

Multiple Sclerosis and Chronic Autoimmune Encephalomyelitis

A Comparative Quantitative Study of Axonal Injury in Active, Inactive, and Remyelinated Lesions

Barbara Kornek,* Maria K. Storch,*[†]
Robert Weissert,[‡] Erik Wallstroem,[‡]
Andreas Stefferl,*[§] Tomas Olsson,[‡]
Christopher Linington,[§] Manfred Schmidbauer,[¶]
and Hans Lassmann*

From the Division of Neuroimmunology, Brain Research Institute, University of Vienna, Vienna, Austria; the Department of Neurology,[†] Karl-Franzens-University, Graz, Austria; the Neuroimmunology Unit,[‡] Center of Molecular Medicine, Karolinska Hospital, Stockholm, Sweden; the Department of Neuroimmunology,[§] Max Planck Institute of Neurobiology, Martinsried, Germany; and the Department of Neurology,[¶] Hospital Lainz, Vienna, Austria*

Recent magnetic resonance (MR) studies of multiple sclerosis lesions indicate that axonal injury is a major correlate of permanent clinical deficit. In the present study we systematically quantified acute axonal injury, defined by immunoreactivity for beta-amyloid-precursor-protein in dystrophic neurites, in the central nervous system of 22 multiple sclerosis patients and 18 rats with myelin-oligodendrocyte glycoprotein (MOG)-induced chronic autoimmune encephalomyelitis (EAE). The highest incidence of acute axonal injury was found during active demyelination, which was associated with axonal damage in periplaque and in the normal appearing white matter of actively demyelinating cases. In addition, low but significant axonal injury was also observed in inactive demyelinated plaques. In contrast, no significant axonal damage was found in remyelinated shadow plaques. The patterns of axonal pathology in chronic active EAE were qualitatively and quantitatively similar to those found in multiple sclerosis. Our studies confirm previous observations of axonal destruction in multiple sclerosis lesions during active demyelination, but also indicate that ongoing axonal damage in inactive lesions may significantly contribute to the clinical progression of the disease. The results further emphasize that MOG-induced EAE may serve as a suitable model for testing axon-protective therapies in inflammatory demyelinating conditions. (*Am J Pathol* 2000, 157:267–276)

Multiple sclerosis (MS) is a chronic inflammatory disease of the central nervous system characterized by widespread inflammation, focal demyelination, and a variable degree of axonal loss.¹ Axonal damage in MS lesions has recently attracted significant attention, because neuroimaging studies suggest that it may be the major pathological correlate of permanent functional deficit.^{2–13} The presence of axonal degeneration in multiple sclerosis has long been recognized,^{1,14–26} but only recently have efforts been made toward a quantitative assessment of axonal injury with particular emphasis on early, actively demyelinating lesions.^{27,28}

Similarly, the presence of axonal degeneration has been shown in animal models of the human demyelinating disease, such as experimental autoimmune encephalomyelitis (EAE).^{29–31} Detailed, quantitative results which allow a comparison with MS lesions, are so far lacking, however.

In the present study we give a detailed account of acute axonal injury, as revealed by the axonal immunoreactivity for the beta-amyloid-precursor protein (β APP) in myelinated and demyelinated areas of the central nervous system of 22 MS patients. Immunocytochemistry for β APP has been shown to be an early and sensitive marker for axonal damage in various disorders of the central nervous system,^{32,33} including MS.²⁷ Our data show that acute axonal injury occurs at a very high incidence during the phase of active demyelination, at a very early time point of lesion formation. In addition, however, there is a low incidence of axonal damage in inactive demyelinated plaques, which may be responsible for disease progression in the chronic inflammatory inactive stage of the disease. In contrast, remyelination seems to protect axons within MS plaques from further degeneration. The acute phase of axonal destruction in actively demyelinating lesions is closely reflected in chronic autoimmune encephalomyelitis in rats. EAE induced by ac-

Supported by the Austrian Science Foundation Project P 12658-MED, by the Austrian Ministry for Science (GZ 650.223/2-III/2a/99) and the EC Biomed 2-Project BMH 4-97-2027.

Accepted for publication April 3, 2000.

Address reprint requests to Prof. Dr. Hans Lassmann, Division of Neuroimmunology, Brain Research Institute, University of Vienna, Spitalgasse 4, A-1090 Wien, Austria. E-mail: hans.lassmann@univie.ac.at.

Table 1. Number and Characteristics of Patients Included in the Study

Patient no.	Age/sex	Disease duration	Disease course
1	47/F	3.5 months	acute
2	46/F	12 days	acute
3	29/M	1.5 months	acute
4	51/F	7 months	acute
5	46/M	3 months	acute
6	28/F	8 months	acute
7	68/F	1.5 months	acute
8	45/M	3 weeks	acute
9	20/F	4 years	RR/acute
10	35/M	1.5 months	acute
11	35/F	4 years	RR/acute
12	53/M	1 month	RR/acute
13	39/F	3 years	SP
14	43/F	20 years	SP
15	53/F	21 years	SP
16	42/F	11 years	RR
17	45/F	16 years	SP
18	37/F	2 years	PP
19	34/F	13 years	SP
20	33/F	10 years	SP
21	30/M	6 years	SP
22	40/F	10 years	SP

RR, relapsing-remitting MS; SP, secondary progressive MS; PP, primary progressive MS.

tive sensitization with myelin-oligo-dendrocyte glycoprotein (MOG) thus may serve as a good model for testing axon-protective therapeutic strategies in inflammatory demyelination.

Material and Methods

MS Patients and Controls

The study was performed on autopsy tissue from 20 and biopsy tissue from 2 MS patients. Biopsy material was included in this study to determine the reliability of the immunocytochemical techniques in the autopsy cases. In addition, 9 control brains were included in this series from patients (age, 63.56 ± 12.4 years; male/female, 5/4) without evidence of neurological disease or neuropathological alterations. Clinical background data as well as the number and characteristics of lesional areas studied in each patient are summarized in Table 1. All material was fixed in 4% paraformaldehyde and embedded in paraffin wax.

Table 2. Acute Axonal Injury in Multiple Sclerosis Lesions

	Control white matter	Early active	Late active	Inactive + active edge	Inactive	Remyelination
β APP-positive axons/ 0.01 mm ²	0.05 ± 0.02	41.26 ± 5.59	39.85 ± 7.72	5.38 ± 0.91	0.47 ± 0.12	0.07 ± 0.01
-fold increase compared to control WM		825.2	797	107.6	9.4	1.4
<i>P</i> values		CO/EA: < 0.0001	CO/LA: < 0.0001	CO/IA + A: < 0.0001	CO/IA: <0.01	CO/RM: n.s.

EAE Material

Lewis 1N rats were kindly provided by Prof. H. Hedrich.³⁴ EAE was induced in 18 rats by active sensitization with 20 to 100 micrograms of recombinant MOG as described in detail before.³⁵ Three healthy rats of the same strain served as controls. Animals were weighed and examined daily for clinical signs of EAE. At various time points after sensitization (days 11–61), animals were sacrificed and perfused via the aorta with 4% paraformaldehyde. Brains and spinal cords were dissected and routinely embedded in paraffin wax.

Neuropathology and Immunocytochemistry

Serial sections 2 to 4 μm thick were stained with hematoxylin/eosin (HE), Luxol fast blue (LFB)/Periodic acid-Schiff (PAS), and Bielschowsky silver impregnation to assess inflammation, demyelination, and axonal pathology, respectively. Immunohistochemistry was performed on adjacent serial sections using an avidin-biotin or an alkaline phosphatase/anti-alkaline phosphatase technique. Primary antibodies were used against the following targets: Myelin-oligodendrocyte glycoprotein (anti-MOG; Department of Biochemistry, Cardiff, UK), proteolipid protein (anti-PLP; Serotec, Oxford, UK), 2'-3'-cyclic nucleotide phosphodiesterase (anti-CNPase, Affinity Research Products, Ilkeston, UK), myelin basic protein (anti-MBP, BioGenex, San Ramon, CA), human T cells (anti-CD3, Serotec), human macrophages (anti-CD68, Dako, Glostrup, Denmark), human common leukocyte antigen (anti-CD 45, Pharmingen, San Diego, CA), rat T cells (anti-CD43-equivalent, clone W3/13; Harlan Sera-Lab, Loughborough, UK), macrophages/activated microglia in rats (anti-ED 1; Serotec), early activated human hematopoietic macrophages (anti-MRP 14, BMA Biomedicals, Augst, Switzerland; anti-27E10; BMA Biomedicals), beta-amyloid precursor protein (anti-APP, Boehringer Mannheim, Mannheim, Germany). Control sections were incubated in the absence of primary antibody. *In situ* hybridization for PLP mRNA was performed according to Breitschopf et al.³⁶

Selection of Demyelinated Plaques and Definition of Lesional Staging

In both MS and EAE, areas of normal white matter, periplaque white matter, actively demyelinating plaques,

Table 3. Acute Axonal Injury in Normal and Periplaque White Matter of Multiple Sclerosis

	Control white matter	Periplaque white matter (active MS)	Normal white matter (active MS)	Periplaque white matter (inactive MS)	Normal white matter (inactive MS)
β APP-positive axons/0.01 mm ² -fold increase compared to control WM	0.05 ± 0.02	1.07 ± 0.17 21.4	0.27 ± 0.09 5.4	0.07 ± 0.02 1.4	0.05 ± 0.02 1
<i>P</i> values		CO/PP: <0.0001	CO/NWM: < 0.01	CO/PP: n.s.	CO/NWM: n.s.

inactive demyelinated plaques, and remyelinated shadow plaques were selected for further analysis of axonal pathology. Normal white matter (NWM) was defined as an area that showed no evidence of demyelination by macroscopic inspection and histology within the area and the surrounding tissue. Periplaque white matter (PP) represented a strip of tissue of 5 mm adjacent to the border of active or inactive plaques.

The following categories for demyelinated plaques were defined:

Early active lesions (EA): These lesions were heavily infiltrated by T cells and macrophages. Myelin sheaths were in the process of disintegration and macrophages contained degradation products, which were stained by Luxol fast blue and were immunoreactive for all myelin proteins, including MOG or CNPase.

Late active lesions (LA): In these lesions myelin was already destroyed and removed from axons. Macrophages contained degradation products reactive for major myelin proteins, such as PLP, but were negative for MOG or CNPase.

Inactive lesions with active border (IA + A): The inactive center of radially expanding lesions still showed pronounced inflammation and macrophage infiltration. The macrophages revealed empty vacuoles and showed no immunoreactivity for myelin proteins.

Inactive lesions (IA): These lesions showed no evidence for ongoing myelin destruction at their borders. Although some of these lesions too contained T cells and macrophages, their number was much lower compared to active ones.

Remyelinated shadow plaques (RM): These lesions were characterized by myelin pallor, due to abnormally thin myelin sheaths and a pronounced expression of PLP mRNA in oligodendrocytes. Similar to inactive lesions, residual inflammation was present.

Patient Groups

Patients (*n* = 22) were divided into groups according to the following criteria (see Results and Tables 3 and 4). Those classified as active MS (*n* = 17) besides inactive

lesions, had at least one actively demyelinating lesion present in the CNS; those classified inactive MS (*n* = 5) had only inactive and/or remyelinated lesions and no evidence for ongoing demyelinating activity in the whole CNS.

Patients were considered to have either acute or chronic MS (Table 4) as follows:

Acute MS (malignant MS; *n* = 12) patients exhibited a rapidly progressing disease course, leading to significant disability in multiple neurological systems and often to death.³⁷

Classical or chronic MS (*n* = 10) patients exhibited relapsing-remitting, secondary progressive, primary progressive disease, with duration ranging from 3 to 21 years.

Quantitative Determination of Acute Axonal Injury and Inflammation

Camera lucida drawings of demyelinating lesions were made in order to define precisely the pattern of myelin destruction for each lesional area. In the selected area, the demyelinating activity was determined by the presence or absence of myelin degradation products within macrophages immunoreactive for MOG, proteolipid protein (PLP), and CNPase as described above. On adjacent serial sections the number of β APP- and common leukocyte antigen (CLA)-positive elements stained per square unit of tissue was counted. A 0.01-mm² field, defined by an ocular morphometric grid, taken throughout the middle of each lesional area was selected. In this field β APP-positive fibers were counted under a 100× objective. In inactive demyelinated lesions, shadow plaques as well as in periplaque and normal white matter for each lesional area the average number of β APP-positive axons in 10 adjacent fields of 0.01 mm² were taken for quantification because of the much lower density of injured axons.

To assess the extent of inflammation, CLA-positive elements were counted in a 0.1 mm² field in the respective lesions. Total number of MS lesional areas analyzed: *n* = 240; among those: *n* = 36 early active (EA), *n* = 13

Table 4. Acute Axonal Injury in Acute versus Chronic Multiple Sclerosis

		Active (early + late active)	Inactive + active edge	Inactive	Periplaque white matter	Normal white matter
β APP-positive axons/0.01 mm ²	Acute MS	47.27 ± 6.02	7.6 ± 1.43	1.48 ± 0.17	1.3 ± 0.21	0.41 ± 0.14
	Chronic MS	26.6 ± 4.29	3 ± 0.72	0.23 ± 0.05	0.19 ± 0.04	0.06 ± 0.01
<i>P</i>		<0.05	<0.05	<0.01	<0.0001	<0.001

late active (LA), $n = 29$ inactive center with active border (IA + A), $n = 21$ inactive (IA), $n = 26$ remyelination (RM), $n = 36$ normal white matter (NWM) of cases with active lesions (active MS), $n = 41$ periplaque white matter (PP) of cases with active lesions (active MS), $n = 15$ normal white matter (NWM) of cases with only inactive and/or remyelinating lesions (inactive MS), $n = 23$ periplaque white matter (PP) of cases with only inactive and/or remyelinating lesions (inactive MS); $n = 27$ white matter of controls (CO). Because there were no regional or inter-individual differences in β APP reactivity in normal white matter of controls, the same controls were used for all comparisons.

The total number of EAE lesional areas analyzed: $n = 55$; among those, $n = 16$ early active (EA), $n = 12$ late active (LA), $n = 11$ inactive plus active border (IA + A), $n = 4$ remyelination with superimposed demyelinating activity (RM-A), $n = 12$ white matter of control animals (CO).

Mann-Whitney U test and chi-square test were used for statistical analysis.

Results

Acute Axonal Injury in MS and EAE

Axonal pathology was qualitatively assessed by Bielschowsky's silver impregnation method. This staining technique revealed many swollen and distorted axons as well as axonal spheroids in actively demyelinating lesions of MS and EAE (Figures 1 and 2). In inactive lesions and in shadow plaques without concomitant active demyelination silver stained axonal spheroids were found only occasionally. For the quantitative assessment of acute axonal injury in MS and EAE, immunohistochemistry for β APP protein was performed. β APP serves as a marker for disturbance of axonal transport.^{32,38-40} As described previously,²⁷ β APP reactivity was found in some axons of normal axonal caliber, being most prominent in focal axonal swellings and in terminal ovoids (Figures 1 and 2).

Incidence of β APP-Positive Axons in Human Control Brain Tissue

Brain tissue of nine individuals without macroscopic or microscopic evidence for CNS disease served as control tissue. In each individual three distinct areas of 10×0.01 mm² were selected. The areas studied were located on brain regions where MS plaques are frequently found, such as the frontal, parietal, temporal, or occipital periventricular white matter or the subcortical white matter. β APP-positive axons were observed only exceptionally (0.05 ± 0.02 mean \pm SE) and there were no differences noted

between different brain areas. Therefore the data were pooled and used as controls for all comparisons.

Incidence of β APP-Positive Axons in Actively Demyelinating Lesions of MS

The highest incidence of β APP-positive axons was observed in areas of early and late active demyelination. Although in total β APP levels in early active lesions were slightly higher compared to late active lesions, some early active lesions showed only minor axonal injury (EA: 41.26 ± 5.59 range: 1-136 β APP-positive axons/0.01 mm², LA: 39.85 ± 7.72 range: 13-101 β APP-positive axons/0.01 mm²). The difference to late active lesions was not statistically significant (CO/EA: $p < 0.0001$; CO/LA: $p < 0.0001$, EA/LA: n.s.; Table 2).

Incidence of β APP-Positive Axons in Inactive Demyelinated Plaques of MS

β APP-positive axons were also abundant in the inactive center of lesions with ongoing demyelinating activity at their border (CO/IA + A, $P < 0.0001$). Their number, though, was significantly lower compared to early and to late active lesions (EA/IA + A, $p < 0.0001$; LA/IA + A, $P < 0.0001$). In addition, 15 completely inactive lesions of five patients were analyzed where no area of ongoing demyelination was found in the whole CNS. Even in these chronic inactive lesions a significant number of β APP-positive axons was found, when compared to the control areas (CO/IA, $P < 0.01$), but it was significantly lower compared to inactive lesions with an active edge (IA/IA + A, $P < 0.0001$; Table 2).

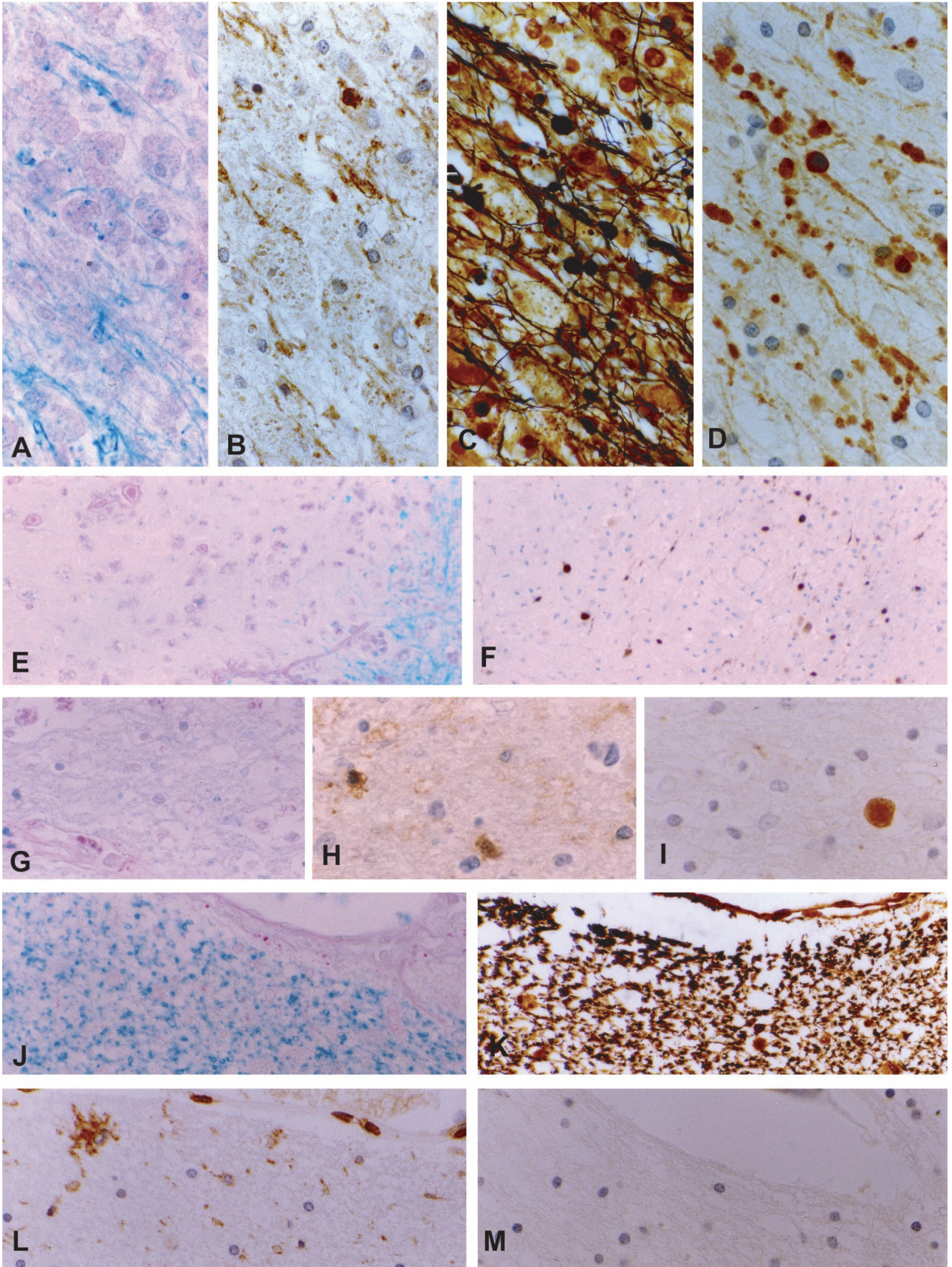
Incidence of β APP-Positive Axons in Remyelinated Shadow Plaques

There was no evidence for active demyelination in all shadow plaques analyzed, though inflammation of some T cells, macrophages, and, in particular, activated microglia, was still present and comparable to that in inactive demyelinated lesions (Figure 1). However, inactive demyelinated and remyelinated plaques differed in β APP reactivity; whereas significant axonal injury was found in inactive demyelinated lesions, axonal damage was minor in shadow plaques (CO/RM, n.s.; IA/RM, $P < 0.05$; Table 2).

Incidence of β APP-Positive Axons in Periplaque and Normal White Matter

In comparison to controls, there was also a low, but significant number of injured axons found in periplaque

Figure 1. Axonal pathology in multiple sclerosis. **A-D:** Early active MS lesion. Macrophages containing myelin degradation products (**A, B**), immunoreactive for all myelin proteins including MOG (**A:** Luxol fast blue (LFB), **B:** immunocytochemistry for MOG). **C:** Bielschowsky silver impregnation for axons reveals apparently normal axons as well as axonal swellings and spheroids. **D:** Immunocytochemistry for β APP shows numerous injured axons and axonal end-bulbs. **E-F:** Inactive demyelinated plaque with ongoing demyelinating activity at the border. Injured axons are found at the plaque edge as well as in the demyelinated plaque center (**E:** LFB, **F:** immunocytochemistry for β APP). **G-I:** Inactive demyelinated plaque without any evidence for active demyelination, but residual inflammation reveals a singular axonal spheroid by immunocytochemistry for β APP (**G:** LFB, **H:** CLA, **I:** β APP). **J-M:** Remyelinated shadow plaque as revealed by the presence of thin myelin pallor (**J:** LFB). **K:** Bielschowsky silver impregnation shows the absence of axonal swellings and spheroids. Residual inflammation, in particular activated microglia is still found (**L:** CLA), but no evidence for acute axonal injury (**M:** β APP). Original magnifications, $\times 484$ (**A-D, G-M**) and $\times 123$ (**E** and **F**).



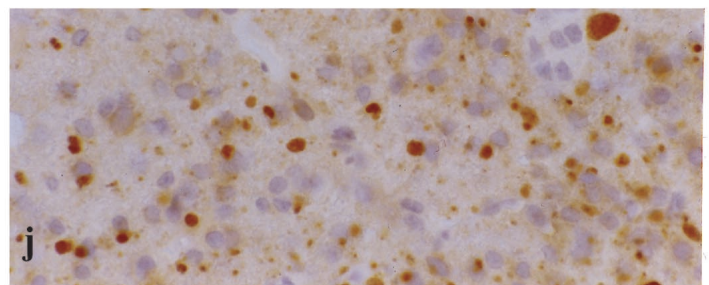
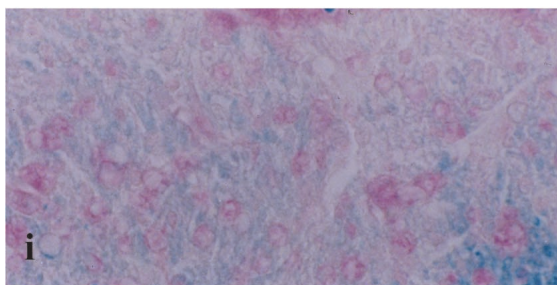
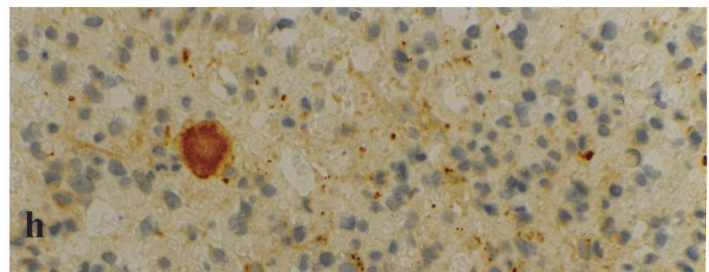
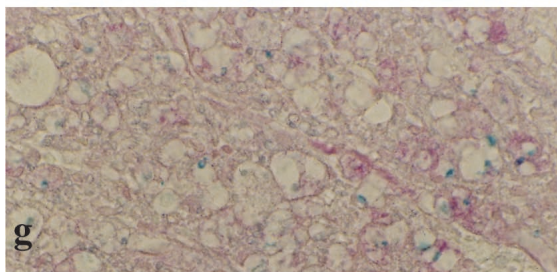
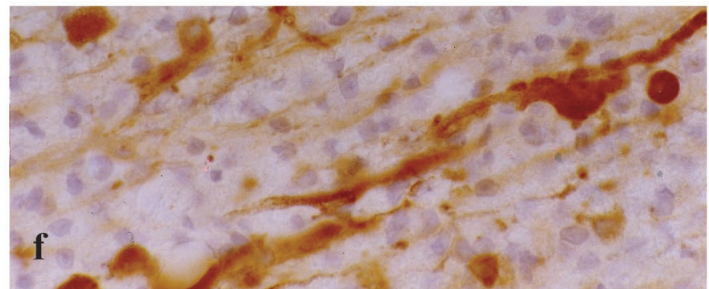
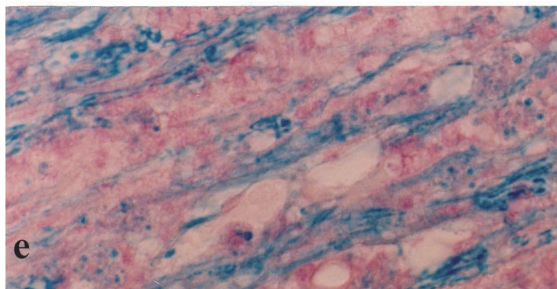
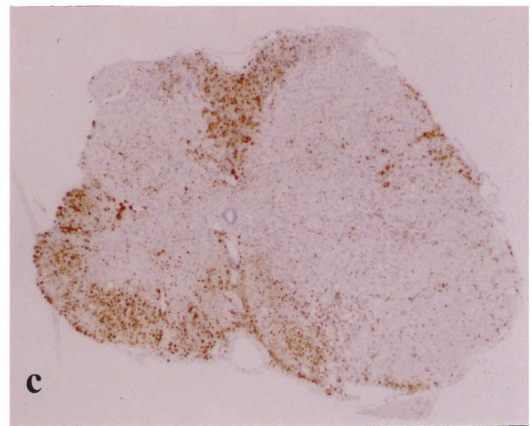
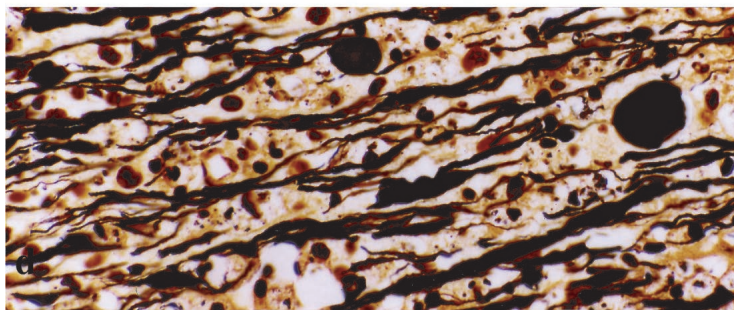
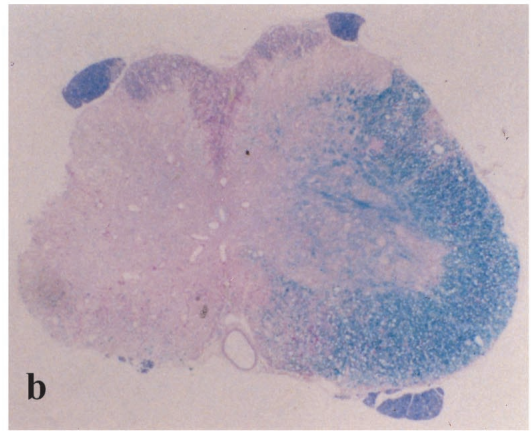
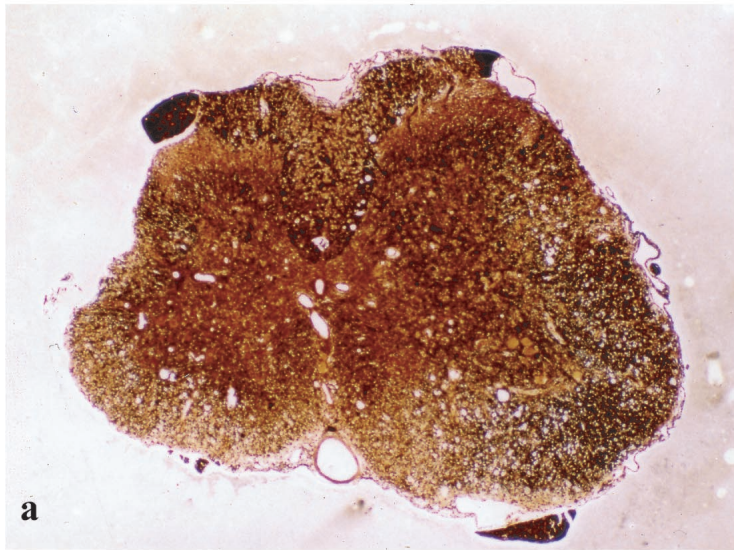


Table 5. Inflammation in Multiple Sclerosis Lesions

	Early active	Late active	Inactive + active edge	Inactive	Remyelination
CLA-positive cells/mm ²	1582.86 ± 117.79	1097 ± 166.33	740 ± 171.68	380.5 ± 44.81	274.29 ± 53.02

white matter of actively demyelinating lesions (active MS; CO/PP, $P < 0.0001$). In these "active" cases significant axonal damage was even found in normal white matter, far distant from established plaques (active MS, CO/NWM, $P < 0.01$, Table 3).

In order to determine whether acute axonal injury in the normal and periplaque white matter was restricted to cases with active demyelination, we also analyzed axonal β APP reactivity in cases with only inactive and/or remyelinated lesions (inactive MS; Table 3). In such inactive cases some increase in the number of β APP-positive axons was also found in the periplaque white matter, but it did not reach statistical significance. β APP immunoreactivity in normal white matter of inactive MS cases was comparable to that in control white matter (inactive MS, CO/PP, n.s.; CO/NWM, n.s., Table 3).

Incidence of β APP in Acute versus Chronic MS

Acute MS is characterized by a fulminant onset and rapid clinical deterioration leading to significant disability or even death within a short time period.³⁷ In all lesions of patients who presented with acute MS, the number of β APP-positive axons was significantly higher compared to the respective lesions of chronic MS cases. A significant difference was also noted in normal and periplaque white matter (Table 4).

Relation between Acute Axonal Injury and Inflammation

Inflammation was assessed by the presence of CLA-positive cells within and around the MS lesions. As expected, inflammation was most prominent in MS plaques with evidence for active demyelination, but still present in inactive demyelinated and remyelinated lesions (Table 5). Similar to a previous study,²⁷ a significant correlation between CLA-positive cells and β APP-positive axons was found in the lesions of MS ($P < 0.0001$). When the different types of lesions were analyzed separately, a highly significant correlation was found for late active and inactive lesions, but only a weak correlation was found for remyelinated lesions and none for early active lesions. The latter finding may be due to the fact that in some very early lesions inflammation was pronounced, whereas ax-

onal injury was not yet fully developed (CLA/APP; EA: n.s.; LA: $P < 0.01$; IA + A: $P < 0.01$; IA: $P < 0.0001$; RM: $P < 0.05$).

Inactive demyelinated and remyelinated plaques revealed similar values for inflammation (Table 5), but differed in β APP reactivity. Whereas significant axonal injury was found in inactive demyelinated lesions, axonal damage was minor in shadow plaques (Table 2).

Incidence of β APP in Chronic EAE

Acute axonal injury in chronic active EAE lesions was qualitatively and quantitatively similar to that in active MS plaques (Figure 2 and Table 6). Also, axonal damage occurred most prominently during a short time window when myelin was in the process of disruption (CO/EA, $P < 0.0001$; CO/LA, $P < 0.0001$). Although β APP levels were generally higher in early active lesions, there was no statistically significant difference to late active lesions (EA/LA: n.s.).

In contrast to MS, no completely inactive lesions were present in the sample of EAE material. Such as in multiple sclerosis lesions the number of β APP-positive axons in inactive lesions with an active edge was significantly increased compared to control white matter, but also significantly lower compared to early and late active lesions (CO/IA + A, $P < 0.0001$; EA/IA + A, $P < 0.0001$; LA/IA + A, $P < 0.0001$). In addition, most remyelinated shadow plaques in EAE showed evidence of recurrent demyelinating activity (Figure 2). This may explain the high incidence of axonal β APP reactivity in remyelinated EAE plaques (CO/RM-A, $P < 0.01$).

Discussion

The present study confirms some aspects of axonal damage reported in previous quantitative studies. As reported before, a high incidence of axonal damage apparently occurs in actively demyelinating lesions.^{27,28} There are, however, several aspects of our present study that shed new light on the patterns of axonal damage in inflammatory demyelinating lesions.

For the detection of acute axonal injury, immunohistochemistry for β APP was performed. β APP is a mem-

Figure 2. Axonal pathology in chronic experimental autoimmune encephalomyelitis. **a-c:** Spinal cord atrophy in an animal sacrificed on day 48 post-immunization. **a:** Bielschowsky silver impregnation for axons showing reduction of axonal density on one side of the spinal cord in comparison to the opposite side, where damage to myelin is only minor. **b:** LFB myelin stain reveals demyelination of the atrophic side of the spinal cord A. **c:** Massive macrophage infiltration within the demyelinated area. (ED1) **d:** Axonal swellings and spheroids in an actively demyelinating EAE lesion (Bielschowsky). **e-j:** Acute axonal injury in lesions of different stages of myelin degradation as revealed by immunocytochemistry for β APP. **e** and **f:** Same area of active demyelination as shown in **d** infiltrated by macrophages containing myelin debris (**e**, LFB) and numerous axonal swellings and spheroids immunoreactive for β APP (**f**, β APP). **g** and **h:** Demyelinated plaque with residual activity at the border (**g**, LFB) showing only one β APP-positive axonal spheroid beside some glial reactivity for β APP (**h**, β APP). **i** and **j:** Shadow plaque with thin myelin pallor indicating remyelination, infiltrated by myelin degrading macrophages (**i**, LFB) and many β APP-reactive axons (**j**, β APP). Original magnifications, $\times 40$ (**a**), $\times 51$ (**b** and **c**), $\times 389$ (**d-j**).

Table 6. Acute Axonal Injury in EAE Lesions

	Control white matter	Early active	Late active	Inactive + active edge	Remyelination + active demyelination
β APP-positive axons/0.01 mm ²	0.2 ± 0.07	87.38 ± 16.59	67.08 ± 16.15	7 ± 2.09	30.25 ± 15.59
β -fold increase compared to control WM		436.9	335.4	35	151.25
P values		CO/EA: <0.0001	CO/LA: <0.0001	CO/IA + A: <0.0001	CO/RM-A: <0.01

brane-spanning glycoprotein and a normal constituent of neuronal cells.^{32,38} It is transported by fast axonal transport. Normal levels of β APP in axons are not detectable by standard immunohistochemistry in formalin-fixed tissue. Its accumulation at sites of injury is probably due to the disturbance of axonal transport.^{32,38} Therefore, β APP has been suggested to be the immunocytochemical marker of choice for the detection of injured axons.³⁹ As has been demonstrated in fatal head injury, β APP stains damaged axons within 2 hours after injury³² and remains detectable in axons and bulbs for 10 to 14 days.⁴⁰ With a survival time longer than 2 weeks, β APP reactivity disappears from the injured axons as well as from bulbs.⁴⁰ β APP, therefore, may serve as a marker for early axonal damage, while more advanced stages of axonal degeneration may not be detected by this method.

By correlating acute axonal injury with the presence of myelin degradation products in our study, we can more precisely define the time window during which axonal damage occurs in MS lesions. Because myelin proteins are rapidly degraded when taken up by macrophages, the high incidence of axonal APP reactivity in lesional areas, which contain macrophages with degradation products immunoreactive for MOG, CNPase, and PLP, suggests that the most pronounced acute axonal injury occurs immediately within this short time window of plaque formation, eg, during early and late active demyelination.^{41,42} According to the time course of myelin degradation by macrophages *in vitro* and *in vivo* in EAE,⁴³ (Lassmann H, unpublished data), our data suggest that the time window of active demyelination lasts for approximately 1 to 2 weeks after initiation of the demyelinating process. This observation may be important for the design of axon-protective therapeutic strategies in MS. In some early active lesions only minor axonal injury was found. This may be due to the fact that very early, incipient lesions, where myelin disruption and possibly also axonal damage were incomplete, were included in the study. On the other hand, an interindividual variability in terms of tissue destruction may account for this phenomenon.

As reported before,²⁸ axonal injury in MS is not only restricted to demyelinated lesions, but also affects the periplaque area as well as the normal white matter, far distant from established demyelinated plaques. This diffuse axonal injury may be reflected in abnormalities of N-acetyl aspartate levels in the NWM in magnetic resonance spectroscopy.⁶⁻⁸ Since significant acute axonal injury in the periplaque and normal white matter in our study was only found in cases with ongoing active demyelination, and since the changes were more pro-

nounced in the periplaque, compared to the normal white matter, it is likely that these alterations occur secondary to axonal destruction in actively demyelinating plaques. To what extent the inflammatory process, which is much more widely distributed in the CNS compared to demyelination,⁴⁴ contributes to axonal injury in the so-called normal white matter remains to be determined.

Recent work of axonal changes after trauma suggests that some axonal damage may be repaired.⁴⁵ This may also be reflected in the normalization of NAA levels within MS lesions and NAWM six months after the acute phase of lesion formation in early MS.⁴⁶ Therefore, reactive axonal changes as seen by immunocytochemistry for β APP may be reversible unless they have not led to the formation of fully developed terminal ovoids. Intervention at an early time point of plaque formation, therefore, might not only reduce acute axonal injury, but also reduce the number of injured axons that undergo definite degeneration.

Besides acute axonal injury during active demyelination, we also found significant low-burning axonal damage even in plaques, completely devoid of active myelin destruction in cases where no active lesion was detected (Table 2 and Figure 1). This finding may explain, at least in part, clinical progression in cases where no MRI-activity can be detected.⁴⁷ The mechanisms of axonal degeneration, however, remain speculative. It has to be noted that ongoing axonal destruction in inactive MS lesions occurs on the background of a residual inflammatory process, reflected by infiltration of the lesions by some T cells, macrophages, and activated microglia. Thus, the inflammatory process may play an important role in the ongoing axonal destruction. Alternatively, ongoing axonal destruction may be due to a lack of trophic support provided by myelin and oligodendrocytes.

In remyelinated lesions inflammation was still found, but acute axonal injury was significantly less pronounced in remyelinated shadow plaques compared to that in inactive demyelinated lesions. This observation further supports the view that myelination exerts a protective effect against axonal damage.⁴⁸ This could be due to the protective effect of the myelin sheath against the inflammatory environment, as has been shown for nitric oxide.⁴⁹ On the other hand, trophic supply by oligodendrocytes may protect axons against gradual degeneration. Such a mechanism of axonal damage has been suggested in experimental models deficient for myelin proteins such as myelin-associated glycoprotein (MAG-/- animals)⁵⁰ or proteolipid protein (PLP-DM20-/- mice).⁵¹

In conclusion, our data suggest that extensive axonal damage occurs during plaque formation in a very short time window after onset of demyelination. During active demyelination, significant axonal injury is also found in the periplaque and normal white matter. Second, there is ongoing low-burning axonal destruction in inactive demyelinated lesions. This process, in contrast to acute axonal injury in active lesions, may be continued for prolonged time periods and may explain the observed profound reduction of axonal density in established MS plaques.^{20,52,53} Finally, our results show that axonal pathology in inflammatory demyelinating lesions of chronic MOG-induced autoimmune encephalomyelitis closely reflects that found in MS. This particular model of MOG-induced EAE in Lewis N rats was selected because it is pathologically characterized by focal, very large plaques of demyelination.³⁵ In contrast to MS, however, the development of lesions in this model is condensed to a time period of 1 to 2 months. Thus, most of the lesions studied here still present demyelinating activity at least at their borders to the adjacent periplaque white matter. Furthermore, in contrast to most shadow plaques in MS, remyelinating lesions in this model generally show signs of recurrent active demyelination. Thus, this model closely reflects axonal pathology in actively demyelinating MS cases, but does not, so far, allow study of the slow axonal degeneration in chronic MS cases. Despite these differences in disease activity, MOG-induced EAE may serve as an ideal model for testing the effectiveness of axon-protective therapeutic strategies for chronic inflammatory demyelinating diseases.

Acknowledgments

We thank Helene Breitschopf, Angela Kury, Marianne Leisser, Jutta Wakley-Neuninger, and Petra Tassotti for expert technical assistance and Drs. Christine Stadelmann and Helmut Rauschka for carefully reading the manuscript and for discussion.

References

1. Charcot JM: Histologie de la sclérose en plaques. *Gazette des Hôpitaux civils et militaires* 1868, 140:554–555 and 141:557–558 and 143:566
2. Arnold DL, Riess GT, Matthews PM, Francis GS, Collins DL, Wolfson C, Antel JP: Use of proton magnetic resonance spectroscopy for monitoring disease progression in multiple sclerosis. *Ann Neurol* 1994, 36:76–82
3. Barnes D, Munro PMG, Youl BD: The longstanding MS lesion: a quantitative MRI and electron microscopic study. *Brain* 1991, 114:1271–1280
4. Davie CA, Hawkins CP, Barker GJ, Brennan A, Tofts PS, Miller DH, McDonald WI: Serial proton magnetic resonance spectroscopy in acute multiple sclerosis lesions. *Brain* 1994, 117:49–58
5. Davie CA, Barker GJ, Webb S, Tofts PS, Thompson AJ, Harding AE, McDonald WI, Miller DH: Persistent functional deficit in multiple sclerosis and autosomal dominant cerebellar ataxia is associated with axonal loss. *Brain* 1995, 118:1583–1592
6. Davie CA, Barker GJ, Thompson AJ, Tofts PS, McDonald WI, Miller DH: ¹H magnetic resonance spectroscopy of chronic cerebral white matter lesions and normal appearing white matter in multiple sclerosis. *J Neurol Neurosurg Psychiatr* 1997, 63:736–742
7. De Stephano N, Matthews PM, Fu L, Narayanan S, Stanley J, Francis GS, Antel JP, Arnold DL: Axonal damage correlates with disability in patients with relapsing-remitting multiple sclerosis: results of a longitudinal magnetic resonance spectroscopy study. *Brain* 1998, 121:1469–1477
8. Fu L, Matthews PM, De Stefano N, Worsley KJ, Narayanan S, Francis GS, Antel JP, Wolfson C, Arnold DL: Imaging axonal damage of normal appearing white matter in multiple sclerosis. *Brain* 1998, 121:103–113
9. Losseff NA, Wang L, Lai HM, Yoo DS, Gawne-Cain ML, McDonald WI, Miller DH, Thompson AJ: Progressive cerebral atrophy in multiple sclerosis: a serial MRI study. *Brain* 1996, 119:2009–2019
10. Losseff NA, Webb SL, O'Riordan JI, Page R, Wang L, Barker GJ, Tofts PS, McDonald WI, Miller DH, Thompson AJ: Spinal cord atrophy and disability in multiple sclerosis: a new reproducible and sensitive MRI method with potential to monitor disease progression. *Brain* 1996, 119:701–708
11. van Walderveen MAA, Barkhof F, Hommes OR, Polman CH, Tobi H, Frequin STFM, Valk J: Correlating MRI and clinical disease activity in multiple sclerosis: relevance of hypointense lesions on short-TR/short-TE (T1-weighted) spin echo images. *Neurology* 1995, 45:1684–1690
12. van Walderveen MAA, Kamphorst W, Scheltens P, van Waesberghe JHTM, Ravid R, Polman CH, Barkhof F: Histopathologic correlate of hypointense lesions on T1-weighted spin-echo MRI in multiple sclerosis. *Neurology* 1998, 50:1282–1288
13. van Walderveen MAA, Barkhof F, Pouwels PJW, van Schijndel RA, Polman CH, Castelijns JA: Neuronal damage in T1-hypointense multiple sclerosis lesions demonstrated in vivo using proton magnetic resonance spectroscopy. *Ann Neurol* 1999, 46:79–87
14. Bielschowsky M: Zur Histologie der multiplen Sklerose. *Neurol Centralblatt* 1903, 22:770–777
15. Bielschowsky M: Die marklosen Nervenfasern in den Herden der multiplen Sklerose: eine Antwort an Herrn Strähuber. *Neurol Centralblatt* 1904, 16:59–62
16. Marburg O: Die sogenannte "akute multiple Sklerose" (Encephalomyelitis peraxialis scleroticans). *Jahrb Neurol Psychiatr* 1906, 27:211–312
17. Doinikow B: Über De- und Regenerationserscheinungen an Achsenzylindern bei der multiplen Sklerose. *Z Gesamt Neurol Psychiatr* 1915, 27:151–178
18. Fraenkel M, Jakob A: Zur Pathologie der multiplen Sklerose mit besonderer Berücksichtigung der akuten Formen. *Z Gesamt Neurol Psychiatr* 1913, 14:565–603
19. Siemerling E, Raecke J: Beiträge zur Klinik und Pathologie der multiplen Sklerose mit besonderer Berücksichtigung ihrer Pathogenese. *Archiv für Psychiatrie und Nervenkrankheiten* 1914, 53 (Heft II): 385–564
20. Putnam TJ: Studies in multiple sclerosis. *Arch Neurol Psychiatry* 1936, 35:1289–1308
21. Greenfield JG, King LS: Observations on the histopathology of the cerebral lesions in disseminated sclerosis. *Brain* 1936, 59:445–458
22. Lumsden CE: The neuropathology of multiple sclerosis. *Handbook of Clinical Neurology* 1970, Vol. 9: Multiple Sclerosis and Other Demyelinating Diseases. Edited by PJ Vinken, GW Bruyn. New York, Elsevier, pp 217–309
23. Prineas JW, Connel F: The fine structure of chronically active multiple sclerosis plaques. *Neurology* 1978, 28 (suppl.): 68–75
24. Dahl D, Perides G, Bignami A: Axonal regeneration in old multiple sclerosis plaques. *Acta Neuropathol (Berl)* 1989, 79:154–159
25. Shintaku M, Hirano A, Llena JF: Increased diameter of demyelinated axons in chronic multiple sclerosis of the spinal cord. *Neuropathol Appl Neurobiol* 1988, 14:505–510
26. Kornek B, Lassmann H: Axonal pathology in multiple sclerosis: a historical note. *Brain Pathol* 1999, 9:651–656
27. Ferguson B, Matyszak MK, Esiri MM, Perry VH: Axonal damage in acute multiple sclerosis. *Brain* 1997, 120:393–399
28. Trapp BD, Peterson J, Ransohoff RM, Rudick R, Mörk S, Bö L: Axonal transection in the lesions of multiple sclerosis. *N Engl J Med* 1998, 338:278–285
29. Madrid RE, Wisniewski HM: Axonal degeneration in demyelinating disorders. *J Neurocytol* 1977, 6:103–117
30. Raine CS, Mokhtarian F, MacFarlin DE: Adoptively transferred chronic

- relapsing experimental autoimmune encephalomyelitis in the mouse: neuropathologic analysis. *Lab Invest* 1984, 51:534–546
31. Raine CS, Cross AH: Axonal dystrophy as a consequence of long-term demyelination. *Lab Invest* 1989, 60:714–725
 32. Gentleman SM, Nash MJ, Sweeting CJ, Graham DI, Roberts GW: Beta-amyloid precursor protein as a marker for axonal injury after head injury. *Neurosci Lett* 1993, 160:139–144
 33. An SF, Giometto B, Groves M, Miller RF, Beckett AAJ, Gray F, Tavolato B, Scaravilli F: Axonal damage revealed by accumulation of β -APP in HIV-positive individuals without AIDS. *J Neuropathol Exp Neurol* 1997, 56:1262–1268
 34. Hedrich HJ, ed: *Genetic Monitoring of Inbred Strains of Rats*. Stuttgart, New York, Gustav Fischer, 1990
 35. Storch MK, Stefferl A, Brehm U, Weissert R, Wallstroem E, Kerschensteiner M, Olsson T, Lington C, Lassmann H: Autoimmunity to myelin oligodendrocyte glycoprotein in rats mimics the spectrum of multiple sclerosis pathology. *Brain Pathol* 1998, 8:681–694
 36. Breitschopf H, Suchanek G, Gould RM, Colman DR, Lassmann H: In situ hybridization with digoxigenin-labeled probes: sensitive and reliable detection method applied to myelinating rat brain. *Acta Neuropathol (Berl)* 1992, 84:581–587
 37. Lublin FD, Reingold SC: Defining the clinical course of multiple sclerosis: results of an international survey. *Neurology* 1996, 46:907–911
 38. Sheriff FE, Bridges LR, Sivaloganathan S: Early detection of axonal injury after human head trauma using immunocytochemistry for β -amyloid precursor protein. *Acta Neuropathol (Berlin)* 1994, 87:55–62
 39. Sheriff FE, Bridges LR, Gentleman SM, Sivaloganathan S, Wilson S: Markers of axonal injury in post mortem human brain. *Acta Neuropathol (Berlin)* 1994, 88:433–439
 40. Geddes JF, Vowles GH, Beer TW, Ellison DW: The diagnosis of diffuse axonal injury: implications for forensic practice. *Neuropathol Appl Neurobiol* 1997, 23:339–347
 41. Brück W, Schmied M, Suchanek G, Brück Y, Breitschopf H, Poser S, Piddlesden S, Lassmann H: Oligodendrocytes in the early course of multiple sclerosis. *Ann Neurol* 1994, 35:65–73
 42. Brück W, Porada P, Poser S, Rieckmann P, Hanefeld F, Kretschmar HA, Lassmann H: Monocyte/macrophage differentiation in early multiple sclerosis lesions. *Ann Neurol* 1995, 38:788–796
 43. Lassmann H, Wisniewski HM: Chronic relapsing experimental allergic encephalomyelitis: morphological sequence of myelin degradation. *Brain Res* 1979, 169:357–368
 44. Lassmann H: The pathology of multiple sclerosis and its evolution. *Phil Trans Roy Soc London (B)* 1999, 354:1635–1640
 45. Gennarelli TA: The spectrum of traumatic axonal injury. *Neuropathol Appl Neurobiol* 1996, 22:509–513
 46. DeStefano N, Narayanan S, Matthews PM, Francis GS, Antel JP, Arnold DL: In vivo evidence for axonal dysfunction remote from focal cerebral demyelination of the type seen in multiple sclerosis. *Brain* 1999, 122:1933–1939
 47. Truyen L, van Waesberghe JHTM, van Walderveen MAA, van Oosten BW, Polman CH, Hommes OR, Ader HJA, Barkhof F: Accumulation of hypointense lesions (“black holes”) on T1 spin-echo MRI correlates with disease progression in multiple sclerosis. *Neurology* 1996, 47:1469–1476
 48. Duncan ID: Glial cell transplantation and remyelination of the central nervous system (review). *Neuropathol Appl Neurobiol* 1996, 22:87–100
 49. Redford EJ, Kapoor R, Smith KJ: Nitric oxide donors reversibly block axonal function: demyelinated axons are especially susceptible. *Brain* 1997, 120:2149–2157
 50. Yin X, Crawford TO, Griffin JW, Tu PH, Lee VMY, Li C, Roder J, Trapp BD: Myelin-associated glycoprotein is a myelin signal that modulates the caliber of myelinated axons. *J Neurosci* 1998, 18:1953–1962
 51. Griffiths I, Klugmann M, Anderson T, Yool D, Thompson C, Schwab HM, Schneider A, Zimmermann F, McCulloch M, Nadon N, Nave KA: Axonal swellings and degeneration in mice lacking the major proteolipid protein of myelin. *Science* 1998, 280:1610–1613
 52. Mews I, Bergmann M, Bunkowski S, Gullotta F, Brück W: Oligodendrocyte and axon pathology in clinically silent multiple sclerosis lesions. *Mult Scler* 1998, 4:55–62
 53. Lassmann H: The pathology of multiple sclerosis. *McAlpine’s Multiple Sclerosis*, 3rd edition. Edited by A Compston, G Ebers, H Lassmann, I McDonald, B. Matthews, H Wekerle. London, Churchill Livingstone, 1998