SCIENTIFIC REPORT

Response of choroidal blood flow to carbogen breathing in smokers and non-smokers

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Br J Ophthalmol 2004;88:776-781. doi: 10.1136/bjo.2003.031773

Aim: To investigate a potential difference in ocular vascular reactivity during carbogen breathing in optic nerve head, choroid, and retina between healthy smokers and non-smokers.

Methods: 25 (13 smokers and 12 non-smokers) healthy male volunteers participated in this observer masked, two cohort study. During inhalation of carbogen (5% CO₂ and 95% O₂) over 10 minutes measurements were taken using laser Doppler flowmetry to assess submacular choroidal and optic nerve head blood flow, laser interferometry to assess fundus pulsation amplitudes, and retinal vessel analyser (RVA) to assess retinal vessel diameters.

Results: At baseline choroidal blood flow was higher (p=0.018, ANOVA) in smokers than in non-smokers. During administration of carbogen the response in choroidal blood flow was significantly different between the two groups: there was an increase in non-smokers after carbogen breathing (p=0.048) compared with relatively stable blood flow in smokers (p=0.049 between groups, ANOVA). A similar response pattern was seen for fundus pulsation amplitude, which increased notably after carbogen breathing in non-smokers but not in smokers (p<0.001 between groups, ANOVA). Optic nerve head blood flow and retinal vessel diameters were reduced in both groups to a comparable degree during carbogen breathing.

Conclusion: The study indicated abnormal choroidal vascular reactivity in chronic smokers. These early haemodynamic changes may be related to the increased risk to smokers of developing ocular vascular diseases. The specific mechanisms underlying abnormal choroidal vascular reactivity in chronic smokers remain to be characterised.

hronic smoking is known to be a risk factor for a variety of ocular vascular diseases like hypertensive retino-✓ pathy, age related macular degeneration, and anterior ischaemic optic neuropathy.1 The exact pathomechanisms linking smoking to ocular vascular disease are not fully understood. Animal studies showed that chronic smoking causes a significant increase of choroidal vascular resistance.² In humans, one study found higher blood flow velocities in the ophthalmic artery, the central retinal artery, and the posterior ciliary arteries in chronic smokers by means of a colour Doppler device.³ In contrast, two other studies describe decreased blood flow velocities in these vessels due to chronic smoking also using a colour duplex imaging system.4 5 Acute cigarette smoking, on the other hand, was reported to induce a decrease of blood velocity in the optic nerve head and possibly in the choroid by the means of the laser speckle method, suggesting an increase in vascular resistance.⁶

In a previous study using the 133Xe inhalation method, chronic cigarette smoking reduced the cerebrovascular

responsiveness to 5% CO_2 and 100% O_2 ,⁷ indicating that chronic cigarette smokers have both reduced vasodilator and reduced vasoconstrictor capacity when compared with non-smokers. It is, however, not known whether such a difference also holds true for the ocular circulation.

The present study set out to investigate a potential difference in vascular reactivity of optic nerve head, choroid, and retina during carbogen breathing between healthy smokers and non-smokers.

PATIENTS AND METHODS

Subjects

The present study was performed in adherence to the Declaration of Helsinki and the Good Clinical Practice guidelines. After approval of the study protocol by the ethics committee of the Vienna University School of Medicine and after written informed consent was obtained, 25 healthy male volunteers were enrolled in the study (mean age 25 (SD 3) years, range 19-35). Thirteen participants had been smokers for at least two years and regularly smoked between 15 and 25 cigarettes per day. The other 12 volunteers had no history of smoking. All volunteers were drug free for at least 3 weeks before inclusion and passed a prestudy screening during the 4 weeks before the first study day, which included physical examination and medical history, 12 lead electrocardiogram, blood and urine analysis, random urine drug screen, and ophthalmic examination. Subjects were excluded if any clinically relevant abnormality was found as part of the pretreatment screening. In addition, subjects with ametropia of more than 3 diopters, anisometropia more than 1 diopter, or any evidence of eye disease that might interfere with the purpose of the present trial were excluded.

To objectively distinguish between smokers and nonsmokers, it was necessary to determine the level of cotinine in urine. Cotinine, the major metabolite of nicotine, has a half life of 24 hours and is readily detectable in the smoker's urine even several days after the smoker has terminated smoking. This investigation was done with a homogenous immunoassay (EMIT-technique, Diagnostic Reagents Inc, Sunnyvale, CA, USA).⁸ Sensitivity of this test, defined as the lowest concentration detectable, is 50 ng/mL. Moreover, no significant cross reactivities to other substances exist. In addition, the subjects were asked to complete the Fagerstrom tolerance questionnaire.⁹ This questionnaire shows a significant correlation with daily cigarette consumption.¹⁰

Experimental design

This study followed an observer masked design in two cohorts. Subjects were asked to refrain from alcohol and caffeine for at least 12 hours before trial days. Moreover they

Abbreviations: DBP, diastolic blood pressure; FPA, fundus pulsation amplitude; LDF, laser Doppler flowmetry; MAP, mean arterial blood pressure; ONH, optic nerve head; PR, pulse rate; RBC, red blood cell; RVA, retinal vessel analyser; SBP, systolic blood pressure. were asked to desist from smoking for at least 2 hours before measurements to rule out acute effects of cigarette smoking. Dilatation of one pupil was obtained with tropicamide (Mydriaticum "Agepha" Augentropfen, AGEPHA GmbH, Vienna, Austria).

The actual time schedule of the study day is presented in table 1. After a 20 minute resting period in a sitting position, baseline measurements of fundus pulsation amplitude were obtained during breathing of room air. Stable baseline conditions were established and ensured by repeated measurements of blood pressure. Thereafter baseline measurements of choroidal blood flow with laser Doppler flowmetry were done for 2 minutes. The laser Doppler measurements were continued without cessation during breathing of carbogen (Oxymix 5%, AGA, Vienna, Austria (certified gases for human use)) over a period of 10 minutes. Immediately after cessation of the laser Doppler measurements another measurement of fundus pulsation amplitude was done when gas delivery was still on. The gas was delivered through a partially expanded reservoir bag at atmospheric pressure using a two valve system to prevent rebreathing. Thereafter a minimum time span of 30 minutes was scheduled to re-establish baseline conditions. Two more carbogen breathing periods followed. Measurements were now done with laser Doppler flowmetry in the optic nerve head (period 2) and with the Zeiss retinal vessel analyser (period 3) following the time schedule as described above. Blood gas analysis and laser interferometry were done at baseline and at the end of the first breathing period. The time span between two subsequent breathing periods was at least 30 minutes.

Systemic haemodynamics

Systolic, diastolic, and mean arterial blood pressure (SBP, DBP, and MAP) were measured on the upper arm by an automated oscillometric device. Pulse rate (PR) was automatically recorded from a finger pulse oximeter (HP-CMS patient monitor; Hewlett-Packard, Palo Alto, CA, USA). Systemic haemodynamics were measured in 2 minute intervals during carbogen breathing periods and in 10 minute intervals during resting periods. Pulse rate and a real time ECG were monitored continuously.

Laser Doppler flowmetry

Continuous measurements of laser Doppler flowmetry (LDF) choroidal blood flow parameters (ChBVel, ChBVol, and ChBF) were performed.¹¹ A commercially available fundus camera based system was used (Oculix 4000, Oculix Sarl, Arbaz, Switzerland). With this technique, the vascularised tissue is illuminated by coherent laser light, and light scattered by the moving red blood cells (RBCs) undergoes a frequency shift. Based on a model of light scattering in tissue¹² the following flow parameters are calculated: mean RBCs velocity in Hz, blood volume, and blood flow in

arbitrary units. All these flow parameters were then averaged over periods of two minutes.

In the present study laser Doppler flowmetry was performed in the fovea to assess choroidal blood flow,¹¹ and at the neuroretinal rim to assess optic disc blood flow.

Fundus pulsation measurement with laser interferometry

Pulse synchronous pulsations of the eye fundus were assessed by laser interferometry. The method is described in detail by Schmetterer *et al.*¹³ Briefly, the eye is illuminated by the beam of a single mode laser diode along the optical axis. The light is reflected at both the front side of the cornea and the retina. The two remitted waves produce interference fringes from which the distance changes between cornea and retina during a cardiac cycle can be evaluated. The fundus pulsation amplitude (FPA), which is the maximum distance change between cornea and retina during the cardiac cycle, has been shown to estimate pulsatile blood flow in the choroid.¹⁴ Measurements were performed in the fovea to assess pulsatile choroidal blood flow.

Zeiss retinal vessel analyser

Retinal vessel diameters were determined from retinal images recorded with a fundus camera based system. The Zeiss retinal vessel analyser (RVA) is a commercially available system which comprises a fundus camera (Zeiss FF 450, Jena, Germany), a video camera, a real time monitor, and a personal computer with an analysing software for the accurate determination of retinal arterial and venous diameters.15 For this purpose the fundus is imaged onto the charge coupled device chip of the video camera. Because of the absorbing properties of haemoglobin each blood vessel has a specific transmittance profile. The consecutive fundus images are digitised using a frame grabber. In addition, the fundus image can be inspected on the real time monitor. Evaluation of the retinal arterial diameters was done online, and of venous diameters offline from the recorded video tapes. Our previous data showed excellent reproducibility with the Zeiss retinal vessel analyser.16

Blood gas analysis

Blood gas values were determined from capillary blood samples of the earlobe. After applying an ointment (Finalgon, Thomae, Biberach, Germany) locally to the earlobe to induce capillary vasodilatation, a lancet incision was made. The arterialised blood was drawn into a thin glass capillary tube. Arterial oxygen tension (pO_2) , carbon dioxide tension (pCO_2) , and oxygen saturation (SaO_2) were determined with an automatic blood gas analysis system (AVL 995 Hb, Graz, Austria).

At least					
30 minutes	\downarrow	2 minutes	\downarrow	10 minutes	\downarrow
Resting period	Fundus pulsation measurement	Baseline measurements with LDF in the macula	Start of carbogen breathing	LDF measurement in the macula during breathing of carbogen	Fundus pulsation measuremer
Resting period		Baseline measurements with LDF in the optic nerve head	Ū	LDF measurement in the optic nerve head during breathing of carbogen	
Resting period		Baseline measurements with the Zeiss retinal vessel analyser		Retinal vessel diameter measurement during breathing of carbogen	

DATA ANALYSIS

Statistical analysis was done with Statistica for Windows (Statsoft Inc (1997), Tulsa, OK, USA). All outcome parameters were averaged over periods of 2 minutes. Effects of carbogen breathing on haemodynamic parameters were analysed by *t* test and repeated measure ANOVA. A p value of <0.05 was considered the level of significance. For data description, relative values are given as mean (SEM).

RESULTS

The results of the Fagerstrom Tolerance Questionnaire were 0 (0) points for the non-smoking group and 4.0 (0.6) points for the smoking group, indicating nicotine dependence among the smokers. Cotinine levels in urine were 45.1 (11.5) ng/ml in non-smokers and 2025.7 (336.1) ng/ml in smokers. This again clearly indicates that the smokers were adequately selected, because the urine cotinine concentration in non-smokers is normally below 500 ng/ml.¹⁷

At baseline (table 2) a significantly lower diastolic blood pressure (p = 0.041, ANOVA) was seen in non-smokers as compared with smokers. In addition, a lower choroidal blood cell volume (p = 0.026, ANOVA) as well as a lower choroidal blood flow (p = 0.018, ANOVA) were observed in non-smokers. No significant differences in pO₂, pCO₂, or SaO₂ were observed between smokers and non-smokers at baseline conditions.

The effects of 10 minutes of breathing carbogen on systemic and ocular haemodynamic parameters are presented in table 3. Administration of carbogen significantly increased systolic blood pressure (+6.0 (1.5)%, p = 0.002 v baseline) and mean arterial pressure (+4.1 (1.6)%, p = 0.024 v baseline) in smokers but not in non-smokers. This change in systolic and mean blood pressure was, however, not significantly different between the two groups. Diastolic blood pressure and pulse rate remained unchanged during carbogen breathing in both groups.

During administration of carbogen choroidal blood cell volume tended to decrease in smokers whereas it increased in

Parameters	Smokers	Non-smokers	p Value (ANOVA
SBP (mm Hg)	118 (3)*	117 (3)	0.845
DBP (mm Hg)	67 (3)	56 (4)	0.041
MAP (mm Hg)	85 (3)	76 (3)	0.058
PR (beats/min)	71 (2.0)	69 (3.0)	0.553
Choroidal red blood cell velocity (AU)	0.46 (0.02)	0.47 (0.02)	0.763
Choroidal red blood cell volume (AU)	0.29 (0.03)	0.21 (0.02)	0.026
Choroidal blood flow (AU)	11.8 (1.0)	8.6 (0.8)	0.018
ONH red blood cell velocity (AU)	0.35 (0.02)	0.33 (0.02)	0.510
ONH red blood cell volume (AU)	• 0.30 (0.04)	0.25 (0.03)	0.371
ONH blood flow (AU)	9.1 (1.0)	7.1 (0.7)	0.149
Arterial retinal diameter (µm)	125.8 (4.7)	125.2 (4.0)	0.924
Venous retinal diameter (µm)	158.8 (4.5)	139.5 (3.9)	0.091
Fundus pulsation amplitude (um)	3.6 (0.3)	3.8 (0.2)	0.713
PO ₂ (mm Ha)	83.4 (2.2)	83.9 (1.1)	0.824
PCO ₂ (mm Ha)	41.1 (1.0)	42.1 (0.9)	0.438
SaO ₂ (%)	95.6 (0.4)	95.9 (0.2)	0.603

SBP, systolic blood pressure; DBP, diastolic blood pressure; MAP, mean arterial pressure; PR, pulse rate; AU, arbitrary units; ONH, optic nerve head. *Data are presented as mean (SEM). Table 3Effects of 10 minutes of breathing of carbogenon systemic and ocular haemodynamic parameters(% change from baseline, t test, post hoc testing) andsignificant differences between the two groups (ANOVA)

Parameters	Smokers (%)	Non-smokers (%)	p Value (ANOVA)
SBP (mm Hg)	+6.0 (1.5)*	+0.8 (2.5)	0.187
DBP (mm Hg)	+3.9(3.0)	+26.2(17.9)	0.251
MAP (mm Hg)	+4.1(1.6)	+8.8 (4.2) p=0.063	0.667
PR (beats/min)	-3.1(1.5) p=0.067	+0.7 (2.8) p = 0.793	0.919
Choroidal red blood cell velocity (AU)	+2.9(1.5) p=0.079	+1.9(2.5) p=0.469	0.675
Choroidal red blood cell volume (AU)	-9.1(5.2) p=0.103	+6.3 (4.7) p=0.203	0.032
Choroidal blood flow (AU)	−5.7 (5.0) p=0.269	+8.0 (3.6) p=0.048	0.049
ONH red blood cell velocity (AU)	-5.2(4.3) p=0.246	-7.9 (5.1) p=0.154	0.734
ONH red blood cell volume (AU)	–15.3 (5.0) p=0.01	-10.5 (8.3) p=0.237	0.282
ONH blood flow (AU	l)-20.2 (5.4) p=0.003	-21.3 (5.1) p=0.002	0.934
Arterial retinal diameter (um)	-5.6 (1.6) p=0.001	-5.3 (1.7) p=0.013	0.959
Venous retinal diameter (um)	–14.7 (1.4) p< 0.001	–11.0 (1.3) ¤< 0.001	0.104
Fundus pulsation amplitude (um)	-2.5(3.3) p=0.46	+18.1 (2.0) p< 0.001	<0.001
PO ₂ (mm Hg)	+241.5 (36.0) p<0.001	+255.0 (19.0) p< 0.001	0.737
PCO ₂ (mm Hg)	+14.8 (2.6) p<0.001	+10.2 (2.0) p< 0.001	0.149
SaO ₂ (%)	+4.3 (0.1) p< 0.001	+4.1 (0.1) p< 0.001	0.564

non-smokers. This response was significantly different between groups (p = 0.032, ANOVA). The response in choroidal blood to carbogen breathing was also significantly different between the two groups (p = 0.049, ANOVA) with an increase in blood flow in non-smokers (p = 0.048) and no change in smokers (fig 1). Fundus pulsation amplitude markedly increased after carbogen breathing in non-smokers (p<0.001 v baseline) but not in smokers and this difference was highly significant between the two groups (table 3; p<0.001, ANOVA).

Red blood cell volume in the optic nerve head (fig 2) significantly decreased in smokers ($p = 0.01 \nu$ baseline), but not in non-smokers. This effect was, however, not significantly different between the two groups. Red blood cell velocity in response to carbogen breathing tended to decrease in both groups, but this effect was not significant. Accordingly, carbogen reduced optic nerve head blood flow in both groups versus baseline (p < 0.01 each), but no difference between groups was seen.

During administration of carbogen, retinal vessel diameters decreased in both groups to a comparable degree (fig 3).

As expected, both pO_2 and SaO_2 increased notably during carbogen breathing (p<0.001 v baseline for both). The increase of pO_2 and SaO_2 during carbogen breathing was, however, comparable between the two groups. The increase of pCO_2 after carbogen breathing was significant in smokers (p<0.001 v baseline) and non-smokers (p<0.001 v baseline), but not significant between the two groups.



Figure 1 Choroidal red blood cell velocity, volume, and flow during carbogen breathing. Data are presented as mean (SEM) (smokers, n = 13; non-smokers, n = 12). *Significant difference versus baseline (*t* test, post hoc testing). †Significant difference between smokers and non-smokers (ANOVA).

DISCUSSION

The main finding of our study is that breathing of carbogen resulted in an increase of choroidal blood flow parameters in non-smoking volunteers, but not in smokers. No differences in response to carbogen breathing could be observed in systemic haemodynamics, optic nerve had blood flow, retinal vessel diameters, and blood gas values between both groups. This clearly indicates abnormal choroidal vascular responsiveness in smokers, whereas normal response patterns were seen in the optic nerve head and retinal vasculature.

Carbogen, a gas mixture containing 5% CO₂ and 95% O₂, has been used in humans for the treatment of central retinal artery occlusion. The amount of CO₂ is assumed to prevent oxygen induced vasoconstriction and therefore maintain or even increase blood flow while providing the retina with increased oxygenation.¹⁸ The effects of carbogen on retinal circulation have already been investigated by several authors. A recent study of our group showed that carbogen breathing decreases retinal vessel diameter, retinal blood velocity, and



Figure 2 Optic nerve head red blood cell velocity, volume, and flow during carbogen breathing. Data are presented as mean (SEM) (smokers, n = 13; non-smokers, n = 12). *Significant difference versus baseline (*t* test, post hoc testing).

retinal blood flow.¹⁹ This is in accordance with another study also observing reduced retinal blood flow during carbogen breathing.²⁰ In contrast, other groups found increased retinal perfusion, using the blue field entoptic technique and scanning laser ophthalmoscopy respectively.^{21 22} Scanning laser ophthalmoloscopy, however, measures blood cell velocity only and the results cannot necessarily be extrapolated to blood flow. The blue field entoptic system assesses white blood cell flux, and potential differences between this method and retinal blood flow measurements with combined laser Doppler velocimetry and the retinal vessel analyser have been discussed in detail.²³

It is well known that choroidal blood flow is strongly dependent on pCO_2 and only slightly influenced by changes in pO_2 .²⁴ In humans, even pronounced changes in arterial oxygen tension have little impact on CO_2 induced changes in subfoveal choroidal blood flow.²⁴ This is in accordance with the findings of the present study in healthy subjects. In our study carbogen decreased optic nerve head blood flow in both



Figure 3 Retinal venous and arterial diameters during carbogen breathing. Data are presented as mean (SEM) (smokers, n = 13; non-smokers, n = 12). *Significant difference versus baseline (*t* test, post hoc testing).

groups. Harris et al reported that optic nerve head blood flow is increased by hypercapnia by 28% in healthy subjects using the same technique as applied in the present study,²⁵ whereas it is reduced by carbogen, as indicated by our results. Hence it may be speculated that the optic nerve head is more oxygen sensitive or less CO2 sensitive than the choroid. These findings may in part be explained by the fact that the optic nerve head is supplied by the posterior ciliary arteries as well as by the central retinal artery. Our findings in the optic nerve head are, however, in contrast to investigations of optic nerve head blood flow during carbogen breathing using the Heidelberg Retina Flowmeter, where carbogen increased optic nerve head blood flow.26 However, these previous results cannot be directly compared with our data, because laser Doppler flowmetry was applied over the whole optic disc area including visible larger vessels. With both techniques applied for the assessment of choroidal perfusion, an abnormal response to carbogen breathing was evident.

The mechanisms underlying CO₂ induced vasodilation in the eye and the brain are not fully understood. Intra- and extracellular acidosis,²⁴ adenosine,²⁷ nitric oxide,²⁸ ATP sensitive potassium channels,29 and the membrane calcium transport³⁰ are assumed to play a role in hypercapnia induced vasodilation in these vascular beds. Moreover, NO was shown to be involved in hypercapnia induced vasodilatation in the human choroid.³¹ NO also appears to facilitate the action of other vasodilators during hypercapnia in the brain.³² A number of studies in humans tried to characterise which of these pathways are altered in chronic smokers. Smokers not only exhibited an impairment in nitric oxide mediated vasodilation,33 but also showed a defect in the vasoconstrictor response to angiotensine-I.34 Endothelial function is further altered by the free radical burden of cigarette smoke and a dysfunctional nitric oxide synthase III (NOS III) due to tetrahydrobiopterin (BH4) depletion in chronic smokers.35

Hence, one may hypothesise that abnormal endothelial function contributes to the results obtained for the choroid in the present study, although this hypothesis remains to be proved in future studies. In addition, future research is required to clarify why the response to hypercapnia in chronic smokers is altered in the choroid, but not in the optic disc or retina.

When discussing the results of the present study one needs to consider the limitations of each method. Laser Doppler flowmetry measures choroidal blood flow in relative units only, and absolute values are influenced by perfusion values as well as by the scattering properties of tissue. Therefore one must be careful to interpret the higher baseline laser Doppler flowmetry value in smokers as a proof of increasing basal choroidal blood flow in these subjects. This limitation does, however, not apply for relative changes as observed during carbogen breathing. With fundus pulsation amplitude measurements, one needs to consider that only the pulsatile component of choroidal blood flow is measured. However, consistency between laser Doppler flowmetry and fundus pulsation amplitude measurements was acceptable in the present study. We did not assess blood flow in the retina in this study and our data are limited to diameter information. In principle such measurements could have been done with bidirectional laser Doppler velocimetry; however, this study schedule was already extremely exhausting for the participating subjects.

In conclusion, the present study indicates abnormal vascular reactivity in smokers. These early haemodynamic changes may be related to the increased risk to smokers of developing ocular vascular disease. The specific mechanisms—either endothelial or humoral—that are foremost perturbed by chronic smoking, remain to be characterised.

ACKNOWLEDGEMENTS

Financial support by the Medizinisch-wissenschaftlicher Fonds des Bürgermeisters der Bundeshauptstadt Wien. (Project #2190) is gratefully acknowledged.

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Accepted 17 November 2003

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