## Production of Fumonisins by *Fusarium moniliforme* and *Fusarium proliferatum* Isolates Associated with Equine Leukoencephalomalacia and a Pulmonary Edema Syndrome in Swine

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Fumonisin  $B_1$  (FB<sub>1</sub>) and FB<sub>2</sub> were isolated from corn cultures of both *Fusarium moniliforme* and *Fusarium proliferatum*. Respective concentrations in culture materials of FB<sub>1</sub> and FB<sub>2</sub> ranged from 960 to 2,350 and 120 to 320 µg/g for *F. moniliforme* and from 1,670 to 2,790 and 150 to 320 µg/g for *F. proliferatum*. Thin-layer chromatography, gas chromatography-mass spectroscopy, high-performance liquid chromatography, and liquid secondary ion mass spectroscopy were used for detection. Fumonisins from *F. proliferatum* have not previously been reported.

During the 1989 corn harvest season, the National Veterinary Services Laboratories received numerous reports of outbreaks of equine leukoencephalomalacia (ELEM) and a porcine pulmonary edema syndrome (PPE). The PPE outbreaks were generally confined to the central portion of the United States, while ELEM cases in several states, ranging from Arizona to Maryland, were reported. In almost all cases, feed containing corn and/or corn screenings from the 1989 harvest was implicated as the causative factor. Because ELEM is known to be caused by fumonisin  $B_1$  (FB<sub>1</sub>) (4), a mycotoxin produced by Fusarium moniliforme, and because a PPE-like syndrome caused by feeding F. moniliforme culture material (CM) to swine has been reported (3), feed samples were collected for mycological evaluation and chemical analyses. Nine feed samples were obtained from farms in southeastern Iowa: two were associated with an ELEM case (284A and 284B), five were associated with PPE cases (943A, 567, 615, 317A, and 378B), and two were not associated with animal health problems (943B and 317B). All the samples comprised primarily corn and/or corn screenings. F. moniliforme was isolated from all nine samples, and Fusarium proliferatum was isolated from one ELEM sample (284A), one PPE sample (317A), and one nonproblem sample (943B). Reported here are the results of a study to determine the fumonisin-producing potential of the Fusarium isolates, including the discovery that  $FB_1$  and  $FB_2$  are produced by F. proliferatum. Results of chemical analyses of the feeds are described elsewhere (P. F. Ross, L. G. Rice, R. P. Plattner, G. D. Osweiler, T. M. Wilson, D. L. Owens, H. A. Nelson, and J. L. Richard, Mycopathologia, in press).

Ingredients from the feed samples were cultured initially on a modified pentachloronitrobenzene selective medium and then transferred to potato dextrose agar and carnation leaf agar and identified as described by Nelson et al. (7). Isolates were lyophilized and stored at the Fusarium Re-

CMs were analyzed for FB<sub>1</sub> and FB<sub>2</sub> by thin-layer chromatography, gas chromatography-mass spectroscopy, highperformance liquid chromatography, and liquid secondary ion mass spectroscopy, as previously reported (1, 8, 10). All four techniques were in agreement on the presence of FB<sub>1</sub> and FB<sub>2</sub> in the CMs. Qualitatively, CMs and reference standards of FB1 and FB2 (Division of Food Science and Technology, Research Institute for Nutritional Diseases, Pretoria, South Africa) matched by thin-layer chromatography migration and color of spot, liquid secondary ion mass spectra, gas chromatography-mass spectroscopy retention times and mass spectra, and high-performance liquid chromatography retention times. Quantitatively, concentrations were obtained from high-performance liquid chromatography responses compared with standards (Table 1). Concentrations based on gas chromatography-mass spectroscopy

search Center, Pennsylvania State University, University Park. The 12 isolates were grown on autoclaved corn by the following technique. Fifty grams of locally obtained yellow corn with no detectable FB<sub>1</sub> or FB<sub>2</sub> (detection limit,  $5 \mu g/g$ ) along with 50 g of water was added to a 250-ml beaker and allowed to imbibe at room temperature for 1 h. The beakers were covered with foil and autoclaved for 1 h. After the beakers cooled, the foil was removed and the corn was stirred with a sterile spatula to separate the kernels. The beakers were then covered with a layer of cotton sandwiched between two layers of cheesecloth. The covers were secured with heat-resistant tape, and the beakers were autoclaved again for 1 h. After the cooling period, 1 ml of phosphate-buffered saline (pH 7.4) inoculum (a suspension of conidia from a carnation leaf culture) was introduced through the covering with a needle and syringe. The cultures were incubated in the dark for 2 weeks at 27°C followed by another 2 weeks at 15°C. The CM was then autoclaved, dried at 60°C for 2 to 3 days, ground to a uniform consistency with a Stein mill (Fred Stein, Inc., Atchison, Kans.), and stored at 4°C until analyzed.

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| Feed | Disease | Type of feed <sup>a</sup> | Species and isolate <sup>b</sup> | Concn (µg/g of CM) |                 | FB <sub>1</sub> /FB <sub>2</sub> |
|------|---------|---------------------------|----------------------------------|--------------------|-----------------|----------------------------------|
|      |         |                           |                                  | FB <sub>1</sub>    | FB <sub>2</sub> | concn ratio                      |
| 943A | PPE     | S                         | F. moniliforme, M-5939           | 960                | 170             | 5.6                              |
| 943B | None    | S                         | F. moniliforme, M-5954           | 1,310              | 220             | 5.9                              |
|      |         |                           | F. proliferatum, M-5956          | 2,790              | 320             | 8.7                              |
| 567  | PPE     | S                         | F. moniliforme, M5982            | 1,940              | 220             | 8.8                              |
| 615  | PPE     | S                         | F. moniliforme, M-5986           | 2,300              | 350             | 6.6                              |
| 317A | PPE     | S                         | F. moniliforme, M-5990           | 2,350              | 320             | 7.3                              |
|      |         |                           | F. proliferatum, M-5991          | 1,670              | 150             | 11.1                             |
| 317B | None    | С                         | F. moniliforme, M-5996           | 1,590              | 180             | 8.8                              |
| 378B | PPE     | С                         | F. moniliforme, M-6004           | 1,730              | 120             | 14.4                             |
| 284A | ELEM    | F                         | F. moniliforme, M-5958           | 1,520              | 160             | 9.5                              |
|      |         |                           | F. proliferatum, M-5964          | 1,690              | 190             | 8.9                              |
| 284B | ELEM    | F                         | F. moniliforme, M-5972           | 1,420              | 130             | 10.9                             |

TABLE 1. FB<sub>1</sub> and FB<sub>2</sub> concentrations in corn cultures of F. moniliforme and F. proliferatum isolates from feeds

<sup>*a*</sup> S, Screenings; F, mixed feed; C, whole corn. <sup>*b*</sup> Fusarium Research Center identification number.

response for FB<sub>1</sub> were in agreement with the high-performance liquid chromatography results; differences between the two techniques were less than 25% for all 12 CMs tested. The nine F. moniliforme CMs had  $FB_1$  levels ranging from 960 to 2,350  $\mu$ g/g and FB<sub>2</sub> levels from 130 to 350  $\mu$ g/g. These concentrations are similar to the FB<sub>1</sub> and FB<sub>2</sub> levels of 1,000 and 100  $\mu$ g/g, respectively, reported by Gelderblom et al. (2) for a corn culture of F. moniliforme MRC 826, an isolate from South African corn intended for human consumption (6). The F. moniliforme isolates from nonproblem feeds (M-5954 and M-5996) produced FB<sub>1</sub> and FB<sub>2</sub> levels in the same range as the isolates from problem feeds. The  $FB_1/FB_2$ ratio is relatively constant for all nine F. moniliforme isolates, ranging from 5.6 to 14.4. Ratios for all isolates are similar to the ratio of 10 for MRC 826 and are similar to values for naturally contaminated feedstuffs of 6.0 for an ELEM case from Arizona (10).

The three F. proliferatum CMs had FB<sub>1</sub> concentrations ranging from 1,670 to 2,790  $\mu$ g/g and FB<sub>2</sub> concentrations from 150 to 320  $\mu$ g/g. M-5956, an isolate from a nonproblem feed, was the greatest FB<sub>1</sub> producer of all isolates of F. moniliforme and F. proliferatum that were tested. The FB<sub>1</sub>/FB<sub>2</sub> ratio (8.7 to 11.1) for F. proliferatum isolates is similar to that of the F. moniliforme isolates.

The production of fumonisins by F. proliferatum has not previously been documented. In fact, F. proliferatum has not previously been associated with human or animal toxicoses (6). Its close relationship to F. moniliforme suggests that other close relatives should be tested for fumonisinproducing potential. The fumonisin-producing ability of F. proliferatum suggests a link with ELEM and a possible relationship with PPE. The long-held association of ELEM and F. moniliforme (5, 9) must now be expanded to include F. proliferatum and possibly other Fusaria species. Further study is required to determine the role of fumonisins in PPE.

The production of high levels of fumonisins by isolates from both problem and nonproblem feeds suggests potential for fumonisin contamination in any feed containing F. moniliforme and/or F. proliferatum. No attempt was made to correlate fumonisin-producing potential of the isolates and the levels of fumonisins in the feed samples collected here. Those fumonisin levels, along with levels from 219 other feed samples, have been reported elsewhere (Ross et al., in press).

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