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Synthesis and Evaluation of Isosteres of N-Methyl Indolo[3, 2-b]-quinoline (Cryptolepine) as New Antiinfective Agents.

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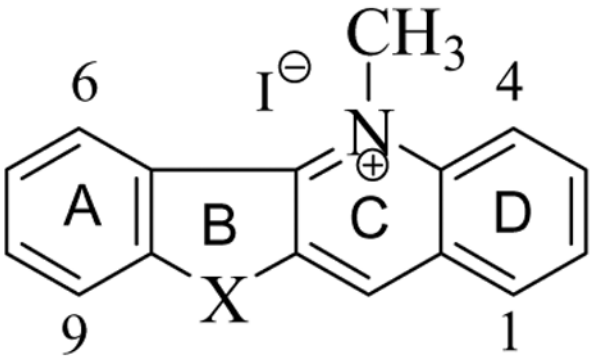
Abstract

Isosteres of cryptolepine (**1**) were synthesized and evaluated for their antiinfective activities. Overall, the sulfur isostere, 5-methyl benzothieno[3,2-b]quinoline (**5b**) was equipotent to **1** and has shown no cytotoxicity at 23.8 μg/ml. Compound **5b** was also found to have a broad spectrum of activity. Both the carbon and oxygen isosteres were less potent than cryptolepine. A limited library of 2-substituted analogs of **5b** has been synthesized and evaluated in antifungal screens but did not show increase in potency compared to the unsubstituted **5b**. Similarly, evaluation of tricyclic benzothieno [3,2-b]pyridines while showing promise in individual screens did not produce an overall increase in potency. Overall, the evaluation of the activities of **5b** compared with standard antifungal/anti-protozoal agents suggests that the benzothienoquinoline scaffold could serve as a lead for optimization.

Opportunistic infections are caused by pathogens that take advantage of a suppressed immune system. Such conditions as HIV AIDS disease, organ transplantation and long-term use of corticosteroids for example, cause either immune suppression or some disruption in the immune system.[1] With an estimated 40.3 million people living with AIDS around the globe[2] and the increasing development of resistance to current therapies, there is a continuing need for new antiinfective agents against opportunistic infections.

Previous studies in our laboratories[3,4] and those of others[5] have indicated that cryptolepine and other alkyl-substituted indolo[3,2-b]quinolines possess interesting biological activities including antiinfective activity against several opportunistic infectious organisms (OIs). These include *Candida albicans* (Ca), *Cryptoccocus neoformans (Cn)*, and *Aspergillus fumigatus (Af)*. These actions of the indoloquinolines appear to operate through intercalation into DNA, binding preferentially at GC-rich sequences and stimulating DNA cleavage by human topoisomerase II. On the basis of these initial observations, we embarked on Structure-Activity Relationship (SAR) studies to probe and identify structural features that enhance the potency of the indoloquinoline scaffold.



Compd 1; 5-Methyl indolo[3,2-b]quinolinium iodide; X = NH

Compd 5b; 5-Methyl benzothieno[3,2-b]quinolinium iodide; X = S

Several publications from our laboratories have indicated that alkylation of the nitrogen at the 5 position is required for the antiinfective activities associated with the quindoline scaffold.[6,7] In particular, an ω-cycloalkyl pentyl group was associated with increased potency as antifungal, antibacterial and anti-protozoal agents. It was also observed that the absence of ring D appears to have little effect on antimicrobial activity[8] but may alter the overall mode of action as both toxicity and spectrum of activity appeared to have increased with the tricyclic analogs. Overall however, increasing potency appears to be associated with increasing cytotoxicity.[9] This observation has spurred our desire to modify the indoloquinoline scaffold so as to separate potency from cytotoxicity. In this paper, we report on the effect of isosteric replacements at the N-10 position and the results of further SAR investigation associated with these modifications.

Chemistry

The syntheses of cryptolepine (**1**) and other indoloquinolines were previously reported.[10] 5-Methyl 11H-indeno[3,2-b]quinolinium iodide (**2**), the carbon isostere of cryptolepine, was obtained by a series of reactions beginning with the reaction of anthranilic acid with 1-indanone to yield indenoquinolone (Scheme 1).[11] The resulting indenoquinolone was treated with phosphorus oxychloride ($POCl_3$) followed by hydrogenation with palladium on carbon to produce 11H-indeno[3,2-b]quinoline, which was methylated (MeI) to produce the desired product, **2**. Attempts to alkylate with 5-phenylpentyl iodide were unsuccessful and often led to decomposition.

The preparation of benzothienoquinoline and benzofuroquinoline was achieved using modifications of a previously published procedure[10,12] outlined in Scheme 2. Briefly, substituted anthranilic acids, available from commercial sources, were acylated with acyl chlorides obtained by alkylating phenol or thiophenol. In a double intramolecular cyclization reaction with polyphosphoric acid (PPA), the acylated intermediates were converted to substituted-11-quinolones (**3a–3f**), which were subsequently converted to benzofuroquinoline (**4a**) and benzothienoquinolines (**4b–4h**) by chlorination with phosphorous oxychloride ($POCl_3$) and hydrogenation on Pd/C to remove the chlorine. The targeted final products (**5a–5h**) were then obtained by methylation with methyl iodide. Compound **4b** was also alkylated with methyl triflate and 5-phenylpentyl iodide to form 5-methylbenzothieno[3,2-b] quinolinium triflate (**6**) and 5-(5-phenylpentyl)benzothieno[3,2-b]quinolinium iodide (**7**) respectively.

Construction of the tricyclic benzothieno[3,2-b]pyridine ring (**10a–c**) proved to be more challenging than we first thought. Eventually, 3-bromopyridine N-oxide[13] was coupled with 4-nitrobenzene diazonium tetrafluoroborate[14] to obtain 3-bromo-1-(4-nitrophenoxy) pyridinium tetrafluoroborate (**8**) in good yield (68%) and then converted to 3-bromo-2-(2-hydroxy-5-nitrophenyl) pyridine (**9a**). Carbamoylation with dimethylthiocarbamoyl chloride produced **9b,** which when heated at 190 °C underwent rearrangement and subsequent cyclization in ethylene glycol to form 8-nitro-benzo[4,5]thieno[3,2-b]pyridine (**10a**). Attempts to obtain the unsubstituted benzothieno[3,2-b]pyridine using the same approach were unsuccessful. However, the unsubstituted compound (**11c)** was obtained using the amino derivative, **10b** as the starting material. Diazotization of **10b** and treatment of the resulting diazonium ion with hypophosphorous acid (H_2PO_3) yielded **10c**. Compound **10a** was methylated to **11a** but was resistant to alkylation with 5-phenylpentyl iodide, apparently as a result of decreased pKa of the N atom. Compound **10c** was similarly alkylated to produce the target compound, **11c.** The amine substituent **11b**, was prepared by reducing compound **10a** with $SnCl_2$ to yield the primary amine (**10b**) which was subjected to alkylation with methyl iodide to form **11b**. Compound **11d** was obtained by alkylation of amine **10c** with 5-phenylpentyl iodide. The overall yields for these reactions were rather low although no attempts were made to optimize yields.

Results and Discussion

Previous studies on N-alkylated δ-carbolines and their carbon isostere, azafluorenes,[8] suggest that antifungal activity is modulated by isosteric replacement. While cryptolepine may be a useful lead for drug design, it has shown considerable cytotoxicity in Vero cell lines, prompting us to look for ways to attenuate its cytotoxicity. As part of that desire, we have embarked on various SAR approaches including isosteric replacement of the N-10 atom. Three isosteres were synthesized and evaluated first against two common opportunistic pathogens, *C. albicans* and *C. neoformans* and subsequently against other microorganisms. The microbes were selected for evaluation because of their known pathogenicity.[15] Compounds which show inhibition (IC_{50}) of the microorganism at 20 μg/ml or better were so evaluated and the results are recorded in Tables 1 and 2. The results indicate that cryptolepine is more active than its carbon (**2**) and oxygen (**3**) isosteres against *C. albicans* and *C. neoformans*. The sulfur isostere or 5-methyl benzothieno[3,2-b]quinolinium iodide (**5b**) was found to have similar potency as cryptolepine. All compounds except **7** were further evaluated against other pathogens including *A. fumigatus* and *M. intracellulare*. As shown in Table 2, most of the compounds have no activity against *A. fumigatus* or *M. intracellulare* (activity below 20 μg/ml). The compounds were also evaluated against leishmania, an infectious species causing infection in about 1.5 million people a year.[16] All the compounds show significant activities against leishmania as compared to the standard pentamidine. In fact, **5a** and **5b** are twice as potent as pentamidine. In addition, because previous studies[17] indicated compound **1** had activity against malaria parasites, the isosteres were also evaluated for activity against *P. falciparum*. Moderate activities were associated with the isosteres as shown in Table 2.

The potential for selective toxicity of the compounds was also evaluated in Vero cell lines. All the compounds except for cryptolepine and **5g** showed no cytotoxicity in Vero cell lines up to a concentration of 23.8 μg/ml. Compound **5g** was less cytotoxic than **1** (IC_{50} of 7.0 and 3.2 μg/ml respectively). Finally, compounds **1** and **5b** were further evaluated for cytotoxicity against several cell lines and, the results recorded in Table 3 showed compound **5b** to be significantly less toxic than compound **1** in all cell lines.

The broad spectrum of activity shown by compound **5b** and its lower cytotoxicity spurred us on to evaluate a limited library of substituted analogs for their antiinfective properties. Position 2 which is *para* to the quaternary N-5 atom was of interest because of the effect substituents

would have on the positive charge on the N-atom at position 5. The results of the evaluation are reported in Tables 1 and 2. Five such compounds (**5c–5i**) were synthesized to test the effect of electron withdrawing and electron donating substituents at the 2-position. These effects could be inductive or mesomeric. The results show that activities of the compounds (**5c–5i)** did not change substantially when the substituted 5-methyl benzothienoquinolines were compared with the unsubstituted compound (**5b**). Similarly, removal of ring D from **5b** to form benzothieno[3,2-b]pyridines (**11a–11c**) did not result in much improvement in potency. However, as observed with the indoloquinolines, activity increased substantially when an ω-phenyl pentyl or an ω-cyclohexyl pentyl moiety was introduced onto the pyridine nitrogen in both rings (**7** and **11d**). There was also a general observation that by replacing nitrogen with a sulfur atom in the indole ring, cytotoxicity to Vero cells was attenuated. Compounds **5b** and **6** enabled a comparison of the effect of different co-anions on activity. In terms of potency, there was little difference in the use of the triflate as compared to iodide. However, the use of different anions might affect solubility.

In a search for an explanation as to why isosteres might have different activities, we evaluated the charge density on the N-5 atom of each isostere. The rationale for focusing on the N^+ atom is 2-fold. First, we have shown that N-alkylation which leads to a quaternary nitrogen atom formation is required for activity.[4,6 & 7] Secondly, Aymani[18] and others have reported a role for the N^+ charge in its binding to DNA fragments. The ^1H and ^{13}C NMR chemical shift (δ) of the methyl group on each quaternary N atom was used in this evaluation since the density of the positive charge on the N-5 atom should have a deshielding effect on the methyl group. In addition, the proton attached to C-11 may reflect the overall electron density in the C ring as a result of isosteric replacement of the N-10 atom. The results of this investigation are recorded in Table 4. Evaluation of the results shows that despite its electron donating capacity, which was thought to reduce the positive charge density on N^+-5, the N-Me in 5-methyl benzo [3,2-b]thienoquinolinium iodide had the highest chemical shift. It is possible that the presence of sulfur may lead to increased aromaticity and a dispersal of the electron density in **5b** than in other isosteres, i.e., N, O or carbon and thus, explain the higher chemical shift value. The trend of increasing proton chemical shift values appears to roughly correlate with the observed activities against *C. albicans* and *C. neoformans* and may thus reflect the positive nature of the N atom. On the other hand, the ^{13}C-NMR chemical shift values of the N-methyl group do not correlate well with the activities. Overall though, the role of the positive charge in modulating activity may be confounded by other factors including lipophilicity and topology of the tetracyclic ring.

Conclusion

The results of the current investigations have shown that carbon (indenoquinoline) and oxygen (benzofuroquinoline) isosteres are significantly less potent as antiinfectives when compared with the parent nitrogen (indoloquinolines) isostere. The sulfur isostere (benzothienoquinoline) on the other hand, appears is as potent as the indoloquinolines. In addition to potency, the benzothienoquinolines have a broad spectrum of activity against not only fungi but also bacteria and protozoa. More importantly, they have significantly lower cytotoxicity than the indoloquinolines in several cell lines. In general, all isosteres have significant activity against leishmania, and hence further development of these compounds against this microorganism is warranted. At the current time, it is unclear to us why there are such significant differences in the activity profiles of the isosteres. The possibility that the positive charge density on the nitrogen might correlate with potency is not currently completely supported by the data. Further investigation to understand the effect of isosteric replacement in the indoloquinoline scaffold is also warranted.

Experimental

Melting points were determined on a Gallenkamp (UK) apparatus and are uncorrected. NMR spectra were obtained on a Varian 300 MHz Mercury or a Bruker 270 MHz NMR Spectrometer. Elemental analyses were carried out by Atlantic Microlab, Inc., Norcross, GA and are within 0.4% of theory unless otherwise noted. Flash chromatography was performed with Davisil grade 634 silica gel. N,N-Dimethylformamide was distilled from $CaSO_4$ and stored over 4 Å molecular sieves. Sulfolane was dried over 4 Å molecular sieves. 5-Cyclohexylpentyl bromide and 5-phenylpentyl bromide were prepared by treatment of the corresponding alcohols with PBr_3. The remaining alkyl halides were obtained from either Sigma–Aldrich Chemicals or Fisher Scientific and were used without further purification. Quindoline (10H-indolo[3,2-b] quinoline), the starting material in several of the syntheses below was obtained as previously reported.[4]

5.1. General method for the synthesis of N-10 alkylated quindolines

5-Methyl-11H-indeno[1,2-b]quinolin-5-ium iodide, 2: Method A A mixture of anthranilic acid (5.0 g, 36.5 mmol), 1-indanone (7.5 g, 56.8 mmol) was heated with stirring for 3 h at 185 °C. The resulting mixture was filtered to obtain a solid which was washed with pyridine (25 mL), Et_2O (2× 25 mL) to give yellow solid, 5.65 g, 66%. The yellow solid (1.0 g, 4.3 mmol) was dissolved in $POCl_3$ (15 mL) and was refluxed for 3 h. The excess $POCl_3$ was removed in vacuo. The residue was neutralized with saturated $NaHCO_3$ and the aqueous solution was extracted with CH_2Cl_2 (300 mL). The organic layer was washed with brine, dried (Na_2SO_4), and filtered. The filtrate was concentrated in vacuo and the residue was purified on silica gel with 20% EtOAc-hexane to provide a white solid, 0.88 g, yield 82%. The white solid (340 mg) was dissolved in THF (50 mL), the solution was charged with 10% Pd/C (100 mg) and was hydrogenated in H_2 atmosphere (50 psi) for 6 h to yield the desired product, 11H-Indeno[1,2-b]quinoline, 251 mg, 85%. ^{1}H NMR (300 MHz, $CDCl_3$): δ8.31 (1H, brs), 8.20 (1H, m), 8.14 (1H, s), 7.75 (1H, d, J = 8.1 Hz), 7.64 (1H, t, J = 8.7 Hz), 7.54 (1H, m), 7.43 (3H, m), 3.96 (2H, s). *Calcd for*: $C_{16}H_{11}N$: C 88.45, H 5.10, N 6.45; *Found*: C 88.55, H 5.12, N 6.43.

Method B—A mixture of 11H-Indeno[1,2-b]quinoline (100 mg, 0.46 mmol) and MeI (0.2 mL) in tetramethylene sulfone (TMS) (0.5 mL) was heated in a sealed tube for 12h at 110 °C. After cooling to room temperature, EtOAc was added to induce precipitation. The precipitate was collected by filtration followed by recrystalization from MeOH-Et_2O to afford 5-Methyl-11H-indeno[1,2-b]quinolin-5-ium iodide **2**, 112 mg, 68%. Mp 251-252 °C. ^{1}H NMR (300 MHz, DMSO-d_6: δ 9.24 (1H, s), 8.71 (2H, m), 8.42 (1H, d, J = 8.4 Hz), 8.21 (1H, t, J = 8.7 Hz), 7.98 (2H, m), 7.89 (1H, t, J = 7.5 Hz), 7.76 (1H, t, J = 7.5 Hz), 4.88 (3H, s), 4.43 (2H, s). *Calcd for* $C_{17}H_{14}IN{\cdot}0.3H_2O$: C 56.00, H 3.87, N 3.84; *Found*: C 55.75, H 3.64, N 3.98.

Synthesis of Benzofuro[3,2-b]quinolinium Iodide 5a and Benzo[b]thieno[3,2-b]quinolinium Iodides, 5b–5h

5-Methyl-benzo[*b*]thieno[3,2-b]quinolin-5-ium iodide 5b: General Procedure Method C A mixture of anthranilic acid (7.4 g, 54 mmol), sodium hydroxide (4.32 g, 108 mmol), and phenylthioacetyl chloride (10.0 g, 54 mmol) in H_2O (300 mL) was stirred at 0 °C for 1 h, and then at room temperature for 3 h. The mixture was acidified (pH = 4–5) with 2% aq. HCl and then extracted with EtOAc (2×500 mL). The combined organic layer was dried (Na_2SO_4), filtered and solvent was evaporated in vacuo. The residue was crystallized in EtOAc to give a white solid, 13.5 g, 86%. The white solid (7.7 g, 26.6 mmol) was mixed with polyphosphoric acid (PPA, 120 g) and heated at 130–135 °C for 3 h. After cooling to rt, the mixture was poured onto ice/water (200 mL), neutralized with saturated solution of Na_2CO_3 and the solid was collected. The solid was washed with H_2O, EtOAc, and Et_2O to give quinolone **3b**, 5.75 g, 86%. A suspension of **3b** (5.75 g, 26.6 mmol) in $POCl_3$ (15 ml) was

refluxed with stirring at 120 °C for 3 h. After evaporating the $POCl_3$, the reaction mixture was poured onto ice/water (50 mL) and then neutralized with 10 % NaOH. The aqueous solution was extracted with CH_3Cl (3× 200 mL) and the combined organic layer was washed with brine, dried (Na_2SO_4), and then filtered. The filtrate was concentrated in vacuo to give a solid, 5.30 g, 74%. The solid (3 g, 11.15 mmol) was dissolved in AcOH (300 mL) and hydrogenated (60–65 psi) on Pd/C (10%, 1.5 g) at rt, to afford benzo[b]thieno[3,2-b]quinoline **4b**, 2.0 g, 85%. Mp: 172–173 °C. ^{1}H NMR (300 MHz, $CDCl_3$): δ 8.81 (1H, d, *J* = 7.2 Hz), 8.61 (1H, s), 8.41 (1H, d, *J* = 8.4 Hz), 7.86 (1H, d, *J* = 8.1 Hz), 7.76 (2H, m), 7.57 (3H, m). *Calcd for*: $C_{15}H_9NS$: C 76.57, H 3.86, N 5.95; *Found*: C 76.54, H 3.90, N 5.88.

Methylation was achieved using method B and starting with compound **4b** to afford **5b**, 68%. Mp: 215–216 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 10.00 (1H, s), 9.10 (1H, d, *J* = 8.5 Hz), 8.86 (1H, d, *J* = 9.2 Hz), 8.49 (1H, dd, *J* = 4.8, 7.5 Hz), 8.34 (1H, t, *J* = 7.5 Hz), 8.05 (2H, m), 7.85 (1H, t, *J* = 7.8 Hz), 5.09 (3H, s). *Calcd for* $C_{14}H_{15}IN_2S$: C 50.94, H 3.21, N 3.71; *Found*: C 51.07, H 3.13, N 3.64.

5-Methyl-benzo[4,5]furo[3,2-b]quinolin-5-ium iodide, 5a—Compound **4a,** benzo[4,5] furo[3,2-b]quinoline, was obtained using Method C above; 23%. Mp: 154–155 °C. ^{1}H NMR (300 MHz, $CDCl_3$): δ 8.18 (1H, d, *J* = 8.1 Hz), 8.09 (1H, d, *J* = 8.4 Hz), 7.94 (1H, s), 7.73 (1H, d, *J* = 8.1 Hz), 7.52 (1H, m), 7.40 (2H, m) 7.25 (1H, m), 7.11 (1H, m). *Calcd for*: $C_{15}H_9NO$: C 82.18, H 4.14, N 6.39; *Found*: C 82.08, H 4.20, N 6.33. Compound **4a** was methylated using the general procedure Method B to obtain **5a**; 81%. Mp: 204–205 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 9.65 (1H, s), 8.90 (1H, d, *J* = 8.4 Hz), 8.84 (1H, d, *J* = 9.3 Hz), 8.57 (1H, d, *J* = 8.4 Hz), 8.30 (1H, t, *J* = 9.0 Hz), 8.10 (3H, m), 7.81 (1H, t, *J* = 8.4 Hz), 5.01 (3H, s). *Calcd for* $C_{16}H_{12}INO$: C 53.21, H 3.35, N 3.88; *Found*: C 53.21, H 3.32, N 3.89.

2-Chloro-5-methyl-benzo[*b*]thieno[3,2-b]quinolin-5-ium iodide, 5c—Using method C and starting with 5-chloroanthranilic acid and phenylthio acetyl chloride, 2-Chloro benzo[b] thieno[3,2-b]quinoline**, 4c**, was obtained in a yield of 16%. Without further purification, **4c** was methylated using method B to yield the desired compound **5c**, 15%. Mp 185–186 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 9.87 (1H, s), 9.07 (1H, d, *J* = 8.5 Hz), 8.88 (1H, d, *J* = 9.6 Hz), 8.65 (1H, d, *J* = 2.3 Hz), 8.47 (1H, d, *J* = 8.1 Hz), 8.33 (1H, dd, *J* = 2.4, 9.6 Hz), 8.04 (1H, t, *J* = 7.5 Hz), 7.85 (1H, t, *J* = 7.5 Hz), 5.08 (3H, s). *Calcd for* $C_{16}H_{11}ClINS{\cdot}0.2H_2O$: C 46.18, H 2.66, N 3.37; *Found*: C 46.02, H 2.67, N 3.32.

2-Cyano-5-methyl-benzo[*b*]thieno[3,2-b]quinolin-5-ium iodide, 5e—Using method C and 5-iodoanthranilic acid as starting material, 2-iodo benzo[b]thieno[3,2-b]quinolinone was obtained and subsequently converted to 2-cyano benzo[b]thieno[3,2-b]quinoline **4e**, 32%. Mp: 244–245 °C. ^{1}H NMR (300 MHz, $CDCl_3$): δ 8.78 (1H, d, *J* = 8.1Hz); 8.67 (1H, s); 8.45 (1H, d, *J* = 10.8 Hz); 8.34 (1H,s); 7.89 (2H, dd, *J* = 8.1Hz & 2.7Hz); 7.71 (1H, t, *J* = 8.1Hz); 7.61 (1H, t, *J* = 8.1Hz); *Calcd for*: $C_{16}H_{18}N_2S$: C, 73.84; H, 3.07; N, 10.77; *Found*: C, 73.52; H, 3.16; N, 10.51. Compound **5e** was obtained from compound **4e** in 74% yield. Mp: 204–205 ° C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 9.35 (1H, s), 9.25 (1H, d, *J* = 1.8 Hz), 8.96 (1H, d, *J* = 8.7 Hz), 8.85 (1H, d, *J* = 8.1Hz), 8.47 (1H, dd, *J* = 1.8, 8.1Hz), 8.01 (1H, dt, *J* = 1.8, 8.1Hz), 7.90 (1H, d, *J* = 8.1Hz), 7.57(1H, dt, *J* = 1.8, 8.1Hz), 5.05 (3H,s). *Calcd for*: $C_{17}H_{11}IN_2S{\cdot}2.5H_2O$: C 45.65, H 3.61, N 6.26; *Found*: C 45.61, H 3.55, N 6.25.

2-Methoxy-5-methyl-benzo[*b*]thieno[3,2-b]quinolin-5-ium iodide, 5f—Using method C and 5-iodoanthranilic acid as starting material, 2-iodo benzo[b]thieno[3,2-b] quinolinone was obtained and subsequently converted to 2-methoxy benzo[b]thieno[3,2-b] quinoline **4f**, 32%. ^{1}H NMR (300 MHz, $CDCl_3$): δ8.60 (1H, d, *J* = 6.9 Hz), 8.44 (1H, s), 8.16 (1H, d, *J* = 9.3 Hz), 7.81 (1H, d, J = 7.2 Hz), 7.58 (1H, dt, *J* = 2.7, 7.2 Hz), 7.53 (1H, dt, *J* =

2.7, 7.2 Hz), 7.41 (1H, dd, *J* = 2.7, 9.3 Hz), 7.11 (1H, d, *J* = 2.7 Hz), 3.95 (3H, s). Starting from compound **4f**, and using method B, **5f** was obtained in 51% yield. Mp: 216–217 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 9.77 (1H, s), 9.02 (1H, d, *J* = 8.4 Hz), 8.78 (1H, d, *J* = 9.8 Hz), 8.43 (1H, d, *J* = 8.1 Hz), 7.96 (2H, m), 7.82 (2H, m), 5.06 (3H, s), 4.04 (3H, s). *Calcd for* $C_{17}H_{14}INOS{\cdot}0.5H_2O$: C 49.05, H 3.63, N 3.36; *Found*: C 49.01, H 3.60, N 3.30.

2-Carboxy-5-methyl-benzo[*b*]thieno[3,2-b]quinolin-5-ium iodide, 5g—A mixture of 2-cyano benzo[b]thieno[3,2-b]quinoline, **4e** (0.302 g, 1.16 mmol), conc. H_2SO_4 and H_2O (1:1 mixture, 4 mL) was stirred at 55 °C overnight. After cooling to rt, H_2O (80 mL) was added. A solid was collected, and washed with water. Further purification was conducted on silica gel to afford a solid **4g**, 200 mg, 65%; eluent: CH_2Cl_2:EtOAc (4/2). The crude product was used for alkylation without further purification. Using Method B, compound **4g** was converted to **5g**, 64%. Mp: decomposes at 203–204 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 13.08 (1H, brs), 9.08 (1H, d, *J* = 9.0Hz), 8.95 (1H, d, *J* = 3.0 Hz), 8.84 (1H, d, *J* = 9.0 Hz), 8.65 (1H, dd, *J* = 9.0 Hz & 3.0 Hz), 8.04 (1H, t, *J* = 9.0 Hz), 7.91 (1H, d, *J* = 3.0 Hz), 7.86(2H, m), 5.10 (3H, s). *Calcd for*. $C_{17}H_{12}INO_2S$: C 48.47, H 2.87, N 3.32; *Found*: C 48.72, H 2.61, N 3.22.

2-Carbamoyl-5-methyl-benzo[*b*]thieno[3,2-b]quinolin-5-ium iodide, 5h—A mixture of 2-cyano benzo[b]thieno[3,2-b]quinoline, **4e** (0.2 g, 0.76 mmol) and polyphophoric acid (30 g) was heated at 110 °C for 3 h. After cooling to rt, the mixture was poured onto ice/water (100 mL) and neutralized with saturated solution of $NaHCO_3$. A solid was obtained, washed with H_2O and EtOAc to yield 180 mg, 84%. The crude product, **4h** was used for alkylation without further purification. Using Method B, compound **4h** was converted to **5h**, 53%. Mp: 234–235 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 10.05 (1H, s), 9.12 (1H, d, *J* = 9.0 Hz), 8.96 (1H, d, *J* = 3.0 Hz), 8.93 (1H, s), 8.55 (1H, dd, *J* = 3.0, 9.0, Hz), 8.54 (1H, s), 8.51 (1H, d, *J* = 3.0 Hz), 8.05 (1H, t, *J* = 3.0 Hz), 7.87 (2H, m); 5.12 (3H, s). *Calcd for*: $C_{17}H_{13}IN_2OS{\cdot}1.25H_2O$: C 46.11, H 3.53, N 6.33; *Found*: C 46.40, H 3.16, N 6.19.

5-Methyl-benzo[*b*]thieno[3,2-b]quinolin-5-ium triflate, 6—A mixture of benzo[*b*]thieno[3,2-b]quinoline **4b** (80 mg, 0.34 mmol) in toluene (5 mL) and MeOTf (0.2 mL) was stirred for 12h at rt. A yellow solid appeared and was collected by filtration. Recrystalization from MeOH-Et_2O afforded 5-Methyl benzo[*b*]thieno[3,2-b]quinolin-5-ium triflate **6**, 114 mg, 84%. Mp: 237–238 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 9.99 (1H, s), 9.10 (1H, d, *J* = 8.4 Hz), 8.87 (1H, d, *J* = 9.0 Hz), 8.50 (1H, d, *J* = 3.6 Hz), 8.48 (1H, d, *J* = 3.6 Hz), 8.34 (1H, t, *J* = 8.4 Hz), 8.09 (1H, d, *J* = 7.8 Hz), 8.02 (1H, d, *J* = 7.8 Hz), 7.86 (1H, t, *J* = 8.1 Hz), 5.10 (3H, s). *Calcd for* $C_{17}H_{12}F_3NO_3S_2{\cdot}0.2H_2O$: C 50.66, H 3.00, N 3.49; *Found*: C 50.58, H 2.89, N 3.52.

5-(5′-Cyclohexyl-pentyl)-benzo[*b*]thieno[3,2-b]quinolin-5-ium iodide, 7—Using Method B, compound **4b** was converted to **7** in 30% yield. Mp: 182–184 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 10.02 (1H, s), 8.64 (1H, d, *J* = 9.0 Hz), 8.65 (1H, d, *J* = 8.4 Hz), 8.51 (2H, d, *J* = 8.2 Hz), 8.36 (1H, t, *J* = 7.5 Hz), 8.10 (1H, d, *J* = 7.2 Hz), 8.04 (1H, d, *J* = 8.0 Hz), 7.92 (1H, t, *J* = 7.2 Hz), 5.50 (2H, t, *J* = 7.8 Hz), 2.19 (2H, m), 1.66 (7H, m), 1.44 (2H, m), 1.21 (6H, m), 0.86 (2H, m). *Calcd for* $C_{26}H_{30}INS{\cdot}2.5\ H_2O$: C 55.71, H 6.29, N 2.50; *Found*: C 55.68, H 5.88, N 2.29.

Synthesis of Benzothieno[3,2-b]pyridines 10a–d

3-Bromo-1-(4-nitro-phenoxy)-pyridinium tetrafluoroborate, 8—To a solution of 3-bromopyridine N-oxide in (4 g, 23 mmol) in anhydrous CH_3CN (20 mL) was added 4-nitrobenzene diazonium tetrafluoroborate (5.5 g, 23 mmol) with vigorous stirring. The mixture was allowed to stir at rt for 48 hours. EtOAc (50 mL) was added, and the solution was allowed

to sit for an additional 24 hours, the salt proceeded to crystallize out of solution. The crystals where collected by filtration and washed with MeOH to yield pure 3-bromo-1-(4-nitro-phenoxy)-pyridinium tetrafluoroborate, **8**, 6.1 g, 68%. Mp: 149–150 °C (lit. 155–156 °C). ^{1}H NMR (DMSO-d_6): δ 10.27 (1H, s), 9.77 (1H, d, *J* = 6.4 Hz), 9.13 (1H, d, *J* = 8.3 Hz), 8.39 (1H, dd, *J* = 6.4, 8.4 Hz), 8.36 (2H, d, *J* = 9.27 Hz), 7.52 (2H, d, *J* = 8.2 Hz).

3-Bromo-2-(2-hydroxy-5-nitrophenyl) pyridine, 9a—A solution of 3-bromo-1-(4-nitrophenoxy)-pyridinium tetrafluoroborate, **8** (20 g, 0.052 mol) in anhydrous CH_3CN was refluxed for 30 min, after which Et_3N (5 mL) was added dropwise. The mixture was allowed to reflux for an additional hour, after which the solvent was removed *in vacuo*. The remaining residue was purified by chromatography on silica gel with 20–50 % EtOAc/Hexane to yield 2-(3-bromo-pyridin-2-yl)-5-nitrophenol **9a**, 10 g, 65%. mp: 157–159 °C (lit. 165 °C); ^{1}H NMR (DMSO-d_6): δ 11.40 (1H, s), 8.63 (1H, dd, *J* = 1.0, 4.4 Hz), 8.27 (1H, dd, *J* = 2.9, 9.0 Hz), 8.17 (1H, d, *J* = 8.4 Hz), 8.05 (1H, d, *J* = 2.8 Hz), 7.39 (1H, dd, *J* = 4.8, 8.0 Hz), 7.10 (1H, d, *J* = 7.90 Hz).

O-[2-(3-bromopyridine)-4-nitrophenyl] dimethylthiocarbamate, 9b—To a solution of 3-bromo-2-(2-hydroxy-5-nitrophenyl) pyridine, **9a** (5g, 0.017 mol) and KOH (2.54 g) in H_2O (90 ml) cooled to 5 °C, was added dropwise, a solution of dimethylthiocarbamoyl chloride (3.86 g, 0.031 mol) in anhydrous THF (40 mL) over a period of 15 minutes. The mixture was allowed to stir at rt for 45 minutes, after which it was poured into an aqueous solution of 10% KOH and extracted with EtOAc (5 x 200 ml). The extracts were combined, washed with brine, dried with $MgSO_4$, and evaporated *in vacuo*. The crude product was poured unto a column of silica gel and eluted with EtOAc/Hexane (1:4) to give o-[2-(3-bromopyridine)-5-nitrophenyl] dimethylthiocarbamate, **9b**, as a light brown solid, 4.82 g, 75%. mp: 102–103 °C (lit. 107–108 °C). ^{1}H NMR (300 MHz, DMSO-d_6): 8.64 (1H, dd, *J* = 1.3, 4.7 Hz), 8.37 (1H, dd, *J* = 3.2, 8.8 Hz), 8.31 (1H, d, *J* = 2.83 Hz), 8.22 (1H, d, *J* = 1.3 Hz), 7.55 (1H, d, *J* = 8.8 Hz), 7.42 (1H, dd, *J* = 4.5, 8.1 Hz), 3.14 (3H, s), 3.10 (3H, s).

8-Nitro-benzo[4,5]thieno[3,2-b]pyridine, 10a—Compound **9b** (2 g, 5.23 mmol) was heated under nitrogen for 1 hour at 190–195 °C. After cooling to room temperature, a solution of potassium hydroxide (1.10 g) in water (3 mL) and ethylene glycol (15 mL) was added and the resulting solution was refluxed for 45 minutes, and then allowed to cool to room temperature. On cooling, 8-nitro-benzo[4,5]thieno[3,2-b]pyridine **10a** precipitated out of solution and was collected by filtration, washed with EtOAc and dried under vacuum, 1 g, 83%. mp: 256–258 °C (lit.[#] 258–260 °C). ^{1}H NMR (DMSO-d_6): δ 9.07 (1H, s), 8.85 (1H, d, *J* = 4.4 Hz), 8.65 (1H, d, *J* = 8.2 Hz), 8.45 (2H, m), 7.65 (1H, m).

8-Amino-benzo[4,5]thieno[3,2-b]pyridine, 10b—To a solution of $SnCl_2$ (7.2 g, 38 mmol) in 6N HCl (72 mL) was added 7-nitro benzo[4,5]thieno[3,2-b]pyridine, **10a** (1.2 g, 5 mmol), and the solution was heated to boiling under reflux for 3.5 hours. On cooling, the solution was brought to pH 10 with 30% NaOH and extracted with EtOAc (3 x 200 ml). The combined extracts was dried with $MgSO_4$ and concentrated *in vacuo*. The residue was purified on a column of silica gel eluted with EtOAc to yield 7-amino-benzo[4,5]thieno[3,2-b]pyridine, **10b**, 0.8 g, 80%. mp: 183–185 (lit. 186–187 °C); ^{1}H NMR (DMSO-d_6): δ 8.65 (1H, m), 8.37 (1H, d, *J* = 8.2 Hz), 7.66 (1H, d, *J* = 8.5 Hz), 7.56 (1H, d, *J* = 2.2 Hz), 7.42 (1H, dd, *J* = 4.5, 8.0 Hz), 6.93 (1H, dd, *J* = 2.4, 8.5 Hz), 5.83 (2H, brs).

Benzo[4,5]thieno[3,2-b]pyridine, 10c—To a mixture of conc. HCl (5 mL) and H_2O (5 mL) was added 8-amino-benzo[4,5]thieno[3,2-b]pyridine, **10b** (1 g, 5 mmol). The mixture was cooled to 0 °C, and a cold solution of $NaNO_2$ (1 g, 14.5 mmol) was added slowly with the

temperature being kept at 0–5 °C. The diazotized solution was filtered and cooled to 0 °C, and a cold solution of H_3PO_2 (30 ml) was added at 0 °C. The solution was allowed to stir at rt for 48 hours, after which the pH was made alkaline (9–12) using KOH pellets and the solution was extracted with EtOAc (3 x 100 mL). The combined extracts was dried with $MgSO_4$ and concentrated *in vacuo*. The residue was then dried under vacuum to yield benzo[4,5]thieno [3,2-b]pyridine, **10c** as a light brown solid, 610 mg, 65%. mp: 80–81 °C (lit. 81–82 °C); ^{1}H NMR ($CDCl_3$): δ 8.74 (1H, dd, *J* = 1.4, 4.7 Hz), 8.52 (1H, m), 8.17 (1H, dd, *J* = 1.4, 8.2 Hz), 7.6 (3H, m), 7.37 (1H, dd, *J* = 4.7, 8.2 Hz).

General procedure for the N-alkylation of benzothieno[3,2-b]pyridine

1-Methyl-benzo[4,5]thieno[3,2-b]pyridin-1-ium iodide 11c: (Method D) To a solution of benzothieno[3,2-b]pyridine, **10c** (200 mg, 1.08 mmol) in TMS (0.2 mL) was added CH_3I (0.2 mL). The mixture was heated at 110 °C for 12 hours in a sealed pressure tube. After cooling to rt, EtOAc (10 mL) was added to precipitate a solid. The precipitate was collected, washed with additional EtOAc and recrystalized from MeOH-Et_2O as a yellow powder, 1-methyl-benzo[4,5]thieno[3,2-b]pyridin-1-ium iodide, **11c**, 252 mg, 71%. mp 194–196 °C; ^{1}H NMR (300 MHz, DMSO-d_6): δ 9.3 (1H, d, *J* = 8.0 Hz), 9.19 (1H, d, *J* = 6.0 Hz), 8.85 (1H, d, *J* = 8.4 Hz), 8.45 (1H, d, *J* = 8.0 Hz), 8.18 (1H, dd, *J* = 6.0, 8.1 Hz), 7.94 (1H, t, *J* = 7.5 Hz), 7.81 (1H, t, *J* = 7.5 Hz), 4.91 (3H, s). *Calcd for* $C_{12}H_{10}INS$: C 44.05, H 3.08, N 4.29; *Found*: C 44.30, H 3.07, N 4.13.

1-Methyl-8-nitro-benzo[4,5]thieno[3,2-b]pyridin-1-ium iodide, 11a—Using method D and **10a**, compound **11a,** was obtained in 78% yield. Mp: 254–255 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 9.49 (1H, d, *J* = 8.2 Hz), 9.46 (1H, d, *J* = 1.5 Hz), 9.34 (1H, d, *J* = 6.0 Hz), 8.760 (1H, d *J* = 9.0 Hz), 8.71 (1H, dd, *J* = 1.8, 9.0 Hz), 8.31 (1H, dd, *J* = 6.0. 8.3 Hz), 5.03 (3H, s). *Calcd for* $C_{13}H_9IN_2O_2S$: C 38.73, H 2.44, N 7.53; *Found*: C 38.70, H 2.47, N 7.50.

8-Dimethylamino-1-methyl-benzo[4,5]thieno[3,2-b]pyridin-1-ium iodide, 11b— Using method D and starting material **10b** (100 mg, 0.5 mmol), a mixture of compounds 8-amino-1-methyl-benzo[4,5]thieno[3,2-b]pyridin-1-ium iodide and 8-methylamino-1-methyl-benzo[4,5]thieno[3,2-b]pyridin-1-ium iodide was obtained. The above mixture was treated with MeI (0.2 mL) and $NaHCO_3$ (1.0 g, 11.9 mmol) with stirring for 12 h at room temperature. Solid was filtered off, and filtrate was concentrated *in vacuo*, to dry. The residue was crystallized from MeOH-Et_2O to give compound **11b**, 120 mg, 65%. Mp: 207–208 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ9.26 (1H, d, *J* = 8.1 Hz), 9.13 (1H, d, *J* = 6.0 Hz), 8.19 (1H, d, *J* = 9.0 Hz), 8.08 (1H, dd, *J* = 6.0, 8.1 Hz), 7.76 (1H, d, *J* = 2.1 Hz), 7.47 (1H, dd, *J* = 2.1, 9.0 Hz), 4.94 (3H, s), 3.32 (3H, s), 3.25 (3H, s). *Calcd for* $C_{14}H_{15}IN_2S\cdot 3.8H_2O$: C 38.33, H 3.45, N 6.39; *Found*: C 38.27, H 3.77, N 6.24.

1-(5′-Phenyl-pentyl)-benzo[4,5]thieno[3,2-b]pyridin-1-ium iodide, 11d—Using method D, a mixture of compound **10c** (70 mg, 0.38 mmol), 5-phenylpentyl iodide (0.2 ml) produced **11d,** 80 mg, 82%. Mp: 158–160 °C. ^{1}H NMR (300 MHz, DMSO-d_6): δ 9.40 (1H, d, *J* = 8.1 Hz), 9.22 (1H, d, *J* = 5.7 Hz), 8.57 (1H, d, *J* = 8.4 Hz), 8.47 (1H, d, *J* = 7.9 Hz), 8.20 (1H, dd, *J* = 6.0, 8.1 Hz), 7.94 (1H, t, *J* = 7.8 Hz), 7.84 (1H, t, *J* = 7.8 Hz), 7.23 (2H, m), 7.14 (3H, m), 5.24 (2H, t, *J* = 7.1 Hz), 2.55 (2H, t, *J* = 7.6 Hz), 2.07 (2H, m), 1.64 (2H, m), 1.51 (2H, m). *Calcd for* $C_{22}H_{22}INS\cdot 1.1H_2O$: C 55.14, H 4.63, N 2.92; *Found*: C 55.19, H 4.69, N 2.99.

Biological Testing

Antifungal and Antibacterial Testing

All organisms were obtained from the American Type Culture Collection (Manassas, VA) and include *Candida albicans* ATCC 90028, *Cryptococcus neoformans* ATCC 90113, *Aspergillus fumigatus* ATCC 90906 and *Mycobacterium intracellulare* ATCC 23068. Susceptibility testing was performed using a modified version of the NCCLS methods.[19] *M. intracelluare* is tested using a modified method of Franzblau, et al.[20] DMSO solutions of samples were serially-diluted in saline, and transferred in duplicate to 96 well microplates. Microbial suspensions were diluted in broth to afford desired colony forming units/mL according to the 0.5 McFarland Standard [*C. albicans*: either Saboraud Dextrose broth (SDB) or RPMI 1640, *C. neoformans*: SDB, *A. fumigatus*: either YM broth (for MICs) or RPMI-1640 + 5% Alamar Blue (for IC_{50} determination), *M. intracellulare*: Middlebrook 7H9 broth with OADC enrichment + 5% Alamar Blue.] After adding microbial cultures to the samples affording a final volume of 200μL and final test concentration starting with 20μg/mL, plates were read prior to and after incubation using either fluorescence at 544ex/590em (*M. intracellulare, A. fumigatus*) using the Polarstar Galaxy Plate Reader (BMG LabTechnologies, Germany) or optical density at 630nm using the EL-340 Biokinetics Reader (Bio-Tek Instruments, Vermont). Growth (saline only), solvent and blank (media only) controls were included on each test plate. Drug controls [Ciprofloxacin (ICN Biomedicals, Ohio) for bacteria and Amphotericin B (ICN Biomedicals, Ohio) for fungi] are included in each assay. Percent growth is calculated and plotted versus test concentration to afford the IC_{50} (sample concentration that affords 50% growth of the organism). The minimum inhibitory concentration (MIC) was determined by visually inspecting the plate, and is defined as the lowest test concentration that allows no detectable growth (for Alamar Blue assays, no color change from blue to pink).

Screening for in vitro Antimalarial Activity

The screening for antimalarial activity was performed as described earlier [21]. For the assay a suspension of red blood cells infected with D6 or W2 strains of *P. falciparum* (200 μL, with 2% parasitemia and 2% hematocrit in RPMI 1640 medium supplemented with 10% human serum and 60 μg/mL amikacin) is added to the wells of a 96-well plate containing 10 μl of test samples diluted in medium at various concentrations. The plate is incubated at 37 °C, for 72 h in a modular incubation chamber flushed with a gas mixture of 90% N_2, 5% O_2, and 5% CO_2 . Parasitic LDH activity is determined by using Malstat™ reagent (Flow Inc., Portland, OR). Briefly, 20 μl of the incubation mixture is mixed with 100 μl of the Malstat™ reagent and incubated at room temperature for 30 min. 20 μl of a 1:1 mixture of NBT/PES (Sigma, St. Louis, MO) is then added, and the plate is further incubated in the dark for 1 h. The reaction is stopped by the addition of 100 μl of a 5% acetic acid solution. The plate is read at 650 nm on an EL-340 Biokinetics Reader (Bio-Tek Instruments, Vermont). Percent growth inhibition is calculated and IC_{50} values are computed from the dose response curves. Artemisinin and chloroquine are included as the drug controls. DMSO (0.25%) is used as vehicle control.

Assay for *in vitro* antileishmanial activity

Antileishmanial activity was tested *in vitro* against a culture of *Leishmania donovani* promastigotes, grown in RPMI 1640 medium supplemented with 10% fetal calf serum (Gibco Chem. Co.) at 26 ° C. A 3 day-old culture was diluted to 5×10^5 promastigotes/mL. Drug dilutions were prepared directly in cell suspension in 96-well plates. Plates were incubated at 26 ° C for 48 h and growth of leishmania promastigotes was determined by Alamar Blue assay as described earlier[21]. Fluorescence was measured on a Fluostar Galaxy plate reader (BMG Lab Technologies) at excitation wavelength of 544 nm and emission wavelength of 590 nm. Pentamidine and amphotericin B were used as the standard antileishmanial agents. IC_{50} and

IC_{90} values were computed from dose curves generated by plotting percent growth versus drug concentration.

Cytotoxicity Assay

The *in vitro* cytotoxicity was determined against mammalian kidney fibroblast (VERO) cells. The assay is performed in 96-well tissue culture-treated microplates and compounds were tested up to a highest concentration of 23.8 μg/ml as described earlier.[22] In brief, cells (25,000 cells/well) were seeded to the wells of the plate and incubated for 24 h. Samples were added and plates were again incubated for 48 h. The number of viable cells was determined according to neutral red assay as previously described.[22] IC_{50} values were determined from dose curves of growth inhibition versus concentration. Doxorubicin was used as a positive control, while DMSO was used as the negative (vehicle) control.

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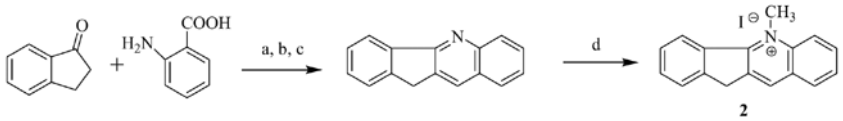
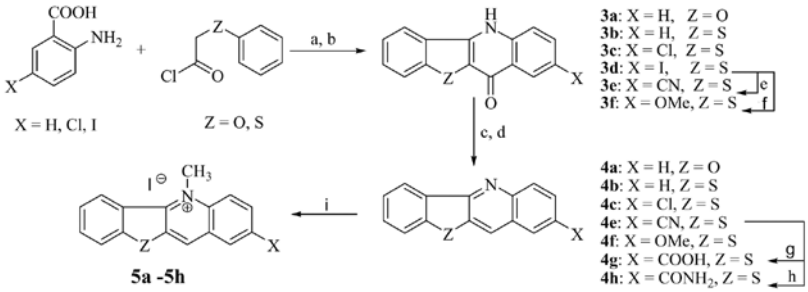


2

Reagents and conditions: a) neat, 185 °C, 3 h; b) $POCl_3$, 110 °C, 3 h; c) 10% Pd/C, H_2, 6 h; d) MeI, TMS, 110 °C, 12h.

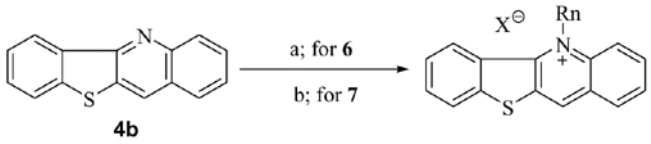
Scheme I.
Preparation of 5-Methyl-11H-indeno[3,2-b]quinolinium Iodide, **2**



Reagents and Conditions: a) NaOH, H_2O, rt. b) PPA, 130 °C, 3 h. c) $POCl_3$, 110 °C, 24 h. d) 10% Pd/C, MeOH, H_2. e) CuCN, DMF, 160 °C, 6 h. f) MeONa, CuI, HMPA. g) H_2SO_4-H_2O (v/v = 1:1), 55 °C, 12 h. h) PPA, 110 °C, 3h. i) CH_3I, TMS, 110 °C, Reflux for 12h.

Scheme 2.

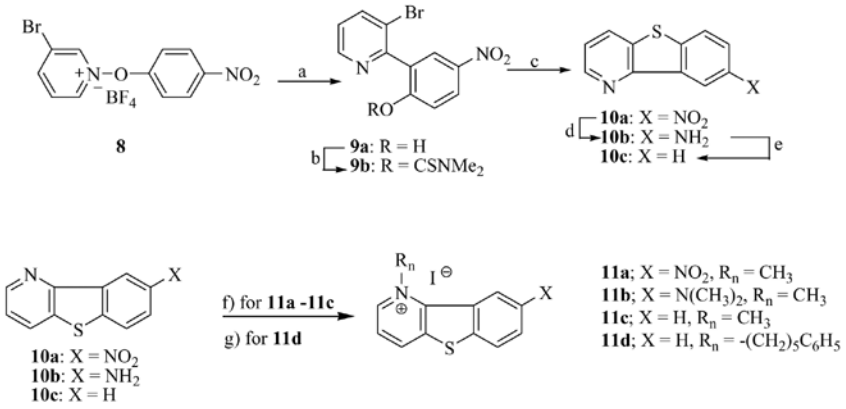
Synthesis of Benzothienoquinolinium Iodides and Benzofuroquinolinium Iodide



6: Rn = -CH_3 X = OTf
7: Rn = -$(CH_2)_5C_6H_{11}$ X = I

Reagents and conditions: a) MeOTf, Toluene, rt, 12 h.; b) 5-Cyclohexylpentyl iodide, TMS, 110 ºC, 12 h.

Scheme 3.
N-5 Alkylation of Benzothieno[3,2-b]quinoline.



Reagents and conditions: a) Et_3N, CH_3CN, reflux, 1.5 h, 65%; b) dimethylthiocarbamoyl chloride, NaOH, THF-H_2O, 0 °C-rt, 48 h, 75%; c) neat 195 °C, 1 h; KOH, ethylene glycol-H_2O, 45 min; 83%; d) $SnCl_2$, 6N HCl, reflux, 3.5h, 80%; e) Conc HCl, $NaNO_2$, 0 °C; H_3PO_2, 60%.; f) MeI, TMS, 110 °C, 12h; g) 5-Ph$(CH_2)_5$-I, TMS, 110 °C, 12h.

Scheme V.

Synthesis of Benzothieno[3,2-b]pyridinium Iodides **11a–d**

Table 1

Physicochemical Data and Antifungal Activities of Synthetic Compounds

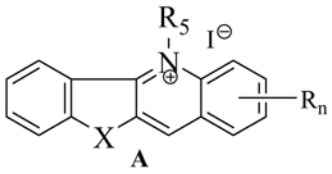
Compd	yield %	MP (°C)	formula	IC_{50} (μg/mL)	
				Cn	Ca
1				2–15.6[b]	2.- >20
2	68%	251–252	$C_{17}H_{14}IN{\cdot}0.3H_2O$	>20	>20
5a	81%	204–205	$C_{16}H_{12}INO$	>20	>20
5b	68%	215–216	$C_{14}H_{15}IN_2S$	2.5*	1.0*
5c	15%	185–186	$C_{16}H_{11}ClINS{\cdot}0.2H_2O$	15	2.0
5e	74%	204–205	$C_{17}H_{11}IN_2S{\cdot}2.5H_2O$	>20	>20
5f	51%	216–217	$C_{17}H_{14}INOS{\cdot}0.5H_2O$	8.0–15>20	
5g	64%	203–204	$C_{17}H_{12}INO_2S$	>20	>20
5h	53%	234–235	$C_{17}H_{13}IN_2OS{\cdot}1.25H_2O$	>20	>20
6	84%	237–238	$C_{17}H_{12}F_3NO_3S_2{\cdot}0.2H_2O$	>20	10.0
7	30%	182–184	$C_{26}H_{30}INS{\cdot}2.5H_2O$	0.5*	1.0*
11a	78%	254–255	$C_{13}H_9IN_2O_2S$	>20	>20
11b	65%	207–208	$C_{14}H_{15}IN_2S{\cdot}3.8H_2O$	>20	>20
11c	71%	194–196	$C_{12}H_{10}INS$	>20	>20
11d	82%	158–160	$C_{22}H_{22}INS{\cdot}1.1H_2O$	20	>20
Amphotericin B				0.95	0.25

*Data (MIC, μg/ml) provided by Bristol Myers Squibb.

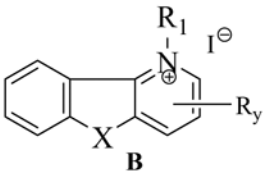
[b] = Previously reported in ref [4] &[7]

Table 2

Antiinfective Activities of Synthetic Compounds



A



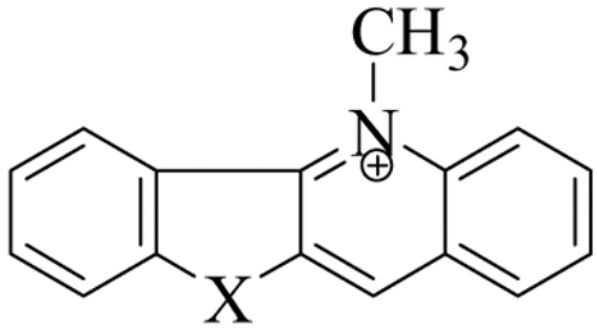
B

STRUCTURE		IC_{50} (μ g/ml)			*P. falciparum* (IC_{50} μg/ml)		*Cytotox (vero cells)*
Compd.	**A/B X, R5/R1, Rn/Ry**	**Af**	**Mi**	**Leish**	**D6 Clone**	**W2 Clone**	**IC_{50}**
1	A, NH, CH_3, H	>20[a]	15	0.23	0.18	0.35	3.2
2	A, CH_2, CH_3, H	>20[a]	>20	NT	NT	NT	NT
5a	A, O CH_3, H	15	>20	2.2	0.28	0.28	>23.8
5b	A, S CH_3, H	>20[a]	>20	0.35	1.6	2.0	>23.8
5c	A, S CH_3, 2-Cl	4.0[a]	>20	0.42	4.0	2.8	>23.8
5e	A, S CH_3, 2-CN	20[a]	>20	2.6	NA	NA	>23.8
5f	A, S CH_3, 2-OMe	20[a]	>20	0.6	0.74	0.74	>23.8
5g	A, S CH_3, 2-CO_2H	10[a]	20	0.39	NA	4.5	7.0
5h	A, S CH_3, 2-$CONH_2$	>20[a]	>20	40	NA	NA	>23.8
6	**5a** with OTf co-anion	6.5	15	0.27	0.34	0.30	>23.8
11a	B, S CH_3, 8-NO_2	>20[a]	>20	>40	0.4	0.46	>23.8
11b	B, S CH_3, 8-$N(Me)_2$	>20[a]	15	3.0	0.18	0.20	>23.8
11c	B, S CH_3, H	>20[a]	>20	NT	1.5	1.2	>23.8
11d	B, S, $(CH_2)_5$Ph, H	20[a]	1.5	1.2	0.27	0.22	20.0
	Amphotericin B	0.85	NT	NT	NT	NT	6.5
	Ciprofloxacin		0.35				NT
	Pentamidine			0.7			NT
	Chloroquine/ Artemisinin				0.02/0.02	0.14/0.02	>4.7/>4.7

[a] = MIC values; Abbreviations: NA = Not active up to 4.76 μg/ml; NT = Not tested. Af = Aspergillus fumigatus; MI = Mycobacterium intracellulare; Leish = Leishmaniasis

Table 3

Toxicity of Key Compounds



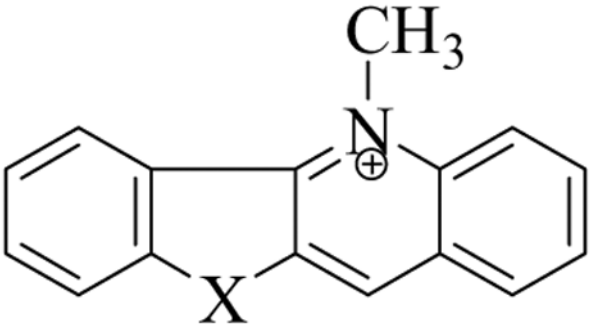
STRUCTURE		**Toxicity expressed as IC_{50} (μ g/ml)**					
Compd	X	Murine M109[a]	Human Hep2[a]	Human HuVec[a]	Vero Cells	L6 Cytotox[b]	BrdU Cytotox[b]
Compd **1**	NH	0.44	<0.18	1.38	3.2	39.3	<0.78
Compd **5b**	S	1486	355	662	>4.7	>100	4.6
Amph B		NT	NT	NT	6.5	NT	NT

[a] = Data supplied by Bristol Myers Squibb.

[b] = Data supplied by Eli Lilly & Co.

Table 4

^{1}H and ^{13}C NMR chemical shift (δ) of the methyl groups on the N-5 atom of Isosteres.



STRUCTURE		^{1}H Chemical Shift (δ)		^{13}C NMR Chemical Shift (δ)
Compound	X	N^{+}-CH_3	-$C_{11}H$	N^{+}-CH_3
Compd **1**	NH	5.05	9.25	40.14
Compd **2**	CH_2	4.88	9.24	41.42
Compd **5a**	O	5.01	9.65	41.28
Compd **5b**	S	5.09	10.00	43.28