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Fluorous synthesis of sclerotigenin-type benzodiazepine-quinazolinones

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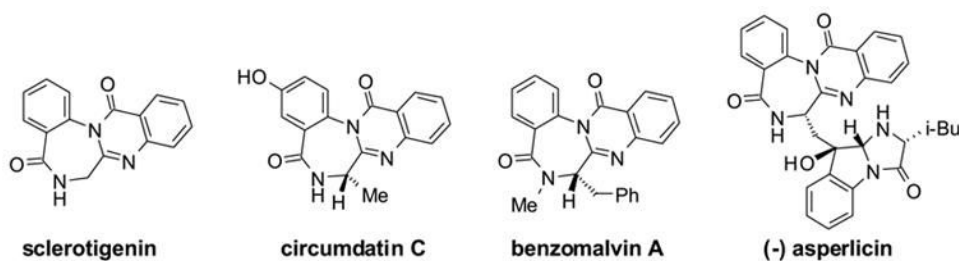
Abstract

A new synthetic protocol for sclerotigenin-type benzodiazepine-quinazolinone library scaffold is introduced. A fluorous benzyl protecting group is used for synthesis of 4-benzodiazepine-2,5-dione intermediate and also as a phase tag for fluorous solid-phase extraction (F-SPE).

Keywords

sclerotigenin; benzodiazepine-quinazolinone; 1,4-benzodiazepine-2,5-dione; fluorous synthesis; solid-phase extraction

Sclerotigenin was isolated from the sclerotia of *Penicillium sclerotigenum* and has shown promising antiinsectan activity.¹ It is the simplest member of the benzodiazepine-quinazolinone natural alkaloid family. Other members in this family such as circumdatins A–G isolated from terrestrial fungus *Aspergillus ochraceus*² and benzomalvins A–C isolated from fungus *Penicillium* sp also possess interesting biological activities.³



Privileged 1,4-benzodiazepine-2,5-dione ring systems are the key intermediates for synthesis of benzodiazepine-quinazolinone alkaloids.⁴ As part of our continuous effort on the development of fluorous synthetic protocols, we have employed a series of fluorous protecting groups for library synthesis.^{5,6} Reported here is a new approach to synthesize benzodiazepinedione scaffold using fluorous benzyl as a protecting group and also as a phase tag for fluorous solid-phase extraction (F-SPE).⁷ Further derivatization of benzodiazepinediones leads to formation of sclerotigenin ring skeleton.

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Taking the advantage that numbers of conventional solution-phase and solid-phase synthetic methods for benzodiazepine have been reported in literature,⁸ we adopted Ellman's solid-phase method for fluoros synthesis (Scheme 1).⁹ Fluorous benzaldehyde **1** prepared by reaction of a hydroxybenzaldehyde with a fluoros alcohol was used as the starting material. Compound **2** was produced by reductive amination of **1** with an amino ester. Compound **2** was reacted with an anthranilic acid in the presence of 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDCI) and *N*-methylpyrrolidine (NMP). The 1,4-benzodiazepine-2,5-dione ring formation was accomplished by base-promoted cyclization of **3**. Compounds **2**, **3**, and **4** generated in this reaction sequence were purified by simple workup or F-SPE with FluoroFlash® cartridges.¹⁰ In F-SPE, the first wash with 80:20 MeOH-H₂O eluted the non-fluorous components. The desired fluoros compound was eluted with 100% MeOH. A total of nine analogs of compound **4** with substitution variations (R¹ and R²) were prepared.¹¹

With nine different benzodiazepinediones **4** in hand, we then conducted parallel synthesis to construct the quinazolinone ring skeleton (Scheme 2).^{1c} Compound **4** was acylated with 2-nitrobenzoyl chloride in the presence of *t*-BuN=P(NMe₂)₃ as a base to give compound **5** (Table 1). If substituted 2-nitrobenzoyl chloride was employed for acylation, the third diversity point (R₃) could be introduced. Compounds **5** were purified by automated RapidTrace F-SPE.¹² The nitro group of **5** was reduced with zinc dust in acetic acid under sonication conditions. Resulted amino group simultaneously underwent cyclization to form quinazolinone ring of **6**. The parallel sonication reactions of **5** gave the reduction/cyclization products **6** in a broad range of yield (21–73%). Since some reactions had low yields, F-SPE was not sufficient for purification. Reverse-phase chromatography was applied to purify compounds **6**. The capability to purify fluoros compounds by non-fluorous technique is a useful option. It could be a difficult task in solid-phase synthesis to separate resin-bound impurities. At the last step, F-benzyl tag of compounds **6** was removed by treated with 90:5:5 TFA-H₂O-dimethylsulfide (DMS) under microwave radiation, followed by F-SPE on RapidTrace® workstation to give the final product **7** with the sclerotigenin ring skeleton.¹³

In summary, we have developed a new approach for the synthesis of fluoros 1,4-benzodiazepine-2,5-diones. The key intermediates can be readily converted to sclerotigenin ring skeleton. The new method which produces the library scaffold with substitution variation coupled with the simple F-SPE separation is an alternative way for solution-phase parallel synthesis of benzodiazepine-quinazolinone analogs.

Acknowledgements

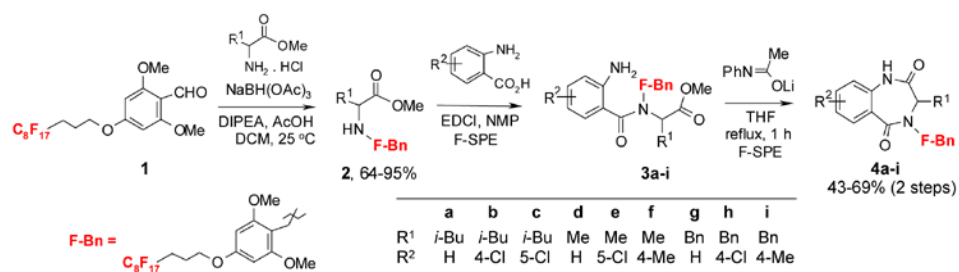
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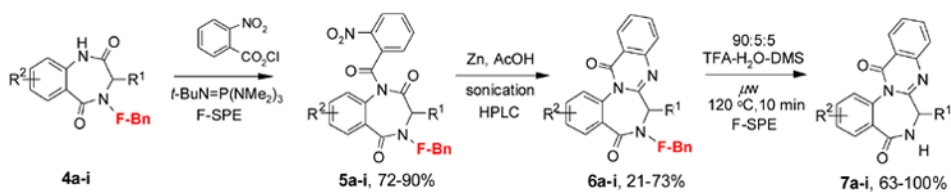
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 10. FluoroFlash® SPE cartridges are available from Fluororous Technologies, Inc. (www.fluororous.com)
 11. A general procedure for the synthesis of compounds **2** and **4**. To a solution of leucine methyl ester hydrochloride (7.6 g, 42 mmol), 2,6-dimethoxy-4-[3-(perfluorooctyl)propyloxy]benzaldehyde **1** (26 g, 40 mmol), and *N,N*-diisopropylethylamine (7 mL, 0.04 mol) in CH₂Cl₂ (0.3 L) was added 4 Å molecular sieves (3 g) at 23 °C. NaBH(OAc)₃ (13 g, 60 mmol) was added after 4 h, then water was added after additional 3 h. The CH₂Cl₂ layer was washed with aq. NH₄Cl and brine. After most of the solvent was removed using a rotary evaporator, the residue was passed through a pad of silica gel (50 mL). The product was eluted with hexanes–EtOAc (1:1, 300 mL). The concentrated product was further triturated with hexanes–Et₂O to give the desired compound **2** (R¹ = *i*-Bu, 3.9 g, 95% yield). ¹H NMR (270 MHz, CDCl₃) δ 6.08 (s, 2H), 4.02 (t, 2H, *J* = 5.8 Hz), 3.78 (s, 6H), 3.59 (s, 3H), 3.26 (t, 1H, *J* = 7.1 Hz), 2.45–2.00 (m, 5H), 1.82–1.35 (m, 3 H), 0.88 (d, 3H, *J* = 6.5 Hz), 0.81 (d, 3H, *J* = 6.4 Hz). ¹³C NMR (67.5 MHz, CDCl₃) δ 176.4, 159.3, 108.9, 90.7, 66.3, 59.0, 55.5, 51.3, 42.8, 39.9, 28.2, 27.9, 27.5, 24.8, 22.6, 22.0, 20.5. LC-MS (APCI+) *m/z* 772 [M+1]⁺. To a solution of **2** (R¹ = *i*-Bu, 4.3 g, 5.6 mmol) in *N*-methylpyrrolidine (30 mL), 4-chloroanthranilic acid (1.9 g, 11 mmol) and EDCI-HCl (2.1 g, 11 mmol) were added as solid at 23 °C. The same amounts of the acid and EDCI-HCl were added after 2 h and 4 h. One day after the final addition, the reaction mixture was diluted with DMSO (300 mL), and was loaded onto an F-SPE cartridge (50 g), and the flask was rinsed with DMSO (100 mL), and was loaded to the silica gel. The nonfluororous components were eluted with MeCN–H₂O (1:1, 300 mL, and 4:1, 200 mL), and then most of the solvent was drained from the cartridge. The amide coupling product was eluted with MeCN (0.4 L). The MeCN solution was concentrated in a rotary evaporator, and the residue was treated with a solution of lithium acetanilide (0.33 M in THF, 30 mL). The mixture was refluxed for 1 h. After cooling, AcOH (0.6 mL) was added, and the solvent was removed in a rotary evaporator. MeOH (30 mL) was added to the residue, and it was heated until the solvent started to boil. The mixture was left at 23 °C for 1 d, and product **4b** (R¹ = *i*-Bu, R² = 4-Cl) was collected as a solid by filtration (3.5 g, 69% yield based on the amount of **2**). ¹H NMR (270 MHz, CDCl₃) δ 9.55 (s, 1H), 7.99 (d, *J* = 8.5 Hz, 1H), 7.16 (dd, *J* = 7.1, 1.9 Hz 1.), 6.90 (d, *J* = 1.8 Hz, 1H), 6.09 (s, 2H), 5.23 (d, *J* = 13.8 Hz, 1H), 4.56 (d, *J* = 13.8 Hz, 1H), 4.15–3.85 (m, 3H), 3.75 (s, 6H), 2.45–2.00 (m, 4H), 1.60–1.45 (m, 1H), 1.35–1.15 (m, 2H), 0.80 (d, *J* = 6.4 Hz, 3H), 0.73 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (67.5 Hz, CDCl₃) δ 173.5, 165.1, 160.6, 160.1, 137.7, 136.2, 133.3, 125.5, 124.6, 119.4, 104.1, 90.6, 66.3, 59.5, 55.5, 42.0, 38.3, 27.9 (t, *J* = 22 Hz), 25.2, 22.3, 22.1, 20.5. LC-MS (APCI +) *m/z* 893 [M+1]⁺.
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 13. A general procedure for the synthesis of compounds **5**, **6**, and **7**. To a solution of **4** in CH₂Cl₂ was added *t*-butylimino-tris(dimethylamino)phosphorane (10 equiv) and 2-nitrobenzoic acid (2 equiv). The reaction mixture was stirred for 10 min and then concentrated in a rotary evaporator. The residue was dissolved in DMF (1 mL) and purified on RapidTrace® SPE workstation with 2 g cartridges to afford **5** in 72–90% yield. A solution of **5** in acetic acid (1 mL) was added Zn dust (20 equiv) and

sonicated at room temperature for 2 h. The Zn was filtered and the filtrate was diluted with EtOAc and washed with NaHCO₃ and brine. The EtOAc solution was dried and concentrated in a rotary evaporator. The residue was dissolved in MeCN and purified by C18 HPLC to afford **6** in 21–73% yields. A solution of **6** in TFA-H₂O-DMS (90:5:5) was stirred for 3 days before being concentrated in a rotary evaporator. The residue was dissolved in DMF (1 mL) and purified by RapidTrace® SPE workstation to afford **7** in 63–100% yields. Analytical data for compound **7b** (R¹ = *i*-Bu, R² = 4-Cl): ¹H NMR (275 Hz, CDCl₃) δ 0.90 (d, *J* = 6.5 Hz, 3H), 1.00 (d, *J* = 6.5 Hz, 3H), 1.80–2.05 (m, 2H), 2.05–2.25 (m, 1H), 4.10–4.35 (m, 1H), 6.66 (d, *J* = 6.2 Hz, 1H), 7.45–7.59 (m, 2H), 7.67 (d, *J* = 1.9 Hz, 1H), 7.70–7.85 (m, 2H), 7.91 (d, *J* = 8.4 Hz, 1H), 8.31 (dd, *J* = 1.4, 8.0 Hz, 1H); ¹³C NMR (67.5 Hz, CDCl₃) δ 22.0, 23.1, 24.3, 38.0, 52.4, 121.3, 127.5, 127.8, 127.9, 128.7, 128.9, 129.5, 131.0, 134.3, 135.2, 137.4, 146.0, 154.1, 161.5, 167.1; LCMS (APCI+) 368 [M+1]⁺.



Scheme 1.
Fluorous synthesis of benzodiazepinedione **4**



Scheme 2.
Parallel synthesis of nine benzodiazepine-quinazolinones **7**

Table 1Yields for analogs of compounds **5**, **6**, and **7**

	a	b	c	d	e	f	g	h	i
R ¹	<i>i</i> -Bu	<i>i</i> -Bu	<i>i</i> -Bu	Me	Me	Me	Bn	Bn	Bn
R ²	H	4-Cl	5-Cl	H	5-Cl	4-Me	H	4-Cl	4-Me
5a-i	82%	80%	90%	75%	94%	90%	75%	72%	81%
6a-i	44%	50%	67%	21%	51%	62%	70%	65%	73%
7a-i	83%	86%	91%	91%	63%	71%	100%	89%	97%