

Allele and genotype frequencies of *CYP2C9* in a Korean population

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Aims

To determine the frequencies of the variant alleles and the genotypes of *CYP2C9* in a Korean population.

Methods

Three hundred and fifty-eight healthy Korean subjects were studied. *CYP2C9* alleles were detected by polymerase chain reaction-restriction fragment length polymorphism assays and direct sequencing assays.

Results

The allele frequencies were 0.934 for *CYP2C9**1, 0.060 for *CYP2C9**3 and 0.006 for *CYP2C9**13. The *CYP2C9**2, *4, *5 and *11 alleles were not detected. The frequencies of the *CYP2C9**1/*1, *1/*3 and *1/*13 genotypes were 0.869, 0.120 and 0.011, respectively.

Conclusion

The frequency of the *CYP2C9**3 allele in the Korean population studied was significantly higher than reported elsewhere, and a novel allele, *CYP2C9**13, was found at a frequency of 0.006 (95% confidence interval 0, 0.012). Only three genotypes of *CYP2C9*, *CYP2C9**1/*1, *1/*3 and *1/*13 were observed in this Korean population.

Introduction

Genetic polymorphisms of drug metabolizing enzymes are one of the major determinants of interindividual variability in drug response. The cytochrome P450 enzyme *CYP2C9* is primarily responsible for the oxidative metabolism of drugs with a narrow therapeutic index such as warfarin, tolbutamide and phenytoin, other commonly used drugs such as glibenclamide, glimepiride, glipizide, losartan, irbesartan and torasemide, as well as many anti-inflammatory drugs [1, 2]. Twelve allelic variants of the *CYP2C9* gene have been reported, and recently a novel variant, *CYP2C9**13, was identified in

a Chinese population [3, 4]. Three alleles, *CYP2C9**1, *2 and *3, are present in most ethnic populations and decreased *CYP2C9* function has been reported in individuals with the *CYP2C9**2 and *3 mutant alleles [2, 5]. In addition, several other alleles may cause impaired metabolism [6], which may give rise to drug toxicity. Therefore, the doses of these drugs may need to be adjusted according to *CYP2C9* genotype. Because the frequencies of these variant alleles vary according to ethnic group, the aim of the present study was to determine the frequencies of the variant alleles and genotypes of the *CYP2C9* gene in the Korean population.

Methods

Subjects

Three hundred and fifty-eight unrelated healthy Korean subjects (290 males and 68 females; mean age 24.7 ± 2.3 years, range 20-32 years) were enrolled. Written informed consent was obtained from all subjects, and the institutional ethics committee approved the protocol.

Genotyping

For each subject, a 10-ml sample of venous blood was collected in an EDTA tube. The genomic DNA was isolated from the peripheral blood leucocytes. Genotyping for the *CYP2C9**2 (Arg144Cys, C430T) and *CYP2C9**13 (Leu90Pro, T269C) polymorphisms was performed using a previously published and validated polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) assay [4, 7]. Genotyping of the *CYP2C9**3 (Ile359Leu, A1075C), *CYP2C9**4 (Ile359Thr, T1076C), *CYP2C9**5 (Asp360Glu, C1080G) and *CYP2C9**11 (Arg335Thr, C1003T) alleles was performed using direct sequencing. PCR amplification of the *CYP2C9**2 and *13 alleles was carried out using the forward and reverse primer (5'-TACAAATACA ATGAAAATATCATG-3' and 5'-CTAACAACCAGA CTCATAATG-3') in the genomic DNA [4, 7]. Amplifi-

cation was carried using a Mastercycler Gradient (Effen-dorf, Hamburg, Germany). After amplification, the DNA was digested with specific restriction enzymes such as *Ava*II for *CYP2C9**2 and *Psp*GI for *CYP2C9**13. The digested PCR products were separated by electrophoresis on 3% agarose gels and stained with ethidium bro-mide. The variant genotypes identified using PCR-RFLP were confirmed by sequence analysis.

Sequencing of CYP2C9 exon

The genomic DNA was amplified using the intron-specific primers for *CYP2C9* exon 7 (containing *CYP2C9**3, *4, *5 and *11 allele) and *CYP2C9* exon2-exon3 (containing *CYP2C9**2 and *13 allele) [7, 8]. The specific primers coding exon 7 of the *CYP2C9* gene were 5'-CTGAATTGCTACAACAAATGTG-3' as the forward primer and 5'-GATACTATGAATTTGGGAC TTC-3' as the reverse primer [8]. The PCR products were purified and sequenced on an ABI 377 automatic sequencer using a BigDye Terminator Cycle Sequencing Ready Reaction Kit (Applied Biosystems Inc., Foster City, CA, USA).

Statistical analysis

The data were compiled according to the genotype and allele frequencies, which were compared using a χ^2 test.

Table 1

Allele (A) and genotype (B) frequencies for *CYP2C9* in a Korean population. Expected frequencies were calculated using the Hardy-Weinberg equation

A

Alleles	N	Frequency	95% CI
<i>CYP2C9</i> *1	669	0.934	0.916, 0.952
<i>CYP2C9</i> *2	0	–	–
<i>CYP2C9</i> *3	43	0.060	0.043, 0.077
<i>CYP2C9</i> *4	0	–	–
<i>CYP2C9</i> *5	0	–	–
<i>CYP2C9</i> *11	0	–	–
<i>CYP2C9</i> *13	4	0.006	0.000, 0.012

B

Genotype	Number of subjects	Observed frequency	95% CI	Expected frequency
<i>CYP2C9</i> *1/*1	311	0.869	0.833, 0.904	0.872
<i>CYP2C9</i> *1/*3	43	0.120	0.086, 0.154	0.112
<i>CYP2C9</i> *1/*13	4	0.011	0.000, 0.022	0.011
<i>CYP2C9</i> *3/*3	0	0	–	0.004
<i>CYP2C9</i> *3/*13	0	0	–	0
<i>CYP2C9</i> *13/*13	0	0	–	0

Table 2
Allele frequencies for *CYP2C9* in different Asian populations

Populations	n	CYP2C9 allele frequency				Reference
		*1	*2	*3	*13	
Korean	358	0.934 (0.916, 0.952)	0	0.060 (0.043, 0.077)	0.006	Present study
Korean	574	0.989 (0.983, 0.995)	0	0.011 (0.005, 0.017)	–	[9]
Sum of Korean	932	0.968 (0.960, 0.976)	0	0.030 (0.022, 0.038)	0.002 (0, 0.004)	
Chinese	115	0.983	0	0.017	–	[10]
Chinese	102	0.951	0	0.049	–	[11]
Chinese	394	0.963	0.001	0.036	–	[12]
Chinese	98	0.974	0	0.026	–	[7]
Chinese	133	0.959	0	0.041	–	[13]
Sum of Chinese	842	0.965 (0.956, 0.974)	0.001 (–0.001, 0.003)	0.034 (0.025, 0.043)		
Japanese	140	0.982	0	0.018	–	[14]
Japanese	123	0.955	0	0.045	–	[13]
Japanese	218	0.979	0	0.021	–	[15]
Japanese	86	0.983	0	0.017	–	[16]
Japanese	134	0.989	0	0.011	–	[17]
Sum of Japanese	701	0.978 (0.970, 0.986)	0	0.022 (0.014, 0.030)		

Values in parentheses represent the 95% confidence intervals] n, number of subjects. Differences between gene frequencies were calculated using the χ^2 test.

$P < 0.001$ vs. the previous data in Korean [9] (95% CI on the difference 0.023, 0.075).

$P < 0.01$, vs. sum of Chinese (95% CI on the difference –0.001, 0.053).

$P < 0.001$, vs. sum of Japanese (95% CI on the difference 0.011, 0.065).

A P -value < 0.05 was considered significant. The Hardy-Weinberg equilibrium was determined by comparing the genotype frequencies with the expected values using a contingency table χ^2 test.

Results

The frequencies of the allelic variants and genotypes of the *CYP2C9* gene in a Korean population are summarized in Table 1. The observed genotype frequency distribution did not show a significant deviation from the Hardy-Weinberg equilibrium. The *CYP2C9**1 allele was the most common [0.934, 95% confidence interval (CI) 0.916, 0.952]. The most frequently identified mutant allele was *CYP2C9**3 (0.060, 95% CI 0.043, 0.077). The frequency of the *CYP2C9**1/*3 genotype in this study was more than five times higher than that reported elsewhere [9] (Table 2). The frequency of the *CYP2C9**13 allele in our population was 0.006 (95% CI 0, 0.012).

There were 311 subjects (0.869, 95% CI 0.833, 0.904) with the *CYP2C9**1/*1 genotype, 43 (0.120,

95% CI 0.086, 0.154) with the *CYP2C9**1/*3 genotype and four (0.011, 95% CI 0.000, 0.022) with the *CYP2C9**1/*13 genotype (Table 1). None of the subjects had the *CYP2C9**2, *4, *5 or *11 allele, or was homozygous for the *CYP2C9**3 or *13 allele. The genotype results obtained by PCR-RFLP were confirmed by sequencing (data not shown).

Discussion

The allelic variants, *CYP2C9**2 and *CYP2C9**3, code for enzymes with approximately 10–40% and 5–15% of the activity of the wild-type form *CYP2C9**1, respectively [18]. The *CYP2C9**2 allele is the most common mutant allele among caucasians with a frequency of approximately 0.125 [18]. In contrast, *CYP2C9**2 has not been detected in Asian populations [18]. The *CYP2C9**2, *CYP2C9**4, *CYP2C9**5 and *CYP2C9**11 alleles were also not detected in the 358 Koreans studied in the present work.

The functional importance of *CYP2C9**3 is greater

than that of *CYP2C9*2* because the former allele appears to confer the largest decrease in enzyme activity *in vitro*, whereas the *CYP2C9*2* allele produces an intermediate fall in activity compared with *CYP2C9*1* [19]. Caucasians exhibit a wide range (0.033–0.162) in *CYP2C9*2* allele frequency, whereas the *CYP2C9*3* allele has a much lower frequency [19]. The frequency of the *CYP2C9*3* allele in the present Korean population was 0.060, which was above five times higher than that reported elsewhere (0.011) [9]. Such a discrepancy can be seen in other studies of Japanese and Chinese populations (Table 2). Although a frequency of 0.060 for the *CYP2C9*3* is significantly higher ($P < 0.01$ and $P < 0.001$, respectively) than those in the Japanese (0.022) and Chinese populations (0.034) (Table 2), the mean value (0.030) from the two Korean studies published to date [present, 9] was similar to that in the Chinese population. In the present study, all the subjects with the *CYP2C9*3* allele had the heterozygous genotype, *CYP2C9*1/*3*.

The *CYP2C9*4* allele has been detected only in a Japanese epileptic patient [20]. The *CYP2C9*5* allele has been found in African-Americans with a low frequency of 0.017 [2] and in Africans [21]. The *CYP2C9*11* allele has been detected in caucasians and Africans [22]. However, very little is known about the functional effect of these alleles. As expected, no *CYP2C9*4*, **5* and **11* alleles were identified in the present study.

A novel allele, *CYP2C9*13*, was recently identified in a Chinese population, and a large decrease in enzyme activity was observed in those with the *CYP2C9*3/*13* genotype [4]. In the present study, four out of 358 Korean subjects possessed the *CYP2C9*1/*13* genotype, but no other genotypes, including the *CYP2C9*13* allele, were found. Currently, there is no information available on the metabolic activity of the enzyme coded by *CYP2C9*11*13*.

In summary, the frequency of the *CYP2C9*3* allele in the Korean population was significantly higher than reported elsewhere, and a novel allele, *CYP2C9*13*, was found at a frequency of 0.0056 (95% CI 0, 0.011) in the Korean population. Only three genotypes of *CYP2C9*, *CYP2C9*1*1*, **1*3* and **1*13*, were detected.

Competing interests: None to declare.

References

- Miners JO, Birkett DJ. Cytochrome P4502C9: an enzyme of major importance in human drug metabolism. *Br J Clin Pharmacol* 1998; 45: 525–38.
- Schwarz UI. Clinical relevance of genetic polymorphisms in the human CYP2C9 gene. *Eur J Clin Invest* 2003; 33 (Suppl. 2): 23–30.
- Ingelman-Sundberg M, Daly AK, Nebe DW, eds. Human Cytochrome P450 (CYP) Allele Nomenclature Committee Web Site. Available from: <http://www.imm.ki.se/CYPalleles/cyp2c9.htm> Accessed 26 December 2004.
- Si D, Guo Y, Zhang Y, Yang L, Zhou H, Zhong D. Identification of a novel variant CYP2C9 allele in Chinese. *Pharmacogenetics* 2004; 14: 465–9.
- Aithal GP, Day CP, Kesteven PJ, Daly AK. Association of polymorphisms in the cytochrome P450 CYP2C9 with warfarin dose requirement and risk of bleeding complications. *Lancet* 1999; 353: 717–9.
- Blaisdell J, Jorge-Nebert LF, Coulter S, Ferguson SS, Lee SJ, Chanas B, Xi T, Mohrenweiser H, Ghanayem B, Goldstein JA. Discovery of new potentially defective alleles of human CYP2C9. *Pharmacogenetics* 2004; 14: 527–37.
- Sullivan-Klose TH, Ghanayem BI, Bell DA, Zhang ZY, Kaminsky LS, Shenfield GM, Miners JO, Birkett DJ, Goldstein JA. The role of the CYP2C9-Leu359 allelic variant in the tolbutamide polymorphism. *Pharmacogenetics* 1996; 6: 341–9.
- de Morais SM, Schweikl H, Blaisdell J, Goldstein JA. Gene structure and upstream regulatory regions of human CYP2C9 and CYP2C18. *Biochem Biophys Res Commun* 1993; 194: 194–201.
- Yoon YR, Shon JH, Kim MK, Lim YC, Lee HR, Park JY, Cha IJ, Shin SG. Frequency of cytochrome P450 2C9 mutant alleles in a Korean population. *Br J Clin Pharmacol* 2001; 51: 277–80.
- Wang SL, Huang J, Lai MD, Tsai JJ. Detection of CYP2C9 polymorphism based on the polymerase chain reaction in Chinese. *Pharmacogenetics* 1995; 5: 37–42.
- Gedigk A, Casley WL, Tyndale RF, Sellers EM, Jurima-Romet M, Leeder JS. Cytochrome P-4502C9 (CYP2C9) allele frequencies in Canadian Native Indian and Inuit populations. *Can J Physiol Pharmacol* 2001; 79: 841–7.
- Yang JQ, Morin S, Verstuyft C, Fan LA, Zhang Y, Xu CD, Barbu V, Funck-Brentano C, Jaillon P, Becquemont L. Frequency of cytochrome P450 2C9 allelic variants in the Chinese and French populations. *Fund Clin Pharmacol* 2002; 17: 373–6.
- Xie HG, Prasad H, Landau R, Kim RB, Cai WM, Leiri I, Smiley RM, Wilkinson GR, Stein CM, Wood AJJ. Frequency of the defective CYP2C9 variant alleles in different ethnic groups. *Clin Pharmacol Ther* 2002; 71: P102 [Abstract].
- Kimura M, leiri I, Mamiya K, Urae A, Higuchi S. Genetic polymorphism of cytochrome P-450s, CYP2C19, and CYP2C9 in a Japanese population. *Ther Drug Monitor* 1998; 20: 243–7.
- Nasu K, Kubota T, Ishizaki T. Genetic analysis of CYP2C9 polymorphism in a Japanese population. *Pharmacogenetics* 1997; 7: 405–9.
- Takahashi H, Kashima T, Nomoto S. Comparisons between in-vitro and in-vivo metabolism of (S)-warfarin: catalytic activities of cDNA-expressed CYP2C9, its Leu359 variant and their mixture

- versus unbound clearance in patients with the corresponding CYP2C9 genotypes. *Pharmacogenetics* 1998; 8: 365–73.
- 17 Mamiya K, Ieiri I, Shimamoto J, Yukawa E, Imai J, Ninomiya H, Yamada H, Otsubo K, Higuchi S, Tashiro N. The effects of genetic polymorphisms of CYP2C9 and CYP2C19 on phenytoin metabolism in Japanese adult patients with epilepsy: studies in stereoselective hydroxylation and population pharmacokinetics. *Epilepsia* 1998; 39: 1317–23.
- 18 Scordo MG, Caputi AP, Arrigo CD, Fava G, Spina E. Allele and genotype frequencies of CYP2C9, CYP2C19 and CYP2D6 in an Italian population. *Pharmacol Res* 2004; 50: 195–200.
- 19 Lee CR, Goldstein JA, Pieper JA. Cytochrome P450 2C9 polymorphisms: a comprehensive review of the in-vivo and human data. *Pharmacogenetics* 2002; 12: 251–63.
- 20 Imai J, Ieiri I, Mamiya K, Miyahara S, Furuumi H, Nanba E, Yamane M, Fukumaki Y, Ninomiya H, Tashiro N, Otsubo K, Higuchi S. Polymorphism of the cytochrome P450 (CYP) 2C9 gene in Japanese epileptic patients: genetic analysis of the CYP2C9 locus. *Pharmacogenetics* 2000; 10: 85–9.
- 21 Yasar U, Aklillu E, Canaparo R, Sandberg M, Sayi J, Roh HK, Wennerholm A. Analysis of CYP2C9*5 in Caucasian, Oriental and black-African populations. *Eur J Clin Pharmacol* 2002; 58: 555–8.
- 22 Allabi AC, Gala JL, Horsmans Y, Babaoglu MO, Bozkurt A, Heusterspreute M, Yasar U. Functional impact of CYP2C9*5, CYP2C9*6, CYP2C9*8, and CYP2C9*11 in vivo among black Africans. *Clin Pharmacol Ther* 2004; 76: 113–8.