References

- Bauer F, Urdaci M, Aigle M, Crouzet M (1993) Alteration of a yeast SH3 protein leads to conditional viability with defects in cytoskeletal and budding patterns. Mol Cell Biol 13:5070-5084
- Crouzet M, Urdaci M, Dulau L, Aigle M (1991) Yeast mutant affected for viability upon nutrient starvation: characterization and cloning of the RVS161 gene. Yeast 7:727-743
- David C, Solimena M, De Camilli P (1994) Autoimmunity in Stiff-Man syndrome with breast cancer is targeted to the Cterminal region of human amphiphysin, a protein similar to the yeast proteins, Rvs167 and Rvs161. FEBS Lett 351:73-79
- De Camilli P, Thomas A, Cofiell R, Folli F, Lichte B, Piccolo G, Meinck H-M, et al (1993) The synaptic vesicle-associated protein amphiphysin is the 128-kD autoantigen of Stiff-Man syndrome with breast cancer. J Exp Med 178:2219–2223
- Inglehearn CF, Carter SA, Keen TJ, Lindsey J, Stephenson AM, Bashir R, Al-Maghtheh M, et al (1993) A new locus for autosomal dominant retinitis pigmentosa on chromosome 7p. Nat Genet 4:51-53
- Inglehearn CF, Keen TJ, Al-Maghtheh M, Gregory CY, Jay MR, Moore AT, Bird AC, et al (1994) Further refinement of the location for autosomal dominant retinitis pigmentosa on chromosome 7p (RP9). Am J Hum Genet 54:675–680
- Kremer H, Pinckers A, van den Helm B, Deutman AF, Ropers H-H, Mariman CM (1994) Localization of the gene for dominant cystoid macular dystrophy on chromosome 7p. Hum Mol Genet 3:299-302
- Lichte B, Veh RW, Meyer HE, Kilimann MW (1992) Amphiphysin, a novel protein associated with synaptic vesicles. EMBO J 11:2521-2530
- Yamamoto R, Li X, Winter S, Francke U, Kilimann MW (1995) Primary structure of human amphiphysin, the dominant autoantigen of paraneoplastic Stiff-Man syndrome, and mapping of its gene (AMPH) to chromosome 7p14p13. Hum Mol Genet 4:265-268

Address for correspondence and reprints: Dr. Manfred W. Kilimann, Institut für Physiologische Chemie, Ruhr-Universität Bochum, D-44780 Bochum, Germany.

Am. J. Hum. Genet. 57:972-975, 1995

Localization of the Candidate Gene D-Amino Acid Oxidase Outside the Refined I-cM Region of Spinocerebellar Ataxia 2

To the Editor:

Spinocerebellar ataxia 2 (SCA2) is one form of the neurodegenerative autosomal dominant cerebellar ataxias (for review, see Harding 1983) and has been linked to chromosome 12q (Gispert et al. 1993*b*) in 25 previously

described and 13 new families from a founder collective of >500 patients in Holguín, Cuba (Auburger et al. 1990; Gispert et al. 1993a). Although SCA2 in most patients cannot be distinguished from other spinocerebellar ataxias by clinical criteria, in some patients it exhibits a particular phenotype with early neuropathy/ late slow saccades and late myoclonus (Orozco et al. 1990). Autopsy in 11 patients demonstrated olivoponto-cerebellar atrophy with a selective sparing of the dentate nucleus (R. Estrada, J. Galarraga, G. Orozco, A. Nodarse, and G. Auburger, unpublished data). Complete allelic association within the Holguín population was established with the microsatellite D12S105 (Hernández et al. 1995), and the candidate region was determined to be within a 6- cM region distal to the marker D12S84, contrasting previous reports by Pulst et al. (1993) and Lopes-Cendes et al. (1994) and according to preliminary data between D12S84 and D12S1329 (Allotey et al. 1994).

The D12S105 sequence (hs262xb9.seq) including 342 bp representing the region of maximal allelic association in the Cuban SCA2 founder effect was subjected to sequence homology analysis at the European Molecular Biology Laboratories database and yielded an almost perfect match with 99.70% similarity with intron 1 of the human D-Amino acid Oxidase (DAmOx) gene, which has previously been shown to be linked to all SCA2 pedigrees worldwide with no recombination (Hernández et al. 1995). The small sequence differences were the result of length variations in the four primitive repeat motifs contained in this intron. DAmOx has previously been mapped by chromosome specific hybrids to chromosome 12 and pseudogenes have not been detected (Fukui and Miyake 1992). Primers designed from exons 1 and 10 of the DAmOx cDNA-sequence (Momoi et al. 1988) gave specific PCR products in the CEPH-YAC838f5 corresponding to the locus D12S105, confirming the physical localization and providing the investigation with an excellent candidate gene for SCA2, since DAmOx is expressed specifically in spinocerebellar tissue (Horiike et al. 1985) without its function being known. A mutation in this DAmOx gene would fit well with previous hypotheses on the pathomechanism of spinocerebellar degeneration, since oral loading tests with glutamate in such patients have demonstrated a decreased metabolism of amino acids glutamate and aspartate (Plaitakis 1982) and since accumulation of the excitotoxic neurotransmitter glutamate is known to lead to cerebellar Purkinje neuron death (Meldrum and Garthwaite 1990), matching well the morphological changes observed in SCA2.

Within intron 1 of the DAmOx gene a second $(AC)_n$ microsatellite is found that we termed D12S105a (AFM-262xb9a). With flanking primers 5'-AGCAGTTGA-GAGGATTGAGAGG-3' and 5'-GCAAGCTTGGAG-

^{© 1995} by The American Society of Human Genetics. All rights reserved. 0002-9297/95/5704-0033\$02.00

				6-36									
MABe	PL	Fi	Ry	Es	Ve	LB	In	Su	AI	Mu	Pu	So	Microsatellite
Q	4	4	S←S	ND+5	S	4	4	S	Z	S	4	4	IGF1
Q	7	đ	5←2	ND+10	ŝ	2	2	80	2	2	2	1	D12S78
QZ	4	4	4+4	ND+8	4	4	6	4	4	4	4	4	PAH-VNTR-Bglil
1	1	1	ND+1	ND+1	1	٦	1	1	1	1	1	1	D12S338
4	4	QZ	7+4	ND+7	4	4	4	4	4	4	4	4	D12S353
ŝ	Q	ę	ND+3	P-4	4	ŝ	3	æ	4	4	4	4	D12S1331
80	8	8	ND-8	ND+8	œ	œ	œ	8	8	œ	œ	œ	D12S330
9	9	9	ND+5	9-QN	9	9	9	9	9	9	9	9	D12S317
1	1	1	1←2	9+1	-1	1	1	1	1	1	1	1	D12S84
S	S	S	5←5	NI←S	S	S	S	S	5	S	5	5	D12S105
S	S	q	6←5	4 ←5	QZ	S	S	S	S	S	S	S	D12S105a
ę	ŝ	ę	4+-3	ę	ŝ	e	ŝ	ę	ŝ	÷	ę	ę	D12S1328
													SCA2
6→∠	6	6	6	6	6	6	6	6	6	6	6	6	D12S1329
6←11	11	QZ	11	11	11	Ð	11	11	11	11	11	11	D12S1333
3←3	ŝ	ŝ	ę	æ	£	ę	ŝ	ŝ	ŝ	ę	ŝ	ŝ	D12S1332
4←5	6←5	S	S	S	£	S	S	S	S	5	S	5+6	D12S1330
4+4 4-4	9-4 4	4	4	4	4	4	4	4	4	4	4	5←3	D12S354
5←5	3←5	ŝ	ŝ	S	9	S	S	S	S	5	S	5←5	D12S79
5←2	4←2	S	7	Ð	£	2	7	2	2	2	2	2←2	D12S369
QZ	QZ	9	5	ŝ	£	2	9	S	Q	Q	Q	Ð	D12S366
QZ	4 ~ 4	4	2	4	4	Ð	QZ	Ð	4	4	Ī	Ð	D12S349
QZ	3+3	£	4	3	3	5	3	7	7	4	ŝ	6→8	D12S86

Haplotypes in the Ancestors of Each of 13 Large Cuban SCA2 Families Together with Intrafamilial Recombinant Haplotypes (Written Always to the Left of the Ancestral Haplotype)

Table I

NOTE.—ND = no data; and NI = not investigated.

TATGTATCC-3' we performed genotyping in 25 Cuban SCA2 families and 2 families from the French Caribbean (Martinique) with probable SCA2 (Belal et al. 1994). Seven alleles were observed vielding a heterogeneity index of .79. In addition, six recently developed microsatellites in the SCA2 candidate interval of 6 cM between D12S84 (AFM116xb8) and D12S79 (AFM067yc5) and eight further new microsatellites were genotyped in these families: D12S129 (Montgomery et al. 1993), D12S338 (AFM291wd9), D12S353 (AFM304wg5), D12S1331 (AFM340xg1), D12S330 (AFM086xd7), D12S317 (AFM065ye9), D12S1328 (AFM240we1), D12S1329 (AFM291xe9), D12S1333 (AFM154tc5), D12S1332 (AFMa128yf1), D12S1330 (AFM312yb1), D12S354 (AFM304wh5), D12S369 (AFM142zc5), D12S366 (AFM351tb1), D12S349 (AFM299zd5) (see Gyapay et al. 1994; CEPH database). This analysis identified recombinant events in families Rs and Es and placed the SCA2 gene to the telomeric side of D12S1328 (see table 1 and fig. 1a), excluding conclusively the DAmOx gene from the SCA2 region with a distance of ~ 1 cM between the D12S105a microsatellite and SCA2. Fine mapping of the SCA2 locus was helped further by the clinical characterization of a previously undescribed Cuban SCA2 family named Be, which consists of 4 branches ABe, MABe, HBe, and CoBe and contains 155 affected members. Within the branch MABe a crossing-over event was identified that places the SCA2 mutation centromeric to D12S1329, thus reducing the candidate region to ~ 1 cM (see fig. 1b) between flanking markers D12S1328 and D12S1329 (map according to Généthon, unpublished data). It is important to note that further analysis of these Rs and MABe recombinants by using newly generated microsatellites from the 1-cM candidate region will be able to reduce the candidate region even further.

In contrast, the investigation of potential distantly related SCA2 families seemed to be of limited value for genetic mapping: Whereas the ancestral haplotype D12S105, D12S105a, D12S1328, D12S1329, D12S1333, and D12S1332 (see table 1) is well preserved among the founders of the 12 published families and 1 new family presented in table 1 and of additional 15 smaller families from Holguín (supporting the notion of a Cuban founder effect), the Cuban D12S1329 allele was not preserved in SCA2 families from France, Tunisia, Canada, or the United States, and the Cuban D12S1328 allele was preserved in only two of six non-Cuban SCA2 families, thus giving little support for a common origin of SCA2 families.

Suzana Gispert,¹ Astrid Lunkes,¹ Nieves Santos,¹ Guillermo Orozco,² Duc Ha-Hao,¹ Tim Ratzlaff,¹ Jorge Aguiar,¹ Isis Torrens,¹ Luis Heredero,¹ Alexis Brice,⁴



Figure 1 *a*, Haplotype analysis in two meioses representative of two branches of the Rs family from the Holguín SCA2 population. Both the partial and the complete disease haplotype represent typings in five affected individuals of the two family branches. *b*, Haplotype analysis in a recombinant branch of MABe (*left*) and a nuclear pedigree representative of the rest of family Be containing 155 patients (*right*).

Géraldine Cancel,⁴ Giovanni Stevanin,⁴ Jean-Claude Vernant,⁶ Alexandra Dürr,⁴ Agnes Lepage-Lezin,⁶ Samir Belal,⁷ Mongi Ben-Hamida,⁷ Stephan Pulst,⁸ Guy Rouleau,⁹ Jean Weissenbach,⁵ Denis LePaslier,⁵ Raju Kucherlapati,¹⁰ Kate Montgomery,¹⁰ Kiyoshi Fukui,¹¹ And Georg Auburger¹

¹Department of Neurology, University Hospital, Düsseldorf; ²Department of Neurology, Lenin Hospital, Holguín, Cuba; ³National Center of Medical Genetics, Havana; ⁴Unité INSERM 289, Hôpital de la Salpetrière, and ⁵Unité de Génétique Moléculaire Humaine, CNRS URA 1445, Institut Pasteur, Paris; ⁶Hôpital Pierre Zobda-Quitman, Fort-de-France, Martinique; ⁷National Institute of Neurology, Tunis; ⁸Cedars-Sinai Medical Center, Los Angeles; ⁹Centre de Recherche en Neurosciences, McGill University, Montreal; ¹⁰Albert Einstein College of Medicine, Bronx; and ¹¹Department of Biochemistry, National Cardiovascular Center Research Institute, Osaka

Acknowledgments

We are grateful to Nicola Bergmann and Dorothea Humbert for their excellent technical assistance and to O. Ritter (DKFZ, Heidelberg) for the help with EMBL database. S.G., A.L. and N.S. were supported by the Deutsche Forschungsgemeinschaft and the Fritz Thyssen Stiftung, J.A. and I.T. by the Deutsche Akademische Austauschdienst and the Deutsche Heredo-Ataxie Gesellschaft, K.F. by special coordination funds for promoting science and technology from STA of Japan and a grant-in-aid for Scientific Research from MESC of Japan. We thank the Cuban Ministry of Health for continuous help and the family members for their collaboration.

References

- Allotey R, Twells R, Orozco G, Cemal C, Heredero L, Weissenbach J, Williamson R, et al (1994) Location of the spinal cerebellar ataxia 2 locus to a 1 cM interval on chromosome 12q23-24.1. Am J Hum Genet Suppl 55:A179
- Auburger G, Orozco Diaz G, Ferreira Capote R, Gispert Sanchez S, Paradoa Perez M, Estrada del Cueto M, Garcia Meneses M, et al (1990) Autosomal dominant ataxia: genetic evidence for locus heterogeneity from a Cuban foundereffect population. Am J Hum Genet 46:1163-1177
- Belal S, Cancel G, Stevanin G, Hentati F, Khati C, Ben Hamida C, Auburger G, et al (1994) Clinical and genetic analysis of a Tunisian family with autosomal dominant cerebellar ataxia type I linked to the SCA2 locus. Neurology 44:1423–1426
- Fukui K, Miyake Y (1992) Molecular cloning and chromosomal localization of a human gene encoding D-amino-acid oxidase. J Biol Chem 267:18631-18638
- Gispert S, Nothers C, Orozco G, Auburger G (1993a) Search of the chromosome locus of autosomal dominant cerebellar ataxia from Holguín, Cuba: exclusion from candidate regions on chromosome 4 and 11q. Hum Hered 43:12-30
- Gispert S, Twells R, Orozco G, Brice A, Weber J, Herdero L, Scheufler K (1993b) Chromosomal assignment of the second locus for autosomal dominant cerebellar ataxia (SCA2) to chromosome 12q23-24.1. Nat Genet 4:295-299
- Gyapay G, Morisette J, Vignal A, Dib C, Fizames C, Millasseau P, Marc S (1994) The 1993-94 Généthon human genetic linkage map. Nat Genet 7:246-339
- Harding AE (1983) Classification of the hereditary ataxias and paraplegias. Lancet 1:1151–1155
- Hernández A, Magarino C, Gispert S, Santos N, Lunkes A, Orozco G, Heredero L, Auburger G (1995) Genetic mapping of the spinocerebellar ataxia 2 (SCA2) locus on chromosome 12q23-24.1. Genomics 25:433-435

- Horiike K, Arai R, Tojo H, Yamano T, Nozaki M, Maeda T (1985) Histochemical staining of cells containing flavoenzyme D-amino acid oxidase based on its enzymatic activity: application of a coupled peroxidation method. Acta Histochem Cytochem 18:539-550
- Lopes-Cendes, Andermann E, Rouleau GA (1994) Evidence for the existence of a fourth dominantly inherited spinocerebellar ataxia locus. Genomics 21:270–274
- Meldrum B, Garthwaite J (1990) Excitatory amino acid neurotoxicity and neurodegenerative disease. Trends Pharmacol Sci 11:379-387
- Momoi K, Fukui K, Watanabe F, Miyake Y (1988) Molecular cloning and sequence analysis of cDNA encoding human kidney D-amino acid oxidase. FEBS Lett 238:180-184
- Montgomery K, LeBlanc J, Tsai P, McNinch J, Ward D, de-Jong P, Kucherlapati R, et al (1993) Characterization of two chromosome 12 cosmid libraries and development of STSs from cosmids mapped by FISH. Genomics 17:682-699
- Orozco G, Nodarse A, Cordoves R, Auburger G (1990) Autosomal dominant cerebellar ataxia: clinical analysis of 263 patients from a homogeneous population in Holguín, Cuba. Neurology 40:1369-1375
- Plaitakis A, Berl S, Yahr MD (1982) Abnormal glutamate metabolism in adult-onset degenerative neurological disorder. Science 216:193-196
- Pulst SM, Nechiporuk A, Starkman S (1993) Anticipation in spinocerebellar ataxia type 2. Nat Genet 5:8-10

Address for correspondence and reprints: Dr. Georg Auburger, Department of Neurology, University Hospital, P.O. Box 101007, 40001 Düsseldorf, Germany. © 1995 by The American Society of Human Genetics. All rights reserved. 0002-9297/95/5704-0034\$02.00

Am. J. Hum. Genet. 57:975-977, 1995

Association Analysis of the Monoamine Oxidase A Gene in Bipolar Affective Disorder by Using Family-Based Internal Controls

To the Editor:

It is well accepted that association studies are a major tool in investigating the contribution of single genes to the development of diseases that do not follow simple Mendelian inheritance pattern (so-called complex traits) (e.g., Lander and Schork 1994). Such major psychiatric diseases as bipolar affective disorder and schizophrenia clearly fall into this category of diseases.

Lim et al. (1994) have recently observed a significant overall association between bipolar affective disorder and alleles of a microsatellite polymorphism at the monoamine oxidase A (MAOA) locus in bipolar patients (n = 57) and controls (n = 59) of western European extraction. The finding of an overall association was replicated by Kawada et al. (1995) in Japanese patients (n = 58) and controls (n = 68), although individ-