

The inflammatory response: friend or enemy for muscle injury?

H Toumi, T M Best

Limiting certain aspects of inflammation may be a useful new treatment for sport related muscle injury

Muscle injury can occur through diverse mechanisms such as mechanical injury, muscular dystrophies, infectious diseases, and biochemical toxicities. Several types of skeletal muscle injury fall into the broad category of sport and exercise induced muscle injury. When exercise involves eccentric muscular contractions, it is associated with overloading of the contractile elements and connective tissues—that is, the force requirement of the muscle exceeds the habitual requirements—and can result in injury to skeletal muscle. It has traditionally been felt that the events following the initial injury, including inflammation, are necessary for optimal repair. The inflammatory response to eccentric exercise as well as stretch injury consist of neutrophilia, neutrophil activation, and the accumulation of neutrophils within

the injured muscle as early as one to two hours. In this early inflammatory stage, cellular debris is removed by the infiltrating neutrophils and is followed by a regenerative response during which satellite cells proliferate to replace the previously damaged and phagocytosed muscle.¹⁻⁴ In addition to phagocytosis, neutrophil invasion and activation can lead to the release of oxygen free radicals and proteases which potentially cause injury.⁵ Neutrophils contain more than 40 hydrolytic enzymes and toxic molecules in their granules and can generate various oxidants such as superoxide anion, hydrogen peroxide, and hypochlorous acid. The NADPH oxidase complex located on activated neutrophils and macrophages can initiate a “respiratory burst” which leads to production of superoxide anion, which can quickly be converted into hydrogen peroxide. In

addition, myeloperoxidase, an enzyme present in neutrophils and macrophages, can generate hypochlorous acid, a highly reactive oxidising agent. Although the exact time course of the appearance of inflammatory cells and secondary muscle damage is debated, recent studies of single stretch injury have highlighted that peak damage occurs at the same time as maximum neutrophil invasion of the injured tissue,^{6,7} prompting the idea that neutrophils may somehow participate in causing injury (fig 1).

We will examine the relation between muscle injury and change in neutrophil concentration after both eccentric exercise and acute stretch injury. We will present evidence to suggest that invading leucocytes—that is, neutrophils and macrophages—exacerbate the initial mechanical damage. Although the clinical implications are not clear at this time, it is conceivable that limiting certain aspects of inflammation will present new treatment strategies for sport related muscle trauma. Perspectives for rational approaches to handle the development of muscle injury during neutrophil inflammation are considered.

MUSCLE INFLAMMATION AND CHANGE IN NEUTROPHIL CONCENTRATION AFTER REPEATED ECCENTRIC EXERCISE

There is a ubiquitous host response to various types of skeletal muscle injury.^{8,9} Neutrophils are the first subpopulation of leucocytes to appear at the injury site.¹⁰ They are produced in the bone marrow and circulate in the bloodstream, representing 50–60% of the total circulating leucocytes. Neutrophils constitute the first line of defence against infectious agents or non-self substances that penetrate the body's physical barriers.

Studies on whether neutrophils participate only in phagocytosis or also promote muscle injury after modified musculoskeletal loading use different injury methods and rely on various variables to assess such injuries. Nevertheless, inflammatory cell infiltration after repeated eccentric contractions has been well established in both human and animal models.^{2,11,12} Stretch of activated skeletal muscles—that is, eccentric or lengthening contraction induced muscle injury—involves a variety of histopathological changes, including swelling of muscle cells, loss of the intermediate filament proteins desmin and dystrophin, and inflammatory cell infiltration.¹³ Repeated eccentric contractions of skeletal muscle can lead to an immediate loss of isometric force production that is the result of excitation/contraction uncoupling and damage to force producing or transmitting structures.¹⁴ This initial strength loss is

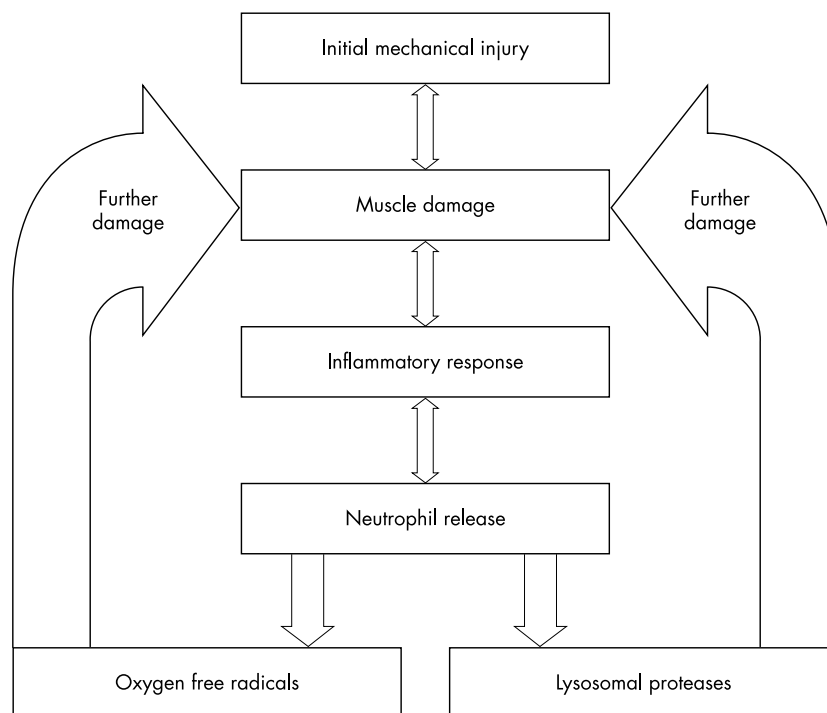


Figure 1 Proposed mechanism of the relation between the inflammatory response to mechanical injury and further muscle damage. The initial mechanically induced damage produces myofibre tearing and inflammatory cell infiltration. Neutrophils may promote further damage through the release of oxygen free radicals and lysosomal proteases and elastases.

followed by the so called “secondary damage” phenomenon,⁹ which many now consider to result from, among several possibilities, inflammatory cell mediated processes.

Clinical studies have supported the notion that there is an associated neutrophilia and secondary damage associated with repeated eccentric contractions. Saxton *et al*¹⁵ showed that the total leucocyte count was increased within four hours of repeated eccentric contraction and bench stepping exercise, but had returned to pre-exercise levels 24 hours later. This increase in total leucocyte count was attributed to changes in the numbers of circulating neutrophils. Previously Pizza *et al*¹⁶ measured blood neutrophil concentrations in humans during early recovery (<24 hours) from two bouts of eccentric exercise. The exercise bouts were separated by four weeks. Neutrophil concentrations were significantly higher three, six, and nine hours after exercise for the first bout than the second bout. The isometric strength deficit was significantly greater for bout 1 than bout 2 six, nine, and 24 hours after exercise. These data suggest that the neutrophilia associated with novel eccentric arm exercise precedes secondary changes in isometric strength and, furthermore, point to the possibility that invading neutrophils are linked to the secondary damage process which has been observed both clinically and in animal studies.

MUSCLE INFLAMMATION AND CHANGE IN NEUTROPHIL CONCENTRATION AFTER STRETCH INJURY

In animal models of single stretch injury, there appears to be a decline in muscle function that persists for 24 hours after injury.¹⁷ Recently, it has been shown using monoclonal antibodies that peak neutrophil invasion of stretch injured muscle occurs over the time course.⁷ This observation is true regardless of the amount of initial mechanical damage.⁷ Moreover, this study noted for the first time an absence of monoclonal antibody specific macrophages, suggesting that, in certain animal models, macrophages may not be involved in muscle regeneration and repair.⁷ In another study from the same laboratory, peak levels of oxygen free radicals were measured within the muscle 24 hours after injury.⁶ Similarly, muscle fibre tearing increases locally at the site of injury over the first 24 hours.⁶ Collectively, these studies suggest that a relation exists between neutrophil infiltration and degree of damage to stretch injured skeletal muscle. Moreover, these observations suggest that a valid target for attenuating muscle fibre damage may be to block neutrophil derived oxygen free radical production.

DO NEUTROPHILS CAUSE EXERCISE ASSOCIATED MUSCLE INJURY?

In the last decade, major advances have been achieved in understanding the possible mechanisms of neutrophil mediated tissue damage. Muscle injury by inflammatory cells has been examined most thoroughly in experimental models of muscle ischaemia followed by reperfusion, in which neutrophils have been clearly shown to promote muscle fibre damage during the reperfusion phase.¹⁸⁻²⁰ It is clear from non-exercise models, in particular ischaemia-reperfusion studies, that increased neutrophil adhesiveness to the endothelium is a critical early step in the sequence of events leading to muscle damage.²¹ Furthermore, there is very strong evidence that polymorphonuclear leucocytes (particularly neutrophils) and oxygen free radicals are key mediators of ischaemia related tissue injury.¹⁸⁻²⁰ After ischaemia, local leucosequestration of activated neutrophils occurs, with generation of reactive oxygen species,²² leading to the hypothesis that endothelial and subendothelial damage are caused by neutrophil derived oxidants.²³ Animals rendered leucopenic develop less damage and less oxidant production during the reperfusion phase.²⁴

On the basis of the observed time course for neutrophil infiltration and degree of injury,⁷ the hypothesis that neutrophils play a role in stretch injury has been investigated. Using an antibody that blocks the neutrophil's respiratory burst, it has been shown that the degree of myofibre damage can be considerably reduced 24 hours after injury.²⁵ Moreover, there is preservation of the intermediate filaments desmin and dystrophin, suggesting that oxygen free radicals may operate directly on these proteins. Although the consequences of these observations are unknown for human muscle injury, they parallel similar in vitro experiments showing that neutrophils can injure skeletal myotubes and may indicate that neutrophils exacerbate injury¹⁶ and or/delay regeneration.²⁶ These findings may be species specific, however, as rat ankle dorsiflexor muscles submitted to in situ lengthening contractions do not show evidence of accumulation of neutrophils after activity.²⁷ Furthermore, others have shown that passive stretch and isometric contractions can elevate neutrophil concentrations without causing overt signs of injury.¹⁰

CLINICAL IMPLICATIONS

From the above discussions, it appears that blocking neutrophil recruitment may limit the amount of damage that occurs in various models of muscle injury. Although we do not understand

the precise mechanism, it appears that neutrophils may cause damage by release of oxygen free radicals.²⁵ Furthermore, we now have evidence (unpublished results) that neutrophil derived free radicals target the proteins desmin and dystrophin, which are intermediate filament proteins important in regulation of muscle contraction.

To manipulate the host inflammatory response and attenuate the potentially negative consequences of inflammation, investigators have used animal models to examine the effects of non-steroidal anti-inflammatory drugs on muscle injury. Early studies suggested that they may reduce the decline in the muscle's tensile strength after injury.²⁸ In a similar study, rabbits treated with piroxicam showed an earlier recovery of muscle contractile force; however, no significant differences were observed at any other time period between treated and untreated animals.²⁹ According to Mishra *et al*,³⁰ flurbiprofen administration to rabbits produced more complete functional recovery three and seven days after repeated bouts of eccentric contractions. However, the same animals showed a deficit in torque and force generation at 28 days. In addition, flurbiprofen administration resulted in preservation of the intermediate filament protein desmin. Early in the recovery period, there was a dramatic increase in the regenerative response, which persisted until seven days. It was argued that flurbiprofen may have delayed the muscle's regenerative response. Although a definitive study has not been performed, these data argue that non-steroidal anti-inflammatories, which can attenuate neutrophil activity, may lead to delayed recovery and functional losses in animal models of muscle injury.

CONCLUSIONS

Neutrophils and macrophages play a role in muscle damage after repeated eccentric exercise and acute stretch injury. However, contrary to conventional thinking, it is possible that certain aspects of neutrophil function cause damage to healing muscle or delay its regenerative capabilities. Because neutrophils can release oxygen free radicals during phagocytosis, it is possible that neutrophil derived oxidants exacerbate pre-existing muscle injury in vivo by damaging previously uninjured muscle.¹⁰ These findings suggest the possibility that innovative treatment strategies directed at specific functions of the neutrophil are theoretically possible to improve recovery from muscle injury. Pharmacological intervention may be better targeted against specific aspects of neutrophil function such as free radical production, while maintaining the steps necessary for phagocytosis

and removal of cellular debris. This possibility is being investigated.

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Authors' affiliations

H T Toumi, T T Best, Departments of Orthopedics and Rehabilitation and Family Medicine, University of Wisconsin, Madison, WI, USA

Correspondence to: Dr Toumi, 501 North Henry Street (appart 812), Madison, WI 53703, USA; htoumi@wisc.edu

REFERENCES

- Smith LL. Acute inflammation: the underlying mechanism in delayed onset muscle soreness? *Med Sci Sports Exerc* 1991;**23**:542–51.
- Tidball JG. Inflammatory cell response to acute muscle injury. *Med Sci Sports Exerc* 1995;**27**:1022–32.
- Fielding RA, Manfredi TJ, Ding W, et al. Acute phase response in exercise. III. Neutrophil and IL-1 beta accumulation in skeletal muscle. *Am J Physiol* 1993;**265**:R166–72.
- Pizza FX, Mitchell JB, Davis BH, et al. Exercise-induced muscle damage: effect on circulating leukocyte and lymphocyte subsets. *Med Sci Sports Exerc* 1995;**27**:363–70.
- Best TM, Fiebig R, Corr DT, et al. Free radical activity, antioxidant enzyme, and glutathione changes with muscle stretch injury in rabbits. *J Appl Physiol* 1999;**87**:74–82.
- Brickson S, Hollander J, Corr DT, et al. Oxidant production and immune response after stretch injury in skeletal muscle. *Med Sci Sports Exerc* 2001;**33**:2010–15.
- Schneider BS, Sannes H, Fine J, et al. Desmin characteristics of CD11b-positive fibers after eccentric contractions. *Med Sci Sports Exerc* 2002;**34**:274–81.
- Carlson BM, Faulkner JA. The regeneration of skeletal muscle fibers following injury: a review. *Med Sci Sports Exerc* 1983;**15**:187–98.
- Armstrong RB, Warren GL, Warren JA. Mechanisms of exercise-induced muscle fibre injury. *Sports Med* 1991;**12**:184–207.
- Pizza FX, Koh TJ, McGregor SJ, et al. Muscle inflammatory cells after passive stretches, isometric contractions, and lengthening contractions. *J Appl Physiol* 2002;**92**:1873–8.
- Fielding RA, Violan MA, Svetkey L, et al. Effects of prior exercise on eccentric exercise-induced neutrophilia and enzyme release. *Med Sci Sports Exerc* 2000;**32**:359–64.
- Clarkson PM. Exercise-induced muscle damage: animal and human models. *Med Sci Sports Exerc* 1992;**24**:510–11.
- Friden J, Sjoström M, Ekblom B. Myofibrillar damage following intense eccentric exercise in man. *Int J Sports Med* 1983;**4**:170–6.
- Warren GL, Ingalls CP, Shah SJ, et al. Uncoupling of in vivo torque production from EMG in mouse muscles injured by eccentric contractions. *J Physiol (Lond)* 1999;**515**:609–19.
- Saxton JM, Claxton D, Winter E, et al. Peripheral blood leucocyte functional responses to acute eccentric exercise in humans are influenced by systemic stress, but not by exercise-induced muscle damage. *Clin Sci (Lond)* 2003;**104**:69–77.
- Pizza FX, McLoughlin TJ, McGregor SJ, et al. Neutrophils injure cultured skeletal myotubes. *Am J Physiol Cell Physiol* 2001;**281**:C335–41.
- Nikolaou PK, Macdonald BL, Glisson RR, et al. Biomechanical and histological evaluation of muscle after controlled strain injury. *Am J Sports Med* 1987;**15**:9–14.
- Korthuis RJ, Grisham MB, Granger DN. Leukocyte depletion attenuates vascular injury in postischemic skeletal muscle. *Am J Physiol* 1988;**254**:H823–7.
- Cambria RA, Anderson RJ, Dikdan G, et al. Leukocyte activation in ischemia-reperfusion injury of skeletal muscle. *J Surg Res* 1991;**51**:13–17.
- Jolly SR, Kane WJ, Hook BG, et al. Reduction of myocardial infarct size by neutrophil depletion: effect of duration of occlusion. *Am Heart J* 1986;**112**:682–90.
- Rosen GM, Pou S, Ramos CL, et al. Free radicals and phagocytic cells. *FASEB J* 1995;**9**:200–9.
- Welbourn CR, Goldman G, Paterson IS, et al. Neutrophil elastase and oxygen radicals: synergism in lung injury after hindlimb ischemia. *Am J Physiol* 1991;**260**:H1852–6.
- Kearns S, Moneley D, Murray P, et al. Oral vitamin C attenuates acute ischaemia-reperfusion injury in skeletal muscle. *J Bone Joint Surg [Br]* 2001;**83**:1202–6.
- Crinnion JN, Homer-Vanniasinkam S, Parkin SM, et al. Role of neutrophil-endothelial adhesion in skeletal muscle reperfusion injury. *Br J Surg* 1996;**83**:251–4.
- Brickson SL, Ji LiL, Schell K, et al. M1/70 attenuates blood-borne neutrophil activation and increases plasma interleukin-8 following stretch injury. *J Appl Physiol* 2003; in press.
- Seale P, Rudnicki MA. A new look at the origin, function, and "stem-cell" status of muscle satellite cells. *Dev Biol* 2000;**218**:115–24.
- Lapointe B, Frenette J, Côté C. Lengthening contraction-induced inflammation is linked to secondary damage but devoid of neutrophil invasion. *J Appl Physiol* 2002;**92**:1995–2004.
- Almekinders LC, Gilbert JA. Healing of experimental muscle strains and the effects of nonsteroidal antiinflammatory medication. *Am J Sports Med* 1986;**14**:303–8.
- Obremsky WT, Seaber AV, Ribbeck BM, et al. Biomechanical and histologic assessment of a controlled muscle strain injury treated with piroxicam. *Am J Sports Med* 1994;**22**:558–61.
- Mishra DK, Friden J, Schmitz MC, et al. Anti-inflammatory medication after muscle injury. A treatment resulting in short-term improvement but subsequent loss of muscle function. *J Bone Joint Surg [Am]* 1995;**77**:1510–19.

Group email for BASEM members

Following the last Executive meeting, it was decided that the Communications Officer, Dan Lane, should attempt to put all BASEM members with email on a group list in order that they can communicate more readily on both clinical and non-clinical matters. It will not mean that members will receive hundreds of irrelevant emails, but it will enable vastly improved communication throughout the Association.

If you would like to join the list, please forward your email address to either Dan Lane or Patrick Milroy (BASEM Secretary) at Patrick.Milroy@gp-n81030.nhs.uk