

Genes, oxidative stress, and the risk of chronic obstructive pulmonary disease

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Introductory article

Association between polymorphism in gene for microsomal epoxide hydrolase and susceptibility to emphysema

CAD Smith, DJ Harrison

Background. The first-pass metabolism of foreign compounds in the lung is an important protective mechanism against oxidative stress. We investigated whether polymorphisms in the gene for microsomal epoxide hydrolase (mEPHX), an enzyme involved in this protective process, had any bearing on individual susceptibility to the development of chronic obstructive pulmonary disease (COPD) and emphysema. **Methods.** We designed PCR-based genotyping assays to detect variant forms of mEPHX that confer slow and fast activity. We used these assays to screen 203 blood-donor controls and groups of patients with asthma (n=57), lung cancer (n=50), COPD (n=68), and emphysema (n=94), who were attending specialised clinics in Edinburgh, UK. **Findings.** The proportion of individuals with innate slow mEPHX activity (homozygotes) was significantly higher in both the COPD group and the emphysema group than in the control group (COPD 13 [19%] vs control 13 [6%]; emphysema 21 [22%] vs 13 [6%]). The odds ratios for homozygous slow activity versus all other phenotypes were 4.1 (95% CI 1.8–9.7) for COPD and 5.0 (2.3–10.9) for emphysema. **Interpretation.** Genetic polymorphisms in xenobiotic enzymes may have a role in individual susceptibility to oxidant-related lung disease. Epoxide derivatives of cigarette-smoke components may be the cause of some of the lung damage characteristic of these diseases. (Lancet 1997;350:630–33)

Chronic obstructive pulmonary disease (COPD) is one of the major causes of premature death in industrialised countries. While its primary pathology is pulmonary emphysema together with narrowing and obliteration of airways, COPD remains a clinical diagnosis characterised by chronic airflow limitation which progresses slowly over a period of years and is largely irreversible.¹ The majority of cases are a consequence of chronic cigarette smoking and are thus preventable. However, only a relatively small proportion of smokers develop symptomatic disease.² As a consequence, there has been considerable interest in identifying those who are most susceptible, and the mechanisms of their susceptibility. A number of studies indicate that genetic factors contribute to the risk of COPD. Alpha₁-antitrypsin deficiency is already well recognised and twin studies have suggested the presence of other undetermined genetic factors.^{3–6} Identification of these genetic components could provide useful insights into the pathogenesis of COPD in the same way as did recognition of the association between alpha₁-antitrypsin deficiency and COPD.⁷ This association, together with the fact that

emphysema can be produced experimentally by intratracheal instillation of papain, led to the protease-anti-protease hypothesis of pulmonary emphysema.

The introductory article contributes further to the debate. It suggests that genetic susceptibility to oxidative stress may also confer a risk for the development of COPD. This implies that oxidant-antioxidant imbalance, like protease-antiprotease imbalance, may be important in its pathogenesis.⁸ Indeed, a number of investigators have implicated oxidant-antioxidant interaction in the pathogenesis of smoking induced COPD.^{9,10} In brief, smoking increases alveolar oxidants, in part because cigarette smoke itself contains an enormous number of free radicals and in part because it increases the number of inflammatory cells in alveoli which spontaneously release oxidants. These oxidants inactivate alpha₁-antitrypsin and other protease inhibitors such as secretory leukoprotease inhibitor. Furthermore, the recruited inflammatory cells also increase the protease burden, thus tipping the protease-antiprotease balance further towards the protease side. Moreover, oxidants in cigarette smoke can directly damage com-

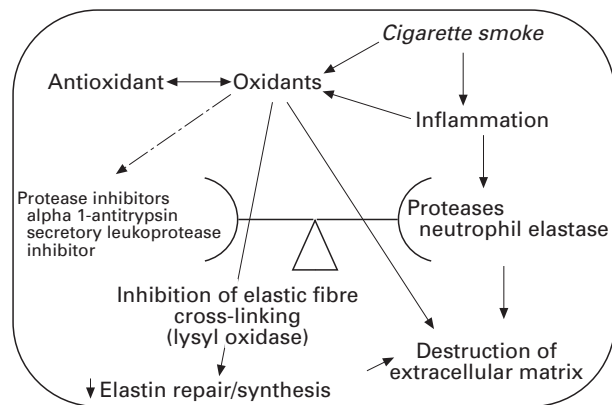


Figure 1 Protease-antiprotease balance and oxidant-antioxidant balance.

ponents of the lung extracellular matrix such as elastin and collagen, or modify the matrix to make it more susceptible to protease attack (fig 1).

The systemic effects of these changes can be detected in a number of ways. Morrow *et al* have recently found increased levels of prostaglandin F_2 like compounds (F_2 -isoprostanes) in the plasma of smokers, indicating that smoking causes lipid peroxidation *in vivo*.¹¹ Rahman *et al* found a decreased antioxidant capacity of plasma in smokers, the decrease being most marked one hour after smoking. Thus, cigarette smoke may deplete the antioxidant capacity.¹² There is consequently good theoretical and experimental evidence, as well as some clinical support, pointing to the importance of the oxidant-antioxidant imbalance in the pathogenesis of pulmonary emphysema. In addition, there have been suggestions that genetic susceptibility to oxidative stress may have a role in the pathogenesis of other diseases such as diabetes mellitus¹³ and its vascular complications,¹⁴ and Parkinson's disease.¹⁵

We shall review the introductory article in the context of all the known genetic associations of COPD.

Introductory article

Smith and Harrison investigated the genetic polymorphism of an important xenobiotic metabolising enzyme, microsomal epoxide hydrolase. There are four common microsomal epoxide hydrolase alleles, depending on the presence or absence of two point mutations in the coding gene. The presence of one mutation results in a "slow" allele, while the presence of the other gives rise to a "fast" allele. The rare occurrence of both mutations together produces the enzyme with normal activity. The homozygous state for the slow allele results in a phenotype of very slow microsomal epoxide hydrolase activity.

Using a PCR-based genotyping assay, Smith and Harrison investigated whether polymorphisms in the gene for microsomal epoxide hydrolase were linked with susceptibility to COPD and emphysema. The mechanism proposed was that, since this enzyme system is involved in the metabolism of highly reactive epoxide intermediates, slow metabolisers might experience greater oxidative stress from cigarette smoke and so develop lung disease. Four groups were studied: blood donor controls, asthma controls, patients with COPD, and patients with lung cancer. The group with lung cancer was then subdivided into those with and those without emphysema by examination of the resection specimens from the pathology archives of Edinburgh

University. No clinical data were available for the lung cancer patients but lung function was presumably quite well preserved or they would not have undergone surgery.

This rather odd study design suggests a possible change of direction during the course of the research. It may be that the investigators started by seeking a genetic susceptibility to lung cancer (reactive epoxides are carcinogenic) with negative results, but stumbled on an association between slow metabolisers and COPD. Whatever the primary aim, the outcome was interesting. In brief, the COPD and "emphysema" groups contained significantly more homozygous mutants for the exon 3 (slow) polymorphism than the controls.

Some caution is needed over the interpretation of these findings. While they may indicate a true and novel causal association, the study population was small and the findings could be due to chance. They clearly need to be confirmed in other study populations. Furthermore, the "emphysema" group was unusual, being defined from the morbid anatomy of lung samples resected for cancer. While this is the "purest" way to make such a diagnosis, it provides no clinical information. If these subjects simply had histological evidence of mild emphysema, they may not be satisfactorily representative of smokers in general who are susceptible to COPD.

A second concern is whether biochemical pathways affected by microsomal epoxide hydrolase are actually involved in the pathogenesis of emphysema. Cigarette smoke certainly contains free radicals and multiple other chemicals capable of generating reactive epoxides, and some of these can certainly damage nucleic acids, proteins and lipids. It is not clear, however, whether these processes are important *in vivo*, how they affect the protease/antiprotease balance or oxidant/antioxidant balance, nor whether epoxide hydrolase is an important rate limiting step in such a process.

The hypothesis that oxidant/antioxidant imbalance is due to polymorphisms of the gene for microsomal epoxide hydrolase is nevertheless an interesting one which deserves further investigation. There are many possible genetic mechanisms for undue susceptibility to cigarette smoke, and it is worth reviewing the range of these in order to put this new observation in perspective.

Genetic contributions to COPD

XENOBIOTIC METABOLISM

Genetically determined variation in xenobiotic detoxification/biotransformation has attracted interest recently as a possible mechanism for observed differences in susceptibility to various conditions—for example, idiosyncratic reactions to pharmacological agents and smoking induced lung cancers.^{16,17} A detailed discussion of the metabolic process of detoxification of xenobiotics is beyond the scope of this review. Briefly, the major pathway of metabolism involves two main types of enzymes: the phase I cytochrome P-450 mediating oxidative metabolism, and phase II conjugating enzymes such as glutathione S-transferases. A number of xenobiotics, including benzo[a]pyrene, a carcinogen contained in tobacco smoke, are enzymatically metabolised to highly reactive intermediates such as epoxides by the cytochrome P450 system. These resultant metabolites may be cytotoxic, mutagenic, and/or carcinogenic. The enzymatic conversion of these metabolites to inactive intermediates is therefore protective.

Epoxides can be detoxified principally by glutathione S-transferases (GST) or epoxide hydrolases. The rel-

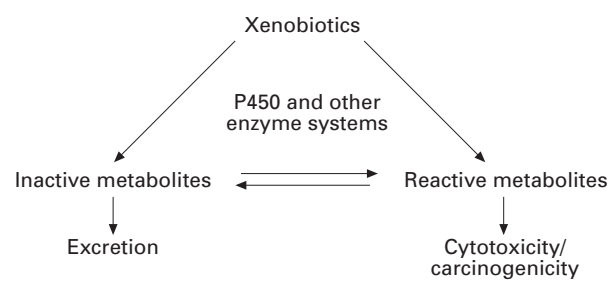


Figure 2 Xenobiotic metabolism: activation-inactivation balance.

active balance between activation and inactivation is thought to underlie susceptibility to the noxious effects of various xenobiotics (fig 2). For example, altered phenotypes and genotypes in the cytochrome P450 isoforms *CYP1A1* have been reported to be associated with smoking-induced lung cancer and other cancers.¹⁸ Defective glutathione S-transferases caused by the *GST M1* null genotype have also been linked to an increased risk of developing lung cancer,¹⁹ although the results of studies conducted in different ethnic groups have not always been consistent. In addition, individuals with the susceptible genotypes of *CYP1A1* with deficient *GST M1* were reported to be at remarkably high risk in a Japanese population.²⁰ This topic has recently been reviewed by Smith and associates²¹ and Raunio *et al.*²² Although the liver plays a major role in xenobiotic metabolism, other organs (including lungs, kidneys and the gastrointestinal tract) have also been shown to be potential sites of metabolism.²³

The introductory article is the first to suggest the involvement of genetic susceptibility to oxidative stress in the development of COPD. Previous studies on oxidant-antioxidant imbalance in COPD have focused largely on reactive oxygen species or oxidative molecules such as superoxide anion, hydrogen peroxide and hydroxyl radical, nitrogen dioxide, and ozone as the offensive agents. Many have also concerned the "classic" antioxidant enzymes such as superoxide dismutase, catalase, and glutathione peroxidase, as well as non-en-

zymatic antioxidants such as α -tocopherol, β -carotene, ascorbate, or glutathione. Since epoxide hydrolases catalyse hydrolysis of epoxides formed as a result of oxidative metabolism mediated by cytochrome P450, it is conceivable that this important enzyme does play a significant role in preventing the tissue injury caused by cigarette smoke, together with other detoxifying enzymes such as glutathione S-transferases. In fact, some researchers, including the authors of the introductory article, have shown possible associations of COPD with polymorphisms in *GST M1*²⁴ or *GST P1*.²⁵

The expression and activity of enzymes are influenced not only by genetic factors but also by various chemicals in the environment through induction and competitive inhibition. Furthermore, the clinical effect will depend not only on the kinetics of metabolism but also on the major site of action. Local expression in lungs might well be as important as hepatic metabolism, and so the relationship of the activity of a given enzyme and its clinical consequences is likely to be complex.

DEFICIENCY OF ANTIPROTEASE SCREEN

*Alpha*₁-antitrypsin deficiency

A number of genetic factors have been proposed to increase the risk for developing COPD, but only a few have been confirmed to be clinically significant. The most important is α ₁-antitrypsin deficiency; the odds ratio for the development of COPD in association with the homozygous ZZ phenotype (presence versus absence) has been calculated at more than 30 (table 1). As α ₁-antitrypsin is a potent inhibitor of neutrophil elastase, the recognition of this association has led to an elastase-antielastase hypothesis (or, more broadly, a protease-antiprotease hypothesis) as a pathogenic explanation for pulmonary emphysema (fig 1). However, as the frequency of this homozygosity is relatively low even in Caucasian populations, the deficiency accounts for less than 1% only of all patients with COPD, although some estimates have been higher.²⁶ Furthermore, α ₁-antitrypsin deficiency with the PiZZ phenotype is not necessarily associated with COPD and many such

Table 1 Frequencies of various phenotypes in controls and patients with COPD

	Total	MM	MS	MZ	ZZ	SS	SZ	Author	Location
Controls	2054	1828	82	85	1	2	4	Gulsvik (1979)	Oslo, Norway
	280	254	17	8	0	0	0	Bencze (1980)	Munich, Germany
	106	94	8	3	0	0	0	Talamo (1972)	Mass, USA
	642	583	42	8	2	0	1	Bartman (1985)	Bonn, Germany
	721	644	57	14	0	0	0	Cox (1976)	Toronto, Canada
	930	835	60	21	0	0	0	Shigeoka (1976)	New York, USA
	91	81	5	2	0	0	0	Barnett (1975)	North Caroline, USA
	114	98	8	6	0	1	0	Kueppers (1977)	Minnesota, USA
	1380	1213.5	110.5	34	0	1	5	Lieberman (1976)	California, USA
Total	6318	5630.5	389.5	181	3	4	10		
COPD	149	123	14	7	1	0	1	Gulsvik (1979)	Oslo, Norway
	76	70	0	2	2	0	0	Bencze (1980)	Munich, Germany
	99	84	8	4	1	0	0	Talamo (1972)	Mass, USA
	526	429	34	31	5	0	18	Bartman (1985)	Bonn, Germany
	123	101	7	6	8	0	0	Cox (1976)	Toronto, Canada
	502	442	35	14	1	0	0	Shigeoka (1976)	New York, USA
	107	87	6	10	2	0	0	Barnett (1975)	North Caroline, USA
	114	97	5	9	3	0	0	Kueppers (1977)	Minnesota, USA
	190	146	10	26	5	0	2	Janus (1988)	Melbourne, Australia
	965	97	74	18	3	3	3	Lieberman (1986)	California, USA
	350	275	22	35	12	3	3	Mitmann (1974)	California, USA
Total	3201	1854	238	218	58	6	27		
OR*		0.59	1.22	2.48	38.85	2.44	7.92		
OR vs MM†			1.10	2.42	40.49	2.67	8.54		

This table was compiled from studies conducted in different populations using different definitions of the disease. Cases and controls were not matched. Reliable odds ratios cannot therefore be calculated.

* OR = odds ratio of each phenotype for COPD against all other phenotypes.

† OR vs MM = odds ratio of each phenotype against the MM phenotype.

people remain healthy in the absence of a smoking history into their sixth and seventh decades.²⁷

The heterozygous state of α_1 -antitrypsin deficiency has also been implicated as a possible risk factor for COPD; the odds ratio for developing COPD in MZ heterozygotes (compared with subjects who are neither heterozygotes nor homozygotes) has been reported by some investigators to range from 1.5 to 5.0 in a review by Stanford *et al*²⁸ (see also table 1). Furthermore, Tarján and colleagues have recently reported accelerated declines in expiratory flow rates and diffusing capacity, as well as increases in total lung capacity and residual volume, in PiMZ heterozygous subjects in a longitudinal study.²⁹ In practice, however, the PiMZ state has not been found to pose a major risk for COPD, especially in the absence of smoking. Indeed, several studies in randomly selected populations have failed to demonstrate a definite association between MZ phenotype and COPD,^{30–34} one having had a power of 95% to detect a difference in FEV₁/FVC% as low as 3%.³⁴

Mutation of the flanking sequence of the α_1 -antitrypsin gene

A mutation of the 3' flanking sequence of the α_1 -antitrypsin gene has been associated with COPD in a few studies.^{35,36} The odds ratios for developing COPD (or emphysema) in association with this mutation (present versus absent) were calculated to be 4.3 and 3.2, respectively. This condition is different from α_1 -antitrypsin deficiency in that the sequence which codes for the protein itself is normal and the basal serum level of α_1 -antitrypsin is not reduced. Kalsheker *et al* suggested that the mutation might diminish the response of the α_1 -antitrypsin gene to interleukin 6 and thus suppress the acute phase response of α_1 -antitrypsin.³⁷ This acute response is thought to counter the increased proteolytic burden at the region of injury induced by inflammation.

Alpha₁-antichymotrypsin deficiency

The deficiency of another serine protease inhibitor, α_1 -antichymotrypsin, has also been shown to be associated with COPD. This counteracts the adverse enzymic effects of neutrophil cathepsin G, mast cell chymase, and chymotrypsin. Poller *et al* detected this deficiency in four of 100 patients with COPD but in none of 100 healthy controls.³⁸ However, there seems to be a wide variation in the prevalence of this deficiency between different populations,³⁹ as is the case for α_1 -antitrypsin

deficiency, and this abnormality is unlikely to account for a large proportion of patients who develop COPD.

TUMOUR NECROSIS FACTOR- α GENE POLYMORPHISM

A recent study from Taiwan has reported an association between chronic bronchitis and a polymorphism at the -308 position of the tumour necrosis factor α (TNF α) gene.⁴⁰ This polymorphism gives rise to two alleles, TNF1 and TNF2. The investigators found that the less common allele, TNF2, which was associated with higher basal and induced expression of TNF α , was more prevalent in patients with chronic bronchitis. They suggested there was an augmented inflammatory process associated with tissue injury due to increased TNF α expression, and hence the development of chronic bronchitis.

The patient group consisted of 42 male adults who had histories of chronic or recurrent productive cough for more than two successive years. There was also airflow limitation defined as FEV₁ of <80% predicted and FEV₁/FVC <69% predicted. Thirteen patients (31%) were non-smokers. No occupational histories were given, and the 13 may have included patients with chronic asthma. Further studies in other populations will be needed to confirm this interesting association.

OTHER FACTORS

Inherited disorders of connective tissue such as Marfan syndrome,⁴¹ Ehlers-Danlos syndrome,⁴² and cutis laxa⁴³ are reported to be associated with a number of pulmonary diseases including emphysema. Abnormal elastic tissue in lungs has been found in necropsy material from infants with Marfan syndrome,⁴¹ and this may explain the association.

Blood group-related phenotypes including ABO blood group, ABH secretor/non-secretor, and Lewis positive/negative status have also been focuses of attention, but study results have been inconclusive.^{44–47}

The sex and race of the subjects have also been implicated as possible factors of relevance. The prevalences of COPD and chronic bronchitis are said to be higher in men than in women, for equivalent cigarette consumption,⁴⁸ and some studies have shown a greater relative loss of lung function in men than in women.⁴⁹ Others suggest that women may be more susceptible than men to the adverse effects of smoking on lung function.⁵⁰ Furthermore, some studies have suggested that white males may be more prone to developing

LEARNING POINTS

- * Only 10–20% of cigarette smokers develop symptomatic COPD, implying undue susceptibility compared with the remainder of the population at large.
- * Alpha₁-antitrypsin deficiency, which is the only fully established genetic risk factor, accounts for less than 1% of all cases of COPD.
- * Polymorphisms for the genes controlling xenobiotic metabolism (hence oxidant-antioxidant balance) may explain some of the observed differences in susceptibility to various conditions caused by environmental factors, including COPD.
- * Elucidation of additional genetic risk factors may provide useful insights into the pathogenesis of COPD, but clinical benefits are not yet apparent.
- * The absence of demonstrable risk factors in the individual for developing COPD or lung cancer should not deter physicians from persuading smokers to quit the habit.

COPD than non-white males,⁵¹ but neither sex nor race are likely to be major genetic determinants of susceptibility to tobacco smoke.

- 1 Siafakas NM, Vermeire P, Pride NB, *et al* on behalf of the Task Force. Optimal assessment and management of chronic obstructive pulmonary disease. *Eur Respir J* 1995;8:1398-20.
- 2 Fletcher C, Peto R. The natural history of chronic airflow obstruction. *BMJ* 1977;1:1645-8.
- 3 Webster PM, Lorimer EG, Man SFP, *et al*. Pulmonary function in identical twins: Comparison of nonsmokers and smokers. *Am Rev Respir Dis* 1979;119:223-8.
- 4 Hankins D, Drage C, Zamel N, *et al*. Pulmonary function in identical twins raised apart. *Am Rev Respir Dis* 1982;125:119-21.
- 5 Hubert HB, Fabsitz RR, Feinleib M, *et al*. Genetic and environmental influences on pulmonary function in adult twins. *Am Rev Respir Dis* 1982;125:409-15.
- 6 Redline S, Tishler PV, Lewitter FI, *et al*. Assessment of genetic and nongenetic influences on pulmonary function: a twin study. *Am Rev Respir Dis* 1987;135:217-22.
- 7 Laurell CB, Eriksson S. The electrophoretic α_1 -globulin pattern of serum in α_1 -antitrypsin deficiency. *Scand J Clin Invest* 1963;15:132.
- 8 Smith CAD, Harrison DJ. Association between polymorphism in gene for microsomal epoxide hydrolase and susceptibility to emphysema. *Lancet* 1997;350:630-3.
- 9 Buhl R, Meyer A, Vogelmeier C. Oxidant-protease interaction in the lung: prospects for antioxidant therapy. *Chest* 1996;110:267-72S.
- 10 Rahman I, MacNee W. Oxidant/antioxidant imbalance in smokers and chronic obstructive pulmonary disease. *Thorax* 1996;51:348-50.
- 11 Morrow JD, Frei B, Longmire AW, *et al*. Increase in circulating products of lipid peroxidation (F₂-isoprostanes) in smokers. *N Engl J Med* 1995;332:1198-203.
- 12 Rahman I, Morrison D, Donaldson K, *et al*. Systemic oxidative stress in asthma, COPD, and smokers. *Am J Respir Crit Care Med* 1996;154:1055-60.
- 13 Kubisch HM, Wang J, Luche R, *et al*. Transgenic copper/zinc superoxide dismutase modulates susceptibility to type I diabetes. *Proc Natl Acad Sci USA* 1994;91:9956-9.
- 14 Ruiz J. Diabetes mellitus and the late complications: influence of the genetic factor. *Diabetes Metabolism* 1997;23(Suppl 2):57-63.
- 15 Ikebe S, Tanaka M, Ozawa T. Point mutations of mitochondrial genome in Parkinson's disease. *Mol Brain Res* 1995;28:281-95.
- 16 West WL, Knight EM, Pradhan S, *et al*. Interpatient variability: genetic predisposition and other genetic factors. *J Clin Pharmacol* 1997;37:635-48.
- 17 Seidegård J, Ekström G. The role of human glutathione transferases and epoxide hydrolases in the metabolism of xenobiotics. *Environ Health Perspect* 1997;105(Suppl 4):791-9.
- 18 Kawajiri K, Eguchi H, Nakachi K, *et al*. Association of *CYP1A1* germ line polymorphisms with mutations of the *p53* gene in lung cancer. *Cancer Res* 1996;56:72-6.
- 19 Zhong S, Howie AF, Ketterer B, *et al*. Glutathione S-transferase mu locus: use of genotyping and phenotyping assays to assess association with lung cancer susceptibility. *Carcinogenesis* 1991;12:1533-7.
- 20 Nakachi K, Imai K, Hayashi S, *et al*. Polymorphisms of the *CYP1A1* and glutathione S-transferase genes associated with susceptibility to lung cancer in relation to cigarette dose in a Japanese population. *Cancer Res* 1993;53:2994-9.
- 21 Smith CAD, Smith G, Wolf CR. Genetic polymorphisms in xenobiotic metabolism. *Eur J Cancer* 1994;30A:1921-5.
- 22 Raunio H, Husgafvel-Pursiainen K, Anttila S, *et al*. Diagnosis of polymorphisms in carcinogen-activating and inactivating enzymes and cancer susceptibility: a review. *Gene* 1995;159:113-21.
- 23 Krishna DR, Klotz U. Extrahepatic metabolism of drugs in humans. *Clin Pharmacokinet* 1994;26:144-60.
- 24 Harrison DJ, Cantlay AM, Rae F, *et al*. Frequency of glutathione S-transferase M1 deletion in smokers with emphysema and lung cancer. *Human Exp Toxicol* 1997;17:356-60.
- 25 Harries LW, Stubbins MJ, Forman D, *et al*. Identification of genetic polymorphism at the glutathione S-transferase Pi locus and association with susceptibility to bladder, testicular and prostate cancer. *Carcinogenesis* 1997;18:641-4.
- 26 Mittman C, Barbela T, Lieberman J. Alpha₁-antitrypsin deficiency as an indicator of susceptibility to pulmonary disease. *J Occup Med* 1973;15:33-8.
- 27 Black LF, Kueppers F. Alpha₁-antitrypsin deficiency in nonsmokers. *Am Rev Respir Dis* 1978;117:421-8.
- 28 Standford AJ, Weir TD, Par PD. Genetic risk factors for chronic obstructive pulmonary disease. *Eur Respir J* 1997;10:1380-91.
- 29 Tarján E, Magyar P, Vácz Z, *et al*. Longitudinal lung function study in heterozygous PiMZ phenotype subjects. *Eur Respir J* 1994;7:2199-204.
- 30 McDonagh DJ, Nathan ST, Knudson RJ, *et al*. Assessment of alpha-1-antitrypsin deficiency heterozygosity as a risk factor in the etiology of emphysema. *J Clin Invest* 1979;63:299-309.
- 31 Morse JO, Lebowitz MD, Knudson RJ, *et al*. A community study of the relation of alpha-antitrypsin levels to obstructive lung diseases. *N Engl J Med* 1975;292:278-81.
- 32 Webb DR, Hyde RW, Schwartz RH, *et al*. Serum α_1 -antitrypsin variants: prevalence and clinical spirometry. *Am Rev Respir Dis* 1973;108:918-25.
- 33 Cole RB, Nevin NC, Blundell G, *et al*. Relation of alpha-1-antitrypsin phenotype to the performance of pulmonary function tests and to the prevalence of respiratory illness in a working population. *Thorax* 1976;31:149-57.
- 34 Bruce RM, Cohen BH, Diamond EL, *et al*. Collaborative study to assess risk of lung disease in PiMZ phenotype subjects. *Am Rev Respir Dis* 1984;130:386-90.
- 35 Kalsheker NA, Hodgson IJ, Watkins GL, *et al*. Deoxyribonucleic acid (DNA) polymorphism of the α_1 -antitrypsin gene in chronic lung disease. *BMJ* 1987;294:1511-4.
- 36 Poller W, Meisen C, Olek K. DNA polymorphisms of the α_1 -antitrypsin gene region in patients with chronic obstructive pulmonary disease. *Eur J Clin Invest* 1990;20:1-7.
- 37 Kalsheker NA, Morgan K. Regulation of the α_1 -antitrypsin gene and a disease-associated mutation in a related enhancer sequence. *Am J Respir Crit Care Med* 1994;150:S183-9.
- 38 Poller W, Faber J-P, Scholz S, *et al*. Mis-sense mutation of α_1 -antitrypsin gene associated with chronic lung disease. *Lancet* 1992;339:1538.
- 39 Samilchuk EI, Chuchalin AG. Mis-sense mutation of α_1 -antitrypsin gene and chronic lung disease. *Lancet* 1993;342:624.
- 40 Huang S-L, Su C-H, Chang S-C. Tumor necrosis factor- α gene polymorphism in chronic bronchitis. *Am J Respir Crit Care Med* 1997;156:1436-9.
- 41 Reye RDK, Bale PM. Elastic tissue in pulmonary emphysema in Marfan syndrome. *Arch Pathol* 1973;96:427-31.
- 42 Cupo LN, Pyeritz RE, Olson JL, *et al*. Ehlers-Danlos syndrome with abnormal collagen fibrils, sinus of Valsalva aneurysms, myocardial infarction, panacinar emphysema and cerebral heterotopias. *Am J Med* 1981;71:1051-8.
- 43 Merten DF, Rooney R. Progressive pulmonary emphysema associated with congenital generalized elastolysis (cutis laxa). *Radiology* 1974;113:691-2.
- 44 Abboud RT, Yu P, Chan-Yeung M, *et al*. Lack of relationship between ABH secretor status and lung function in pulp mill workers. *Am Rev Respir Dis* 1982;126:1089-91.
- 45 Kauffmann F, Kleisbauer JP, Cambon-de Mouzon A, *et al*. Genetic markers in chronic air-flow limitation: a genetic epidemiologic study. *Am Rev Respir Dis* 1983;127:263-9.
- 46 Higgins MW, Keller JB, Becker M, *et al*. An index of risk for obstructive airways disease. *Am Rev Respir Dis* 1982;125:144-51.
- 47 Kauffmann F, Frette C, Q-T Pham, *et al*. Associations of blood group-related antigens to FEV₁, wheezing, and asthma. *Am J Respir Crit Care Med* 1996;153:76-82.
- 48 Tager IB, Speizer FE. Risk estimates for chronic bronchitis in smokers: a study of male-female differences. *Am Rev Respir Dis* 1976;113:619-25.
- 49 Dockery DW, Speizer FE, Ferris BG Jr, *et al*. Cumulative and reversible effects of life-time smoking on simple tests of lung function in adults. *Am Rev Respir Dis* 1988;137:286-92.
- 50 Prescott E, Bjerg AM, Andersen PK, *et al*. Gender difference in smoking effects on lung function and risk of hospitalization for COPD: results from a Danish longitudinal population study. *Eur Respir J* 1997;10:822-7.
- 51 Stebbings JH Jr. A survey of respiratory disease among New York City postal and transit workers: IV. Racial differences in the FEV₁. *Environ Res* 1973;6:147-58.