In Vitro Antibacterial Activity of the Pyrrolopyrazolyl-Substituted Oxazolidinone RWJ-416457

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RWJ-416457, an investigational pyrrolopyrazolyl-substituted oxazolidinone, inhibited the growth of linezolid-susceptible staphylococci, enterococci, and streptococci at concentrations of $\leq 4 \mu \text{g/ml}$, generally exhib**iting two- to fourfold-greater potency than that of linezolid. Time-kill studies demonstrated bacteriostatic effects for both RWJ-416457 and linezolid.**

Gram-positive bacteria are a frequent cause of infection among patients in the hospital and community (4). The emergence of vancomycin-resistant *Staphylococcus aureus* and the increasing incidence of community-acquired methicillin-resistant *S*. *aureus* (MRSA) (6), penicillin- and macrolide-resistant *Streptococcus pneumoniae* (2, 8), and vancomycin-resistant enterococci are indications of the increasing resistance of grampositive bacteria to available antimicrobial agents and provide an impetus to discover novel or more efficacious antimicrobial agents (2–4).

Oxazolidinones are one of the newest classes of antibacterial agents, with linezolid as the only member of this class to gain regulatory approval thus far (14). Linezolid is currently approved to treat skin and respiratory infections caused by grampositive pathogens, including multidrug-resistant staphylococci, enterococci, and streptococci (14). It is currently the only drug that can be used both orally and parenterally to treat infections caused by MRSA and vancomycin-resistant enterococci. In the quest for a new oxazolidinone with enhanced characteristics compared with linezolid, RWJ-416457 (Fig. 1), an investigational, orally active, oxazolidinone, was identified (7a, 11a).

This study describes the in vitro activity of RWJ-416457 against clinically important bacteria using linezolid and other relevant agents as comparators. Targeted organisms included susceptible and multidrug-resistant staphylococci, enterococci, and pneumococci, *Haemophilus influenzae*, and *Moraxella catarrhalis* from the Johnson & Johnson Pharmaceutical Research & Development culture collection and the atypical intracellular respiratory tract pathogens *Chlamydia pneumoniae*, *Legionella pneumophila*, and *Mycoplasma pneumoniae* from Focus Bio-Inova.

MICs were determined by the CLSI (formerly NCCLS) broth microdilution procedure for susceptibility testing for bacteria that grow aerobically (11). *C*. *pneumoniae* susceptibility testing was performed in HEp-2 cells by the method of Hammerschlag et al. (7). Frozen aliquots of *C*. *pneumoniae* were diluted in antibiotic-free *Chlamydia* overlay medium (COM) and inoculated into a 72-h monolayer of HEp-2 cells. After preincubation, COM plus antibiotic were serially diluted in twofold steps. Following a 72-h incubation at 35°C, the monolayer was fixed and stained. The MIC end point was determined as the lowest concentration that showed significant or complete inhibition of inclusion-forming units. *M*. *pneumoniae* susceptibility was determined by the microbroth method of Waites et al. (13). Serial 10-fold dilutions of each isolate were prepared in SP4 glucose broth (Remel) to determine the number of color changing units (CCU). Microtiter plates were prepared with antibiotics serially diluted in antibiotic-free SP4 glucose broth, inoculated (104 CCU/ml), and incubated at 35°C until color changed in control wells. The MIC end point was the lowest concentration at which growth was inhibited on the basis of color change. *L*. *pneumophila* susceptibility was performed by broth microdilution. Briefly, the inoculum was suspended in buffered yeast extract broth and transferred to microtiter wells containing serial dilutions of antibiotic and incubated aerobically at 34°C to 36°C for 48 h. The MIC end point was determined as the lowest concentration that inhibited visible growth.

Time-kill experiments with RWJ-416457 and linezolid were performed at four times the MIC by CLSI methodology (10) against *S*. *pneumoniae* ATCC 6305 and OC 4446, *Enterococcus faecalis* ATCC 29212 and ATCC 51299, and *S*. *aureus* ATCC 29213 and MRSA OC 2878. The limit of detection was 100 CFU/ml.

Oxazolidinones have a unique mechanism of action, inhib-

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Organism	No. of isolates	Compound	MIC range $(\mu g/ml)$	MIC ₅₀ $(\mu g/ml)$	MIC ₉₀ $(\mu g/ml)$
Streptococcus pyogenes (37 macrolide-resistant isolates)	39	RWJ-416457 Linezolid Erythromycin	$0.25 - 0.5$ $1 - 2$ $0.03 - > 16$	0.5 1 8	0.5 $\overline{2}$ >16
S. pneumoniae All	108	RWJ-416457 Linezolid	$0.06 - 1$ $0.25 - 2$	0.5 1	0.5 $\mathbf{1}$
Penicillin-resistant	31	RWJ-416457 Linezolid Penicillin	$0.25 - 1$ $0.5 - 2$ $2 - 4$	0.5 1 $\overline{2}$	0.5 1 $\overline{4}$
Macrolide-resistant	51	RWJ-416457 Linezolid Erythromycin	$0.25 - 1$ $0.5 - 2$ $1 - > 16$	0.5 1 $\overline{4}$	0.5 $\mathbf{1}$ >16
Fluoroquinolone-resistant	20	RWJ-416457 Linezolid Levofloxacin	$0.06 - 1$ $0.5 - 2$ $8 - > 32$	0.5 0.5 16	0.5 1 >16
H. influenzae	$\overline{4}$	RWJ-416457 Linezolid Erythromycin	$8 - > 16$ $4 - 16$ $2 - 8$	NA NA NA	NA NA NA
M. catarrhalis	12	RWJ-416457 Linezolid Erythromycin	$2 - 4$ $4 - 8$ $0.12 - 0.25$	$\mathbf{2}$ 8 0.25	$\overline{4}$ 8 0.25
L. pneumophila	17	RWJ-416457 Linezolid Levofloxacin	$2 - 64$ $1 - 64$ $0.03 - 0.5$	4 $\overline{4}$ 0.12	8 16 0.5
M. pneumoniae	17	RWJ-416457 Linezolid Levofloxacin	$16 - 32$ $64 - > 64$ $0.5 - 1$	16 >64 0.5	32 >64 1
C. pneumoniae	3	RWJ-416457 Linezolid Levofloxacin	$16 - 64$ $64 - > 64$ 0.5	NA NA NA	NA NA NA

TABLE 1—*Continued*

^a Isolates received through the Network on Antimicrobial Resistance in *^S*. *aureus* (NARSA) program. *^b* NA, not applicable.

iting bacterial protein synthesis by binding at the peptidyl transferase center of the 50S ribosomal subunit; studies suggest this binding interferes with ribosomal assembly and formation of the initial peptide bond (1, 5, 9, 12). In a bacterial cell-free transcription-dependent translation assay system, RWJ-416457 inhibited protein synthesis at a concentration twofold lower than linezolid, with 50% inhibitory concentrations of 1.4 and $2.9 \mu M$, respectively (D. Montenegro and A. M. Queenan, unpublished data).

The in vitro antibacterial activities of RWJ-416457 and linezolid against *S*. *aureus* and coagulase-negative staphylococci (CoNS) are shown in Table 1. RWJ-416457 had $MIC₉₀$ values of \leq 2μ g/ml against staphylococci (including methicillinsusceptible *Staphylococcus aureus* [MSSA], MRSA, vancomycin-intermediate *S*. *aureus*, and CoNS) (Table 1), consistently twofold lower than those of linezolid. Against a single vancomycin-resistant *S*. *aureus* isolate, RWJ-416457 was equipotent to linezolid, with an MIC of 1 μ g/ml (B. Bozdogan and P. C. Appelbaum, personal communication). Against vancomycin-susceptible and -resistant *E*. *faecalis* and *Enterococcus faecium*, RWJ-416457 had MIC₉₀ values of 1 μ g/ml, two- and

fourfold lower than those of linezolid, respectively (Table 1). As illustrated by the MIC distributions of RWJ-416457 and linezolid against *S*. *aureus* (MSSA and MRSA), *E*. *faecium*, and *E*. *faecalis* (Fig. 2), individual MICs of RWJ-416457 were generally two- to fourfold lower than those of linezolid against these pathogens.

Against linezolid-resistant staphylococci and enterococci, RWJ-416457 MICs ranged from 4 to 32 μ g/ml, values that were again two- to fourfold lower than those of linezolid. Because concentrations of 16 and 32 μ g/ml are not likely to be achieved clinically on the basis of the anticipated drug levels and pharmacodynamic profile (6a), this compound is not expected to have utility against most linezolid-resistant isolates.

When tested against respiratory pathogens, the $MIC₉₀$ of RWJ-416457 for penicillin-, macrolide-, and fluoroquinoloneresistant *S. pneumoniae* was 0.5 µg/ml, twofold lower than that of linezolid. RWJ-416457 MICs were 2 to $>$ 16 μ g/ml against the gram-negative pathogens *H*. *influenzae* and *M*. *catarrhalis*. RWJ-416457 and linezolid had MIC₉₀ values of 8 and 16 μ g/ ml, respectively, against *L*. *pneumophila* and MICs of \geq 16 and ≥64 µg/ml, respectively, against *M*. *pneumoniae* and *C*. *pneu*-

FIG. 2. (a) MIC distribution of RWJ-416457 and linezolid against enterococci (36 *E*. *faecium* isolates and 42 *E*. *faecalis* isolates). (b) MIC distribution of RWJ-416457 and linezolid against *S*. *aureus* (72 MSSA isolates, 102 hospital-associated MRSA isolates, and 13 community-acquired MRSA isolates). The white bars depict RWJ-416457 values, and the black bars depict linezolid values. The numbers of isolates are indicated above the bars.

FIG. 3. Time-kill analysis of isolates treated with RWJ-416457 or linezolid at four times the MIC. Isolates were grown in control medium (\blacklozenge) , medium with RWJ-416457 (\square), and medium with linezolid (\blacklozenge). Against MRSA OC 2878, the RWJ-416457 MIC was $1 \mu g/ml$ and the linezolid MIC was 2 μ g/ml; against *E. faecalis* ATCC 51922, the RWJ-416457 MIC was 1 μ g/ml and the linezolid MIC was 4 μ g/ml.

moniae. Thus, although RWJ-416457 generally retained at least a twofold advantage in potency over linezolid against the atypical respiratory tract pathogens, MICs of both agents frequently exceeded $8 \mu g/ml$.

Time-kill experiments demonstrated that both RWJ-416457 and linezolid were bacteriostatic against the six *S*. *pneumoniae*, *S*. *aureus*, and *E*. *faecalis* strains tested at four times the MIC levels (Fig. 3 for representative time-kill curves). These observations are consistent with the protein synthesis inhibitory mechanism of action for both agents.

In conclusion, the phase 1 investigational oxazolidinone RWJ-416457 demonstrated inhibitory activity against many bacteria resistant to vancomycin, macrolides, and fluoroquinolones. RWJ-416457 generally had two- to fourfold-lower MICs than those of linezolid against most pathogens tested. Its in vitro microbiological activity warrants further investigation to determine an appropriate dosing regimen, in addition to its safety and efficacy in human trials.

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