# Pathogenic Leptospiras Isolated from Malaysian Surface Waters

## A. D. ALEXANDER,<sup>1\*</sup> L. B. EVANS, M. F. BAKER, H. J. BAKER, D. ELLISON, and M. MARRIAPAN

Walter Reed Army Institute of Research, Department of Veterinary Microbiology, Washington, D.C. 20012, and U.S. Army Medical Research Unit, Institute for Medical Research, Kuala Lumpur, Malaysia

### Received for publication 30 September 1974

Pathogenic leptospiras (1,424) isolated from natural waters and wet soils in Malaysia comprised 29 different serovars (synonym serotypes). All except two of the serovars had been found previously in Malaysia. The exceptional serovars were werrasingha, an Autumnalis serogroup member originally isolated in Ceylon, and a new serovar designated evansi. Serovar evansi had serological affinities with serovar ranarum which was isolated from the kidney of a frog in Iowa. The large variety of serovars found in jungle areas was consistent with similar previous findings of diverse serovar infections in troops who had operated in Malaysian jungles.

McCrumb et al. (8) in a clinical and epidemiological study of 244 cases of leptospirosis in Malaysia directed attention to the jungle environment as "one of the most important natural foci of leptospiral infection." In 1961, Baker (5) initiated studies to determine the infectiousness of waters in the Malaysian rain forest.

For this purpose, Baker and Baker (6) developed a sensitive hamster exposure method for the isolation of pathogenic leptospiras from natural milieu (6). The study was focused principally on the Gombak River and other adjacent streams or rivers located within an area of approximately 20 miles from Kuala Lumpur.

Additional studies were carried out on jungle streams in the State of Pahang and in North Borneo, and on a ricefield and tin mining pool. During the course of these studies, approximately 1,424 pathogenic leptospiras were isolated from waters or wet shore soils. The serological characterization of the isolated strains is described in this report. The epidemiological aspects of the study will be presented separately.

#### **MATERIALS AND METHODS**

**Origin of strains.** Strains were isolated from 1961 to 1966 from five different geographic or ecological sites. Approximately 1,185 strains were isolated from jungle waters and soils within a radius of 20 miles from Kuala Lumpur in the State of Selangor. Fifty-six isolates from ricefields and 17 isolates from pools of water in mining areas were also obtained in the proximity of Kuala Lumpur. The 104 leptospiral

<sup>1</sup> Present address: Department of Microbiology, Chicago College of Osteopathic Medicine, 1122 East 53rd St., Chicago, Ill. 60615. strains from the State of Pahang were derived mostly from shore sand along a jungle stream and river located near Kuala Lipis. Jungle streams in the western part of Sabah were the source of 62 leptospiral isolates. Nearly all of the isolates were recovered from hamsters dying from leptospirosis after their exposure to sample water or sample soil washings. The survey method is described elsewhere (6).

Culture typing. The microscopic agglutination test was the basic means for identification of strains. The procedures for this test and also agglutininadsorption tests are described in detail elsewhere (2). Generally, strains employed as live antigens were tested for cross-agglutination reactions against the following eight screening serovar (synonym serotype) antisera: autumnalis, alexi, medanensis, javanica, bataviae, patane, mankarso, and pooled australis, grippotyphosa, and djasiman. On the basis of initial reactions, strains were subsequently tested against one or more serogroups to determine major antigenic affinities. These tests served to identify strains by serogroup. Representative strains of different patterns of cross-agglutination were then selected for comparative serological tests with type strains of serovars previously found in Malaysia. Isolates were assumed to be serologically homologous with type strains if their cross-reaction patterns were similar. Strains which differed from selected type strains in agglutinogenic characteristics were definitively identified by the use of appropriate agglutinin-adsorption tests (2). The identities of nine other representative isolates were confirmed incidentally for other purposes. The identification of strains was based to large extent on considerable background information obtained in previous studies of pathogenic leptospiras in Malaysia (1, 2).

#### **RESULTS AND DISCUSSION**

The 1,362 strains isolated from soil and water in West Malaysia (Selangor and Pahang) could be divided into 13 serogroups comprising a total of 29 serovars (Table 1). The wide variety of pathogenic serovars in jungle milieu was consistent with the broad array of serovar infections found in soldiers on jungle patrols (8). Within the State of Selangor, the ricefield and jungle areas studied had similar relative distributions of serovars. However, of 17 strains isolated in mining pools, 16 appeared to be related to serovar *australis* and one to serovar *paidjan*.

The relative frequency of recovered serovar strains in the Pahang jungle superficially differed from those in the Selangor jungle. The variable findings may have reflected differences in size and types of samples and time of sampling. In the limited survey in Sabah mainly bataviae and autumnalis and a few icterohaemorrhagiae serovars were found.

All except four of the representative strains were related to Malaysian serovars previously studied in this laboratory (1, 2). Three of the exceptional strains were identified to be serovars sentot, weerasingha, and tarassovi, respectively, on the basis of agglutinin-adsorption tests. Serovar sentot and a member of the Tarassovi serogroup have been reported from Malaysia previously (9). Serovar weerasingha was isolated in Ceylon and identified by Y.

	Serovar	Isolates within locations (%)				
· Serogroup		Selangor jungle	Selangor ricefield	Selangor mining pool	Pahang jungle	Sabah jungle
Icterohaemorrhagiae	mankarso <sup>a</sup>	18.3	23.2		2.9	8.1
	smithii	2.0				
	birkin	1.1			1.0	
Australis	australisª	13.5	7.1	<b>94</b> .1	9.6	
Bataviae	paidjanª	11.6	7.1	5.9	1.9	58.1
	bataviaeª	3.7				9.7
Canicola	jonsis	8.1			1.0	
	schuffneri	4.6	1.8		31.7	
	malayaa	0.5				
	sumneri	0.8	3.8		8.7	
Autumnalis	bangkinang	4.1	14.3		3.8	17.7
	rachmat <sup>a</sup>	4.1	1.8		7.7	
	gurungi	0.6				3.2
	weerasingha	1.0	1.8			1.6
	sentot <sup>a</sup>	0.6	1.8		3.8	1.6
Pyrogenes	zanoni	7.3	8.9		5.8	
	biggis	2.0	1.8		11.5	
	hamptoni	1.5				
	abramis	0.2				
Hebdomadis	wolffiª	7.2	23.2		5.8	
	ricardi	1.5			1.0	
	worsfoldi	0.1				
Grippotyphosa	grippotyphosaª	4.9	1.8		3.8	
Javanica	javanicaª		1.8	•		
Celledoni	whitcombi	0.2				
Pomona	pomona	0.3				
Tarassovi	tarassovi <sup>a</sup>	0.1				
Ranarum	evansiª	0.1				

TABLE 1. Pathogenic leptospiras isolated from Malaysian waters and wet soils

<sup>a</sup> The identity of representative strain was confirmed by agglutinin-adsorption tests.

Chernukha (C. Sulzer, personal communication). It had not heretofore been found in Malaysia. It was found in waters of both East and West Malaysia.

The fourth strain (267-1348) cross-reacted to some but not all of the serovars in the Canicola, Icterohaemorrhagia, Javanica, and Autumnalis serogroups, and to homologous titer with a strain isolated from the kidneys of a frog in Iowa (7), and which Babudieri (4) classified as a new pathogenic serovar *ranarum* in a separate serogroup Ranarum (3). In comparative crossagglutination tests with diverse serovar antisera, the reactions of *ranarum* and strain 267-1348 were essentially identical (Table 2). Reciprocal agglutinin-adsorption tests were carried out between strain 267-1348 and *ranarum* 

 TABLE 2. Cross-agglutination reactions of strains

 267-1348 and ICF (serovar ranarum)

Ar	Reciprocal of titer <sup>*</sup> with:		
Serogroup	Serovar	267-1348	ICF
Icterohaemor-	birkini	400	_
rhagiae	smithu	400	100
Canicola	jonsis	200	100
	broomi	800	200
	bindjei	400	100
	schuffneri	400	200
	benjamin	1,600	800
	malaya	800	400
	galtoni	3,200	800
Pyrogenes	rc binsoni	1,600	800
Javanica	javanica	400	_
	poi	1,600	6,400
	sorex	100	1,600
	ceylonica	-	400
Autumnalis	djasiman	1,600	1,600
	gurungi	1,600	1,600
Ranarum	ranarum (ICF)	3,200	3,200
	evansi (267- 1348)	25,600	3,200

<sup>a</sup> Homologous titers ranged from 1:6,400 to 1:102,400.

<sup>b</sup> Titers of 1:100 or less (—) with the following serovar antisera: sarmin, icterohaemorrhagiae, copenhageni, mankarso, naam, canicola, kabura, kashirski, portland-vere, zanoni, abramis, pyrogenes, manilae, hamptoni, biggis, sofia, anhoa, celledoni, panama, shermani, medanensis, bataviae, tarassovi, pomona, autumnalis, rachmati, fort-bragg, bangkinang, erinacei-auriti, mooris, sentot, lanka, australis, grippotyphosa, cynopteri, canalzonae.

 

 TABLE 3. Cross-agglutinin adsorption studies on strain 267-1348

Antiseru	Reciprocal of titer <sup>a</sup> with:		
Against	Adsorbed with	Homolo- gous strain after adsorption	Adsorbing strain before adsorption
267-1348 (102400) <sup>b</sup> 267-1348 (25600) 267-1348 (25600)	ranarum jonsis broomi benjamin malaya robinsoni javanica poi coxus djasiman gurungi 267-1348 267-1348 267-1348	6,400 25,600 25,600 25,600 25,600 25,600 25,600 25,600 25,600 25,600 25,600 25,600 12,800 25,600 102,400 6,400	12,800 1,600 1,600 1,600 1,600 1,600 1,600 1,600 6,400 1,600 6,400 51,200 400 400 1,600
malaya (102400) robinsoni (25600) javanica (25600) poi (102400) coxus (102400) djasiman (25600) gurungi (25600)	$\begin{array}{r} 267-1348\\ 267-1348\\ 267-1348\\ 267-1348\\ 267-1348\\ 267-1348\\ 267-1348\\ 267-1348\\ 267-1348\end{array}$	$102,400 \\ 25,600 \\ 25,600 \\ 102,400 \\ 102,400 \\ 25,600 $	$1,600 \\ 1,600 \\ 400 \\ 1,600 \\ 1,600 \\ 6,400 \\ 1,600$

<sup>a</sup> Cross-agglutinins against heterologous (adsorbing strain) antigens were undetectable at 1:100 dilution after adsorption with respective heterologous antigens. Homologous titers after adsorption with homologous antigen were 1:100 or less except for *javanica*, *malaya*, and *poi* which were 1:400.

<sup>o</sup> Reciprocals of homologous titers of unadsorbed sera shown in parenthesis.

as well as with various serovar representative strains with which it cross-reacted to relatively high titer. The results of tests are summarized in Table 3. The antibodies responsible for specificity of the test serovars were not removed after adsorption with strain 267-1348 nor was the anti-267-1348 sera significantly reduced after adsorption by any of the test antisera. Accordingly, it represents a new serovar which we designate *evansi*.

#### LITERATURE CITED

- Alexander, A. D., L. B. Evans, A. J. Toussaint, R. H. Marchwicki, and F. R. McCrumb, Jr. 1957. Leptospirosis in Malaya. II. Antigenic analysis of 110 leptospiral strains and other serologic studies. Amer. J. Trop. Med. Hyg. 6:871-889.
- Alexander, A. D., P. W. Wetmore, L. B. Evans, H. Jeffries, and C. A. Gleiser. 1955. Classification of leptospiral isolates from Malaya, Thailand, and North Borneo. Amer. J. Trop. Med. Hyg. 4:492-506.

- Babudieri, B. 1972. List of leptospira strains kept in the WHO/FAO Leptospira Reference Laboratory in Rome. Ann. Inst. Super. Sanita 8:159-196.
- Babudieri, B. 1972. Systematics of a leptospira strain isolated from a frog. Experimentia 28:1252-1253.
- Baker, H. J. 1965. Leptospirosis in Malaysia. Military Med. 130:1101-1102.
- Baker, M. F., and H. J. Baker. 1970. Pathogenic Leptospira in Malaysian surface waters. 1. A method of survey for Leptospira in natural waters and soils. Amer. J. Trop. Med. Hyg. 19:485-492.
- 7. Diesch, S. L., W. F. McCulloch, J. L. Brown, and H. C.

Ellinghausen. 1968. Leptospires isolated from frog kidneys. Nature (London) 209:939-941.

- McCrumb, F. R., Jr., J. L. Stockard, C. R. Robinson, L. H. Turner, D. G. Levis, C. W. Maisey, M. F. Kelleher, C. A. Gleiser, and J. E. Smadel. 1957. Leptospirosis in Malaya. 1. Sporadic cases among military and civilian personnel. Amer. J. Trop. Med. Hyg. 6:238-256.
   Smith, C. E. G., L. H. Turner, J. L. Harrison, and J. C.
- Smith, C. E. G., L. H. Turner, J. L. Harrison, and J. C. Broom. 1961. Animal leptospirosis in Malaya. 1. Methods, zoogeographical background and broad analysis of results. Bull. WHO 24:5-21.