Original articles

High resolution comparative genomic hybridisation in clinical cytogenetics

Maria Kirchhoff, Hanne Rose, Claes Lundsteen

Abstract

High resolution comparative genomic hybridisation (HR-CGH) is a diagnostic tool in our clinical cytogenetics laboratory. The present survey reports the results of 253 clinical cases in which 47 abnormalities were detected. Among 144 dysmorphic and mentally retarded subjects with a normal conventional karyotype, 15 (10%) had small deletions or duplications, of which 11 were interstitial. In addition, a case of mosaic trisomy 9 was detected. Among 25 dysmorphic and mentally retarded subjects carrying apparently balanced de novo translocations, four had deletions at translocation breakpoints and two had deletions elsewhere in the genome. Seventeen of 19 complex rearrangements were clarified by HR-CGH. A small supernumerary marker chromosome occurring with low frequency and the breakpoint of a mosaic r(18) case could not be clarified. Three of 19 other abnormalities could not be confirmed by HR-CGH. One was a Williams syndrome deletion and two were DiGeorge syndrome deletions, which were apparently below the resolution of HR-CGH. However, we were able to confirm Angelman and Prader-Willi syndrome deletions, which are about 3-5 Mb. We conclude that HR-CGH should be used for the evaluation of (1) dysmorphic and mentally retarded subjects where normal karyotyping has failed to show abnormalities, (2) dysmorphic and mentally retarded subjects carrying apparently balanced de novo translocations, (3) apparently balanced de novo translocations detected prenatally, and (4) for clarification of complex structural rearrangements. (J Med Genet 2001;38:740-744)

Keywords: comparative genomic hybridisation; chromosome analysis; chromosome aberrations; dysmorphism

Comparative genomic hybridisation (CGH) is a technique which screens the whole genome for imbalances. The major application of CGH has been in the field of cancer genetics. An increasing number of papers, however, show that CGH can also be successfully applied in clinical cytogenetics.¹⁻⁸ The sensitivity of CGH is usually considered to be relatively low; however, we developed the technique further in order to increase the sensitivity as well as the specificity.^{9 10} This has enabled us to detect deletions as small as 3 Mb.¹¹ We have used the improved high resolution CGH technique (HR-CGH) in cancer projects as well as in our clinical cytogenetics laboratory. Here we present the HR-CGH results of 253 clinical cases performed from February 1997 to March 2001.

The results show that apart from being an excellent tool for clarification or confirmation of abnormal karyotypes, HR-CGH is well suited for investigation of dysmorphic and mentally retarded subjects with normal or apparently balanced karyotypes. Contrary to conventional CGH, HR-CGH enabled us to detect small chromosomal aberrations in a number of these patients. The HR-CGH technique has recently been implemented in Cyto-Vision (Applied Imaging) and is thus commercially available.

Materials and methods

CASES

Most of the cases in this survey were referred to our laboratory for routine CGH analysis; however, some cases (especially apparently balanced translocations) were ascertained by us for research purposes.

Eighty-eight of the patients participated in an ongoing investigation of 100 dysmorphic and mentally retarded children with normal karyotypes. This investigation includes HR-CGH, FISH with telomeric probes, and **S**KY karyotyping (CTS project). The detailed results of this investigation will be published elsewhere.

Cases were obtained as blood samples, amniotic fluid, chorionic villus samples, placental or fetal tissue samples, skin biopsies, or purified DNA. Case 12 was obtained from Coriell Cell Repositories (No GM10607), Coriell Institute for Medical Research, Camden, NJ, USA. All cases were karyotyped by conventional cytogenetics according to standard protocols.

Reference DNA for CGH was obtained from peripheral blood drawn from karyotypically normal males and females. High molecular

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Table 1 Summary of 253 clinical cases analysed by HR-CGH during the period February 1997–March 2001

Indication	No of analyses	No of abnormalities
(1) Dysmorphic and mentally retarded with a normal karyotype	144	16
(2) Physically and mentally retarded with a normal karyotype	24	0
(3) Apparently balanced de novo translocation, prenatal	6	0
(4) Apparently balanced de novo translocation associated with disease	25	6
(5) Clarification of abnormal karyotype	19	12
(6) Confirmation of abnormal karyotype	19	13
(7) Others*	16	0
Total	253	47

*Investigation of parental chromosomes.

Familial balanced translocations associated with disease.

weight genomic DNA was prepared by extractions on Qiagen Genomic Tip columns (Qiagen, Hilden, Germany).

FISH

FISH was performed in order to confirm the CGH findings. The results have been described previously for cases 2, 3, 4, and 7 (cases 3, 10, 4, and 7, respectively, in Kirchhoff *et al*¹²). In case 6, FISH with probes for loci D2S447 and D2Z4 showed only one signal. In case 12, FISH with a probe for locus D5S23 showed only one signal. In case 8, whole chromosome painting probes for chromosomes 4 and 10 were used (Oncor, Gaithersburg, MD, USA).

CGH

CGH was performed as described previously.¹⁰ Briefly, patient DNA and normal reference DNA were labelled with FITC-12-dUTP and Texas Red-5-dUTP (DuPont, Boston, MA), respectively. A total of 400-800 ng of DNA and 20-30 µg Cot1 DNA were hybridised to normal

Table 2 Abnormalities detected in dysmorphic and mentally retarded subjects with normal or apparently balanced karyotypes

Case No	Initial karyotype	HR-CGH analysis	Revised karyotype	Subsequent confirmation by	
				G banding	FISH
1	46,XY	dim(1q22q22)	46,XY,del(1) (q22q22)	Yes	Not done
2	46,XY	dim(2p15p15)	46,XY.rev ish dim(2p15p15)	No	Yes
3	46,XX	enh(9pter→qter)	46,XX/47,XX+9	Yes	Yes
4	46,XY	enh(10q11q11)	46,XY.rev ish enh(10q11q11)	No	Yes
5	46,XX	dim(7p15p15)	46,XX,del(7) (p15p15)	Yes	Not done
6	46,XX	dim(2q37→qter)	46,XX,del(2) (q37→qter)	Yes	Yes
7	46,XX	dim(3q22q24)	46,XX,del(3) (q22q24)	Yes	Not done
8	46,XX	dim(4q35→qter) enh(10p15→pter)	46,XX.rev ish dim($4q35 \rightarrow qter$) enh($10p \rightarrow 15pter$)	No	Yes
9	46,XY,t(1;4)(q31; q21.2)t(3;13) (p14.1;q33)	dim(13q33q33)	46,XY,t(1;4)(q31; q21.2)t(3;13) (p14.1;q33).rev ish dim(13q33q33)	No	Yes
10	46,XX,t(1;6;5) (p13;q14;p13)	dim(6q14q14)	46,XX,t(1;6;5) (p13;q14;p13).rev ish dim(6q14q14)	No	Not done
11	46,XX,t(10;16) (p11;q21)	dim(4q12q12)	46,XX,del(4) (q12q12),t(10;16) (p11;q21)	Yes	Not done
12	46,XY,t(3;5) (p23;p13)	dim(2q24q24) dim(5p13p13)	46,XY,del(2)(q24q24), t(3;5)(p23;p13).rev ish dim(5p13p13)	Yes/no	Not done/yes
13	46,XY,t(1;5;12) (q25;p15;q21)	dim(5p14p14)	46,XY,t(1;5;12) (q25;p15;q21).rev ish dim(5p14p14)	No	Not done

Cases 1, 2, 3, 4, 5, 6, 9, 10, and 12 were published in Kirchhoff *et al*¹² as cases 2, 3, 10, 4, 6, 5, 7, 8, and 9, respectively.

Rev ish: reverse in situ hybridisation. Dim: diminished fluorescence ratio intensity \approx deletion. Enh: enhanced fluorescence ratio intensity \approx duplication.

metaphase chromosomes. Slides were hybridised for three to four days, washed, and 4,6-diamidino-2counterstained with phenylindole. CGH image capture was performed with a CytoVision (Applied Imaging, Sunderland, UK) interfaced to a DM RBE fluorescence microscope (Leica, Heerbrugg, Switzerland) and images were transferred to a Magiscan image analysis system (Applied Imaging, Sunderland, UK). In each case, 10 metaphases were analysed. Aberrations were detected by standard reference intervals as described in Kirchhoff et al.¹⁰ Briefly, along the mean ratio profiles, the 99.5% confidence interval of each mean ratio profile value was compared to a corresponding 99.5% standard reference interval based on an average of 17 normal cases. The standard reference interval is especially wide at profile areas where CGH measurements are known to be unreliable. Where no overlap existed between the two intervals, the corresponding chromosome region was designated "aberrant". The standard reference interval was scaled automatically to fit the individual test case.

RESOLUTION

HR-CGH

The resolution of the present HR-CGH technique is approximately 3 Mb.¹¹ The resolution is, however, expected to show some variation depending on the chromosomal location of the abnormality. Thus, the resolution of telomeric regions may be somewhat lower than 3 Mb owing to wider standard reference intervals in the telomeres compared to the rest of the genome.

G banding

The majority of the conventional cytogenetic analyses which preceded the HR-CGH analyses of the dysmorphic and mentally retarded patients with normal or apparently balanced karyotypes were performed in other laboratories. Accordingly we have few data regarding the resolution of these analyses. We consider the resolution of the G banding analyses to be in the range of 300-600 bands corresponding to $\frac{1}{2}$ -1 band or 5-10 Mb.

Results

A total of 253 clinical cases was analysed by HR-CGH during the period February 1997 to March 2001 and 47 abnormalities were detected. They included 16 (11%) abnormalities in a group of dysmorphic and mentally retarded subjects with a normal karvotype and six abnormalities in five subjects (21%) in a group of dysmorphic and mentally retarded subjects with an apparently balanced karyotype. Twelve abnormalities were found in a group of abnormal karyotypes where HR-CGH was used for clarification, and 13 abnormalities were found in a group of abnormal karyotypes where HR-CGH was used for confirmation. A summary of all cases including indications is shown in table 1.

 Table 3
 Unbalanced abnormalities clarified or not clarified by HR-CGH analysis

Case No	Clinical information	Initial karyotype	CGH analysis	Comments or revised karyotype
Translocat				
14	Turner syndrome	46,X,add(Xp)	$enh(Yq11 {\rightarrow} qter)$	46,X,add(Xp).rev ish der(X)t(X;Y) (p22.3;g11)enh(Yg11 \rightarrow gter)
15	Prenatal diagnosis	46,XX,inv(<u>9</u>),add(<u>9</u> p)	$dim(9p22 \rightarrow pter)$ enh(18q11.1 \rightarrow qter)	46,XX,inv(9),add(9)(p22).rev ish der(9) t(9;18)(p22;g11.1)enh(18g11.1 \rightarrow gter)
16	Mentally retarded male	46,X,add(Xp)	enh(Yp10→pter)	46,X,add(X)(p).rev ish der(X)t(X;Y) (p22.3;p10)enh(Yp10→pter)
Duplicatio	ms			(p22:3,p10)em(1p10 /pter)
17	Morbus cordis and growth retarded	46,XX,add(2p)de novo	enh(2p21p23)	46,XX,add(2p).rev ish enh(2p21p23)
18	Prenatal diagnosis	46,XX,add(18q)	enh(18q11.2→qter)	46,XX,add(18q).rev ish enh(18q11.2→qter)
19	Dysmorphic with failure to thrive	46,XY,+mar. Ish add(5)(wcp5+)	enh(5p15-pter)×2	46,XY,add(5p).rev ish enh(5p15→pter)×2
20	Prenatal diagnosis	46,XX,ins(12;?)(q21;?) de novo	Normal	wcp12 paints the entire chromosome 12, pregnancy resulted in a healthy child
Deletions 21	Dysmorphic with malformations	46,XX,r(13)	dim(13q22-gter)	46,XX,r(13).rev ish dim(13p11.2q22)
21 22	Dysmorphic and mentally retarded	46,XX,del(11q?)	dim(13q22-qter) dim(11q23.2)	46,XX,del(11q?).rev ish dim(11q23.2)
Numerica	l			
23	Prenatal diagnosis	47,XY,+mar	enh(15q12q12)	47,XY,+mar.rev ish enh(15)(q12q12)pat, pregnancy continued
24	Missed abortion, suspected trisomy 18	47,XY,+mar	Normal	The marker is assumed to be inactive and of no clinical significance
25	Prenatal diagnosis	47,XY,+mar	enh(14q12q12)	47,XY,+mar.rev ish enh(14q12q12), pregnancy terminated
26	Prenatal diagnosis	46,XY/47,XY,+mar	Normal	The marker is assumed to be inactive, pregnancy resulte in a healthy child
27	Prenatal diagnosis	46,XY,+mar	Normal	The marker is assumed to be inactive
Others				
28	Missed abortion	46,XX,add(12p)	dim(12p13→pter) enh(12q13→qter)	46,XX,add(12p). rev ish rec(12)dup(12) inv(12)(p13q13)dim(12p12→pter)enh (12q13→qter)mat
29	Induced abortion of severely hydropic fetus	46,XX,add(21p)	Normal	The material on 21q is assumed to be inactive and without clinical significance
30	Prenatal diagnosis	46,XY, abnormal Y?	Normal	Father had a similar Y chromosome
Unbalance	ed abnormalities failed to be clarified by CGH			
31	Dysmorphic?	45,XX,-18/46,XX,r(18)	dim(18pter→qter)	Size of r(18) could not be estimated from the CGH analysis
32	Dysmorphic and delayed development	47,XXY/48,XXY,+mar	enh(Xper→pter)	Marker was only present in 7% of the cells and could no be detected with CGH

Rev ish: reverse in situ hybridisation. Dim: diminished fluorescence ratio intensity ~ deletion. Enh: enhanced fluorescence ratio intensity ~ duplication.

Of the 22 abnormalities found in dysmorphic and mentally retarded subjects with a normal or apparently balanced karyotype (indication groups 1 and 4, table 1), 12 were confirmed by reinspection of the G banded karyotypes, 10 by FISH, three by both techniques, one by microsatellite analysis (a patient from the CTS project), and two abnormalities have not yet been confirmed owing to lack of sample material.

Seventeen of the 22 abnormalities were interstitial, four were terminal, and one was a trisomy.

Table 2 outlines abnormalities detected in dysmorphic and mentally retarded subjects with normal or apparently balanced G banded karyotypes. Patients participating in the CTS project are not included. As indicated in table 2, 10 of the cases were previously reported in Kirchhoff *et al.*¹² The parents of the patients in table 2 were investigated when sample material was available. In case 8, the mother was shown to carry a balanced translocation (see Materials and methods). In eight cases (1, 2, 3, 4, 7, 9, 10, and 11), G banding, CGH, or FISH showed that the abnormalities were of de novo origin.

Table 3 shows 17 of 19 unbalanced abnormalities clarified by HR-CGH and two which could not be clarified, and table 4 describes 16 of 19 other unbalanced abnormalities confirmed by HR-CGH and three which could not be confirmed.

Five findings were not included since they were considered to be either false positive results or normal chromosomal variations. These were three duplications of chromosome 15q12, a duplication of chromosome 9p11, and a deletion of chromosome 16p11.

Discussion

The cases where HR-CGH analysis provided essential information fall into two categories: (1) cases where G banded karyotypes were normal or apparently balanced and chromosomal disease was suspected (table 1, indication groups 1 and 4), and (2) cases with unbalanced G banded karyotypes where clarification or confirmation of the abnormalities by additional techniques was attempted (table 1, indication groups 5 and 6).

We have previously shown that HR-CGH is capable of detecting abnormalities in the former category.¹² However, the present survey shows that abnormalities were found in remarkably high numbers, since chromosomal imbalances were detected in 11% of 144 dysmorphic and mentally retarded subjects with a normal conventional karvotype and in 20% of 25 dysmorphic and mentally retarded subjects carrying an apparently balanced de novo translocation. The two unconfirmed deletions in table 2 (cases 10 and 13) were found at the breakpoints of chromosomes involved in apparently balanced de novo translocations, which strengthens the reliability of these findings. Although these deletions could be false positive results, we have previously shown that false positives are very rare at the

Table 4 Unbalanced abnormalities confirmed or not confirmed by HR-CGH analysis

Clinical information	Initial karyotye	CGH analysis	Comments
uctural and numerical abnormali	ties confirmed by CGH		
Dysmorphic with	47,XX,+der(22)t(11;22)	enh (11q23→qter)	
malformations	(q23;q11.2)		
Turner syndrome?	45, X/46, X, -X, +der(X)	dim(Xpter→qter)	The ratio from Xp11.4→pter was more reduced than
5		dim(Xp11.4-pter)	the rest of the X chromosome
Prenatal diagnosis			
i feliutur diugilesis			
Prenatal diagnosis			
i foliutur diugilosio		ann(sqs1)qter)	
Severely growth retarded		$\dim(11a14,2a23,3)$	
Severely growin retarded		um(11q14.2q29.9)	
Propostal diagnosia		$\dim(7a32)$ (star)	
Fienatai diagnosis		dill(/q32→qter)	
Minne and also		dim (V = 10) = t = r)	
Microcephaly	40, AA/40, A, I(A)(q10)		
Devenated diamagnic	47 XX I man mat		Marker assumed to be inactive
			Marker assumed to be inactive
Mental retardation		Normal	Marker assumed to be inactive
Prenatal diagnosis			Retrospectively, 20% mosaicism of an i(Yp) was four
			on G banding
Prenatal diagnosis	46,XY,9qh+mat	Normal	The light stained 9qh region was divided by a dark
			band like a G band
Phenotypic male	46,XX	46,XX	
bnormalities confirmed by CGH			
Prenatal diagnosis	46,XX,-18,+der(18)	enh(11q25q25)	This cryptic translocation was incidentally detected b
	t(11;18)(q25;q23)pat	dim(18q23q23)	interphase FISH ²⁸
Mentally retarded	46,XX,-11,+der(11)	dim(11q25q25)	
	t(11;18)(q25;q23)mat	enh(18q23q23)	
Angelman syndrome	46,XX.ish del(15)	dim(15q11q13)	
0			
Prader-Willi syndrome		dim(15q11q13)	
	(q11q13)		
phormalities failed to be detected b	v CGH		
		Normal	
210001ge synaronie		1.011101	
Williams syndrome		Normal	
w mans syncrome		Tiorinai	
DiCoorgo gun dromo		Normal	
DiGeorge syndrome	(q11.2q11.2)	inormai	
	Dysmorphic with malformations Turner syndrome? Prenatal diagnosis Prenatal diagnosis Severely growth retarded Prenatal diagnosis Microcephaly Prenatal diagnosis Mental retardation Prenatal diagnosis Prenatal diagnosis Prenatal diagnosis Prenatal diagnosis Phenotypic male <i>mormalities confirmed by CGH</i> Prenatal diagnosis Mentally retarded Angelman syndrome Prader-Willi syndrome	Use of the second symmetric of the second symmetric of the second symmetric of the second symmetric of the symmet	Use to the second stress of the second stre

Rev ish: reverse in situ hybridisation. Dim: diminished fluorescence ratio intensity ~ deletion. Enh: enhanced fluorescence ratio intensity ~ duplication.

level of confidence used in the CGH analyses of this work.¹⁰

In the present study, an amplification of chromosome 9p11 and a deletion of chromosome 16p11 were regarded as false positive results or normal variations and were excluded. Normal variations in these chromosomal regions have previously been described.13 14 We have, however, also noticed that, contrary to the rest of the genome, imbalances are on rare occasions detected in normal subjects in 9p11, 16p11, and 1q21, which may disappear when the analyses are repeated. Generally, the ratio deviations in two or all three of these regions are correlated in the individual analyses, raising considerably suspicion that they may represent technical artefacts. Three duplications of chromosome 15q12 were regarded as normal variations. They were found in analyses of an affected boy and his healthy father and in a girl with an apparently balanced t(3;21). Variation of region 15q11-12 has been described in a number of normal subjects.15-17

So far, it is unclear if the abnormalities detected in dysmorphic and mentally retarded subjects with a normal or apparently balanced karyotype are responsible for the clinical findings in all the patients. Analysis of the parents is obviously important, since de novo imbalances in subjects with parents carrying normal or balanced karyotypes are more likely to be associated with an affected phenotype. Detailed knowledge of the diagnostic implications of all the imbalances is, however, unlikely to be obtained until the particular chromosomal regions can be characterised further or until additional cases are identified.

A total of 7.6% of the 144 dysmorphic and mentally retarded patients with a normal G banded karvotype carried an interstitial abnormality while only 2.8% of the patients had terminal abnormalities. These figures are interesting since subtelomeric abnormalities have been shown in approximately 7.5% of mentally retarded subjects in several investigations with subtelomeric FISH probes.¹⁸⁻²³ Knight et al²⁰ concluded that: "once recognizable syndromes have been excluded, abnormalities that include the ends of chromosomes are the commonest cause of mental retardation in children with undiagnosed moderate to severe mental retardation". The results of our survey indicate that in dysmorphic and mentally retarded patients interstitial imbalances are found in numbers close to the number of abnormalities found in subtelomeric regions by FISH probes.

The present data suggest that chromosomal abnormalities may be detected in approximately 15% (7.5% + 7.6%) of mentally retarded and dysmorphic patients if both subtelomeric screening and HR-CGH are applied. However, it may be that these figures are not additive since the primary criterion for the investigations with subtelomeric probes was mental retardation,²⁰ while dysmorphic features were also required in our cases. Nevertheless, it is likely that for most of the patients both these criteria were fulfilled.

All six imbalances detected in the patients with apparently balanced translocations (table 2) were interstitial and, interestingly, two of them were detected outside the regions involved in the translocations. Since these two abnormalities were retrospectively confirmed by G banding, it is likely that the finding of a translocation in a G banded karyotype may distract the cytogeneticist's attention from other abnormalities.

HR-CGH was also applied to de novo apparently balanced translocations detected prenatally. No abnormalities were detected in the six prenatal cases investigated in this survey (table 1, indication group 3). When, however, analyses of more such cases are performed, imbalances are bound to be found in some of them. Naturally they will not appear as frequently as in the dysmorphic and mentally retarded patients with apparently balanced karyotypes, since the majority of the prenatal cases are not expected to be associated with disease.24 25

The cases listed in tables 3 and 4 show that HR-CGH is well suited to the confirmation or clarification of abnormal G banded karyotypes. An increasing number of papers similarly show that this application of CGH is becoming more widely recognised.^{2 3 8 26 27} The cases that were not confirmed or clarified by HR-CGH had abnormalities that were typically too small (DiGeorge and Williams syndrome deletions, table 4, and probably case 31, table 3) or were present in a low level mosaic state (case 32, table 3).

The present survey shows that HR-CGH is likely to make a significant contribution to the difficult and laborious task of associating chromosome abnormalities with clinical manifestations. Hopefully, an increasing number of laboratories will participate in this work. HR-CGH is commercially available on the CytoVision from Applied Imaging Corporation who has the sole and exclusive rights to the marketing of the HR-CGH software.

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