	0	0	0	21
Spinach	14	47	47	0	19	4	0	16
Orange pepper	4	45	8	37	22	0	8	21
Yellow squash	19	44	44	0	0	0	28	9
Cucumber	16	42	38	4	38	0	0	4
Pea	33	41	41	0	21	0	0	5
Green pepper	29	39	36	3	20	0	0	12
Red grape	27	37	33	4	29	0	1	6
Butternut squash	24	37	37	0	34	0	5	0
Orange juice	28	35	15	20	25	0	3	8
Honeydew	18	35	17	18	0	0	0	48
Celery (stalks, leaves)	12	34	32	2	40	1	13	0
Green grapes	10	31	25	7	52	0	0	7
Brussels sprouts	20	29	27	2	39	0	0	11
Scallions	32	29	27	3	35	4	0	0
Green beans	27	25	22	3	42	0	1	5
Orange	36	22	7	15	12	11	8	11
Broccoli	3	22	22	0	49	0	0	27
Apple (red delicious)	22	20	19	1	23	13	5	17
Mango	52	18	2	16	4	6	0	20
Green lettuce	33	15	15	0	36	0	16	0
Tomato juice	0	13	11	2	2	57	12	16
Peach	20	13	5	8	8	0	10	50
Yellow pepper	86	12	12	0	1	0	1	0
Nectarine	18	11	6	6	23	0	0	48
Red pepper	56	7	7	0	2	8	24	3
Tomato (fruit)	0	6	6	0	0	82	0	12
Carrots	0	2	2	0	0	0	43	55
Cantaloupe	9	1	1	0	0	3	0	87
Dried apricots	2	1	1	0	9	0	0	87
Green kidney beans	72	0	0	0	28	0	0	0

The content of the major carotenoids are given in mole%. The amounts of the carotenoids were shown in seven major groups, as neoxanthins and violaxanthins (neoxanthin, violaxanthin, and their related isomers, lutein 5, 6 epoxide), lutein, zeaxanthin, cryptoxanthins ( $\alpha$  cryptoxanthin,  $\beta$  cryptoxanthins, and related isomers), lycopenes (lycopene and related isomers),  $\alpha$  carotene (all *trans*  $\alpha$  carotene and *cis* isomers), and  $\beta$  carotene (all *trans*  $\beta$  carotene and *cis* isomers). Lutein and zeaxanthin are given combined and as single amounts. The data are sorted by the combined amount of lutein and zeaxanthin.

the mobile phase two solvents were prepared. Solvent A consisted of acetonitrile/methanol (85:15, v/v), with 0.01% (w/v) ammonium acetate and pure isopropanol was solvent B. Ammonium acetate was purchased from Baker, Phillipsburg, NJ, USA. Methanol, isopropanol, and acetonitrile were purchased from EM science, Gibbstown, NJ, USA. During the run a mix of 90% solvent A and 10% of solvent B was used. The initial flow rate was set on 0.5 ml/min. At 10 minutes a 5 minute flow gradient ramp was executed, and a final flow rate of 1.2 ml/min was established. At 22 minutes the flow rate was decreased within a 5 minute ramp to the initial setting. The total time of one run was 27 minutes.

### Results

The content of carotenoids for the various fruits and vegetables is shown in Table 1. The data are sorted by looking for the highest mole% of xanthophylls (lutein and zeaxanthin) of total carotenoids. The highest amounts of lutein and zeaxanthin were found in egg yolk (54 mole% of lutein and 35 mole% of zeaxanthin), and in maize (corn) (60 mole% of lutein and 25 mole% of zeaxanthin). In maize we found the highest mole% of lutein among all fruits and vegetables in our study. Lutein also was also the major carotenoid in kiwi fruit (54 mole%), red seedless grapes (53 mole%), zucchini squash (52 mole%), and pumpkin (50 mole%). Zeaxanthin was the major carotenoid in orange pepper (37 mol%), which was also the food with the highest percentage of this carotenoid among all fruits and vegetables (Table 1). It is interesting that green pepper has 36 mole% of lutein and 3 mole% of zeaxanthin, whereas in orange pepper we find a carotenoid composition with only 8 mole% lutein, but 37 mole% of zeaxanthin. In contrast with the green and orange peppers described above, yellow and red pepper are vegetables with a low amount of lutein (12 mole% and 7 mole%) and no zeaxanthin. In carrots, cantaloupe, dried apricots, and green kidney beans almost no lutein and zeaxanthin were found. In green leafy vegetables the amount of lutein in spinach was 47 mole%, in stalks and leaves of celery 34 mole%, in Brussels sprouts 27 mole%, in scallions 27 mole%, in broccoli 22 mole%, and in green lettuce 15 mole%. The amount of zeaxanthin in these vegetables ranged from 0 to 3 mole%.

### Discussion

Carotenoids have many biological effects,<sup>23</sup> and can function as antioxidants to protect eye tissues against free radicals.<sup>4-6</sup> The only source of carotenoids for humans is food, and the carotenoid availability in plasma is critical in long term maintenance of adequate tissue levels. A correlation exists between carotenoid intake and plasma carotenoid concentrations.<sup>24</sup> However, the individual variability in plasma response to carotenoid intake in humans is large.<sup>25</sup> The mechanisms providing the transport of carotenoids from intestine to the tissues are not clearly understood. Scita *et al*<sup>26</sup> proposed that the uptake of carotenoids in cul-

tured small intestine cells of rats takes place in the absence of any receptor regulation. The postprandial incorporation into lipoproteins could be demonstrated in normolipemic subjects for  $\beta$  carotene by Cornwell *et al* in 1962.<sup>27</sup> The transport of the carotenoids in blood, and the distribution to the different tissues is done by lipoproteins. Clevidence and Bieri reported that low density lipoproteins (LDL) carries most of the total carotenoids in plasma, but they also found that individual carotenoids are not uniformly distributed among lipoproteins.<sup>28</sup> In their population, 67% of  $\beta$  carotene was found in LDL but 53% of lutein and zeaxanthin was found in high density lipoproteins (HDL).<sup>28</sup> It might be possible that the retina is able to use this preselection of carotenoids by the lipoproteins to accumulate lutein and zeaxanthin from the plasma carotenoids. Bernstein *et al* recently found an ocular carotenoid binding protein in the bovine retina,<sup>29</sup> which could also play a role in humans. The major carotenoids in the human eye are lutein and zeaxanthin,<sup>1-3 30</sup> and it is not clear yet by which mechanism these carotenoids accumulate in the ocular tissues out of food. However, the amount of macular pigment can be modulated by the diet.<sup>7-9 31 32</sup>

Our results match those from a large analytical study of Mangels *et al*<sup>21</sup> where data from a large number of articles on carotenoids in food from 1971 to 1991 were summarised. However, typically the content of lutein and zeaxanthin was reported as a single number, and there is no easy access to information about the content of the individual macular pigments in fruits and vegetables.<sup>21</sup> The data in Table 1 show that there is a large variation in the amount of lutein and zeaxanthin in fruits and vegetables. Lutein is present in many fruits and vegetables, whereas zeaxanthin is found only in a small number of fruits and vegetables, and in eggs. For example, in maize 85 mole% of the carotenoids are lutein and zeaxanthin. Interestingly, in most dark green vegetables, such as scallions, green lettuce, celery, spinach, and Brussels sprouts, only traces of zeaxanthin, the most prevalent macular pigment,<sup>1 2 30</sup> were found. In our study, the highest amount of zeaxanthin was found in orange pepper, which was not previously analysed.

The highest mole percentage of both lutein and zeaxanthin (89 mole%) was found in egg yolk. Since eggs have a high cholesterol content, a restricted intake of eggs has been recommended for many years, since cholesterol is a risk factor for coronary artery disease. However, in recent years several studies were published showing that a higher intake of cholesterol through the addition of more eggs in the diet, results not only in an increase of serum cholesterol, but also in an increase of HDL cholesterol.<sup>33 34</sup> Since HDL cholesterol is protective against atherosclerosis, extra egg consumption may not change the risk index for ischaemic heart disease based on the cholesterol levels.<sup>33 34</sup> The consumption of eggs could actually be beneficial in order to obtain a higher intake of lutein and zeaxanthin, and since it has no severe adverse effects on cardiac

risk factors, the exclusion of eggs from the diet could be reconsidered.

The two studies of the Eye Disease Case Control Study Group in recent years showed a direct relation between the exudative form of ARMD and a decreased plasma level of lutein and zeaxanthin.<sup>11 12</sup> It was found that a diet rich in dark green leafy vegetables could decrease the risk of ARMD.<sup>12</sup> These data support the hypothesis that a higher intake of lutein and zeaxanthin can prevent this form of ARMD. However, according to our data, this recommendation may be extended to include fruits and vegetables of other colours using Table 1 as a guideline. Recently, Khachik *et al*<sup>25</sup> reported evidence for conversion between lutein and zeaxanthin through several oxidation intermediates, which would suggest that intake of lutein can also raise macular levels of zeaxanthin.

### Conclusion

Our study demonstrated that consumption of fruits and vegetables of various colours would increase dietary intake of lutein and zeaxanthin, and showed that the options are not limited to dark green leafy vegetables, which were previously recommended. In this study no evidence has been produced to show that lutein and zeaxanthin reduce the prevalence of ARMD, and the evidence from other studies is not incontrovertible.

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- Handelman GJ, Dratz EA, Reay CC, *et al*. Carotenoids in the human macula and whole retina. *Invest Ophthalmol Vis Sci* 1988;**29**:850–5.
- Bone RA, Landrum JT, Fernandez L, *et al*. Analysis of the macular pigment by HPLC: retinal distribution and age study. *Invest Ophthalmol Vis Sci* 1988;**29**:843–9.
- Snodderly DM, Handelman GJ, Adler AJ. Distribution of individual macular pigment carotenoids in central retina of macaque and squirrel monkeys. *Invest Ophthalmol Vis Sci* 1991;**32**:268–79.
- Gerster H. Review: Antioxidant protection of the ageing macula. *Age Ageing* 1991;**20**:60–9.
- Schalch W, Weber P. Vitamins and carotenoids—a promising approach to reducing the risk of coronary heart disease, cancer and eye diseases. *Adv Exp Med Biol* 1994;**366**:335–50.
- Young RW. Solar radiation and age-related macular degeneration. *Surv Ophthalmol* 1988;**32**:252–69.
- Hammond BR, Fuld K, Snodderly DM. Iris color and macular pigment density. *Exp Eye Res* 1996;**62**:293–7.
- Landrum JT, Bone RA, Mark HJ, *et al*. A one year study of the macular pigment: The effect of 140 days of a lutein supplement. *Exp Eye Res* 1997;**64**:311–16.
- Hammond BR, Johnson EJ, Russell RM, *et al*. Dietary modification of human macular pigment density. *Invest Ophthalmol Vis Sci* 1997;**38**:1795–801.
- Jacques PF, Chylack LT Jr. Epidemiologic evidence of a role for the antioxidant vitamins and carotenoids in cataract prevention. *Am J Clin Nutr* 1991;**53**:352S–5S.
- Eye Disease Case-Control Study Group. Antioxidant status and neovascular age-related macular degeneration. *Arch Ophthalmol* 1993;**111**:104–9.
- Seddon JM, Ajani UA, Sperduto RD, *et al*. Dietary carotenoids, vitamins A, C, and E, and advanced age-related macular degeneration. Eye Disease Case-Control Study Group. *JAMA* 1994;**272**:1413–20.
- West SK, Vitale S, Hallfrisch J, *et al*. Are antioxidants or supplements protective for age-related macular degeneration? *Arch Ophthalmol* 1994;**112**:222–7.
- Mares-Perlman JA, Brady WE, Klein R, *et al*. Serum antioxidants and age-related macular degeneration in a population-based case-control study. *Arch Ophthalmol* 1995;**113**:1518–23.
- Snodderly DM. Evidence for protection against age-related macular degeneration by carotenoids and antioxidant vitamins. *Am J Clin Nutr* 1995;**62** (suppl):1448S–61S.
- Landrum JT, Bone RA, Kilburn MD. The macular pigment: A possible role in protection from age related macular degeneration. *Adv Pharmacol* 1997;**38**:537–56.
- Khachik F, Beecher GR. Application of a c-45-b-carotene as an internal standard for the quantification of carotenoids in yellow/orange vegetables by liquid chromatography. *J Agric Food Chem* 1987;**35**:732–8.
- Khachik F, Beecher GR. Separation and identification of carotenoids and carotenol fatty acid esters in some squash products by liquid chromatography. 1. Quantification of carotenoids and related esters by HPLC. *J Agric Food Chem* 1988;**36**:929–37.
- Khachik F, Beecher GR, Lusby WR. Separation, identification, and quantification of the major carotenoids in extracts of apricots, peaches, cantaloupe, and pink grapefruit by liquid chromatography. *J Agric Food Chem* 1989;**37**:1465–73.
- Ong ASH, Tee ES. Natural sources of carotenoids from plants and oils. In: Packer L, ed. *Methods in enzymology*. London: Academic Press, 1992;**213**:142–67.
- Mangels AR, Holden JM, Beecher GR, *et al*. Carotenoid content of fruits and vegetables: an evaluation of analytic data. *J Am Diet Assoc* 1993;**93**:284–96. (Published erratum appears in *J Am Diet Assoc* 1993;**93**:527.)
- Sommerburg O, Zang L, van Kuijk FJGM. Simultaneous detection of carotenoids and vitamin E in human plasma. *J Chromatogr* 1997;**695**:209–15.
- Krinsky NI. Actions of carotenoids in biological systems. *Annu Rev Nutr* 1993;**13**:561–87.
- Forman MR, Lanza E, Yong LC, *et al*. The correlation between two dietary assessments of carotenoid intake and plasma carotenoid concentrations: application of a carotenoid food-composition database. *Am J Clin Nutr* 1993;**58**:519–24.
- Micozzi MS. Evaluation of carotenoid intake. *Methods Enzymol* 1993;**214**:17–21.
- Scita G, Aponte GW, Wolf G. Uptake and cleavage of beta-carotene by cultures of rat small intestinal cells and human lung fibroblasts. *Methods Enzymol* 1993;**214**:21–32.
- Cornwell DG, Kruger FA, Robinson HB. Studies on the absorption of beta-carotene and the distribution of total carotenoid in human serum lipoproteins after oral administration. *J Lipid Res* 1962;**3**:65–70.
- Clevidence BA, Bieri JG. Association of carotenoids with human plasma lipoproteins. *Methods Enzymol* 1993;**214**:33–46.
- Bernstein PS, Tsong ED, Rando RR. Isolation of a carotenoid binding protein from bovine retina. *Invest Ophthalmol Vis Sci* 1995;**36**:S5(Abstract)
- Yeum KJ, Taylor A, Guangwen T, *et al*. Measurement of carotenoids, retinols, and tocopherols in human lenses. *Invest Ophthalmol Vis Sci* 1996;**36**:2756–61.
- Malinow MR, Feeney-Burns L, Peterson LH, *et al*. Diet-related macular anomalies in monkeys. *Invest Ophthalmol Vis Sci* 1980;**19**:857–63.
- Hammond BR, Fuld K, Currencecetano J. Macular pigment density in monozygotic twins. *Invest Ophthalmol Vis Sci* 1995;**36**:2531–41.
- Schnohr P, Thomsen OO, Riis Hansen P, *et al*. Egg consumption and high-density-lipoprotein cholesterol. *J Int Med* 1994;**235**:249–51.
- Ginsberg HN, Karmally W, Siddiqui M, *et al*. A dose-response study of the effects of dietary cholesterol on fasting and postprandial lipid and lipoprotein metabolism in healthy young men. *Arterioscler Thromb* 1994;**14**:576–86.
- Khachik F, Bernstein PS, Garland DL. Identification of lutein and zeaxanthin oxidation products in human and monkey retinas. *Invest Ophthalmol Vis Sci* 1997;**38**:1802–11.